

SUPPORTING INFORMATION

Total Syntheses of Ipomoeassin B and E

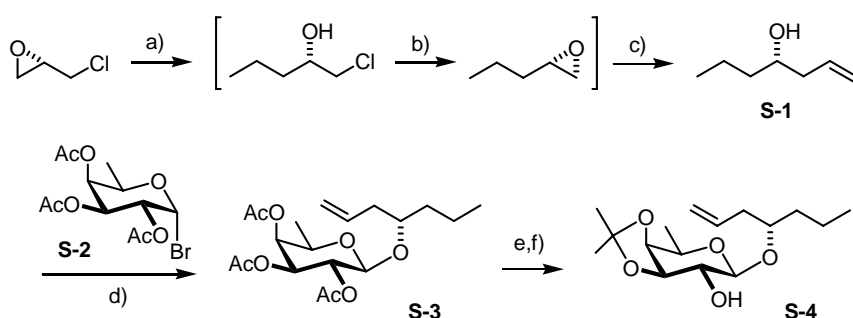
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General. All reactions were carried out under Ar atmosphere. The solvents used were purified by distillation over the drying agents indicated and were transferred under Ar: THF (Mg-anthracene), CH₂Cl₂ (P₄O₁₀), MeCN, Et₃N, DMSO (CaH₂), MeOH (Mg), toluene (Na/K). Flash chromatography: Merck silica gel 60 (230-400 mesh). NMR: Spectra were recorded on a DPX 300, AV 400 or DMX 600 spectrometer (Bruker) in the solvents indicated; chemical shifts (δ) are given in ppm relative to TMS, coupling constants (J) in Hz. The solvent signals were used as references and the chemical shifts converted to the TMS scale (calibration: CDCl₃, $\delta_C = 77.0$, $\delta_H = 7.26$; CD₂Cl₂, $\delta_C = 53.8$, $\delta_H = 5.32$). IR: Nicolet FT-7199 spectrometer, wavenumbers in cm⁻¹. MS (EI): Finnigan MAT 8200 (70 eV), HRMS: Finnigan MAT 95. Melting points: Büchi melting point apparatus (uncorrected). All commercially available compounds were used as received.

Preparation of the Building Blocks



Scheme S1. Conditions: a) EtMgCl, CuCN (10 mol%), THF, $-78 \rightarrow -20^\circ\text{C}$; b) NaOH, Et₂O; c) CH₂=CHMgBr, CuCN (10 mol%), THF, $-78 \rightarrow 0^\circ\text{C}$, 77% (overall), 99% *ee*; d) AgOTf, 2,6-di-*tert*-butylpyridine, MS 4Å, CH₂Cl₂, $0^\circ\text{C} \rightarrow \text{RT}$, 84%; e) KOMe cat., MeOH; f) 2,2-dimethoxypropane, *p*-TsOH·H₂O cat., acetone, 98% (over both steps).

(S)-1-Hepten-4-ol (S-1). A solution of EtMgCl (2 M in THF, 16.2 mL, 32.4 mmol) was added dropwise to a solution of (*S*)-epichlorohydrin (2.00 g, 21.6 mmol) and CuCN (193 mg, 2.16 mmol) in THF (30 mL) at $-78\text{ }^{\circ}\text{C}$. The mixture was warmed to $-20\text{ }^{\circ}\text{C}$ over 3 h before it was poured into sat. aq. NH_4Cl . The organic layer was separated, the aqueous layer was extracted with Et_2O , the combined organic layers were dried over MgSO_4 , filtered and evaporated to afford crude (*S*)-1-chloro-pentan-2-ol, which was used without further purification.

Powdered NaOH (4.80 g, 121 mmol) was added to a solution of the crude (*S*)-1-chloro-pentene-2-ol in Et_2O (30 mL) and the resulting mixture was stirred at room temperature for 22 h before it was poured into water (10 mL). The organic layer was separated, the aqueous layer was repeatedly extracted with Et_2O , the combined organic phases were dried over MgSO_4 , filtered and evaporated to give (*S*)-2-propyloxirane, which was used without further purification.

To a stirred solution of the crude oxirane thus formed and CuCN (193 mg, 2.16 mmol) in THF (15 mL) was added a solution of vinylmagnesium bromide (1 M in THF, 28.1 mL, 28.1 mmol) at $-78\text{ }^{\circ}\text{C}$ over a period of 45 min. The resulting mixture was allowed to warm to $0\text{ }^{\circ}\text{C}$ before the reaction was quenched with sat. aq. NH_4Cl . The aqueous layer was repeatedly extracted with Et_2O , the combined ethereal extracts were washed with brine, dried over MgSO_4 , filtered and evaporated. The residue was purified by flash chromatography on silica gel (hexanes/*tert*-butyl methyl ether, 4/1) to give (*S*)-1-hepten-4-ol (**S-1**) as a pale yellow oil (1.91 g, 77%). The NMR data are in full agreement with those previously reported in the literature. $[\alpha]_{\text{D}}^{20} = -12.8$ (c 0.52, CHCl_3); lit.¹ $[\alpha]_{\text{D}}^{20} = +12.7$ (c 0.54, CHCl_3) for (*R*)-enantiomer (99% *ee*).

Compound S-3. A solution of HBr in HOAc (30% *w/w*, 7.1 mL) was added dropwise to a cold ($0\text{ }^{\circ}\text{C}$) solution of 1,2,3,4-tetra-*O*-acetyl-*D*-fucopyranose² (2.08 g, 6.26 mmol) in CH_2Cl_2 (10 mL) and Ac_2O (0.96 mL) and the resulting mixture was stirred for 0.5 h at room temperature once the addition was complete. The mixture was then concentrated in vacuo and the resulting oil was azeotroped with toluene (3 times) to give crude glycosyl bromide **S-2** which was used in the following step without further purification.

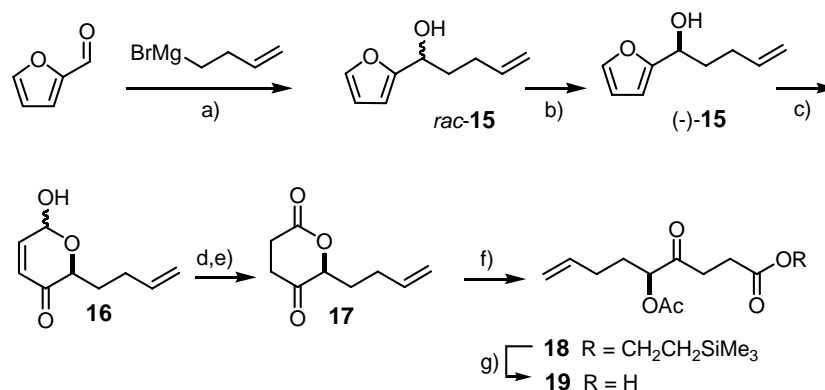
To a suspension of activated MS 4Å in CH_2Cl_2 (50 mL) was added a solution of the

¹ Kang, S.-K.; Park, D.-C.; Rho, H.-S.; Yu, C.-M.; Hong, J.-H. *Synth. Comm.* **1995**, 25, 203-214.

² Nicolaou, K. C.; Hummel, C. W.; Nakada, M.; Shibayama, K.; Pitsinos, E. N.; Saimoto, H.; Mizuno, Y.; Baldenius, K.-U.; Smith, A. L. *J. Am. Chem. Soc.* **1993**, 115, 7625-7635.

crude bromide **S-2** prepared above in CH₂Cl₂ (120 mL) and the resulting mixture was stirred at room temperature for 10 minutes before (*S*)-1-hepten-4-ol (**S-1**) (476 mg, 4.17 mmol), 2,6-di-*tert*-butylpyridine (2.40 g, 12.5 mmol), and AgOTf (1.90 g, 7.32 mmol) were successively added. Stirring was continued for 14 h before the suspension was filtered through a pad of Celite and the filtrate was evaporated. The residue was purified by flash chromatography on silica (hexanes/*tert*-butyl methyl ether, 6/1) to give compound **S-3** as a colorless syrup (1.36 g, 84%). $[\alpha]_D^{20} = -20.1$ (c 0.73, CHCl₃). IR (KAP): $\tilde{\nu} = 3076, 2961, 2937, 2873, 1752, 1641, 1368, 1250, 1223, 1074, 915$. ¹H NMR (400 MHz, CDCl₃): $\delta = 5.77$ (ddt, $J = 7.0, 10.2, 17.2$ Hz, 1 H), 5.21 (dd, $J = 1.1, 3.5$ Hz, 1H), 5.17-4.99 (m, 4H), 4.48 (d, $J = 7.9$ Hz, 1H), 3.76 (dq, $J = 1.0, 6.5$ Hz, 1H), 3.64 (m, 1H), 2.25-2.22 (m, 2H), 2.17 (s, 3H), 2.04 (s, 3H), 1.98 (s, 3H), 1.63-1.34 (m, 4H), 1.20 (d, $J = 6.7$ Hz, 3H), 0.89 (t, $J = 7.3$ Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 170.8, 170.3, 169.4, 134.4, 117.1, 100.4, 79.7, 71.5, 70.4, 69.3, 68.9, 38.4, 36.6, 20.9, 20.7, 20.6, 18.3, 16.1, 14.0$. MS (EI): m/z (%): 273 (27), 157 (17), 153 (25), 115 (13), 111 (24), 83 (12), 55 (23), 43 (100). HRMS: calcd. for C₁₉H₃₀O₈ [Na]⁺ 409.183143, found 409.183288.

Compound S-4. Compound **S-3** (1.92 g, 4.97 mmol) was dissolved in MeOH (20 mL) and treated with KOMe (18 mg, 0.25 mmol) for 3 h. The mixture was neutralized with HCl (1 M) and the solvent was evaporated. The residue was suspended in EtOAc, the mixture passed through a short-pad of silica to remove the inorganic salts, and the filtrate was evaporated. A solution containing the resulting crude product, 2,2'-dimethoxypropane (4.4 mL) and TsOH•H₂O (ca. 20 mg) in acetone (15 mL) was stirred at room temperature for 15 h. For work-up, the solvent was evaporated and the residue purified by flash chromatography on silica (hexanes/EtOAc, 6/1 → 4/1) to give glycoside **S-4** as a colorless syrup (1.46 g, 98%). $[\alpha]_D^{20} = +3.4$ (c 1.18, CHCl₃). IR (KAP): $\tilde{\nu} = 3483, 3076, 2983, 2959, 2935, 2872, 1641, 1380, 1073, 1036, 990, 918$. ¹H NMR (400 MHz, CDCl₃): $\delta = 5.83$ (ddt, $J = 7.2, 10.1, 17.1$ Hz, 1H), 5.13-5.08 (m, 2H), 4.17 (d, $J = 8.2$ Hz, 1H), 4.04-3.98 (m, 2H), 3.83 (dq, $J = 2.2, 6.6$ Hz, 1H), 3.72-3.66 (m, 1H), 3.50 (dd, $J = 7.3, 8.2$ Hz, 1H), 2.35-2.23 (m, 3H), 1.65-1.33 (m, 13H), 0.91 (t, $J = 7.3$ Hz, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 134.9, 117.6, 109.8, 101.6, 78.8, 78.7, 76.3, 73.8, 69.1, 38.7, 37.0, 28.2, 26.3, 18.4, 16.6, 14.0$. MS (EI): m/z (%): 187 (84), 129 (16), 113 (12), 101 (43), 100 (37), 97 (19), 85 (20), 83 (26), 73 (22), 71 (45), 59 (100), 57 (34), 55 (79), 43 (72), 41 (36), 29 (20). HRMS: calcd. for C₁₆H₂₈O₅ [Na]⁺ 323.182872, found 323.182895.



Scheme S2. Conditions: a) 1-bromo-3-pentene, Mg, THF, then 2-furylcarbaldehyde, 82%; b) $\text{Ti}(\text{O}i\text{Pr})_4$, D-(–)-diisopropyltartrate (DIPT), *t*-BuOOH, CH_2Cl_2 , -20°C , 47% (= 94% theoretical yield), > 99 *ee*; c) *t*-BuOOH, $\text{VO}(\text{acac})_2$ (2 mol%), CH_2Cl_2 , 71%; d) CrO_3 , H_2SO_4 , acetone, 0°C ; e) Zn, HOAc, CH_2Cl_2 , 78% (over both steps); f) (i) $\text{HO}(\text{CH}_2)_2\text{SiMe}_3$, *p*-TsOH· H_2O cat., CH_2Cl_2 ; (ii) Ac_2O , DMAP cat., CH_2Cl_2 , 93%, 97% *ee* (over both steps); g) TASF, DMF, 68%.

***dl*-5-Hydroxy-5-(2-furyl)-1-pentene (*rac*-15).** Magnesium turnings (335 mg, 13.8 mmol) in THF (1 mL) were activated with 1,2-dibromoethane (25 μL) before additional THF (7.0 mL) was added. A solution of 1-bromo-3-pentene (1.70 g, 12.5 mmol) in THF (2.0 mL) was added over 30 min and the mixture was stirred at ambient temperature for 1 h. The solution of the resulting Grignard reagent was cooled to 0°C before 2-furylcarbaldehyde (1.0 g, 10.4 mmol) was introduced at that temperature. After 2 h, the mixture was quenched with aq. HCl (1 M), the aqueous layer was repeatedly extracted with Et_2O , and the combined ethereal phases were washed with brine, dried over MgSO_4 , filtered and evaporated. The residue was purified by flash chromatography (hexanes/ EtOAc , 9/1) to give product *rac*-15 as a pale yellow oil (1.30 g, 82%). IR (KAP): $\tilde{\nu} = 3366, 2943, 2863, 1641, 1505, 1149, 1066, 1009, 913, 738, 599$. ^1H NMR (400 MHz, CDCl_3): $\delta = 7.37$ (dd, $J = 0.8, 1.8$ Hz, 1H), 6.33 (dd, $J = 1.8, 3.2$ Hz, 1H), 6.24 (d, $J = 3.2$ Hz, 1H), 5.84 (ddt, $J = 6.6, 10.3, 17.0$ Hz, 1H), 5.08–4.97 (m, 2H), 4.70 (t, $J = 6.7$ Hz, 1H), 2.20–2.12 (m, 2H), 1.98–1.92 (m, 3H). ^{13}C NMR (100 MHz, CDCl_3): $\delta = 156.6, 142.0, 137.8, 115.2, 110.1, 105.9, 67.2, 34.6, 29.7$. MS (EI): m/z (%): 152 (3), 134 (5), 123 (3), 110 (23), 97 (100), 69 (12), 41 (35), 39 (28), 29 (12), 27 (13). HRMS: calcd. for $\text{C}_9\text{H}_{12}\text{O}_2$ 152.083577, found 152.083730.

Kinetic resolution: Preparation of (–)-15. D-(–)-DIPT (2.50 g, 11.8 mmol) was added to a solution of $\text{Ti}(\text{O}i\text{Pr})_4$ (2.90 mL, 9.86 mmol) in CH_2Cl_2 (45 mL) at -20°C . After

stirring for 10 min, the mixture was cooled to $-30\text{ }^{\circ}\text{C}$ and a solution of *rac*-**15** (1.50 g, 9.86 mmol) in CH_2Cl_2 (2 mL) was slowly introduced. After stirring for 30 min, a solution of *tert*-butylhydroperoxide (5 M in decane, 1.18 mL, 5.92 mmol) was added, and the mixture was stirred for 24 h at $-20\text{ }^{\circ}\text{C}$. For work-up, the mixture was filtered through a short pad of silica and the filtrate was evaporated. The residue was purified by flash chromatography on silica (hexanes/EtOAc, 9/1) to give (–)-**15** as a pale yellow oil (701 mg, 47%, >99% *ee*). The *ee* was determined by chiral HPLC (Chiralcel OB-H, Hexane/*i*PrOH = 95/5). The NMR data are identical with those of the racemic sample described above. $[\alpha]_{\text{D}}^{20} = -6.6$ (c 1.12, CHCl_3).

Compound 16. To a solution of alcohol (–)-**15** (333 mg, 2.19 mmol) and $\text{VO}(\text{acac})_2$ (6.00 mg, 0.023 mmol) in CH_2Cl_2 (2.0 mL) was added *tert*-butylhydroperoxide (TBHP, 5 M in decane, 0.44 mL, 2.19 mmol). After stirring for 1 h at room temperature, additional $\text{VO}(\text{acac})_2$ (6.00 mg, 0.023 mmol) and TBHP (0.44 mL, 2.19 mmol) were added and stirring was continued for 2 h. The mixture was then passed through a short pad of silica, the filtrate was evaporated, and the residue purified by flash chromatography on silica (hexanes/EtOAc, 4/1) to give compound **16** as a mixture of diastereomers (262 mg, 71 %, major/minor = 69/31). IR (KAP): $\tilde{\nu} = 3411, 3077, 2926, 1690, 1641, 1089, 1033, 916$. ^1H NMR (400 MHz, CDCl_3): $\delta = 6.94$ (dd, $J = 1.5, 10.3$ Hz, 1H of minor isomer), 6.90 (dd, $J = 3.5, 10.3$ Hz, 1H of major isomer), 6.16 (dd, $J = 1.6, 10.3$ Hz, 1H of minor isomer), 6.11 (d, $J = 10.3$ Hz, 1H of major isomer), 5.87-5.76 (m, 1H), 5.66-5.65 (m, 1H), 5.18-4.98 (m, 2H), 4.59 (dd, $J = 3.8, 8.3$ Hz, 1H of major isomer), 4.11 (ddd, $J = 1.2, 3.8, 8.6$ Hz, 1H of minor isomer), 3.34 (br, 1H of minor isomer), 3.06 (br, 1H of major isomer), 2.24-2.18 (m, 2H), 2.07-2.03 (m, 1H), 1.18-1.74 (m, 1H). ^{13}C NMR (100 MHz, CDCl_3): $\delta = 196.5$ (major), 196.1 (minor), 147.6 (minor), 144.2 (major), 137.7 (major), 137.5 (minor), 128.8 (minor), 127.7 (major), 115.5 (minor), 115.4 (major), 90.9 (minor), 87.7 (major), 78.0 (minor), 73.3 (major), 29.7 (minor), 29.1 (minor), 29.0 (major), 28.8 (major). MS (EI): *m/z* (rel. intensity): 168 (4), 114 (20), 84 (100), 56 (28), 55 (39), 39 (10), 29 (12), 28 (12), 27 (14). HRMS: calcd. for $\text{C}_9\text{H}_{12}\text{O}_3$ 168.078643, found 168.078521.

Compound 17. Jones' reagent (1.7 mL)³ was added dropwise to an ice-cold solution of hemiacetal **16** (413 mg, 2.46 mmol) in acetone (13 mL). The resulting mixture was stirred for 3 h at room temperature. The mixture was diluted with *tert*-butyl methyl ether

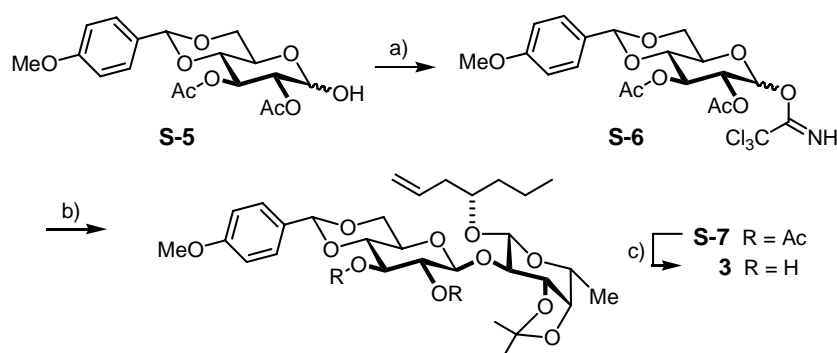
³ Georgiadis, M. P.; Tsekouras, A.; Kotretsou, S. I.; Haroutounian, S. A.; Polissiou, M. G. *Synthesis* **1991**, 929.

(50 mL) and washed with water, the organic phase was dried over Na_2SO_4 , filtered, and evaporated to give the crude oxidation product. To a solution of this material in CHCl_3 (26 mL) and AcOH (17 mL) was added zinc powder (1.2 g). The suspension was stirred for 3 h at room temperature before it was filtered through Celite. The filtrate was evaporated azeotropically with benzene to remove residual HOAc and the crude product was purified by flash chromatography (hexanes/*tert*-butyl methyl ether, 2/1 \rightarrow 1/1) to give compound **17** as a colorless oil (324 mg, 78%). $[\alpha]_{\text{D}}^{20} = -246.3$ (c 1.08, CHCl_3). IR (KAP): $\tilde{\nu} = 3079, 2929, 1759, 1735, 1641, 1267, 1171, 999, 919$. ^1H NMR (400 MHz, CDCl_3): $\delta = 5.77$ (ddt, $J = 6.7, 10.3, 16.9$ Hz, 1H), 5.11-5.02 (m, 2H), 4.68 (dd, $J = 4.0, 8.2$ Hz, 1H), 2.93-2.89 (m, 2H), 2.79-2.64 (m, 2H), 2.34-2.19 (m, 2H), 2.13-2.04 (m, 1H), 1.96-1.86 (m, 1H). ^{13}C NMR (100 MHz, CDCl_3): $\delta = 205.4, 170.0, 136.5, 116.4, 82.2, 33.8, 29.6, 28.6, 28.2$. MS (EI): m/z (%): 168 (5), 114 (72), 98 (3), 86 (5), 56 (100), 55 (27), 42 (10), 41 (12), 39 (13), 29 (18), 28 (53), 27 (22). HRMS: calcd. for $\text{C}_9\text{H}_{12}\text{O}_3$ 168.078649, found 168.078862.

Compound 18. To a solution of ketolactone **17** (100 mg, 0.59 mmol) and 2-trimethylsilylethanol (0.17 mL, 1.18 mmol) in CH_2Cl_2 (1.0 mL) was added *p*-TsOH \cdot H $_2$ O (2 mg). After stirring for 15 h, the mixture was neutralized with triethylamine and passed through a pad of silica which was carefully rinsed with EtOAc. The filtrate was evaporated, the residue dissolved in CH_2Cl_2 (10 mL) and treated with triethylamine (0.50 mL, 3.54 mmol), Ac_2O (0.22 mL, 2.36 mmol) and DMAP (10 mg, 0.08 mmol). After 3 h, the suspension was filtered through a pad of silica which was carefully rinsed with EtOAc. Evaporation of the solvent followed by flash chromatography of the residue (hexanes/EtOAc, 10/1) gave compound **18** as a colorless oil (156 mg, 93%, 97% *ee*). The *ee* was determined by chiral HPLC column (Chiralcel AD, Heptane/*i*PrOH = 98/2). $[\alpha]_{\text{D}}^{20} = -4.3$ (c 0.62, CH_2Cl_2). IR (KAP): $\tilde{\nu} = 3079, 2954, 1732, 1642, 1249, 1235, 1063, 996, 919, 860, 839, 695$. ^1H NMR (400 MHz, CD_2Cl_2): $\delta = 5.81$ (ddt, $J = 6.6, 10.3, 17.0$ Hz, 1H), 5.09-4.98 (m, 3H), 4.14 (m, 2H), 2.78-2.72 (m, 2H), 2.56-2.51 (m, 2H), 2.21-2.11 (m, 5H), 1.92-1.80 (m, 2H), 0.97 (m, 2H), 0.04 (s, 9H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 206.2, 172.7, 170.8, 137.4, 115.9, 78.0, 63.2, 33.7, 30.1, 29.6, 27.9, 20.8, 17.6, -1.5$ (3C). MS (EI): m/z (%): 43 (47), 73 (100), 117 (33), 133 (10), 173 (82). HRMS: calcd. for $\text{C}_{16}\text{H}_{28}\text{O}_5\text{Si}_1$ [$\text{M}^+ + \text{Na}$] 351.159826, found 351.159979.

Acid 19. A solution of compound **18** (50 mg, 0.176 mmol) in DMF (1.0 mL) was added to a solution of TASF (73 mg, 0.264 mmol) in DMF (1.0 mL). After stirring for 3 h, the mixture was filtered through a pad of silica which was rinsed with EtOAc several times,

and the combined filtrates were evaporated. Flash chromatography (hexanes/EtOAc/HOAc, 2/1/0.01) of the residue gave carboxylic acid **19** as a colorless oil (28 mg, 68%). $[\alpha]_D^{20} = -2.1$ (c 0.60, CH₂Cl₂). IR (KAP): ν (cm⁻¹) = 3079, 2928, 1741, 1731, 1713, 1642, 1237, 999, 918. ¹H NMR (400 MHz, CD₂Cl₂): δ = 5.81 (ddt, J = 6.7, 10.3, 17.0 Hz, 1H), 5.09-4.99 (m, 3H), 2.80-2.60 (m, 4H), 2.20-2.12 (m, 5H), 1.92-1.80 (m, 2H). ¹³C NMR (100 MHz, CD₂Cl₂): δ = 206.1, 177.9, 170.9, 137.4, 115.9, 78.0, 33.5, 30.0, 29.6, 27.5, 20.8. MS (EI): m/z (%): 43 (100), 85 (12), 101 (40), 114 (7), 132 (5), 174 (6). HRMS: calcd. for C₁₁H₁₆O₅ [M⁺+Na] 251.088993, found 251.089031.



Scheme S3. Conditions: a) Cl₃CCN, Cs₂CO₃ cat., CH₂Cl₂, 86%; b) compound **S-4**, BF₃·Et₂O cat., CH₂Cl₂/pentane (1:1), -20°C, 77%; c) KOMe cat., MeOH, 84%.

Trichloroacetimidate S-6. To a solution of substrate **S-5**⁴ (1.00 g, 2.62 mmol) in CH₂Cl₂ (4 mL) were added trichloroacetonitrile (0.48 mL, 4.79 mmol) and Cs₂CO₃ (85 mg, 0.26 mmol). The resulting mixture was stirred for 15 h at ambient temperature before it was filtered through a pad of silica and evaporated. The residue was purified by flash chromatography (hexanes/EtOAc, 2/1) to give trichloroacetimidate **S-6** as a mixture of anomers (1.19 g, 86%, α : β = 2:1). $[\alpha]_D^{20} = +25.9$ (c 0.83, CH₂Cl₂). IR (KBr): $\tilde{\nu}$ = 3348, 2940, 1754, 1676, 1616, 1589, 1519, 1236, 1071, 1033, 834. ¹H NMR (400 MHz, CD₂Cl₂): δ = 8.81 (s, 1H of β -anomer), 8.71 (s, 1H of α -anomer), 7.37-7.35 (m, 2H), 6.89-6.86 (m, 2H), 6.53 (d, J = 3.9 Hz, 1H of α -anomer), 6.00 (d, J = 7.7 Hz, 1H of β -anomer), 5.61 (t, J = 9.9 Hz, 1H of α -anomer), 5.49 (s, 1H of α -anomer), 5.48 (s, 1H of β -anomer), 5.35 (t, J = 8.8 Hz, 1H of β -anomer), 5.26 (dd, J

⁴ Fürstner, A.; Radkowski, K.; Grabowski, J.; Wirtz, C.; Mynott, R. *J. Org. Chem.* **2000**, *65*, 8758-8762.

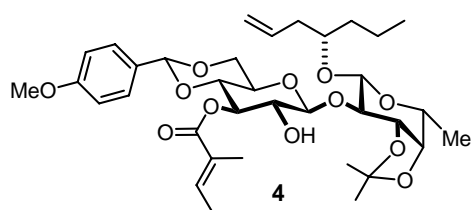
= 7.8, 8.6 Hz, 1H of β -anomer), 5.12 (dd, $J = 3.9, 9.9$ Hz, 1H of α -anomer), 4.37 (dd, $J = 4.3, 9.8$ Hz, 1H of β -anomer), 4.31 (dd, $J = 5.0, 10.4$ Hz, 1H of α -anomer), 4.13-4.05 (m, 1H), 4.88-3.72 (m, 5H), 2.06 (s, 3H of α -anomer), 2.04 (s, 3H of β -anomer), 2.02 (s, 3H of β -anomer), 2.01 (s, 3H of α -anomer). ^{13}C NMR (100 MHz, CD_2Cl_2): α -anomer: $\delta = 170.4, 170.0, 161.3, 160.6, 129.7, 127.8$ (2C), 113.9 (2C), 102.0, 93.9, 78.9, 70.7, 69.0, 68.7, 67.4, 65.5, 55.6, 21.0, 20.6; β -anomer: $\delta = 170.4, 169.4, 161.0, 160.6, 129.7, 127.9$ (2C), 113.9 (2C), 102.0, 96.2, 78.3, 71.8, 71.4, 68.7, 67.4, 65.5, 55.6, 20.9, 20.7. MS (EI): m/z (%): 527 (22), 365 (9), 179 (16), 137 (51), 136 (100), 135 (71), 43 (85). HRMS: calcd. for $\text{C}_{20}\text{H}_{22}\text{Cl}_3\text{N}_1\text{O}_9$ [$\text{M}^+ + \text{H}$] 526.043294, found 526.043271.

Disaccharide S-7. $\text{BF}_3 \cdot \text{Et}_2\text{O}$ (0.25 M in Et_2O , 1.90 mL) was added to a solution of trichloroacetimidate **S-6** (746 mg, 1.41 mmol) and alcohol **S-4** (500 mg, 1.66 mmol) in CH_2Cl_2 /pentane (2.0 mL each) at -20 °C. After stirring at that temperature for 30 min, the reaction was quenched with sat. aq. NaHCO_3 and the mixture diluted with CH_2Cl_2 . The organic layer was separated, dried over Na_2SO_4 , and evaporated, and the residue purified by flash chromatography (hexanes/ EtOAc , 4/1) to give disaccharide **S-7** as a white solid (696 mg, 77%). mp = 129-130 °C. $[\alpha]_{\text{D}}^{20} = -35.1$ (c 0.70, CH_2Cl_2). IR (KAP): $\tilde{\nu} = 3073, 2936, 2873, 1755, 1640, 1616, 1518, 1371, 1244, 1219, 1175, 1099, 1073, 1037, 920, 831$. ^1H NMR (400 MHz, CD_2Cl_2): $\delta = 7.35$ (d, $J = 8.8$ Hz, 2H), 6.87 (d, $J = 8.8$ Hz, 2H), 5.90 (ddt, $J = 7.2, 10.1, 17.2$ Hz, 1H), 5.46 (s, 1H), 5.28-5.23 (m, 1H), 5.10-5.03 (m, 2H), 5.01 (d, $J = 7.6$ Hz, 1H), 4.95-4.91 (m, 1H), 4.33 (dd, $J = 5.0, 10.4$ Hz, 1H), 4.27 (d, $J = 8.0$ Hz, 1H), 4.03-4.00 (m, 1H), 3.96 (dd, $J = 2.1, 5.6$ Hz, 1H), 3.82-3.49 (m, 9H), 2.28-2.24 (m, 2H), 2.05 (s, 3H), 2.02 (s, 3H), 1.52-1.31 (13 H), 0.92 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 170.3, 169.8, 160.6, 135.7, 130.0$ (2C), 127.8, 116.8, 113.8 (2C), 110.0, 101.8, 100.6, 100.1, 80.2, 79.7, 78.7, 78.5, 76.9, 73.0, 72.3, 69.0, 68.8, 66.7, 55.6, 38.9, 37.1, 28.1, 26.4, 21.0, 20.9, 18.7, 16.7, 14.2. MS (EI): m/z (%): 43 (100), 55 (54), 57 (13), 59 (16), 97 (16), 99 (78), 100 (45), 109 (14), 121 (14), 127 (14), 135 (28), 136 (26), 137 (38), 169 (26), 179 (53), 305 (30), 365 (63), 366 (13), 551 (13). HRMS: calcd. for $\text{C}_{34}\text{H}_{48}\text{O}_{13}$ [$\text{M}^+ + \text{H}$] 665.317321, found 665.316665.

Disaccharide 3. A solution of disaccharide **S-7** (686 mg, 1.06 mmol) and KOMe (10 mg) in MeOH (10 mL) was stirred for 4 h before it was filtered through a pad of silica and evaporated. The residue was purified by flash chromatography (hexanes/ EtOAc , 2/1 \rightarrow 0/1) to give disaccharide **3** as a white solid (518 mg, 84%). mp = 68-69 °C. $[\alpha]_{\text{D}}^{20} = -9.9$ (c 1.21, C_2HCl_2). IR (KAP): $\tilde{\nu} = 3459, 3072, 2935, 2872, 1640, 1615, 1589, 1518, 1382, 1250, 1075, 1034, 923, 831$. ^1H NMR (400 MHz, CD_2Cl_2): $\delta = 7.43$ (d, $J =$

8.8 Hz, 2H), 6.92 (d, $J = 8.8$ Hz, 2H), 5.95 (ddt, $J = 7.1, 10.2, 17.2$ Hz, 1H), 5.52 (s, 1H), 5.14-5.07 (m, 2H), 4.69 (d, $J = 7.7$ Hz, 1H), 4.35 (d, $J = 8.2$ Hz, 1H), 4.31 (dd, $J = 4.9, 10.5$ Hz, 1H), 4.17 (dd, $J = 5.5, 7.4$ Hz, 1H), 4.04 (dd, $J = 2.1, 5.4$ Hz, 1H), 3.86-3.44 (m, 12H), 2.80 (br, 1H), 2.32 (m, 1H), 1.66 (br, 1H), 1.56-1.30 (m, 13H), 0.94 (t, $J = 7.3$ Hz, 3H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 160.6, 135.4, 130.2$ (2C), 127.9, 116.9, 113.8 (2C), 110.4, 104.3, 102.1, 100.2, 96.0, 81.0, 79.2, 78.6, 77.0, 76.3, 73.2, 69.0, 68.8, 67.3, 55.6, 38.6, 36.9, 28.0, 26.3, 18.5, 16.6, 14.2. MS (EI): m/z (%): 41 (18), 43 (20), 55 (63), 57 (25), 59 (25), 69 (14), 73 (14), 85 (14), 97 (14), 99 (100), 100 (58), 101 (11), 135 (29), 136 (37), 137 (70), 281 (78), 282 (13), 467 (35). HRMS: calcd. for $\text{C}_{30}\text{H}_{45}\text{O}_{11}$ [$\text{M}^+ + \text{H}$] 581.296842, found 581.296186.

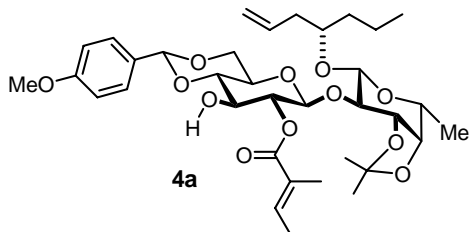
Regioselective Acylation: Preparation of Compound 4. A solution of diol **3** (690 mg, 1.19 mmol), DMAP (73.0 mg, 0.60 mmol) and DCC (293 mg, 1.42 mmol) in CH_2Cl_2 (30 mL) was stirred for 5 min prior to the addition of (*E*)-2-methylbutenoic acid (119 mg, 1.19 mmol). Stirring was continued overnight and the precipitate formed was filtered off through a pad of silica. The insoluble residues were thoroughly washed with EtOAc and the combined filtrates were evaporated. The residue was purified by flash chromatography (hexanes/EtOAc, 4/1 \rightarrow 0/1) to give disaccharide **4** and its regioisomer **4a** (**4/4a** = 9/1). Further purification was performed by preparative HPLC (Nucleodur 100-16-C18/A; MeOH/ H_2O = 4/1; flow rate: 35.0 mL/min; pressure: 4.1 MPa) to give pure **4** (432 mg, 55%) and pure **4a** (45 mg, 6%), respectively. Analytical and spectroscopic data of compound **4**: Colorless solid, mp = 72-73°C. $[\alpha]_{\text{D}}^{20} = -24.7$ (c



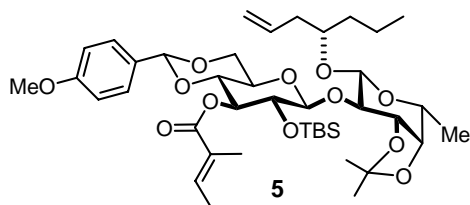
0.42, CH_2Cl_2) IR (KAP): $\tilde{\nu} = 3474, 3072, 2935, 2873, 1718, 1651, 1589, 1518, 1381, 1252, 1175, 1075, 1036, 990, 923, 829$. ^1H NMR (400 MHz, CD_2Cl_2): $\delta = 7.33$ (d, $J = 8.8$ Hz, 2H), 6.91 (m, 1H), 6.86 (d, $J = 8.8$ Hz, 2H), 5.91 (ddt, $J = 7.1,$

10.2, 17.2 Hz, 1H), 5.62 (s, 1H), 5.24 (d, $J = 9.4$ Hz, 1H), 5.11-5.03 (m, 2H), 4.77 (d, $J = 7.7$ Hz, 1H), 4.34-4.29 (m, 2H), 4.15-4.12 (m, 1H), 4.01 (dd, $J = 2.1, 5.4$ Hz, 1H), 3.83-3.50 (m, 9H), 3.36 (d, $J = 2.7$ Hz, 1H), 2.28 (m, 2H), 1.84-1.78 (m, 6H), 1.53-1.33 (m, 14H), 0.91 (t, $J = 7.2$ Hz, 3H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 167.7, 160.5, 138.3, 135.4, 130.1, 128.6, 127.8$ (2C), 117.0, 113.8 (2C), 110.4, 104.4, 101.9, 100.1, 81.0, 79.3, 79.2, 78.4, 77.0, 74.8, 73.5, 69.0, 68.9, 67.3, 55.6, 38.6, 36.9, 28.1, 26.3, 18.6, 16.6, 14.6, 14.2, 12.3. MS (EI): m/z (%): 43 (17), 55 (65), 57 (11), 59 (13), 83 (100), 97 (12), 99 (63), 100 (33), 121 (13), 135 (20), 136 (26), 137 (32), 179 (13), 363 (24), 549 (20). HRMS: calcd. for $\text{C}_{35}\text{H}_{51}\text{O}_{12}$ [$\text{M}^+ + \text{H}$] 663.338772, found 663.338055.

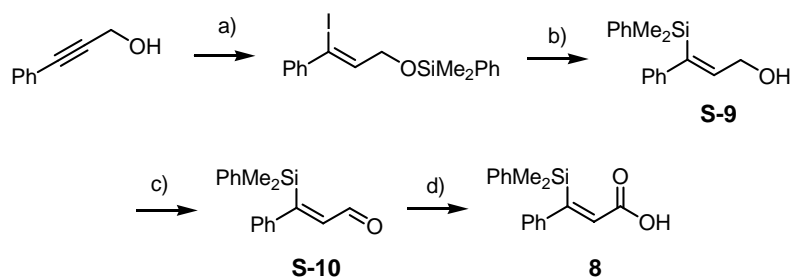
Analytical and spectroscopic data of compound **4a**: mp = 67-68 °C. $[\alpha]_D^{20} = -7.0$ (c 0.83, CH₂Cl₂) IR (KBr): $\tilde{\nu} = 3494, 3074, 2935, 2872, 1722, 1651, 1616, 1589, 1519, 1303, 1174, 921, 830$. ¹H NMR (400 MHz, CD₂Cl₂): $\delta = 7.39$ (d, $J = 8.8$ Hz, 2H), 6.96 (m, 1H), 6.89 (d, $J = 8.8$ Hz, 2H), 5.90 (ddt, $J = 7.1, 10.2, 17.2$ Hz, 1H), 5.51 (s, 1H), 5.09-5.01 (m, 3H), 4.91-4.87 (m, 1H), 4.31 (dd, $J = 5.0, 10.4$ Hz, 1H), 4.25 (d, $J = 7.9$ Hz, 1H), 4.00-3.86 (m, 3H), 3.81-3.71 (m, 4H), 3.65-3.57 (m, 3H), 3.48-3.40 (m, 2H), 2.73 (d, $J = 3.8$ Hz, 1H), 2.27 (m, 2H), 1.86 (m, 3H), 1.81 (m, 3H), 1.51-1.29 (m, 13H), 0.89 (t, $J = 7.2$ Hz, 3H). ¹³C NMR (100 MHz, CD₂Cl₂): $\delta = 167.6, 160.6, 138.6, 135.7, 130.2, 128.6, 127.9$ (2C), 116.8, 113.9 (2C), 109.9, 102.1, 100.3, 100.2, 81.3, 79.8, 79.3, 78.8, 76.8, 75.3, 73.1, 69.1, 68.8, 66.5, 55.6, 38.9, 37.1, 28.0, 26.3, 18.7, 16.7, 14.6, 14.2, 12.3. MS (EI): m/z (%): 549 (8), 363 (98), 219 (19), 209 (12), 179 (19), 137 (18), 136 (18), 135 (16), 100 (24), 99 (41), 83 (100), 55 (28). HRMS: calcd. for C₃₅H₅₀O₁₂ [M⁺+Na] 685.319445, found 685.319537.



Compound 5. TBSOTf (87.0 mL, 0.30 mmol) was added to a solution of compound **4** (100 mg, 0.15 mmol) and 2,6-lutidine (69.0 mL, 0.75 mmol) in CH₂Cl₂ (2 mL) and the resulting mixture was stirred for 2 h. Evaporation of the solvent followed by purification of the residue by preparative TLC (hexanes/EtOAc, 4/1) gave product **5** as a colorless solid (112 mg, 96%). mp = 54-55 °C. $[\alpha]_D^{20} = -11.7$ (c 0.72, CH₂Cl₂). IR (KBr): $\tilde{\nu} = 3073, 2958, 2934, 2859, 1724, 1653, 1616, 1519, 1252, 1181, 1085, 923, 837, 779, 671$. ¹H NMR (400 MHz, CD₂Cl₂): $\delta = 7.20$ (d, $J = 8.8$ Hz, 2H), 6.78 (m, 1H), 6.75 (d, $J = 8.8$ Hz, 2H), 5.80 (ddt, $J = 7.0, 10.2, 17.2$ Hz, 1H), 5.34 (s, 1H), 5.15 (m, 1H), 5.00-4.93 (m, 3H), 4.21-4.17 (m, 2H), 4.06 (dd, $J = 5.7, 6.8$ Hz, 1H), 3.89 (dd, $J = 2.0, 5.5$ Hz, 1H), 3.73-3.66 (m, 6H), 3.59-3.50 (m, 3H), 3.43-3.39 (m, 1H), 2.18 (m, 2H), 1.71-1.66 (m, 6H), 1.43-1.19 (m, 13H), 0.80 (t, $J = 5.0$ Hz, 3H), 0.73 (s, 9H), -0.02 (s, 3H), -0.11 (s, 3H). ¹³C NMR (100 MHz, CD₂Cl₂): $\delta = 167.2, 160.5, 138.1, 135.6, 130.3, 128.8, 127.7$ (2C), 116.8, 113.8 (2C), 110.0, 101.6, 100.6, 100.0, 80.0, 79.6, 78.4, 77.0, 76.2, 74.8, 74.3, 69.1, 68.9, 66.5, 55.6, 38.8, 36.9, 28.0, 26.4, 25.9 (3C), 18.8, 18.3, 16.7, 14.5, 14.2, 12.3, -3.6, -4.8. MS (EI): m/z (%): 719 (10), 663 (12), 477 (7), 283 (13), 211 (18), 183 (18), 179 (21), 158 (10), 157 (79), 136 (17), 135 (13), 121 (35), 99 (23), 97 (19), 83 (100), 73 (29), 59 (14), 55 (79), 43 (14). HRMS: calcd. for C₄₁H₆₄O₁₂Si [M⁺+Na] 799.405930, found 799.405236.



(Z)-3-Dimethyl(phenyl)silyl-2-propenoic Acid
as Protected Cinnamic Acid Surrogate



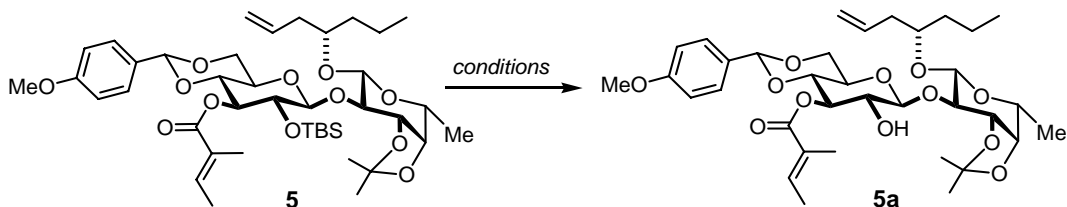
Scheme S4. Conditions: a) (i) $\text{NaAlH}_2(\text{OCH}_2\text{CH}_2\text{OMe})_2$, Et_2O , then I_2 , (ii) PhMe_2SiCl , Et_3N , DMAP cat, CH_2Cl_2 , 87%, cf. ref.; b) BuLi , THF, $-78^\circ\text{C} \rightarrow \text{RT}$, 50%, cf. ref.; c) $(\text{COCl})_2$, DMSO, Et_3N , CH_2Cl_2 , -78°C ; d) NaClO_2 , 2-methyl-2-butene, NaH_2PO_4 , $t\text{BuOH}/\text{H}_2\text{O}$, 93% (over both steps).

(Z)-3-Dimethyl(phenyl)silyl-2-propenoic acid (8). Oxalyl chloride (0.13 mL, 1.49 mmol) was added dropwise at -78°C to a solution of DMSO (0.21 mL, 2.99 mmol) in CH_2Cl_2 (5.0 mL) and the mixture was stirred for 15 min at that temperature. A solution of (Z)-3-dimethyl(phenyl)silyl-2-propene-1-ol (**S-9**)⁵ (267 mg, 1.00 mmol) in CH_2Cl_2 (2.0 mL) was introduced and the resulting suspension was stirred for 45 min, at which point Et_3N (0.56 mL, 3.98 mmol) was added. The reaction mixture was allowed to warm to 0°C over 1.5 h before it was quenched with sat. aq. NH_4Cl . The aqueous layer was repeatedly extracted with ether, the combined organic phases were washed with brine, dried over Na_2SO_4 , and the solvent was evaporated to afford the corresponding aldehyde **S-10** as a pale yellow oil which was used without further purification (239 mg, 90%). Characteristic data: IR (KAP): $\tilde{\nu} = 2839, 2739, 1677, 1489, 1428, 1254, 1111, 814, 783, 733, 701, 641, 472$. ^1H NMR (400 MHz, CD_2Cl_2): $\delta = 9.83$ (d, $J = 8.4$ Hz, 1H), 7.61-7.58 (m, 2H), 7.42-7.30 (m, 6H), 7.16-7.13 (m, 2H), 6.51 (d, $J = 8.4$ Hz, 1H), 0.34 (s, 6H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 192.9, 168.4, 144.6, 143.7$ (2C), 138.2, 134.1 (2C), 130.0, 128.6 (3C), 127.8, 126.7 (2C), 0.3 (2C). MS (EI): m/z (%): 135 (51), 189 (78), 251 (100), 266 (9). HRMS: calcd. for $\text{C}_{17}\text{H}_{18}\text{O}_1\text{Si}$ [$\text{M}^+ + \text{Na}$] 289.101916, found 289.101840.

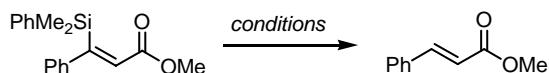
⁵ (a) Ukaji, Y.; Sada, K.; Inomata, K. *Chem. Lett.* **1993**, 1227-1230. (b) See also: Arnold, L. A.; Naasz, R.; Minnaard, A. J.; Feringa, B. L. *J. Am. Chem. Soc.* **2001**, 123, 5841-5842.

A solution of NaH_2PO_4 (73 mg, 0.61 mmol) in water (0.48 mL), 2-methyl-2-butene (0.21 mL), and NaClO_2 (109 mg, 1.2 mmol) were successively added to a solution of the crude aldehyde **S-10** (100 mg, 0.38 mmol) in *t*BuOH (3.0 mL). The mixture was stirred for 2.5 h at ambient temperature, all volatiles were evaporated, and the residue was purified by flash chromatography (hexanes/EtOAc/AcOH, 100:10:0.1) to give acid **8** (99.8 mg, 93%) as a white solid. mp = 77-78 °C. IR (KAP): $\tilde{\nu}$ = 3022, 2954, 1694, 1589, 1489, 1410, 1313, 1251, 815, 701. ^1H NMR (400 MHz, CDCl_3): δ = 7.51-7.49 (m, 2H), 7.30-7.24 (m, 6H), 7.08-7.05 (m, 2H), 6.44 (s, 1H), 0.37 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3): δ = 166.9, 144.8, 138.2, 133.7 (2C), 132.9, 128.7, 128.0 (2C), 127.5 (3C), 126.9, 126.4 (2C), 95.7, -0.9 (2C). MS (EI): m/z (%): 75 (20), 205 (60), 267 (100), 282 (1). HRMS: calcd. for $\text{C}_{17}\text{H}_{17}\text{O}_2\text{Si}$ 281.100333, found 281.100638.

Model Studies Defining the Conditions for the Proto-Desilylation



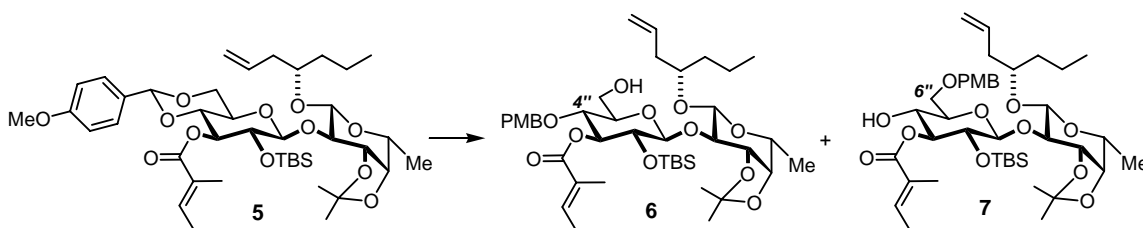
| Nr | Reagent | Conditions | Result |
|----|---------|---|---------------------------|
| 1 | TBAF | THF, 60°C, 3h | cleavage of tiglate ester |
| 2 | TBAF | THF/DMSO, methyl propionate, 80°C, 20 min | cleavage of tiglate ester |
| 3 | AgF | THF/MeOH, rt, 20h | no reaction |
| 4 | TASF | MeCN, rt, 6h | 5a (93%) |



| Nr | Reagent | Conditions | Result |
|----|------------------------------------|----------------------------|------------------------|
| 1 | BF ₃ ·Et ₂ O | CHCl ₃ , rt, 2h | no reaction |
| 2 | AgF ⁶ | THF/MeOH, rt, 5h | methyl cinnamate (32%) |
| 3 | TASF | MeCN, rt, 16h | methyl cinnamate (94%) |

The model studies summarized above suggested that only TBAF would be appropriate for the final deprotection steps in the projected total syntheses of ipomoeassin B and E.

Reductive Cleavage of the 4,6-O-*p*-Methoxybenzylidene Acetal



| Entry | Conditions | Product | Ref. |
|-------|--|----------------------------------|------|
| 1 | BH ₃ ·THF, Cu(OTf) ₂ (15 mol%), THF, RT, 1h | decomposition | 7 |
| 2 | BH ₃ ·THF, Bu ₂ BOTf (1 eq.), CH ₂ Cl ₂ , RT, 4h | decomposition | 8 |
| 3 | Dibal-H, CH ₂ Cl ₂ , 0°C, 5 min | reduction of ester | 9 |
| 4 | PMHS, AlCl ₃ , CH ₂ Cl ₂ /Et ₂ O, RT, 15h | decomposition | 10 |
| 5 | Et ₃ SiH, PhBCl ₂ , MS 4Å, CH ₂ Cl ₂ , -78°C, 1h | decomposition | 11 |
| 6 | NaBH ₃ CN, TMSCl, MS 4Å, MeCN, RT, 15h | 81% (7 : 6 = 4:1) | 12 |

⁶ (a) Fürstner, A.; Radkowski, K. *Chem. Commun.* **2002**, 2182-2183. (b) F. Lacombe, K. Radkowski, A. Fürstner, *Tetrahedron* **2004**, *60*, 7315-7324.

⁷ Shie, C.-R.; Tzeng, Z.-H.; Kulkarni, S. S.; Uang, B.-J.; Hsu, C.-Y.; Hung, S.-C. *Angew. Chem., Int. Ed.* **2005**, *44*, 1665-1668.

⁸ Wu, X.; Schmidt, R. R. *Eur. J. Org. Chem.* **2004**, 2826-2832.

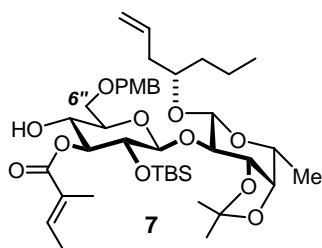
⁹ Crich, D.; Banerjee, A. *Org. Lett.* **2005**, *7*, 1395-1398.

¹⁰ Chandrasekhar, S.; Reddy, Y. R.; Reddy, C. R. *Chem. Lett.* **1998**, 1273-1274.

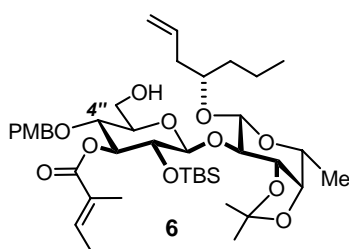
¹¹ Sakagami, M.; Hamana, H. *Tetrahedron Lett.* **2000**, *41*, 5547-5551.

Compounds 6 and 7. Disaccharide **5** (500 mg, 0.643 mmol) was added to a suspension of freshly activated MS 4Å (1.7 g) in CH₃CN (15 mL) and the resulting mixture was stirred for 15 min at that temperature. NaBH₃CN (404 mg, 6.43 mmol) was then introduced before the suspension was cooled to 0 °C and TMSCl (0.82 mL, 6.43 mmol) was added. The mixture was allowed to warm to room temperature and stirring was continued for 3 h. The suspension was filtered through Celite, the filtrate was diluted with Et₂O, the organic phase was washed with sat. aq. NaHCO₃, dried over sodium sulfate, filtered and evaporated. The residue was purified by flash chromatography to give a mixture of the reduction products (308 mg, 62%, **7/6** = 3.5/1). *This mixture does not need to be further purified because compound 9 derived thereof can be isolated in pure form by conventional chromatography at the next step (see below).* For analytical purposes, however, the regioisomers were separated by preparative HPLC (Nucleodur 100-16-C18/A; MeOH/H₂O = 4/1; flow rate: 35.0 mL/min; pressure: 4.1 MPa) to give product **7** (210 mg, 42%) and regioisomer **6** (24 mg, 5%) which showed the following spectroscopic and analytical properties:

Compound **7**: ¹H NMR (400 MHz, CD₂Cl₂): δ = 7.28 (d, *J* = 8.7 Hz, 2H), 6.92 (m, 1H), 6.87 (d, *J* = 8.7 Hz, 2H), 5.90 (ddt, *J* = 7.1, 10.2, 17.2 Hz, 1H), 5.09-4.99 (m, 3H), 4.93 (d, *J* = 7.7 Hz, 1H), 4.54 (d, *J* = 11.5 Hz, 1H), 4.47 (d, *J* = 11.5 Hz, 1H), 4.30 (d, *J* = 8.1 Hz, 1H), 4.14 (m, 1H), 3.98 (dd, *J* = 2.0, 5.6 Hz, 1H), 3.85-3.40 (m, 11H), 2.75 (br s, 1H), 2.28 (m, 2H), 1.84-1.71 (m, 6H), 1.55-1.31 (m, 13 H), 0.89 (t, *J* = 7.2 Hz, 3H), 0.81 (s, 9H), 0.07 (s, 3H), -0.02 (s, 3H). ¹³C NMR (100 MHz, CD₂Cl₂): δ = 168.6, 159.7, 138.7, 135.7, 130.4, 129.9 (2C), 128.7, 116.8, 114.0 (2C), 110.0, 100.1, 99.9, 80.2, 79.3, 78.4, 77.0, 75.9, 74.7, 73.6, 73.3, 71.2, 69.5, 68.9, 55.6, 38.8, 36.9, 28.0, 26.5, 25.9 (3C), 18.8, 18.3, 16.8, 14.5, 14.1, 12.2, -3.5, -4.9. HRMS: calcd. for C₄₁H₆₆O₁₂Si₁ [M⁺+Na] 801.421577, found 801.422035.



Compound **6**: ¹H NMR (400 MHz, CD₂Cl₂): δ = 7.13 (d, *J* = 8.8 Hz, 2H), 6.90 (m, 1H), 6.82 (d, *J* = 8.8 Hz, 2H), 5.91 (ddt, *J* = 7.1, 10.2, 17.2 Hz, 1H), 5.22 (t, *J* = 9.2 Hz, 1H), 5.11-5.03 (m, 2H), 4.95 (d, *J* = 7.6 Hz, 1H), 4.45 (m, 2H), 4.29 (d, *J* = 8.1 Hz, 1H), 4.15 (dd, *J* = 5.7, 6.8 Hz, 1H), 3.98 (dd, *J* = 2.1, 5.6 Hz, 1H), 3.84-3.76 (m, 6H), 3.71-3.65 (m, 2H), 3.61 (t, *J* = 9.5 Hz, 1H), 3.52 (dd, *J* = 7.5, 9.0, 1H), 3.42-3.38 (m, 1H),

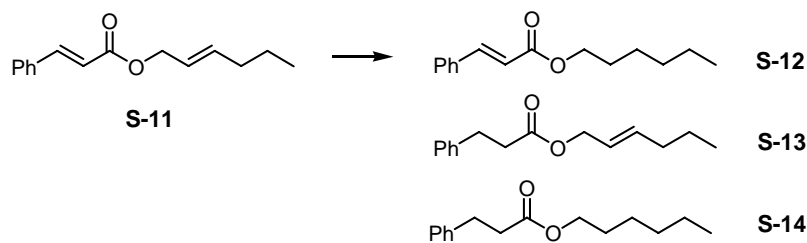


¹² Johansson, R.; Samuelsson, B. *J. Chem. Soc., Perkin Trans. 1* **1984**, 2371-2374.

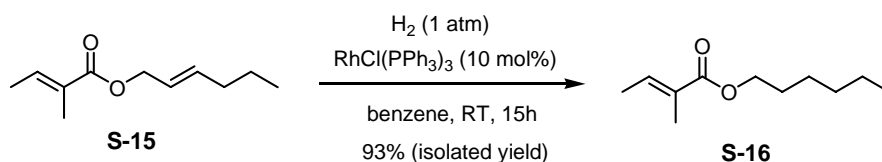
2.28 (m, 2H), 1.84-1.79 (m, 6H), 1.65 (br s, 1H), 1.55-1.32 (m, 13 H), 0.09 (t, $J = 7.2$ Hz, 3H), 0.82 (s, 9H), 0.07 (s, 3H), -0.03 (s, 3H). ^{13}C NMR (100 MHz, CD_2Cl_2): $\delta = 167.1, 159.7, 138.26, 135.6, 130.5, 130.1$ (2C), 129.0, 116.8, 114.0 (2C), 110.0, 100.2, 100.1, 80.0, 78.5, 77.5, 77.0, 76.6, 76.3, 75.5, 74.3, 73.8, 69.0, 62.0, 55.5, 38.8, 36.9, 28.1, 26.3, 25.8 (3C), 18.7, 18.2, 16.7, 14.6, 14.1, 12.3, $-3.6, -4.8$.

The assigned regiochemistry was further corroborated by acylation of both isomers. The corresponding ring proton H-6,6' (m, 2H) in **6** and H-4 (app. t, 1H) in **7** showed the expected acylation shifts.

Model Studies on Selective Hydrogenation

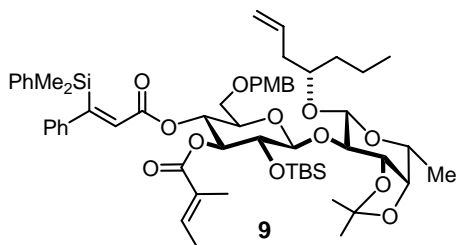


| Conditions | Product Distribution (NMR) | | | |
|---|----------------------------|------|------|------|
| | S-11 | S-12 | S-13 | S-14 |
| H_2 (1 atm), $\text{RhCl}(\text{PPh}_3)_3$ (3 mol%), benzene, RT, 4h | 15% | 52% | 7% | 26% |
| $\text{KOOCN}=\text{NCOOK}$ (2 eq.), AcOH, MeOH, RT, 15h | 44% | 14% | 33% | 9% |



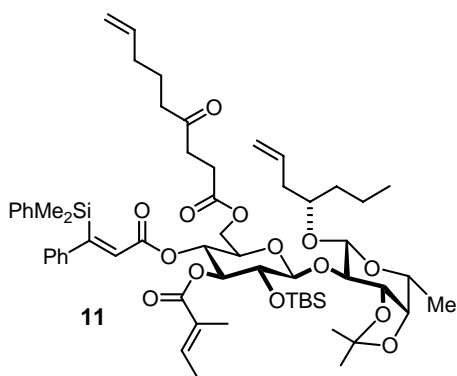
Total Synthesis of Ipomoeassin B

Compound 9. Et₃N (43 μ L, 0.305 mmol) and 2,4,6-trichlorobenzoyl chloride (22 mL, 0.139 mmol) were added to a solution of acid **8**



(39 mg, 0.139 mmol) in toluene (0.6 mL) and the resulting mixture was stirred for 1.5 h at ambient temperature before a solution of the mixture of alcohols **7** and **6** (54 mg, 0.0693 mmol) and DMAP (8.0 mg, 0.0693 mmol) in CH₂Cl₂ (0.5

mL) was introduced. After stirring for 3 h, the mixture was filtered through a pad of silica, the filtrate was evaporated and the residue was purified by flash chromatography (hexanes/EtOAc, 6/1) to give product **9** as a colorless oil (56.8 mg, 79%). $[\alpha]_D^{20} = -4.4$ (c 0.5, CHCl₃). IR (KAP): $\tilde{\nu} = 3070, 2956, 2932, 2904, 2858, 1730, 1652, 1612, 1587, 1513, 1248, 1160, 1074, 915, 839, 780, 701$. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.49-7.46$ (m, 2H), 7.26-7.18 (m, 10 H), 6.97-6.94 (m, 2H), 6.84-6.80 (m, 3H), 6.31 (s, 1H), 5.94 (ddt, $J = 7.1, 10.2, 17.2$ Hz, 1H), 5.27 (t, $J = 9.4$ Hz, 1H), 5.08-4.99 (m, 3H), 4.90 (d, $J = 7.6$ Hz, 1H), 4.41 (d, $J = 11.6$, 1H), 4.39 (d, $J = 11.6$, 1H), 4.31 (d, $J = 7.7$ Hz, 1H), 4.15 (t, $J = 6.1$ Hz, 1H), 3.97-3.90 (m, 2H), 3.78-3.74 (m, 4H), 3.66-3.53 (m, 3H), 3.36-3.34 (m, 2H), 2.29-2.26 (m, 2H), 1.76-1.75 (m, 6H), 1.56-1.24 (m, 11H), 0.86 (t, $J = 7.2$ Hz, 3H), 0.81 (s, 9H), 0.41 (s, 3H), 0.26 (s, 3H), 0.04 (s, 3H), -0.06 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.9, 165.8, 164.8, 159.0, 144.7, 138.3, 137.9, 135.4, 133.9$ (3C), 132.9, 130.3, 129.4 (2C), 128.6, 128.3, 127.8 (2C), 127.4 (2C), 126.8, 126.6 (2C), 116.5, 113.6, 109.7, 100.2, 99.8, 79.6, 78.7, 76.4, 76.0, 75.3, 73.6, 73.3, 73.1, 69.9, 68.5, 68.4, 55.2, 38.5, 36.6, 27.7, 26.3, 25.7 (3C), 18.3, 18.0, 16.7, 14.3, 14.1, 12.0, -0.8, -1.5, -3.9, -5.0. MS (EI): m/z (%): 83 (14), 121 (100), 157 (9), 265 (30), 565 (4), 985 (2). HRMS: calcd. for C₅₈H₈₂O₁₃Si₂ [M⁺+Na] 1065.518622, found 1065.517790.

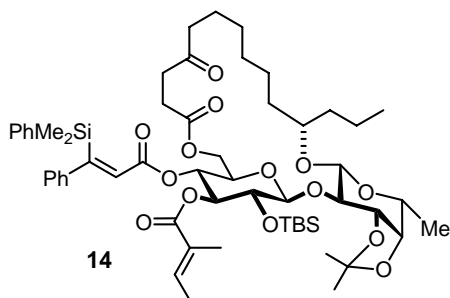


Synthesis of 11. DDQ (49.0 mg, 0.216 mmol) was added to a solution of compound **9** (150 mg, 0.144 mmol) in CH₂Cl₂ (6.0 mL) and water (0.3 mL). The mixture was stirred at room temperature for 16 h before it was filtered through a pad of silica which was carefully rinsed with ethyl acetate. The combined filtrates were evaporated and the residue was purified by flash chromatography (hexanes/EtOAc, 6/1) to give alcohol **10** (150 mg)

which contained *p*-methoxybenzaldehyde as an inseparable impurity.

Et₃N (45 μ L, 0.318 mmol) and 2,4,6-trichlorobenzoyl chloride (23 μ L, 0.148 mmol) were added to a solution of 4-oxo-8-nonenoic acid¹³ (25 mg, 0.0862 mmol) in toluene (1.0 mL) and the mixture was stirred for 1.5 h before a solution of the crude alcohol **10** (75 mg, ca. 0.074 mmol) and DMAP (9.0 mg, 0.074 mmol) in toluene (1.5 mL) was introduced. Stirring was continued for 2 h, the mixture was passed through a pad of silica which was carefully rinsed with EtOAc, the combined filtrates were evaporated, and the residue was purified by flash chromatography (hexanes/EtOAc, 8/1) to give disaccharide **11** as a colorless syrup (61.7 mg, 78%). $[\alpha]_D^{20} = -4.2$ (c 0.43, CH₂Cl₂). IR (KAP): $\tilde{\nu} = 3071, 2956, 2933, 1732, 1651, 1588, 1248, 1157, 914, 839, 780, 730, 701$. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.47-7.44$ (m, 2H), 7.28-7.18 (m, 6H), 6.98-6.95 (m, 2H), 6.81 (m, 1H), 6.33 (s, 1H), 5.93 (ddt, $J = 7.1, 10.1, 17.1$ Hz, 1H), 5.77 (ddt, $J = 6.4, 10.3, 17.0$ Hz, 1H), 5.27 (t, $J = 9.4$ Hz, 1H), 5.08-4.96 (m, 5H), 4.94 (d, $J = 7.6$ Hz, 1H), 4.28 (d, $J = 7.8$ Hz, 1H), 4.14 (t, $J = 5.9$ Hz, 1H), 4.01-3.99 (m, 2H), 3.97 (dd, $J = 2.0, 5.6$ Hz, 1H), 3.89-3.85 (m, 1H), 3.77 (m, 1H), 3.68-3.61 (m, 2H), 3.55 (dd, $J = 7.6, 9.1$ Hz, 1H), 2.67-2.42 (m, 6H), 2.27 (m, 2H), 2.09-2.03 (m, 2H), 1.77-1.75 (m, 6H), 1.59-1.34 (m, 15 H), 0.89 (t, $J = 7.2$ Hz, 3H), 0.81 (s, 9H), 0.39 (s, 3H), 0.27 (s, 3H), 0.04 (s, 3H), -0.06 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 208.6, 172.4, 166.9, 166.6, 164.7, 144.7, 138.2, 138.0, 135.4, 133.8$ (2C), 132.6, 128.7, 128.2, 127.9 (2C), 127.6 (2C), 126.9, 126.6 (2C), 116.5, 115.3, 109.7, 100.3, 99.8, 79.7, 79.1, 77.2, 76.5, 76.0, 75.2, 73.2, 71.5, 69.4, 68.6, 63.0, 41.9, 38.6, 37.1, 36.7, 33.1, 28.0, 27.8, 26.4, 25.7 (3C), 22.8, 18.3, 18.0, 16.7, 14.4, 14.1, 12.1, -0.7, -1.4, -3.8, -4.9. MS (EI): m/z (%): 83 (68), 157 (29), 265 (100), 393 (30), 775 (9), 793 (42), 961 (13), 1017 (7). HRMS: calcd. for C₅₉H₈₆O₁₄Si₂ [M⁺+Na] 1097.544832, found 1097.544163.

Compound 14. The ruthenium carbene complex **12** (5.0 mg, 0.00558 mmol) was added



to a solution of diene **11** (60 mg, 0.0558 mmol) in CH₂Cl₂ (10 mL) and the resulting mixture was refluxed for 4 h before the reaction was quenched with ethyl vinyl ether. Evaporation of all volatile materials followed by flash chromatography of the residue (hexanes/EtOAc, 4/1) gave the corresponding metathesis product **13** as a mixture

of (*E*)- and (*Z*)-isomers.

¹³ Hodgson, D. M.; Stuppel, P. A.; Pierard, F. Y. T. M.; Labande, A. H.; Johnstone, C. *Chem. Eur. J.* **2001**, *7*, 4465.

A solution of cycloalkene **13** thus obtained (42 mg, 0.0398 mmol) and $\text{RhCl}(\text{PPh}_3)_3$ (7.0 mg, 0.0076 mmol) in EtOH (0.6 mL) was stirred under an atmosphere of H_2 (1 atm) overnight. Evaporation of the solvent and flash chromatography of the crude product (hexanes/EtOAc, 4/1) gave compound **14** as a colorless oil (34 mg, 81%). $[\alpha]_{\text{D}}^{20} = +0.6$ (c 0.50, CH_2Cl_2). IR (KAP): $\tilde{\nu} = 3069, 2932, 2858, 1733, 1652, 1588, 1248, 1155, 1074, 839, 780, 730, 701$. ^1H NMR (400 MHz, CDCl_3): $\delta = 7.46\text{--}7.45$ (m, 2H), 7.28–7.17 (m, 6H), 6.97–6.95 (m, 2H), 6.86 (m, 1H), 6.31 (s, 1H), 5.27 (t, $J = 9.3$ Hz, 1H), 5.08 (t, $J = 9.8$ Hz, 1H), 4.93 (d, $J = 7.7$ Hz, 1H), 4.27 (d, $J = 7.8$ Hz, 1H), 4.20–4.13 (m, 2H), 4.00–3.97 (m, 2H), 3.87 (dd, $J = 6.6, 7.7$ Hz, 1H), 3.78 (m, 1H), 3.63 (m, 1H), 3.54 (dd, $J = 7.8, 9.0$ Hz, 1H), 3.51 (m, 1H), 2.81–2.73 (m, 1H), 2.58–2.31 (m, 5H), 1.79–1.24 (m, 29H), 0.90 (t, $J = 7.3$ Hz, 3H), 0.81 (s, 9H), 0.37 (s, 3H), 0.24 (s, 3H), 0.05 (s, 3H), -0.04 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3): $\delta = 209.6, 171.4, 166.8, 166.3, 164.4, 144.7, 138.4, 138.2, 133.8$ (2C), 132.7, 128.6, 128.2, 127.8 (2C), 127.5 (2C), 126.8, 126.6 (2C), 109.7, 101.5, 99.8, 82.0, 79.4, 76.5, 76.4, 75.6, 73.1, 71.7, 68.4, 68.3, 61.7, 42.3, 37.3, 37.0, 33.7, 28.7, 28.3, 28.0, 27.8, 26.3, 25.7 (3C), 24.0, 23.7, 18.4, 18.0, 16.8, 14.4, 14.3, 12.1, $-0.5, -1.6, -3.8, -4.9$. MS (EI): m/z (%): 83 (91), 157 (29), 265 (100), 349 (13), 767 (36), 991 (9). HRMS: calcd. for $\text{C}_{57}\text{H}_{84}\text{O}_{14}\text{Si}_2$ [$\text{M}^+ + \text{Na}$] 1071.529187, found 1071.530087.

Synthesis of Ipomoeassin B (1). A solution of TASF (79 mg, 0.286 mmol) in MeCN (2.0 mL) was added to a solution of compound **14** (30 mg, 0.0286 mmol) in wet MeCN (1.5 mL). After stirring for 4 h, the mixture was filtered through a pad of silica which was carefully rinsed with EtOAc, the combined filtrates were evaporated, and the residue was treated with trifluoroacetic acid (16 μL , 215 mmol) in CH_2Cl_2 (2.0 mL). After stirring for 3 h, the solution was neutralized with Et_3N , the solvent was evaporated, and the residue purified by flash chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 20/1) to afford Ipomoeassin B (**1**) as a colorless syrup which solidifies when kept in the freezer (10 mg, 45%). $[\alpha]_{\text{D}}^{25} = -48.0$ (c 0.36, EtOH); lit.¹⁴ $[\alpha]_{\text{D}}^{25} = -39$ (c 0.3, EtOH). IR (KAP): $\tilde{\nu} = 3365, 3062, 2932, 1744, 1719, 1631, 1371, 1316, 1265, 1249, 1157, 1138, 1073$. ^1H NMR (400 MHz, C_6D_6) and ^{13}C NMR (100 MHz, C_6D_6) data, see Table S1 and S2, respectively. MS (EI): m/z (%): 513 (11), 467 (14), 349 (10), 241 (13), 223 (42), 131 (44), 111 (13), 83 (100), 55 (28). HRMS: calcd. for $\text{C}_{40}\text{H}_{56}\text{O}_{14}$ [$\text{M}^+ + \text{Na}$] 783.356230, found 783.356291.

¹⁴ Cao, S.; Guza, R. C.; Wisse J. H.; Miller, J. S.; Evans, R.; Kingston, D. G. I. *J. Nat. Prod.* **2005**, *68*, 487.

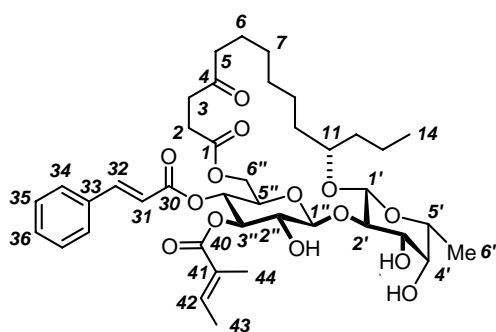


Table S1. Comparison of the published ^1H NMR data (C_6D_6) of Ipomoeassin B (**1**) (500 MHz) with those of the synthetic sample (Bruker dpx300, 300 MHz). Numbering scheme as shown in the insert.

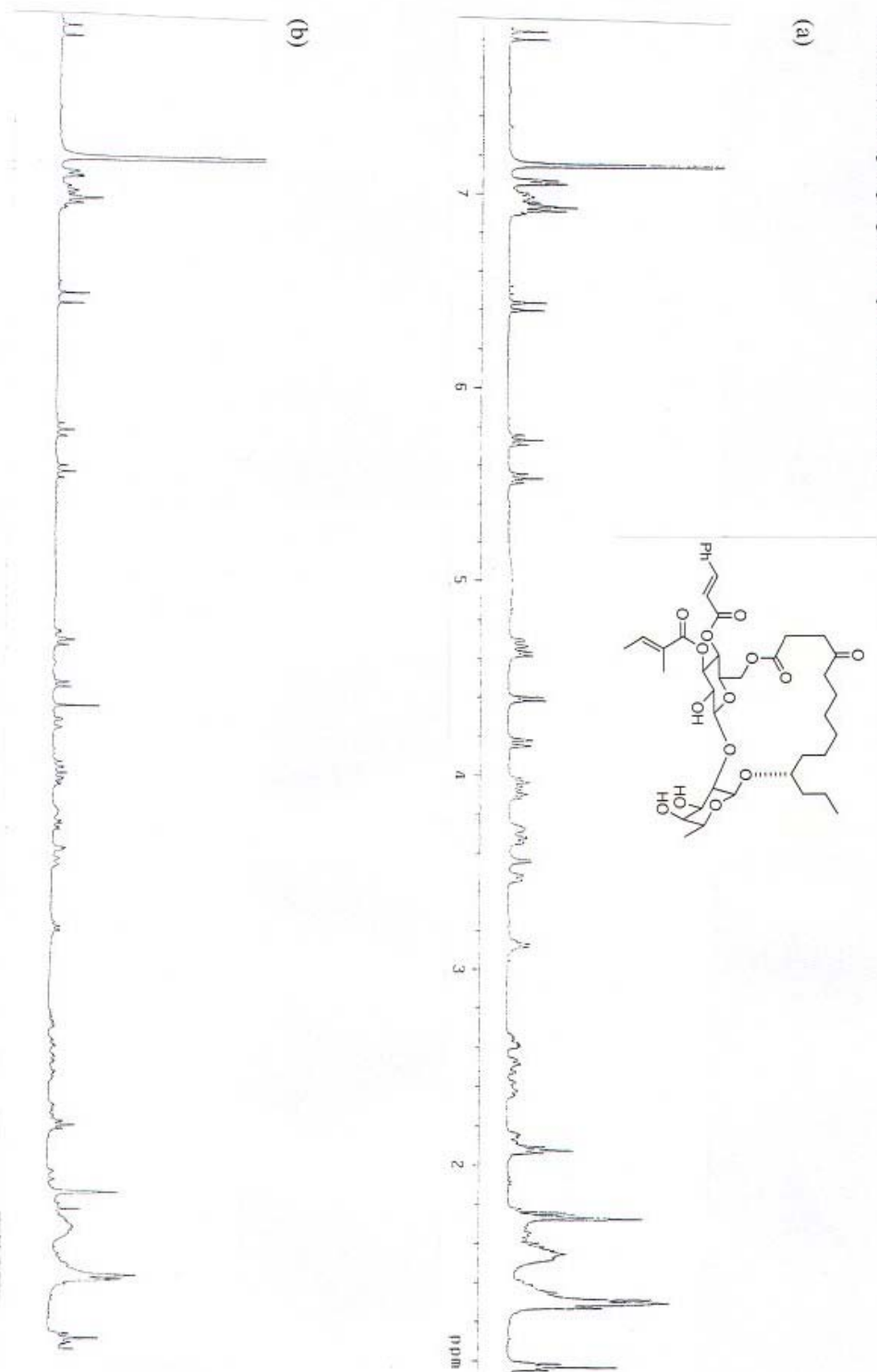
| Position | Ipomoeassin B (<i>J</i> in Hz) | Synthetic Sample (<i>J</i> in Hz) ^a |
|----------|---------------------------------|---|
| 2 | 2.38 ddd (17.4, 9.4, 3.4) | 2.37 ddd (17.1, 9.3, 3.5) |
| | 2.13 ddd (17.4, 7.7, 3.5) | 2.15 ddd (17.1, 7.6, 3.3) |
| 3 | 2.62 ddd (16.1, 7.7, 3.4) | 2.63 ddd (16.1, 7.7, 3.3) |
| | 2.50 ddd (16.1, 9.4, 3.5) | 2.49 ddd (16.1, 9.2, 3.4) |
| 5 | 2.07 t (6.2) | 2.08 t (6.2) |
| 11 | 3.71 m | 3.73 m |
| 14 | 0.96 t (7.1) | 0.97 t (7.0) |
| 1' | 4.38 d (7.6) | 4.39 d (7.6) |
| 2' | 3.88 dd (9.5, 7.6) | 3.88 dd (9.8, 7.5) |
| 3' | 3.65 dd (9.5, 3.3) | 3.65 dd (9.6, 3.3) |
| 4' | 3.53 brs | 3.53 br s |
| 5' | 3.11 brq (6.4) | 3.11 br q (6.3) |
| 6' | 1.29 d (6.4) | 1.29 d (6.4) |
| 1'' | 4.59 d (7.8) | 4.61 d (7.8) |
| 2'' | 3.95 dd (9.7, 7.8) | 3.95 dd (9.6, 8.0) |
| 3'' | 5.50 t (9.7) | 5.51 t (9.6) |
| 4'' | 5.72 t (9.7) | 5.73 t (9.6) |
| 5'' | 3.44 brd (9.7) | 3.46 brm |
| 6'' | 4.66 dd (12.6, 2.1) | 4.64 m |
| | 4.16 brd (12.6) | 4.18 br d (12.4) |
| 31 | 6.40 d (16.1) | 6.41 d (16.0) |
| 32 | 7.81 d (16.1) | 7.82 d (16.0) |
| 34 | 6.88-7.07 | 6.88-7.08 |
| 35 | 6.88-7.07 | 6.88-7.08 |
| 36 | 6.88-7.07 | 6.88-7.08 |
| 42 | 6.95 m | 6.94 m |
| 43 | 1.27 d (7.1) | 1.29 d (7.1) |
| 44 | 1.72 brs | 1.73 brs |

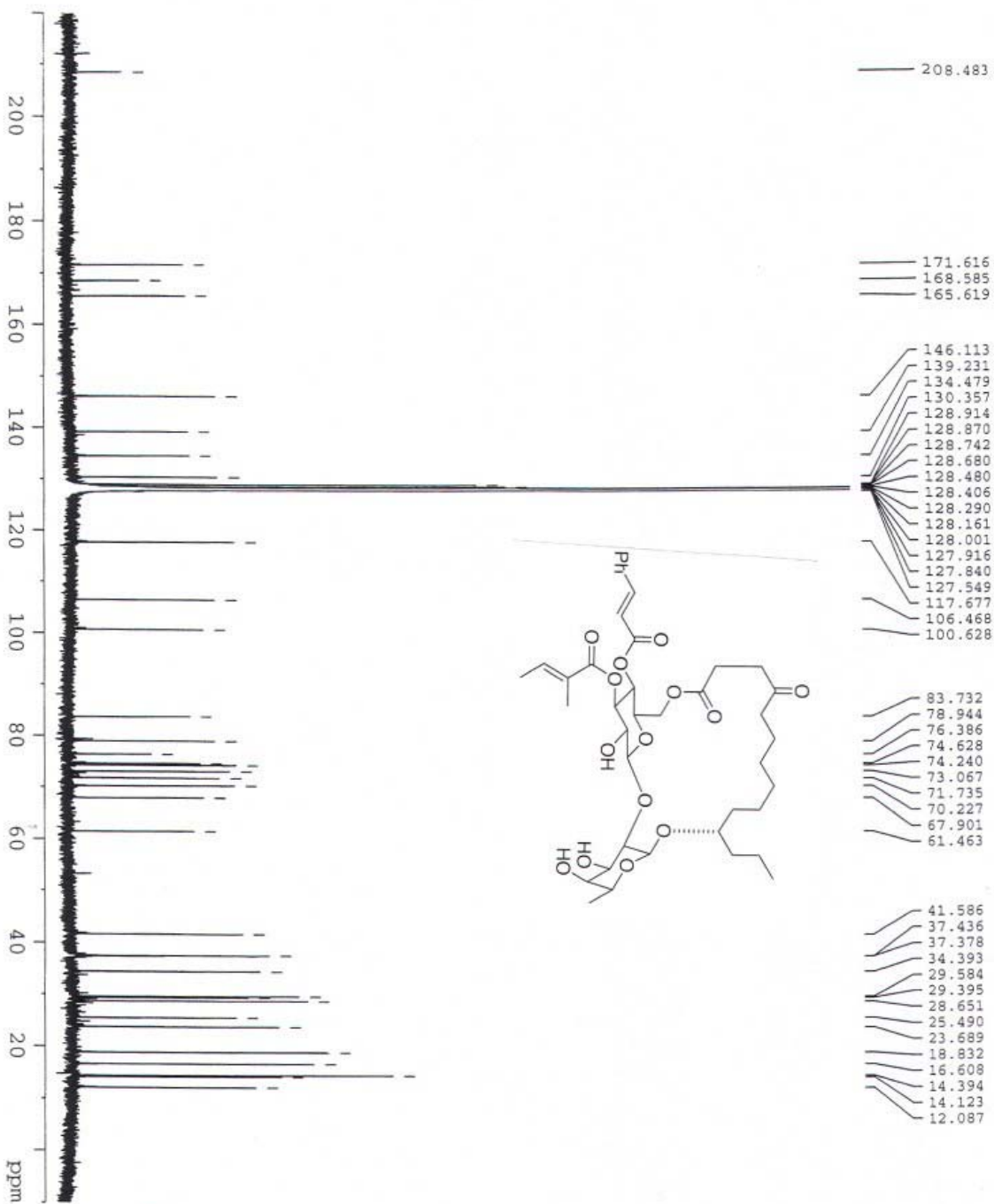
^a The protons at positions 6-10, 12 and 13 appear between δ 1.73-1.26 as complicated multiplet.

Table S2. Comparison of the published ^{13}C NMR data (C_6D_6) of Ipomoeassin B (**1**) (100 MHz) with those of the synthetic samples (Bruker DMX-600, 150 MHz).

| Position | Ipomoeassin B | Synthetic Sample |
|------------|---------------|------------------|
| <i>1</i> | 171.5 | 171.6 |
| <i>2</i> | 37.4 | 37.4 |
| <i>3</i> | 29.5 | 29.6 |
| <i>4</i> | 208.3 | 208.5 |
| <i>5</i> | 41.5 | 41.6 |
| <i>6</i> | 23.6 | 23.7 |
| <i>7</i> | 28.6 | 28.7 |
| <i>8</i> | 29.3 | 29.4 |
| <i>9</i> | 25.4 | 25.5 |
| <i>10</i> | 34.3 | 34.4 |
| <i>11</i> | 78.8 | 78.9 |
| <i>12</i> | 37.4 | 37.4 |
| <i>13</i> | 18.8 | 18.8 |
| <i>14</i> | 14.3 | 14.4 |
| <i>1'</i> | 100.6 | 100.6 |
| <i>2'</i> | 83.7 | 83.7 |
| <i>3'</i> | 74.2 | 74.2 |
| <i>4'</i> | 71.6 | 71.7 |
| <i>5'</i> | 70.2 | 70.2 |
| <i>6'</i> | 14.0 | 14.1 |
| <i>1''</i> | 106.4 | 106.5 |
| <i>2''</i> | 74.6 | 74.6 |
| <i>3''</i> | 76.3 | 76.4 |
| <i>4''</i> | 67.8 | 67.9 |
| <i>5''</i> | 73.0 | 73.1 |
| <i>6''</i> | 61.4 | 61.5 |
| <i>30</i> | 165.5 | 165.6 |
| <i>31</i> | 117.6 | 117.7 |
| <i>32</i> | 146.0 | 146.1 |
| <i>33</i> | 134.4 | 134.5 |
| <i>34</i> | 128.4 | 128.5 |
| <i>35</i> | 128.8 | 128.9 |
| <i>36</i> | 130.3 | 130.4 |
| <i>40</i> | 168.5 | 168.6 |
| <i>41</i> | 128.0 | 127.9 |
| <i>42</i> | 139.1 | 139.2 |
| <i>43</i> | 16.6 | 16.6 |
| <i>44</i> | 12.0 | 12.1 |

¹H NMR spectra of Ipomoeassin B in C₆D₆: (a) natural product reported in literature (500 MHz, Kingston et al. *J. Nat. Prod.* **2005**, 68, 487); (b) sample prepared by us (400 MHz).





- 208.483
- 171.616
- 168.585
- 165.619
- 146.113
- 139.231
- 134.479
- 130.357
- 128.914
- 128.870
- 128.742
- 128.680
- 128.480
- 128.406
- 128.290
- 128.161
- 128.001
- 127.916
- 127.840
- 127.549
- 117.677
- 106.468
- 100.628
- 83.732
- 78.944
- 76.386
- 74.628
- 74.240
- 73.067
- 71.735
- 70.227
- 67.901
- 61.463
- 41.586
- 37.436
- 37.378
- 34.393
- 29.584
- 29.395
- 28.651
- 25.490
- 23.689
- 18.832
- 16.608
- 14.394
- 14.123
- 12.087

C609535

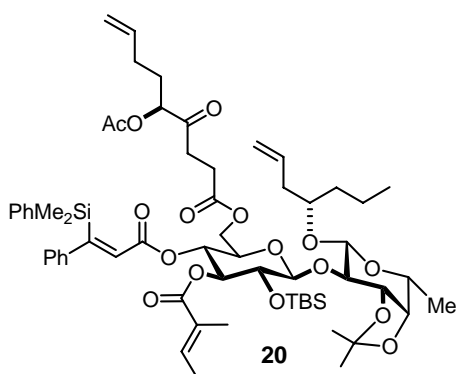
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 PROCNO 11
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 PULPROG zgpg30
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 NS 6424
 DS 200
 SWH 37594.402 Hz
 SFO1 101.626130 MHz
 AQ 0.573645 sec
 FIDRES 0.8716891 Hz
 AQ 13.200 usec
 DE 5.50 usec
 TE 303.0 K
 D1 0.03000000 sec
 D11 0.03000000 sec
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 P1 16.00 usec
 PL1 0.00 dB
 SFO1 150.9413646 MHz
 ===== CHANNEL f2 =====
 GRPRG2 waltz16
 NUC2 13C
 P2 70.00 usec
 PL2 0.00 dB
 PL12 16.31 dB
 SFO2 600.2234213 MHz
 F2 - Processing parameters
 SI 131072
 SF 150.9253900 MHz
 WDW EM
 SSB 0
 GB 1.00 Hz
 PC 1.00

NAG-NB-052-02

DMX-600

Total Synthesis of Ipomoeassin E

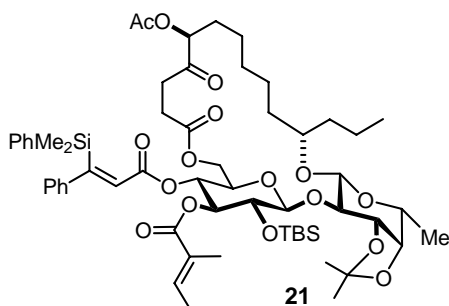
Disaccharide 20. DDQ (19.0 mg, 0.0830 mmol) was added to a solution of compound **9** (57.7 mg, 0.0553 mmol) in CH₂Cl₂ (2.0 mL) and water (0.1 mL), and the resulting mixture was stirred at ambient temperature for 16 h. The suspension was then filtered through a pad of silica which was rinsed with ethyl acetate, the combined filtrates were evaporated, and the residue was purified by flash chromatography (hexanes/EtOAc, 6/1) to give alcohol **10** (45.6 mg) which contained traces of *p*-methoxybenzaldehyde.



Et₃N (26 μL, 0.185 mmol) and 2,4,6-trichlorobenzoyl chloride (13 mL, 0.0862 mmol) were added to a solution of carboxylic acid **19** (19.7 mg, 0.0862 mmol) in toluene (0.5 mL). After stirring for 1.5 h at ambient temperature, a solution of the crude alcohol **10** (45.6 mg) and DMAP (5.0 mg, 0.0431 mmol) in toluene (1.0 mL) was introduced and stirring was continued for 2 h. The mixture was then filtered through a pad of

silica which was carefully rinsed with EtOAc, the combined filtrates were evaporated and the residue was purified by flash chromatography (hexanes/EtOAc, 8/1) to give product **20** as a colorless syrup (54.6 mg, 87% over both steps). $[\alpha]_D^{20} = -3.7$ (c 0.40, CH₂Cl₂). IR (KAP): $\tilde{\nu} = 3072, 2956, 2932, 2858, 1732, 1651, 1642, 1586, 1428, 1248, 1113, 916, 839, 780$. ¹H NMR (400 MHz, CD₂Cl₂): $\delta = 7.47-7.45$ (m, 2H), 7.29-7.19 (m, 6H), 7.00-6.97 (m, 2H), 6.82 (m, 1H), 6.33 (s, 1H), 5.89 (ddt, $J = 6.9, 10.2, 17.2$ Hz, 1H), 5.81 (ddt, $J = 6.6, 10.3, 17.0$ Hz, 1H), 5.25 (t, $J = 9.4$ Hz, 1H), 5.10-4.99 (m, 7H), 4.30 (d, $J = 8.1$ Hz, 1H), 4.16-4.03 (m, 3H), 3.99 (dd, $J = 1.9, 5.5$ Hz, 1H), 3.82-3.65 (m, 4H), 3.60 (m, 1H), 2.83-2.47 (m, 4H), 2.28 (m, 2H), 2.18-2.11 (m, 5H), 1.96-1.74 (m, 8H), 1.55-1.32 (m, 13 H), 0.90 (t, $J = 7.2$ Hz, 3H), 0.81 (s, 9H), 0.37 (s, 3H), 0.28 (s, 3H), 0.07 (s, 3H), -0.04 (s, 3H). ¹³C NMR (100 MHz, CD₂Cl₂): $\delta = 205.8, 172.4, 170.7, 167.1, 166.9, 165.0, 145.1, 138.7, 137.5, 135.6, 134.2$ (2C), 133.0, 129.0, 128.5, 128.2 (2C), 127.9 (2C), 127.2, 126.9 (2C), 116.8, 115.9, 110.0, 100.1, 100.0, 80.2, 78.6, 78.0, 77.1, 76.4, 75.6, 73.5, 72.0, 69.6, 69.0, 63.1, 38.8, 36.9, 33.7, 33.6, 30.1, 29.6, 28.2, 27.5, 26.4, 25.8 (3C), 20.8, 18.8, 18.2, 16.7, 14.6, 14.2, 12.2, -0.7, -1.2, -3.6, -4.8. MS (EI): m/z (%): 83 (63), 157 (29), 265 (100), 265 (100), 451 (32), 833 (10), 851 (49), 1019 (17), 1075 (8). HRMS: calcd. for C₆₁H₈₈O₁₆Si₂ [M⁺+Na] 1155.550316, found 1155.548973.

Product 21. The ruthenium carbene complex **12** (6.0 mg, 0.00653 mmol) was added to



a solution of compound **20** (74 mg, 0.0653 mmol) in CH₂Cl₂ (15 mL) and the resulting mixture was refluxed for 4 h before the reaction was quenched with ethyl vinyl ether. Evaporation of all volatile materials followed by flash chromatography of the residue (hexanes/EtOAc, 4/1) gave the corresponding cycloalkene (*E,Z*-mixture). This product was dissolved in EtOH (1.0 mL) and

stirred overnight in the presence RhCl(PPh₃)₃ (12 mg, 0.0125 mmol) under an atmosphere of H₂ (1 atm). The catalyst was filtered off through a pad of silica, the filtrate was evaporated and the residue was purified by flash chromatography (hexanes/EtOAc, 4/1) to give compound **21** as a colorless syrup (60 mg, 83% over both steps). $[\alpha]_D^{20} = +0.4$ (c 0.77, CH₂Cl₂). IR (KAP): $\tilde{\nu} = 3069, 2931, 2858, 1732, 1652, 1588, 1379, 1371, 1247, 1155, 1074, 839, 780, 731, 702$. ¹H NMR (400 MHz, CDCl₃): $\delta = 7.46-7.44$ (m, 2H), 7.29-7.19 (m, 6H), 6.99-6.97 (m, 2H), 6.81 (m, 1H), 6.31 (s, 1H), 5.28 (t, *J* = 9.2 Hz, 1H), 5.06-5.00 (m, 2H), 4.94 (d, *J* = 7.7 Hz, 1H), 4.25 (d, *J* = 7.8 Hz, 1H), 4.13 (t, *J* = 6.0 Hz, 1H), 4.03-4.02 (m, 1H), 3.98 (dd, *J* = 2.0, 5.8 Hz, 1H), 3.86 (dd, *J* = 6.4, 7.8 Hz, 1H), 3.78 (m, 1H), 3.65 (dt, *J* = 2.7, 10.0 Hz, 1H), 3.54-3.48 (m, 2H), 2.87 (dt, *J* = 6.8, 18.8 Hz, 1H), 2.62 (dt, *J* = 6.8, 18.8 Hz, 1H), 2.49 (m, 2H), 2.14 (s, 3H), 1.77-1.26 (m, 30 H), 0.09 (t, *J* = 7.3 Hz, 3H), 0.81 (s, 9H), 0.37 (s, 3H), 0.23 (s, 3H), 0.05 (s, 3H), -0.04 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 205.9, 170.9, 170.6, 166.8, 166.6, 164.4, 144.6, 138.3, 138.0, 133.6$ (2C), 132.5, 128.7, 128.2, 127.9 (2C), 127.6 (2C), 126.9, 126.6 (2C), 109.7, 101.3, 99.7, 81.9, 79.4, 78.4, 77.2, 76.3, 75.4, 73.1, 71.6, 68.5, 68.3, 61.8, 37.3, 33.9, 33.8, 29.9, 28.5, 28.0, 27.8, 26.3, 25.6 (3C), 24.2 (2C), 20.6, 18.3, 18.0, 16.8, 14.4, 14.3, 12.1, -0.4, -1.6, -3.8, -4.9. MS (EI): *m/z* (%): 83 (66), 157 (26), 265 (100), 567 (11), 825 (41), 1049 (9). HRMS: calcd. for C₅₉H₈₆O₁₆Si₂ [M⁺+Na] 1129.534663, found 1129.535235.

Ipomoeassin E (2). A solution of TASF (75 mg, 0.27 mmol) in MeCN (1.0 mL) was added to a solution of compound **21** (30 mg, 0.027 mmol) in wet MeCN (1.0 mL) and the resulting mixture was stirred for 5 h before it was filtered through a pad of silica which was carefully rinsed with EtOAc. The combined filtrates were evaporated and the residue was treated with trifluoroacetic acid (36 μ L, 0.49 mmol) in CH₂Cl₂ (3.0 mL) for 3 h at ambient temperature. The mixture was neutralized with triethylamine and concentrated in vacuo, and the crude product was purified by preparative HPLC (YMC-PACK ODS A, 5 μ m 12 nm; MeOH/H₂O = 70/30; flow rate: 10 mL/min;

pressure: 3.6 MPa) to afford Ipomoeassin E (**2**) as a colorless syrup which solidifies when kept in the freezer (14 mg, 63% over both steps). $[\alpha]_{\text{D}}^{25} = -32$ (c 0.21, EtOH); lit. $[\alpha]_{\text{D}}^{25} = -24$ (c 0.2, EtOH). IR (KAP): $\tilde{\nu} = 3408, 2933, 2869, 1725, 1636, 1450, 1374, 1309, 1248, 1156, 1073, 768$. ^1H NMR (400 MHz, C_6D_6) and ^{13}C NMR (75 MHz, C_6D_6) data, see Table S3 and S4, respectively. MS (EI): m/z (%): 655 (5), 571 (7), 525 (7), 407 (9), 281 (40), 239 (11), 221 (19), 192 (11), 131 (58), 110 (10), 83 (100), 55 (20), 43 (19). HRMS: calcd. for $\text{C}_{42}\text{H}_{58}\text{O}_{16}$ [$\text{M}^+ + \text{Na}$] 841.36172, found 841.362452.

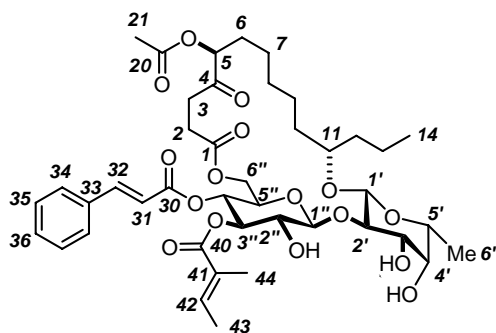


Table S3. Comparison of the ^1H NMR data (C_6D_6) reported for Ipomoeassin E (**2**) (500 MHz) with those of the synthetic material recorded on a Bruker av400 spectrometer (400 MHz). Numbering scheme as shown in the insert.

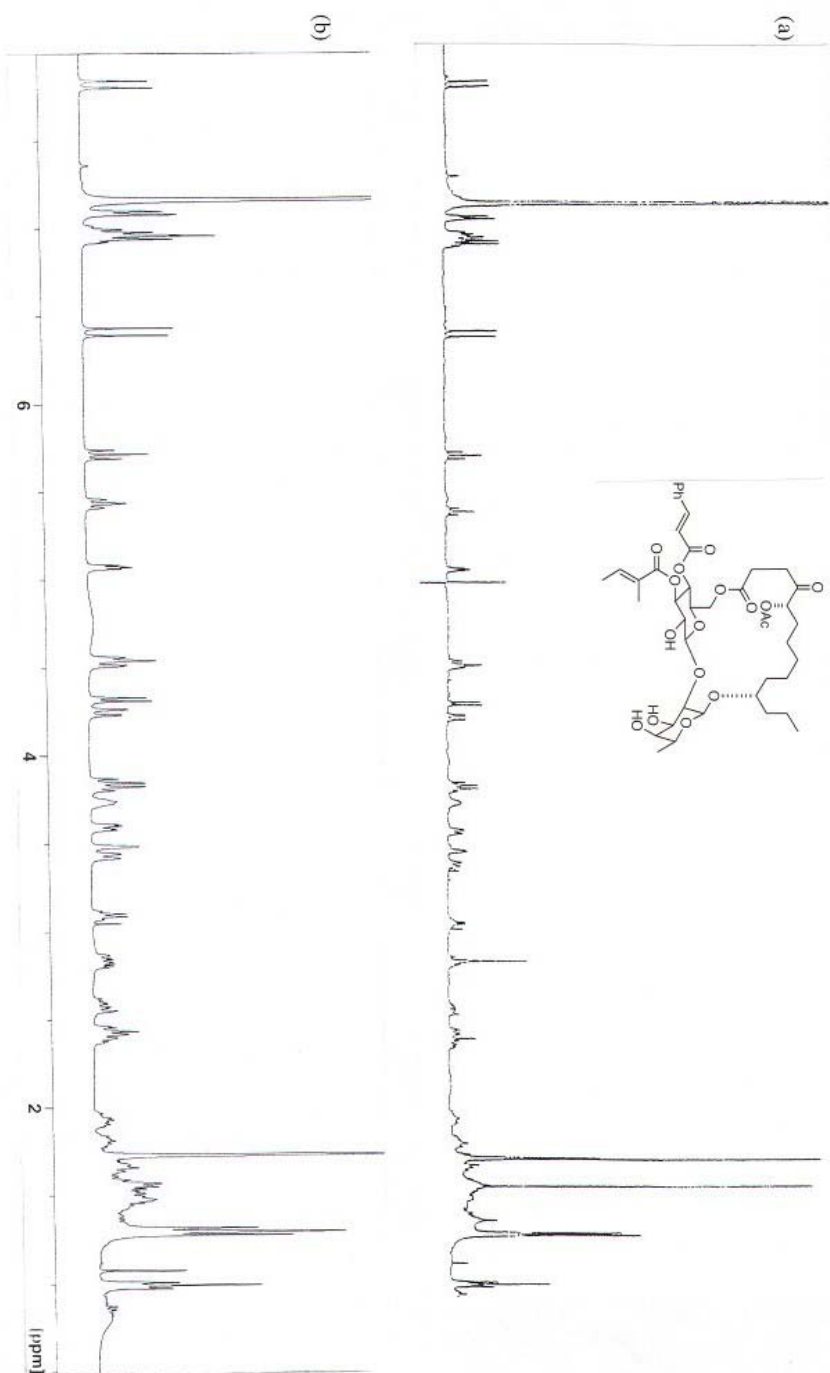
| Position | Ipomoeassin E (J in Hz) | Synthetic Sample (J in Hz) ^{a,b} |
|----------|--|--|
| 2 | 2.55 ddd (18.1, 9.2, 2.8) 2.34 ddd (18.1, 7.8, 3.4) | 2.59 ddd (18.0, 9.2, 3.1) 2.46-2.36 m |
| 3 | 2.81 ddd (16.3, 9.2, 3.4) 2.39 ddd (16.3, 7.8, 2.8) | 2.82 ddd (16.5, 9.2, 3.5) 2.46-2.36 m |
| 5 | 5.04 dd (6.2, 3.9) | 5.07 dd (6.1, 3.9) |
| 11 | 3.71 m | 3.74 brm |
| 14 | 0.97 t (6.9) | 0.99 t (7.0) |
| 1' | 4.28 d (7.6) | 4.32 d (7.6) |
| 2' | 3.80 dd (9.5, 7.6) | 3.82 appt (9.4) |
| 3' | 3.55 dd (9.5, 3.7) | 3.59 dd (9.3, 3.2) |
| 4' | 3.44 brs | 3.48 brs |
| 5' | 3.02 brq (6.4) | 3.09 brq (6.4) |
| 6' | 1.25 d (6.4) | 1.28 d (6.6) |
| 1'' | 4.50 d (7.8) | 4.55 d (8.0) |
| 2'' | 3.81 dd (9.7, 7.8) | 3.84 appt (9.5) |
| 3'' | 5.38 t (9.7) | 5.43 t (9.5) |
| 4'' | 5.70 t (9.7) | 5.71 t (9.7) |
| 5'' | 3.36 brd (9.7) | 3.43 brd (9.8) |
| 6'' | 4.51 dd (11.0, 2.0) 4.20 brd (11.0) | 4.52 dd (11.4, 2.2) 4.24 brd (11.5) |
| 21 | 1.67 brs | 1.73 brs |
| 31 | 6.40 d (16.1) | 6.41 d (16.0) |
| 32 | 7.81 d (16.1) | 7.82 d (16.0) |
| 34 | 6.88-7.03 | 6.91-7.09 |
| 35 | 6.88-7.03 | 6.91-7.09 |
| 36 | 6.88-7.03 | 6.91-7.09 |
| 42 | 6.95 m | 6.95 m |
| 43 | 1.25 d (7.1) | 1.30 d (7.3) |
| 44 | 1.67 brs | 1.73 brs |

^a The chemical shifts depend on the concentration of the sample and the dryness of the C_6D_6 used; see below; the data compiled in this Table were recorded using 3 mg of compound **2** in 0.6 mL of freshly distilled (CaH_2) C_6D_6 . ^b The protons at positions 6-10, 12 and 13 appear between δ 1.96-1.36 as complicated multiplet.

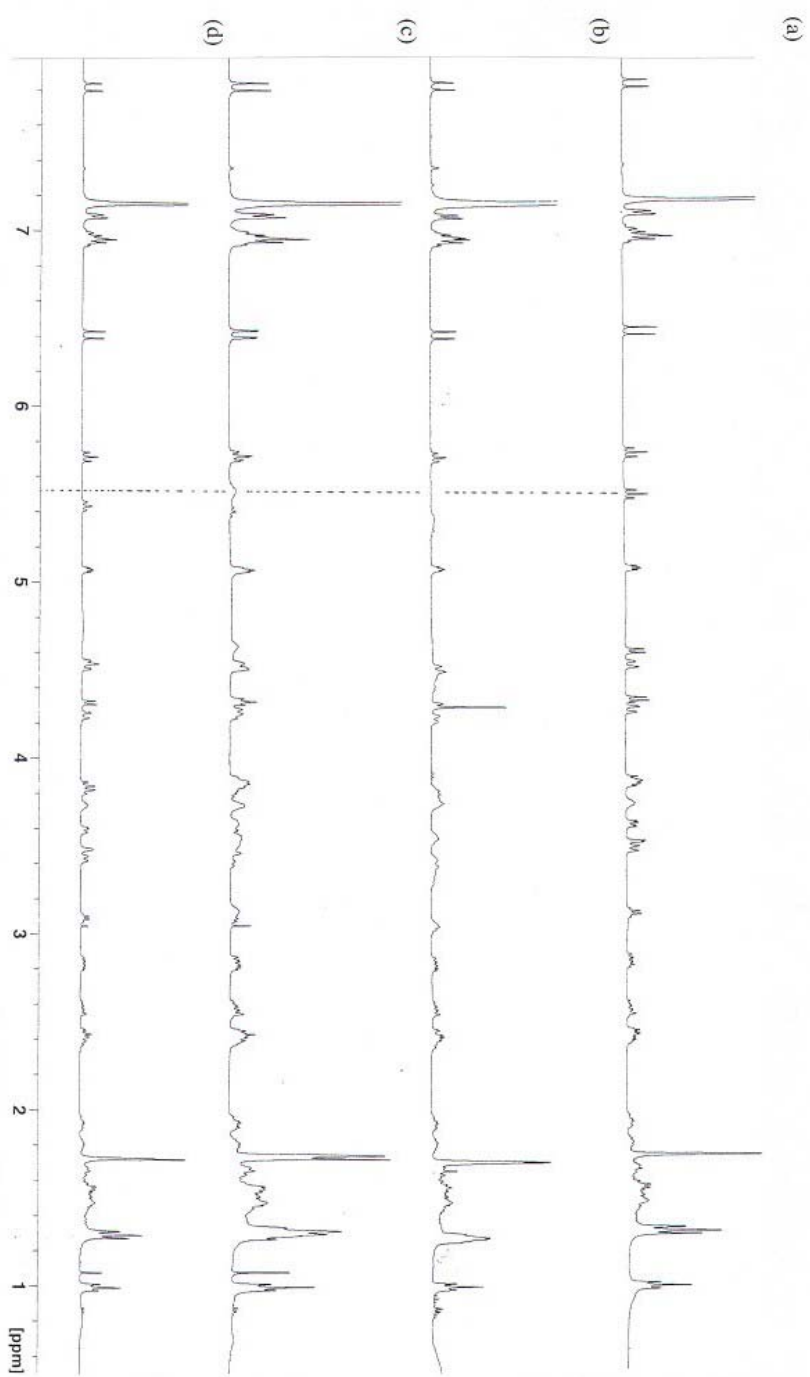
Table S4. Comparison of the ^{13}C NMR data (C_6D_6) reported for Ipomoeassin E (**2**) (125 MHz) with those of the synthetic sample (Bruker AMX-300 spectrometer, 75 MHz).

| Position | Natural Product | Synthetic Sample |
|------------|-----------------|------------------|
| <i>1</i> | 171.4 | 171.4 |
| <i>2</i> | 34.0 | 34.1 |
| <i>3</i> | 28.3 | 28.3 |
| <i>4</i> | 205.8 | 205.7 |
| <i>5</i> | 78.2 | 78.2 |
| <i>6</i> | 30.3 | 30.3 |
| <i>7</i> | 24.0 | 24.0 |
| <i>8</i> | 30.5 | 30.5 |
| <i>9</i> | 25.2 | 25.2 |
| <i>10</i> | 34.0 | 34.1 |
| <i>11</i> | 78.5 | 78.5 |
| <i>12</i> | 37.7 | 37.7 |
| <i>13</i> | 18.9 | 19.0 |
| <i>14</i> | 14.4 | 14.4 |
| <i>1'</i> | 100.5 | 100.4 |
| <i>2'</i> | 84.2 | 84.3 |
| <i>3'</i> | 74.0 | 74.0 |
| <i>4'</i> | 71.6 | 71.6 |
| <i>5'</i> | 70.0 | 70.0 |
| <i>6'</i> | 14.1 | 14.1 |
| <i>1''</i> | 106.7 | 106.7 |
| <i>2''</i> | 74.9 | 74.9 |
| <i>3''</i> | 76.6 | 76.7 |
| <i>4''</i> | 67.7 | 67.7 |
| <i>5''</i> | 72.9 | 73.0 |
| <i>6''</i> | 61.2 | 61.3 |
| <i>20</i> | 169.8 | 169.8 |
| <i>21</i> | 20.3 | 20.3 |
| <i>30</i> | 165.5 | 165.5 |
| <i>31</i> | 117.5 | 117.6 |
| <i>32</i> | 146.2 | 146.2 |
| <i>33</i> | 134.4 | 134.5 |
| <i>34</i> | 128.3 | 128.2 |
| <i>35</i> | 128.5 | 128.3 |
| <i>36</i> | 130.4 | 130.4 |
| <i>40</i> | 168.9 | 168.9 |
| <i>41</i> | 128.0 | 128.0 |
| <i>42</i> | 139.6 | 139.5 |
| <i>43</i> | 16.6 | 16.5 |
| <i>44</i> | 12.0 | 12.0 |

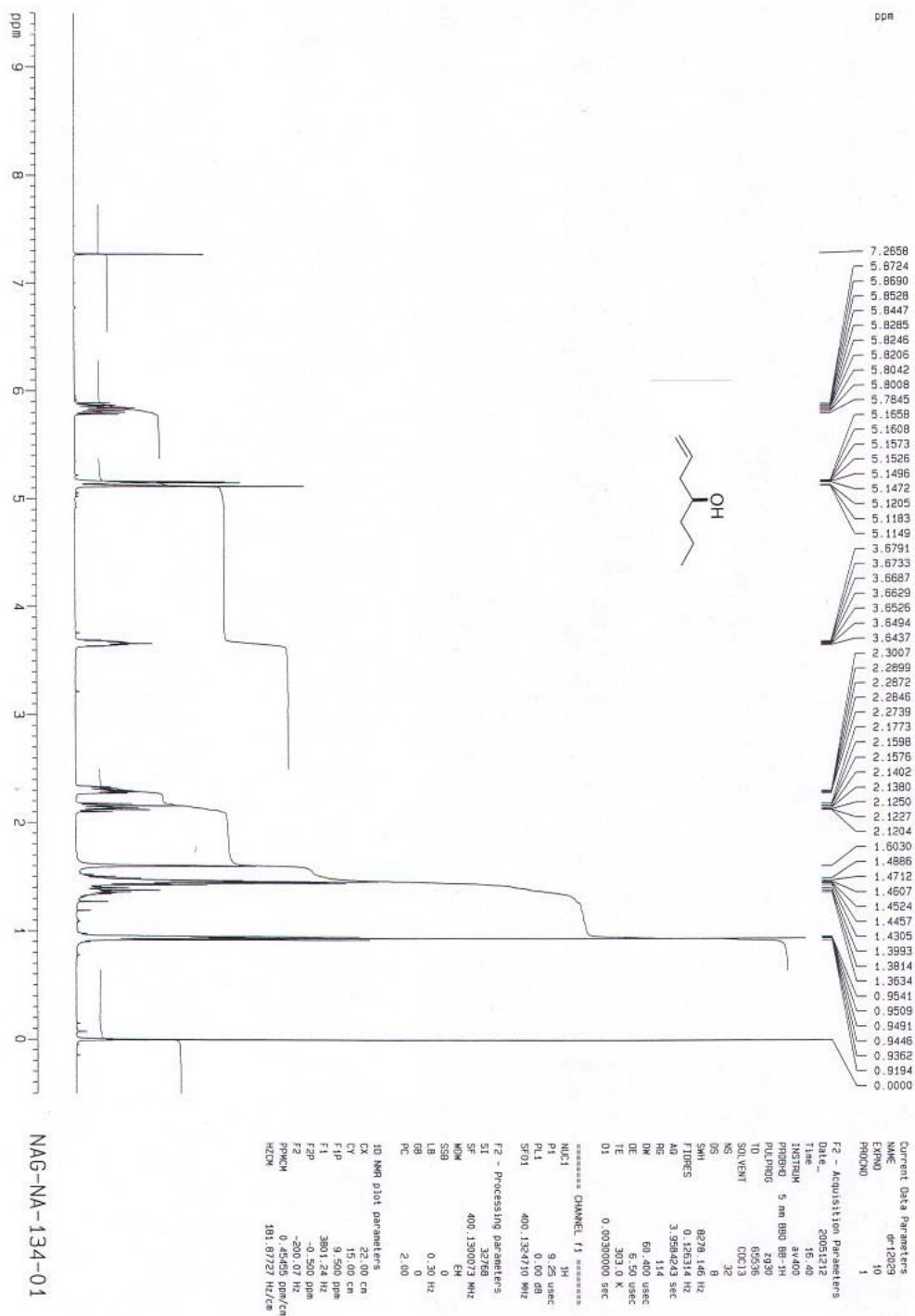
¹H NMR spectra of Ipomoassin E in C₆D₆: (a) natural product reported in literature (500 MHz, Kingston et al. *J. Nat. Prod.* **2005**, 68, 487); (b) sample prepared by us (400 MHz).



¹H NMR spectra of Compound 1 at different concentrations showed remarkable change of chemical shift. (a) 14 mg in 0.6 mL of nondistilled C₆D₆; (b) 4 mg in 0.6 mL of nondistilled C₆D₆; (c) 6 mg in 0.6 mL of distilled C₆D₆; (d) 3 mg in 0.6 mL of distilled C₆D₆.



Spectra of New Compounds





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 PROCNO 1

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 FIDRES 0.505258 Hz
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 RG 16384
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 TE 303.0 K
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 d11 0.03000000 sec

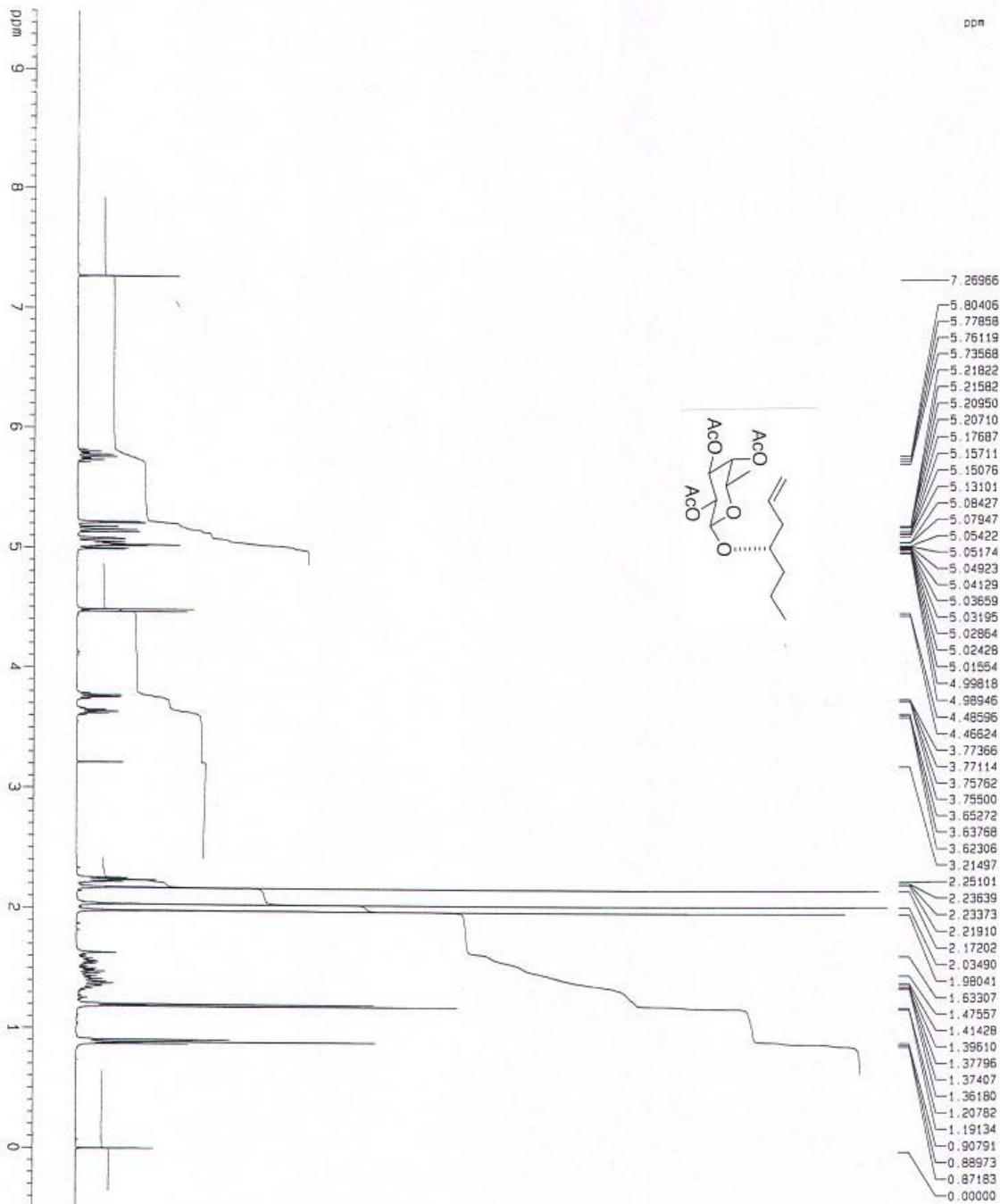
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 PL12 19.75 dB
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F2 - Processing parameters
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 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
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 F2 -3018.38 Hz
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 HZ/CN 1280.55512 HZ/cm

NAG-NA-134-01



Current Data Parameters
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 PROCNO 1

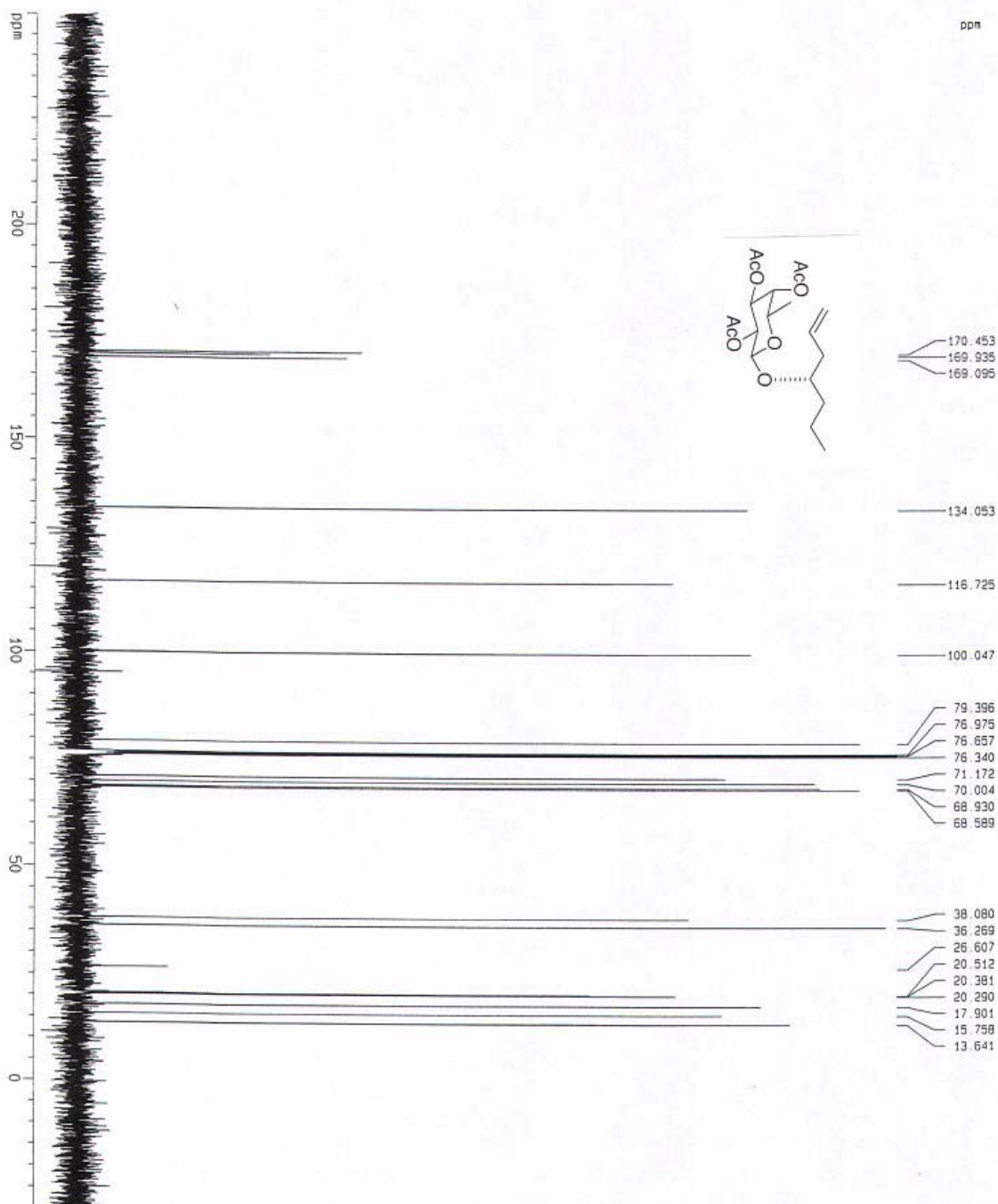
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 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 114
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 DE 303.0 K
 TE 61.30 usec
 D1 0.00300000 sec

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F2 - Processing parameters
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 KW EM
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 LB 0.30 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
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NAG-NA-106-02



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- 169.935
- 169.095
- 134.053
- 116.725
- 100.047
- 79.396
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- 76.340
- 71.172
- 70.004
- 68.930
- 68.589
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- 36.269
- 26.607
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- 20.290
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- 15.758
- 13.641

Current Data Parameters
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 EXPNO 11
 PROCNO 1

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 DS 2
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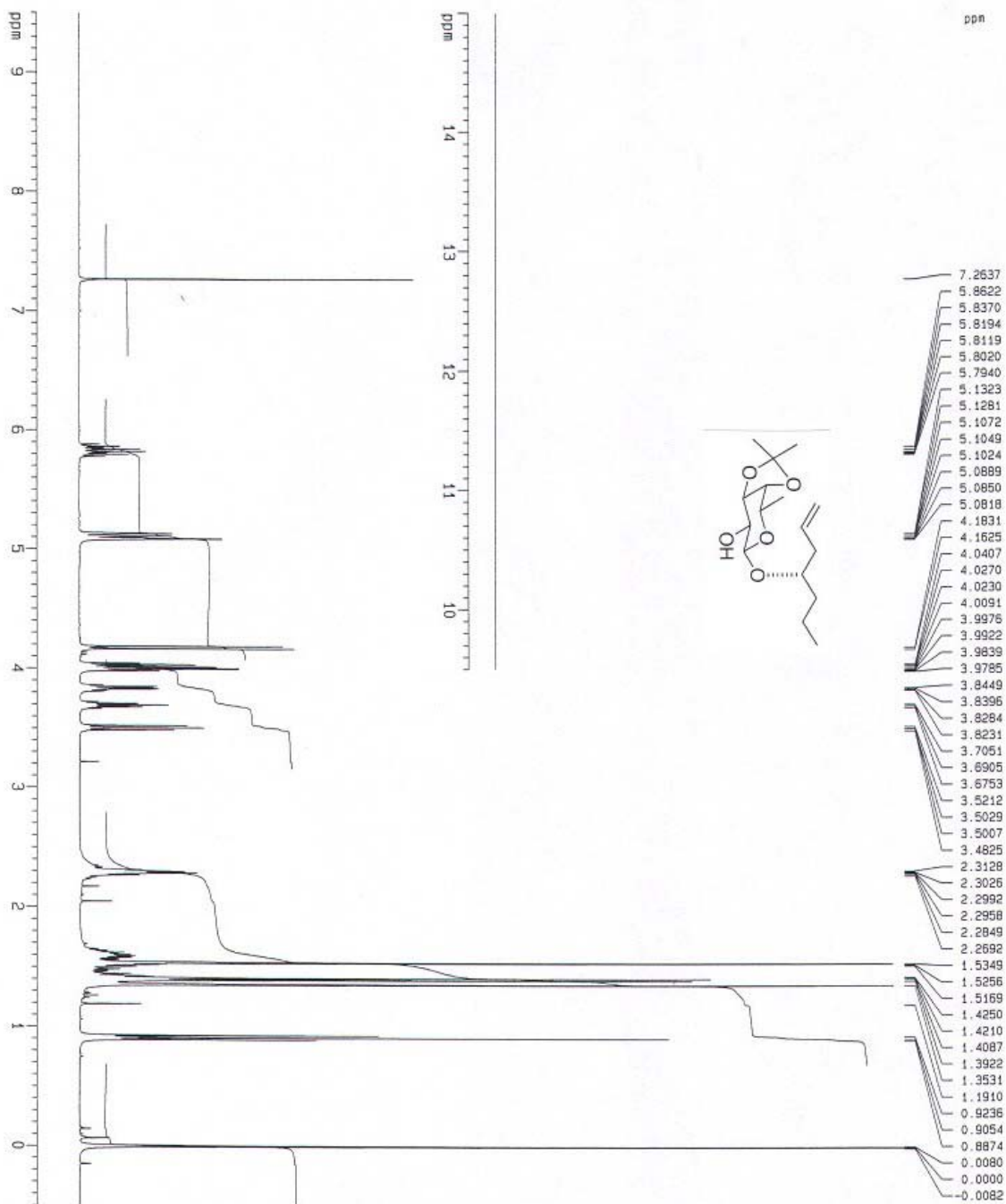
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 PL12 19.75 dB
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F2 - Processing parameters
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 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
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 F2 -3018.38 Hz
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 HZCK 1280.52651 Hz/cm

NAG-NA-106-02



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- 5.8370
- 5.8194
- 5.8119
- 5.8020
- 5.7940
- 5.1323
- 5.1281
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- 5.1024
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- 5.0850
- 5.0818
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- 4.0270
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- 3.9976
- 3.9922
- 3.9839
- 3.9785
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- 3.8396
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- 3.5212
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- 2.2849
- 2.2692
- 1.5349
- 1.5256
- 1.5169
- 1.4250
- 1.4210
- 1.4087
- 1.3922
- 1.3531
- 1.1910
- 0.9236
- 0.9054
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ppm

ppm

Current Data Parameters
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 PROCNO 1

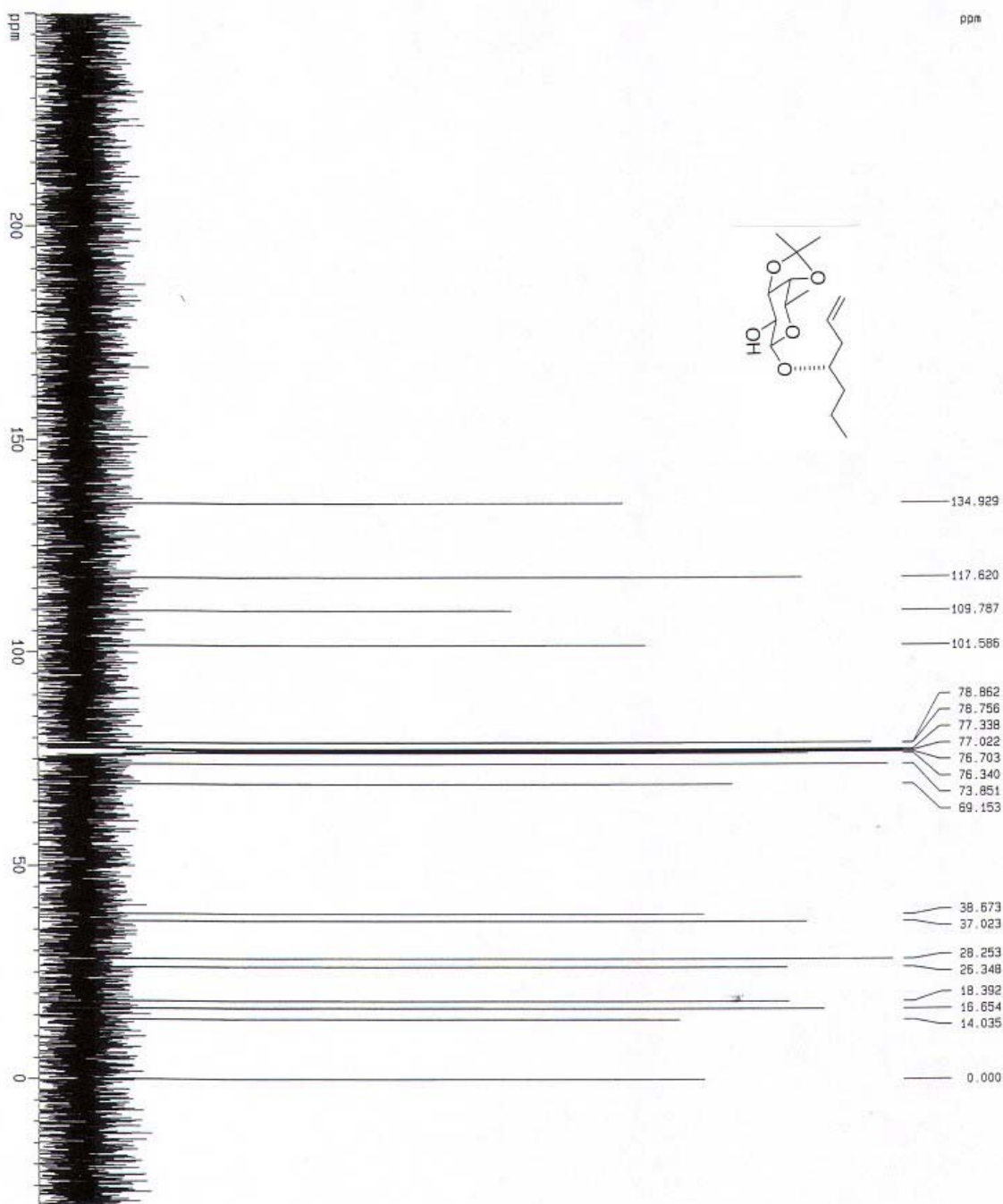
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 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 203.2
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 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
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 LB 0.30 Hz
 GB 0
 PC 2.00

50 MHz plot parameters
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 CY 15.00 cm
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 F2P -0.500 ppm
 F2 -500.07 Hz
 PRACH 0.45455 ppm/cm
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NAG-NA-124-01



Current Data Parameters
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 PROCKO 1

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 NS 1500
 DS 2
 SFO1 33112.982 Hz
 FIDRES 0.562928 Hz
 AQ 0.988445 sec
 RG 16384
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 D11 0.03000000 sec

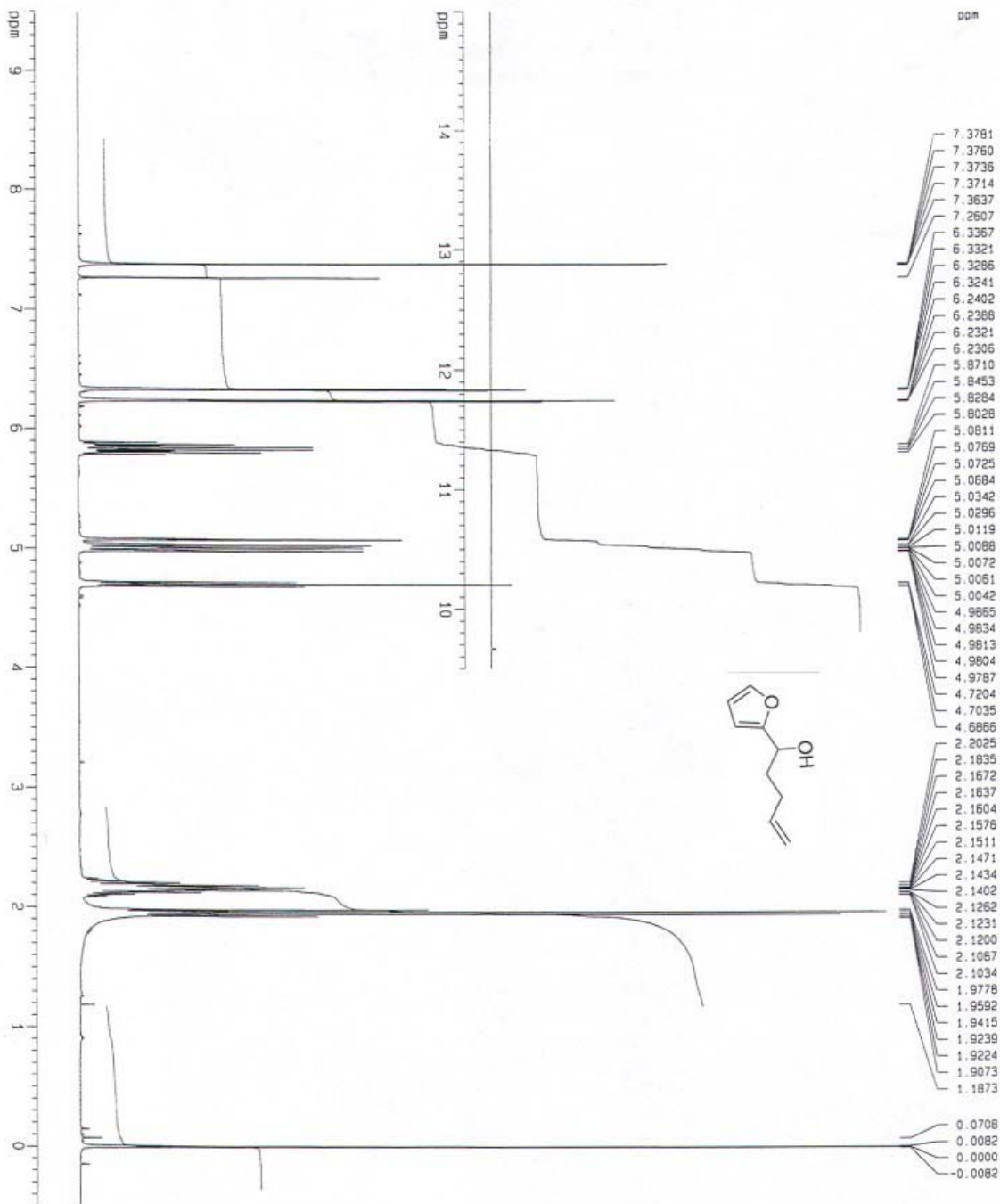
***** CHANNEL f1 *****
 NUCl 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242789 MHz

***** CHANNEL f2 *****
 CPDPRG2 M011216
 NUCl 1H
 PCNO2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127671 MHz
 MDK EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.19 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPHCK 12.72727 ppm/cm
 HZCM 1280.52612 Hz/cm

NAG-NA-124-01



Current Data Parameters

| | |
|--------|---------|
| NAME | 0701010 |
| EXPNO | 10 |
| PROCNO | 1 |

F2 - Acquisition Parameters

| | |
|---------|----------------|
| Date_ | 20051201 |
| Time | 11.51 |
| INSTRUM | av400 |
| PROBHD | 5 mm BBO 90-1H |
| PULPROG | ZG30 |
| TD | 65536 |
| SQ1 | VENT |
| SQ2 | VENT |
| SQ3 | VENT |
| MS | 32 |
| DC | B |
| SH1 | B278.146 Hz |
| SH2 | 0.126316 Hz |
| FIDRES | 3.5984243 sec |
| AQ | 114 |
| RG | 60 |
| DM | 60.400 usec |
| DE | 6.50 usec |
| TE | 303.0 K |
| D1 | 0.00300000 sec |

***** CHANNEL f1 *****

| | |
|-------|-----------------|
| MUCL1 | 1H |
| PI | 9.25 usec |
| PL1 | 0.00 dB |
| SFO1 | 400.1324710 MHz |

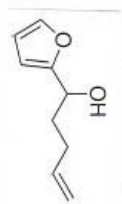
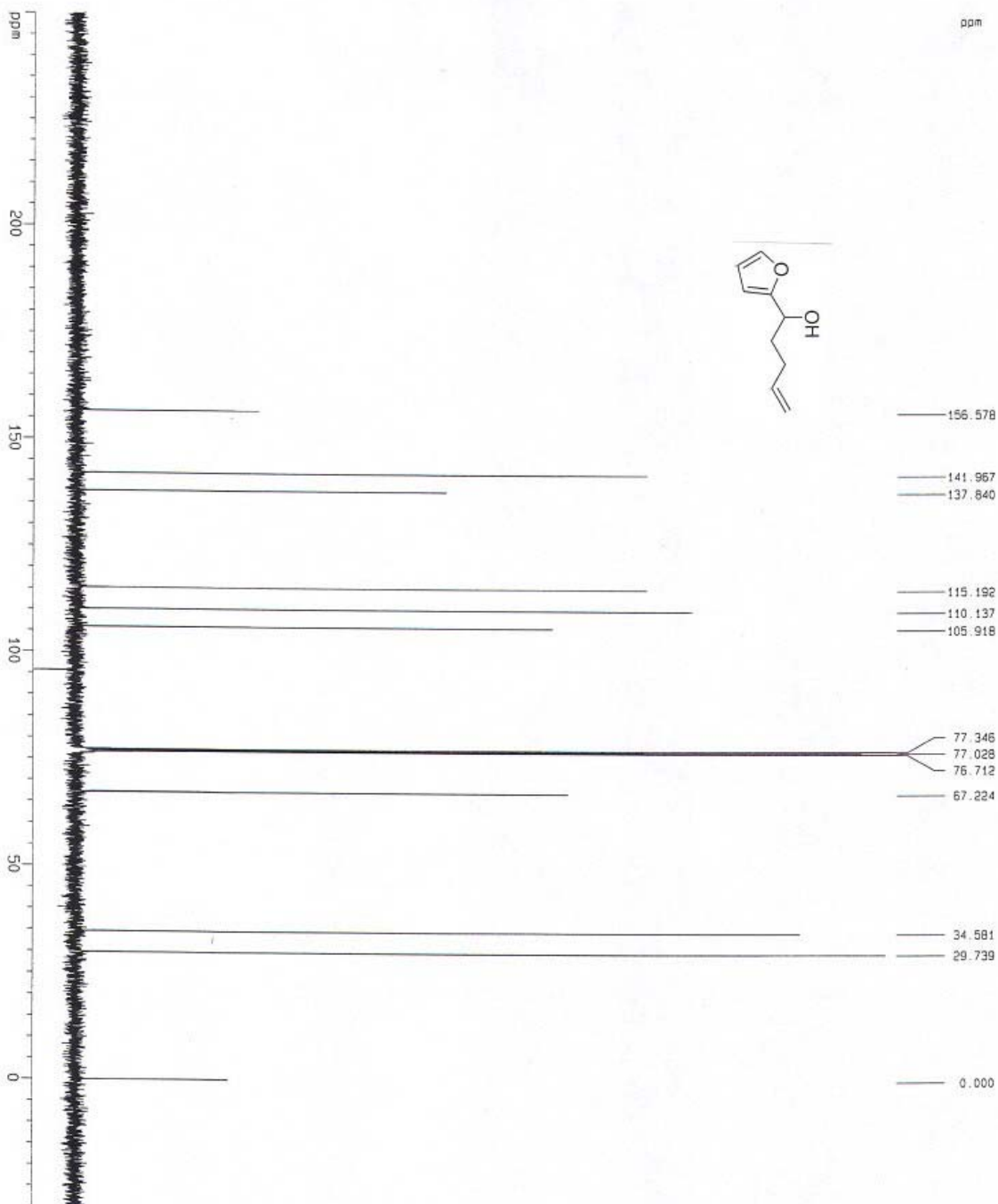
F2 - Processing parameters

| | |
|-----|-----------------|
| SI | 32768 |
| SF | 400.1300093 MHz |
| MDW | EM |
| SSD | 0 |
| LB | 0.30 Hz |
| GB | 0 |
| PC | 2.00 |

10 NMR plot parameters

| | |
|------|-----------------|
| CX | 22.00 cm |
| CV | 15.00 cm |
| F1P | 9.500 DDM |
| F1 | 3801.24 Hz |
| F2P | -0.500 DDM |
| F2 | -200.07 Hz |
| PVCM | 0.454455 ppm/cm |
| HZCM | 181.87727 Hz/cm |

NAG-NA-120-03



Current Data Parameters
 NAME dr01010
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051201
 Time 12:20
 INSTRUM av400
 PROBHD 5 mm BBO BB-4H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2

SH 33112.582 Hz
 FIDRES 0.506236 Hz
 AQ 0.9696436 sec
 RG 16390.4
 DW 15.100 usec
 DE 0.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

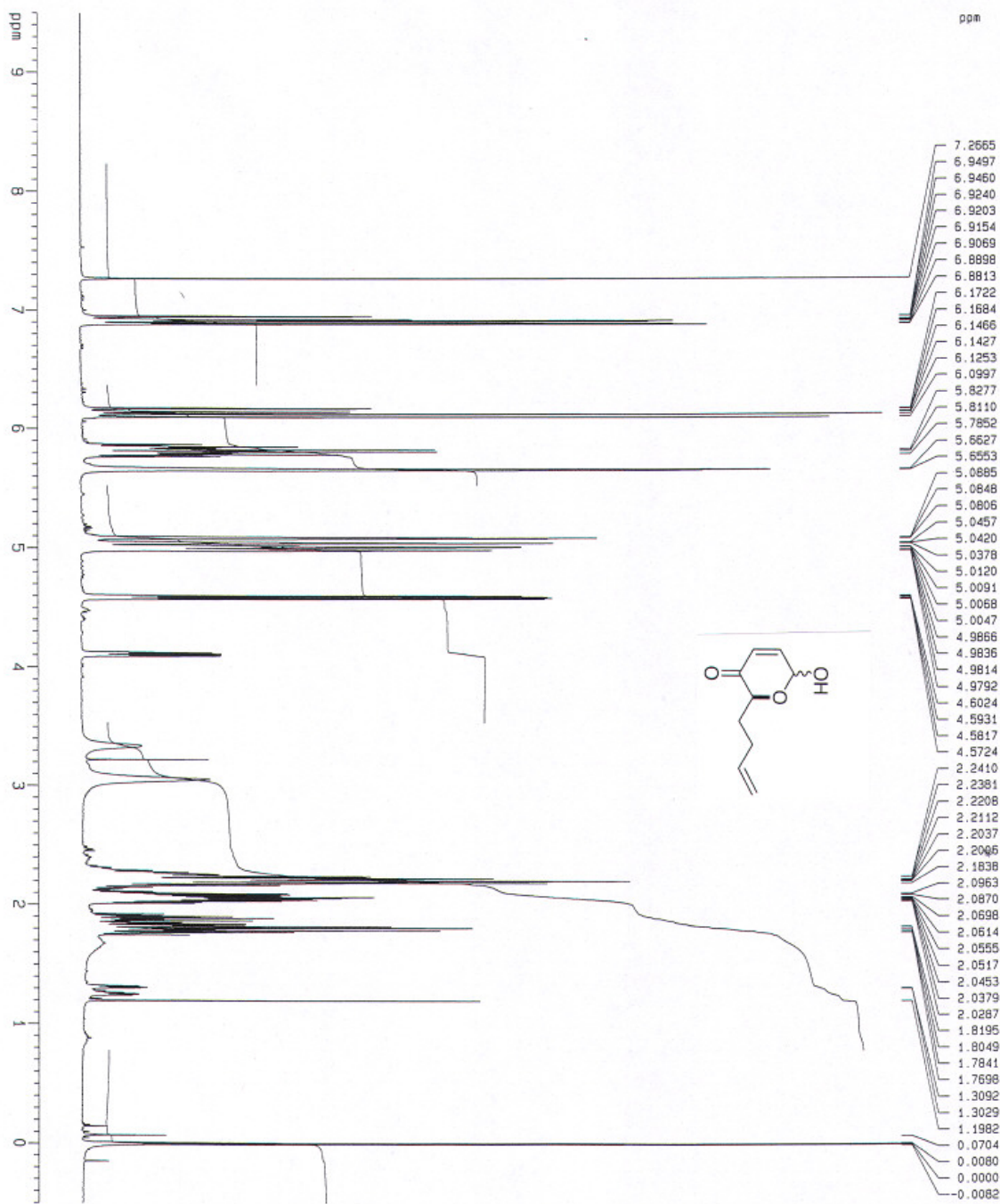
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.04 usec
 PL1 5.00 dB
 SFO1 100.6242189 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127672 MHz
 NQW 0
 SSB 0
 LB 1.00 Hz
 OB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 250.000 ppm
 F1 25153.19 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PRCM 12.2227 ppm/cm
 HZCM 1280.52612 Hz/cm

NAG-NA-120-03



NAG-NA-157-01

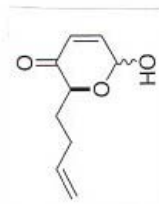
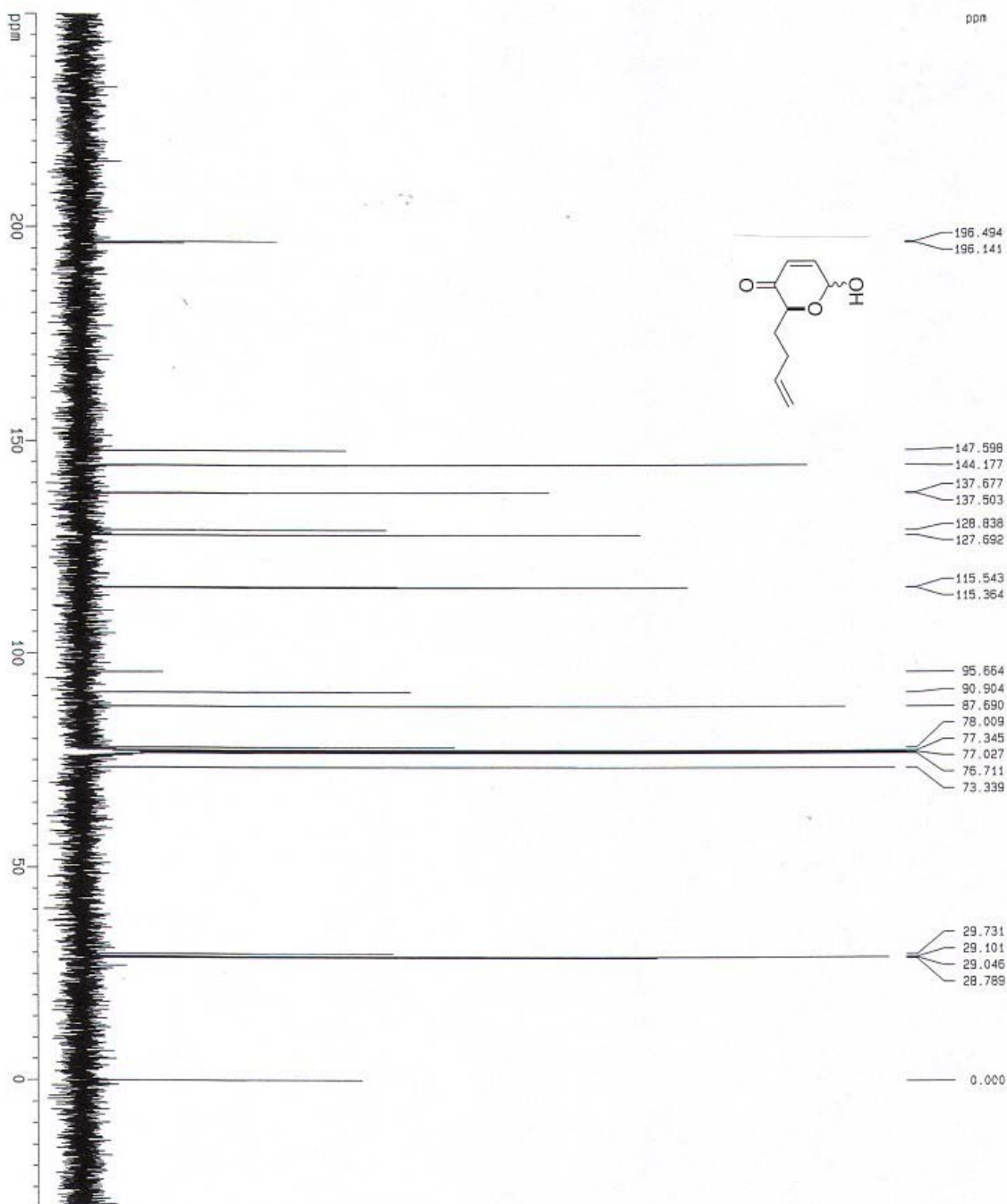
Current Data Parameters
 NAME J120318
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060123
 Time 14.23
 INSTRUM av400
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9564243 sec
 RG 181
 DM 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300070 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 9.500 ppm
 F1 3801.24 Hz
 F2P -200.07 Hz
 F2 0.46655 ppm/cm
 FPMCM 181.81727 Hz/cm
 HZCM



Current Data Parameters
 NAME J1-23018
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060123
 Time 14.54
 INSTRUM av400
 PROBRD 5 mm BBO BB-H1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SM 33112.582 Hz
 FIDRES 0.500528 Hz
 AQ 0.5895436 sec
 RG 16384
 OW 15.100 usec
 OE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

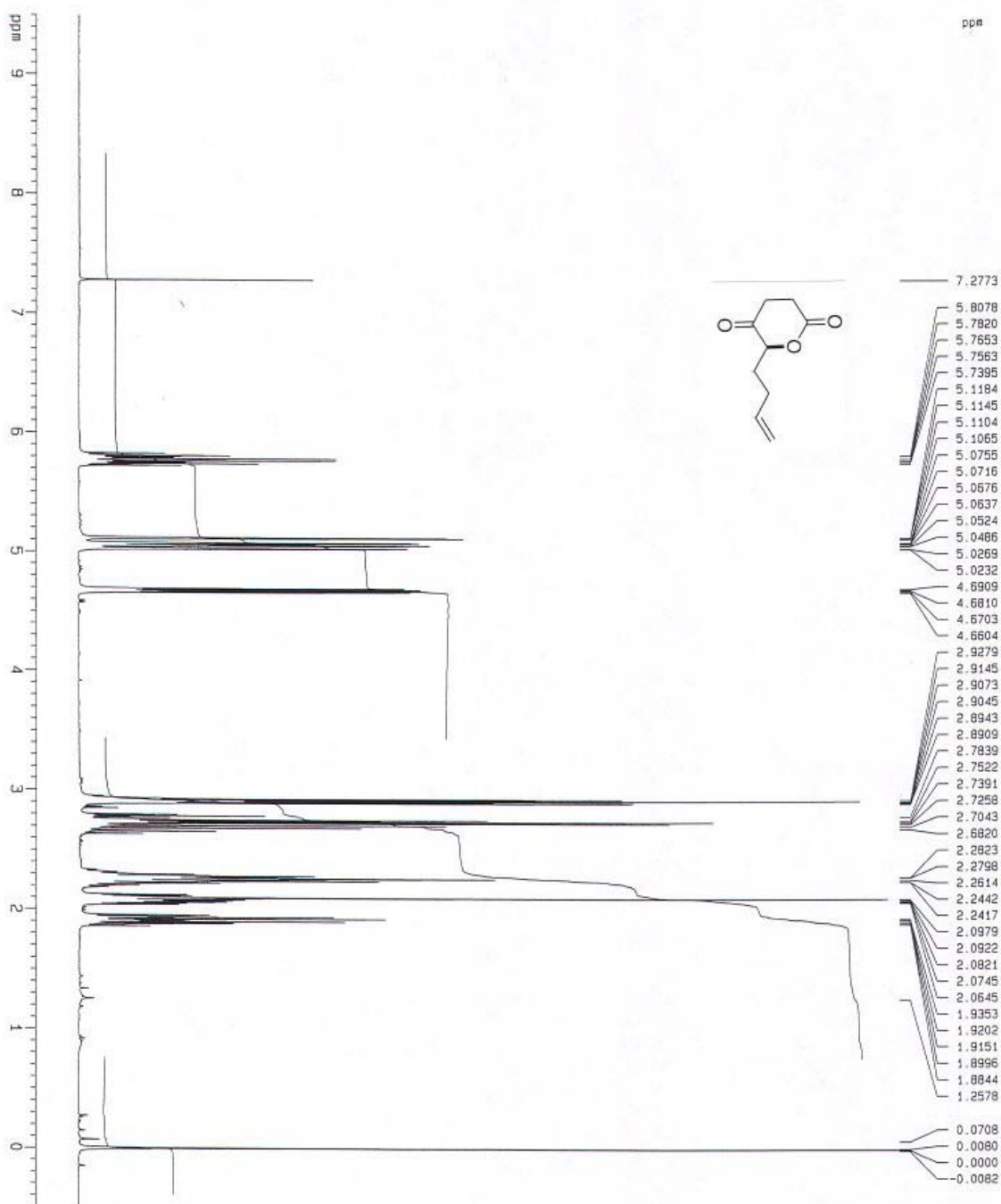
----- CHANNEL f1 -----
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SF01 100.6242789 MHz

----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SF02 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127672 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 250.000 ppm
 F1 26153.19 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPHCN 12.72727 gm/cm
 HZCM 1280.56612 Hz/cm

NAG-NA-157-01



NAG-NA-160-01

```

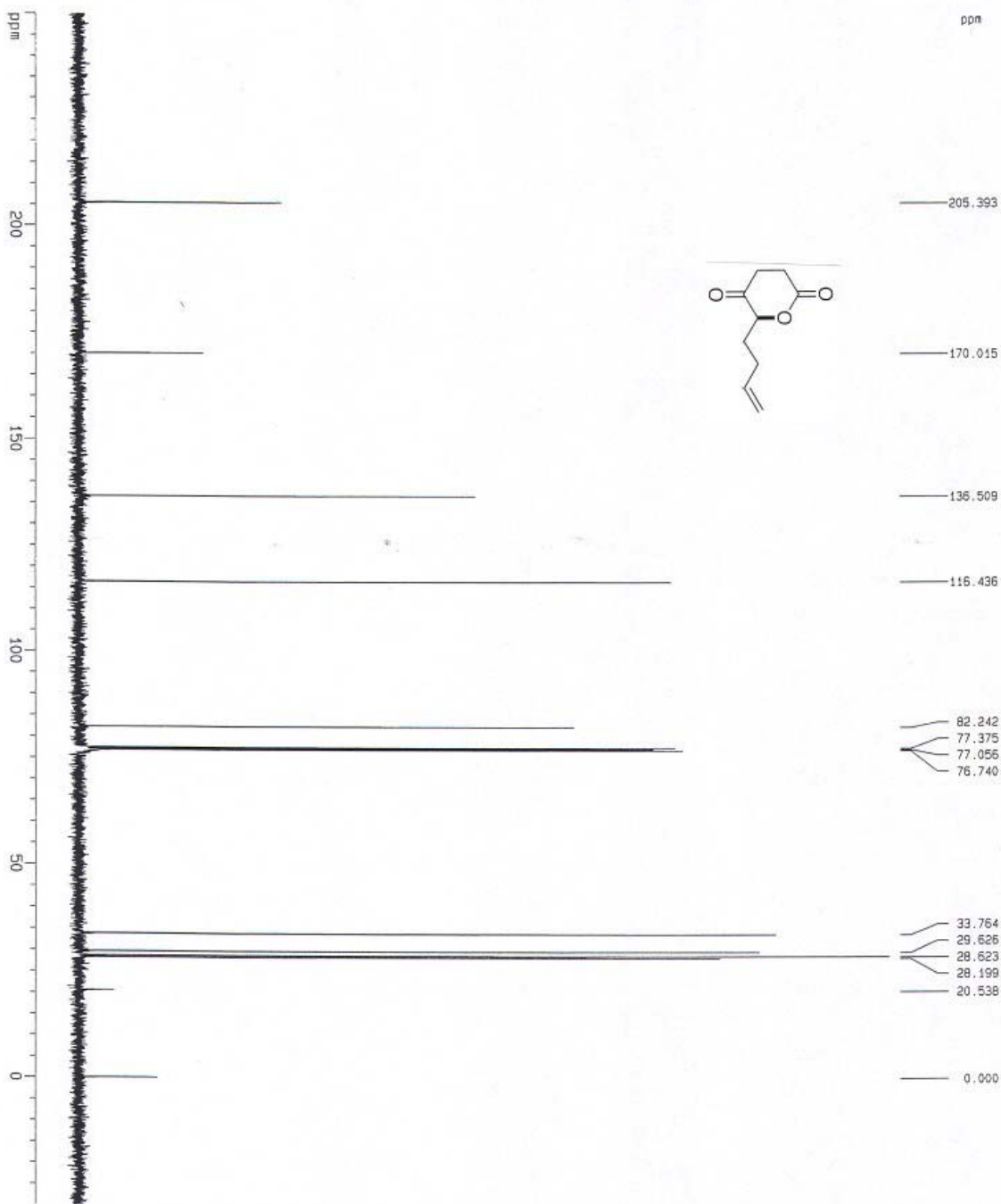
Current Data Parameters
NAME          JY25046
EXPNO         10
PROCNO        1

F2 - Acquisition Parameters
Date_         20060126
Time          14.25
INSTRUM      ay400
PROBHD       5 mm BBO BB-4H
PULPROG      zg30
TD            65536
SOLVENT      CDCl3
NS            32
DS            2
SML          B
FIDRES       0.12614 Hz
AQ           3.998443 sec
RG           114
DM           60.400 usec
DE           6.50 usec
TE           303.0 K
D1           0.00300000 sec

***** CHANNEL f1 *****
NUC1          1H
P1            9.25 usec
PL1           0.00 dB
SFO1         400.1324710 MHz

F2 - Processing parameters
SI            32768
SF           400.1300028 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            2.00

10 MHz pilot parameters
CX           22.00 cm
CY           15.00 cm
F1P          9.500 ppm
F1           3801.24 Hz
F2P          -0.500 ppm
F2           -200.07 Hz
FREQCN      0.46455 ppm/cm
HZCM        181.87727 Hz/cm
    
```

Current Data Parameters
 NAME JF25046
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060126
 Time 14.54
 INSTRUM av400
 PROBRD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 66536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SWH 33112.562 Hz
 FIDRES 0.505259 Hz
 AQ 0.9896436 sec
 RG 16384
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

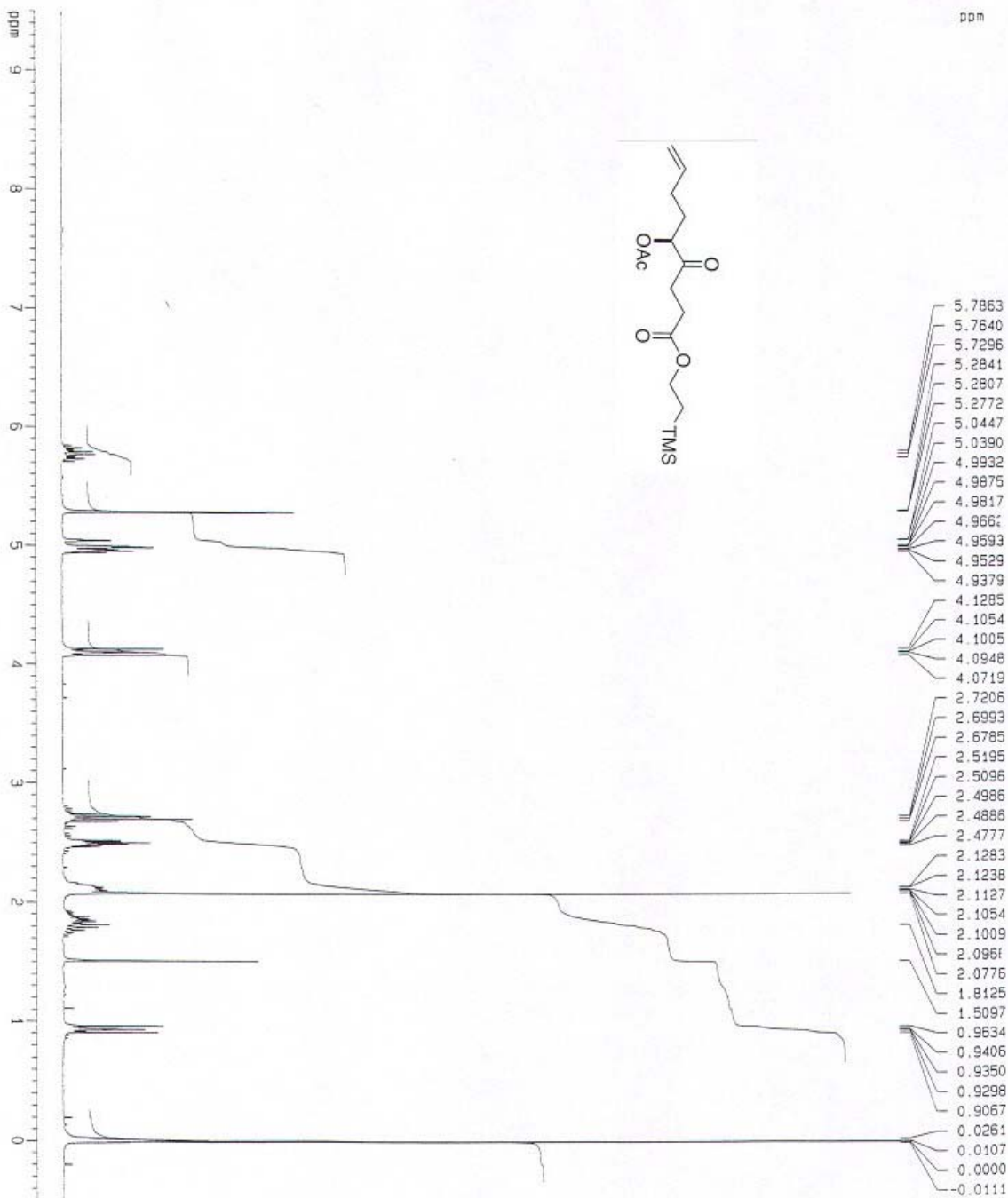
***** CHANNEL f1 *****
 NUC1 13C 13C
 P1 10.34 usec
 PL1 5.00 dB
 SFO1 100.6242199 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.75 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127663 MHz
 KHM 64
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 250.000 ppm
 F1 25153.19 Hz
 F2 -30.000 ppm
 F2 -3016.38 Hz
 SPNCK 12.7222 ppm/cm
 HZCK 1280.52612 Hz/cm

NAG-NA-160-01



NAG-NA-004-02

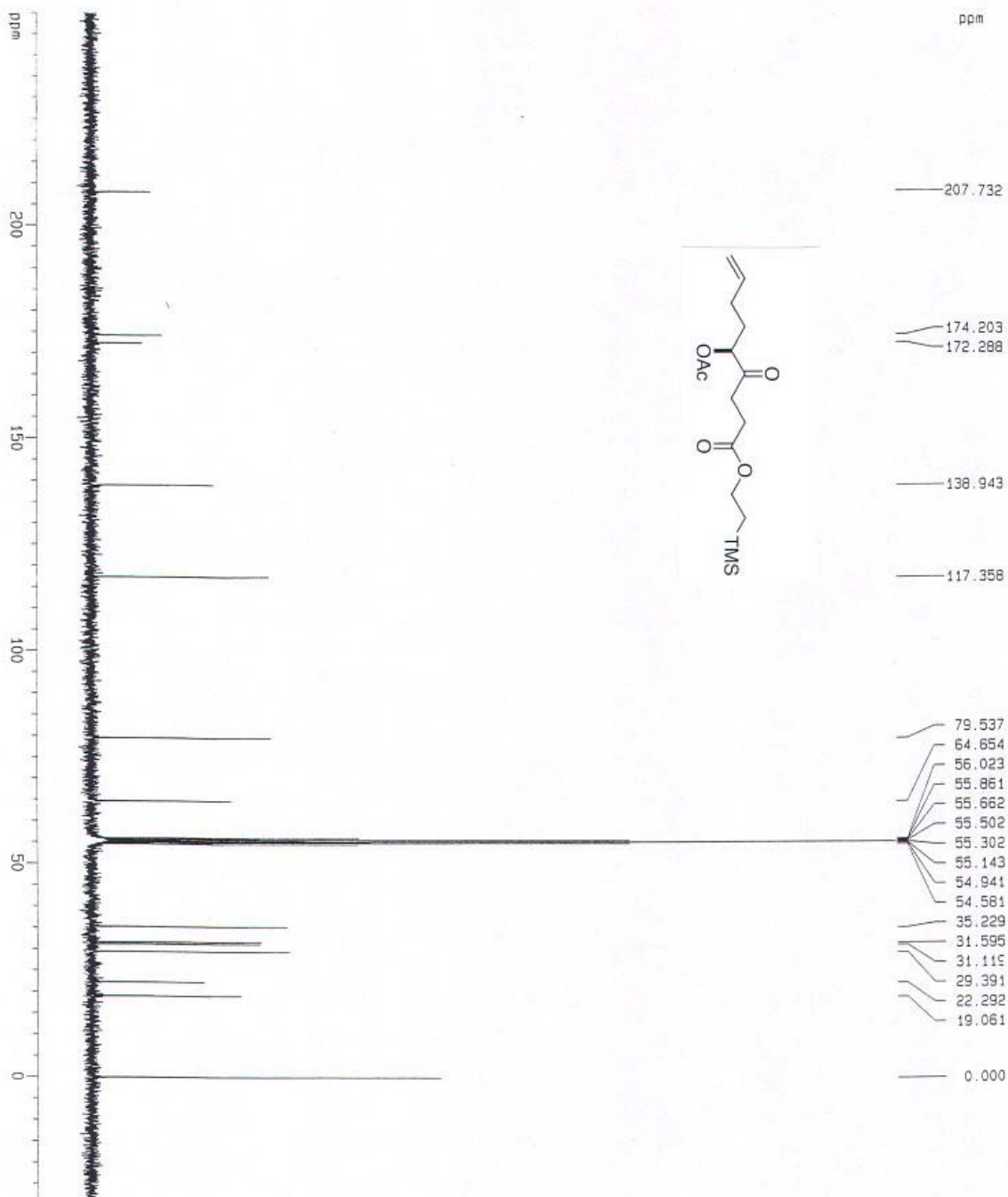
Current Data Parameters
 NAME 1J02019
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060502
 Time 14.15
 INSTRUM dpX300
 PROBDW 5 mm QNP 1H/
 PULPROG zg30
 TO 32768
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 6172.839 Hz
 FIDRES 0.188380 Hz
 AQ 2.6542580 sec
 RG 456.1
 DW 81.000 usec
 DE 4.50 usec
 TE 300.0 K
 D1 2.00000000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.56 usec
 PL1 -6.00 dB
 SF01 300.1318534 MHz

F2 - Processing parameters
 SI 16384
 SF 300.1300224 MHz
 MVM EM
 SSB 0
 LB 0.30 Hz
 BB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 ppm
 F1 2851.24 Hz
 F2P -0.500 ppm
 F2 -150.07 Hz
 PPMCM 0.45455 ppm/cm
 HZCM 136.42274 Hz/cm



Current Data Parameters
 NAME: J002019
 EXPNO: 11
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20060902
 Time: 14.47

INSTRUM: spect
 PULPROG: zgpg30
 TD: 65536
 SFO: 300.1312005
 DS: 2
 SWH: 21231.422 Hz
 FIDRES: 0.323966 Hz
 AQ: 1.5434228 sec
 RG: 4597.6
 DW: 23.590 usec
 DE: 4.50 usec
 TE: 300.0 K
 D1: 0.03000000 sec
 D11: 0.03000000 sec

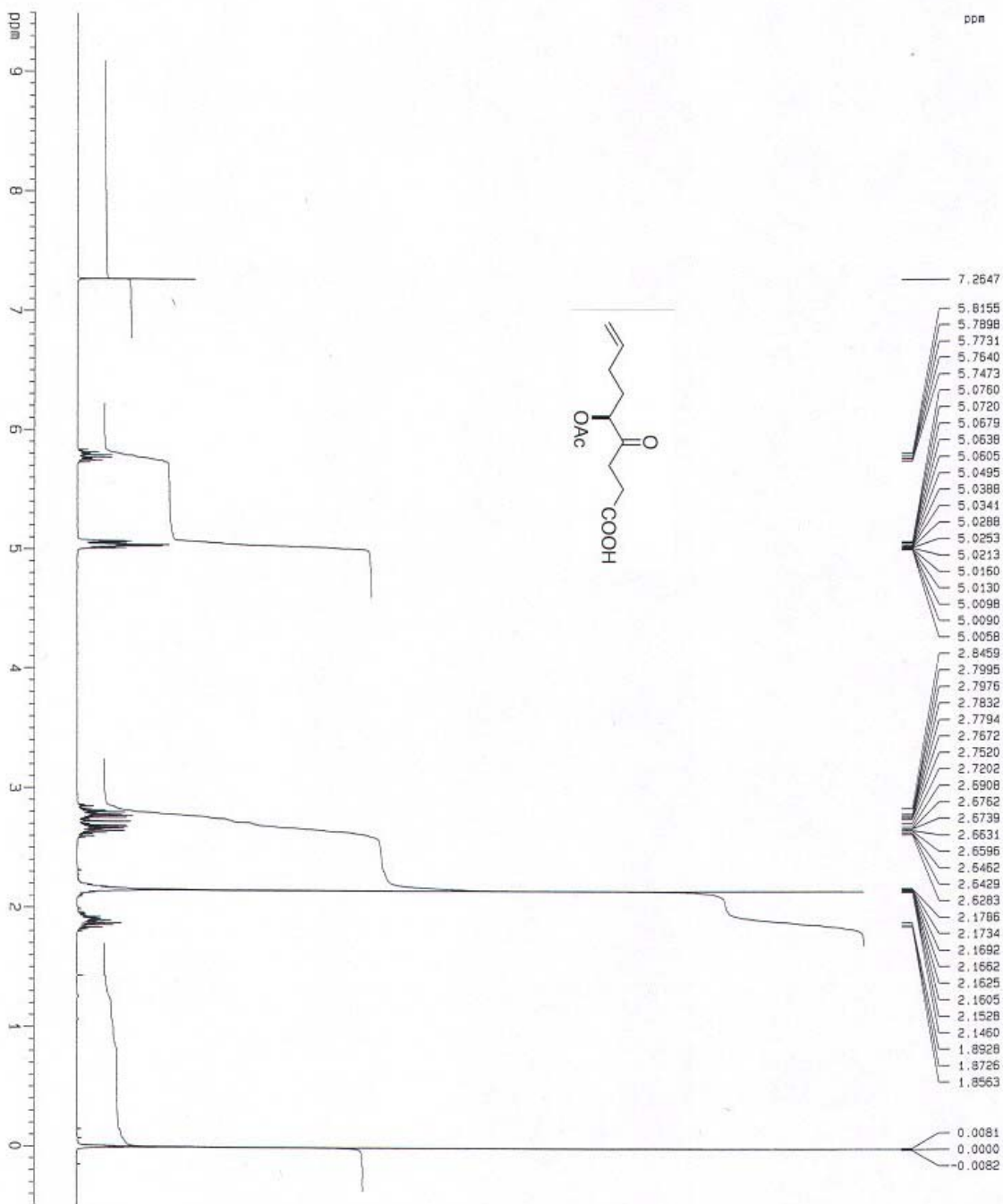
----- CHANNEL f1 -----
 NUC1: 13C
 P1: 6.91 usec
 PL1: -6.00 dB
 SFO1: 75.47560670 MHz

----- CHANNEL f2 -----
 CPDPRG2: waltz16
 NUC2: 1H
 PCPD2: 90.00 usec
 PL2: -6.00 dB
 PL12: 14.49 dB
 SFO2: 300.1312005 MHz

F2 - Processing parameters
 SI: 32768
 SF: 75.4675044 MHz
 WDW: EM
 SSB: 0
 LB: 1.00 Hz
 GB: 0
 PC: 1.40

10 NMR plot parameters
 CX: 22.00 cm
 CY: 15.00 cm
 F1P: 250.000 ppm
 F1: 18886.90 Hz
 F2P: -30.000 ppm
 F2: -2264.03 Hz
 PRNCK: 12.7272 ppm/cm
 HZCM: 960.49883 Hz/cm

MAG-NA-004-02



Current Data Parameters
 NAME J14004
 EXPNO 10
 PROCNO 1

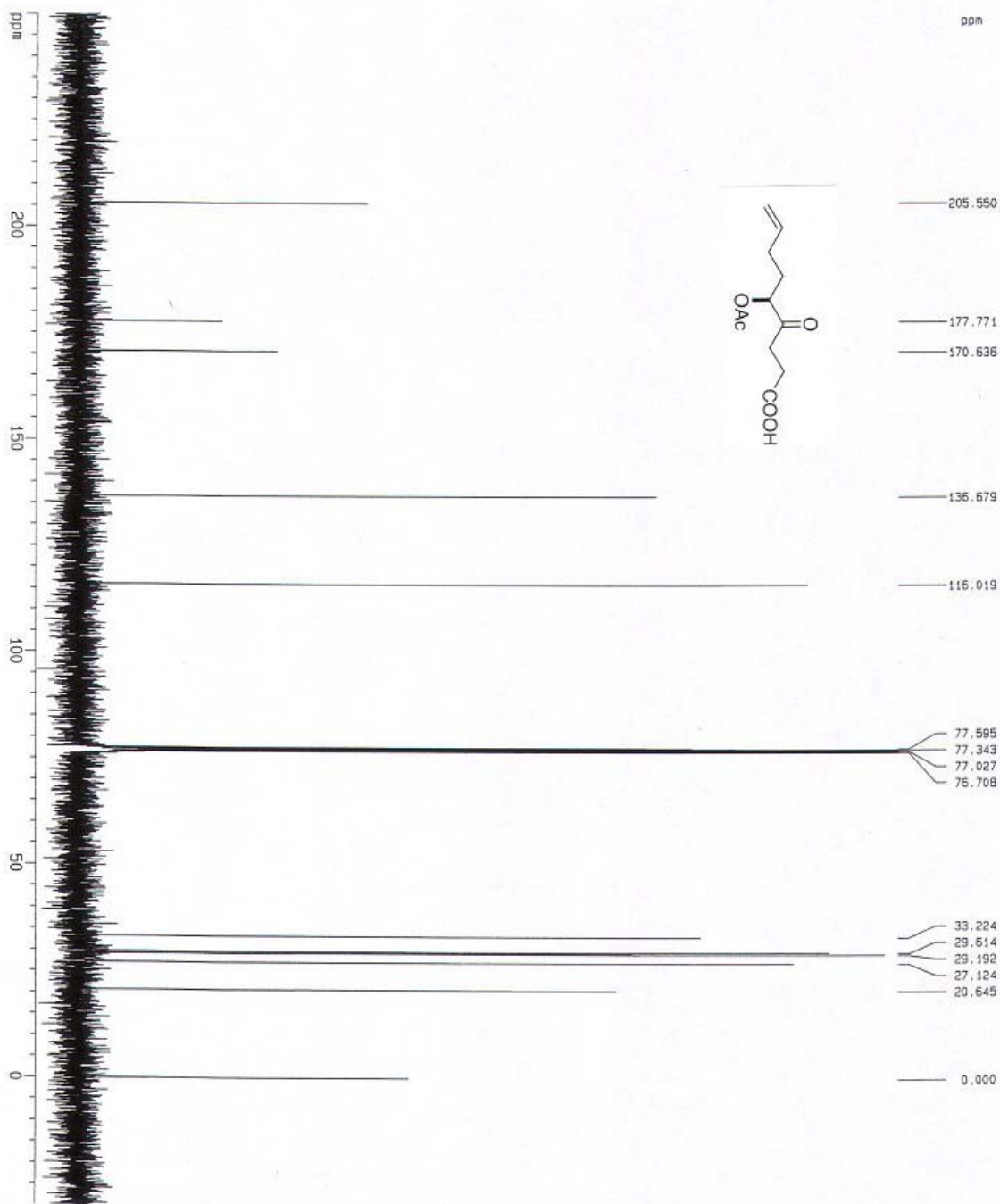
F2 - Acquisition Parameters
 Date_ 20060714
 Time 9:57
 INSTRUM av400
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SWH 8278.145 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 181
 DW 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300078 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 9.500 ppm
 F1 3801.24 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPHCEN 0.45455 ppm/cm
 HZCM 181.87227 Hz/cm

NA6-NB-035-01



Current Data Parameters
 NAME JY14004
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060714
 Time 10.26
 INSTRUM BR-400
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SWH 33112.582 Hz
 FIDRES 0.5062598 Hz
 AQ 0.9696436 sec
 RG 18390.4
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

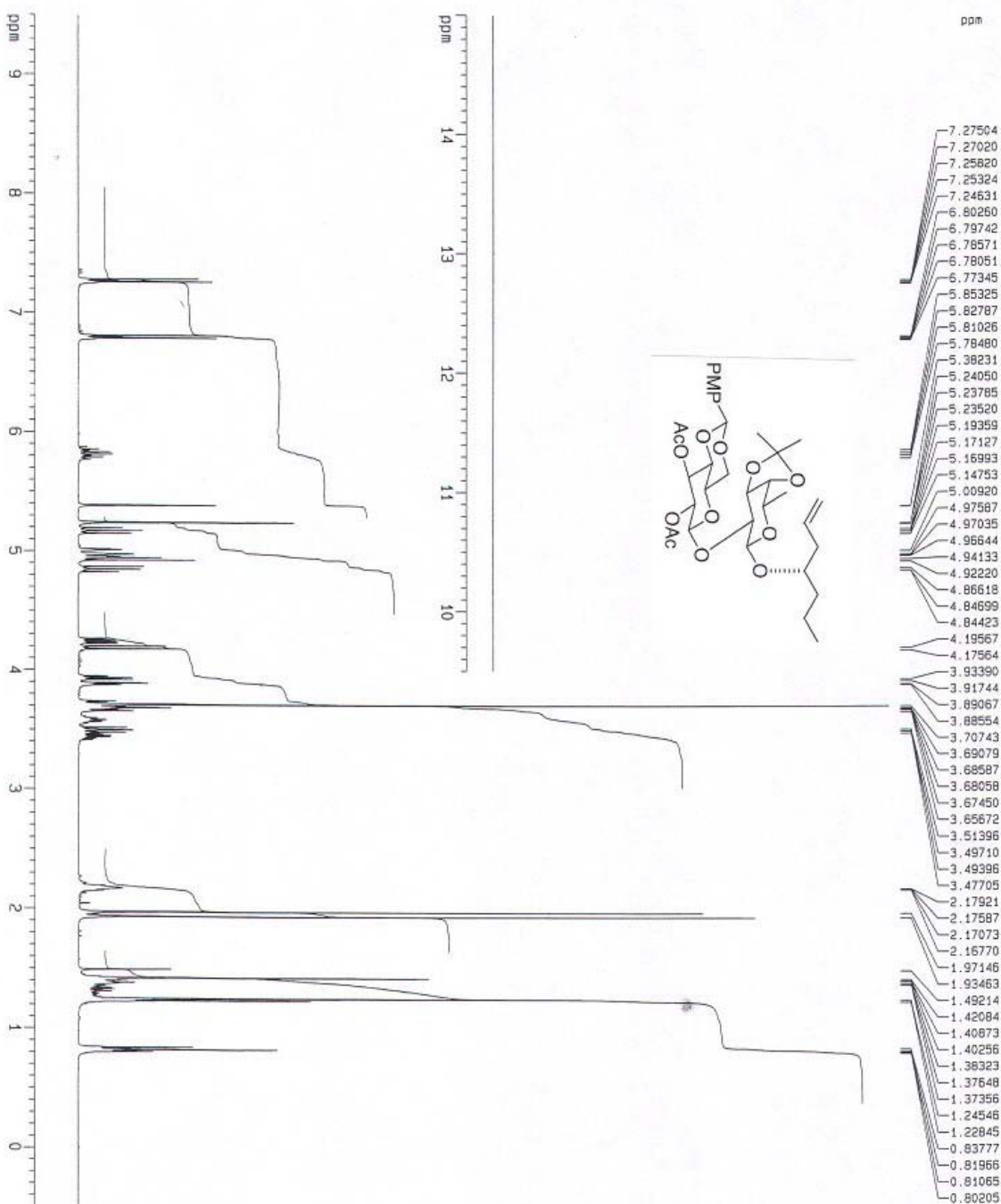
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.34 usec
 PL1 5.00 dB
 SF01 100.6242789 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 P2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SF02 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127956 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 dBm
 F1 20153.19 Hz
 F2P -30.000 dBm
 F2 -3018.38 Hz
 ppm0M 12.7272 ppm/cm
 Hz0M 1280.50612 Hz/cm

NA6-NB-035-01



Current Data Parameters
 NAME J123039
 EXPNO 10
 PROCNO 1

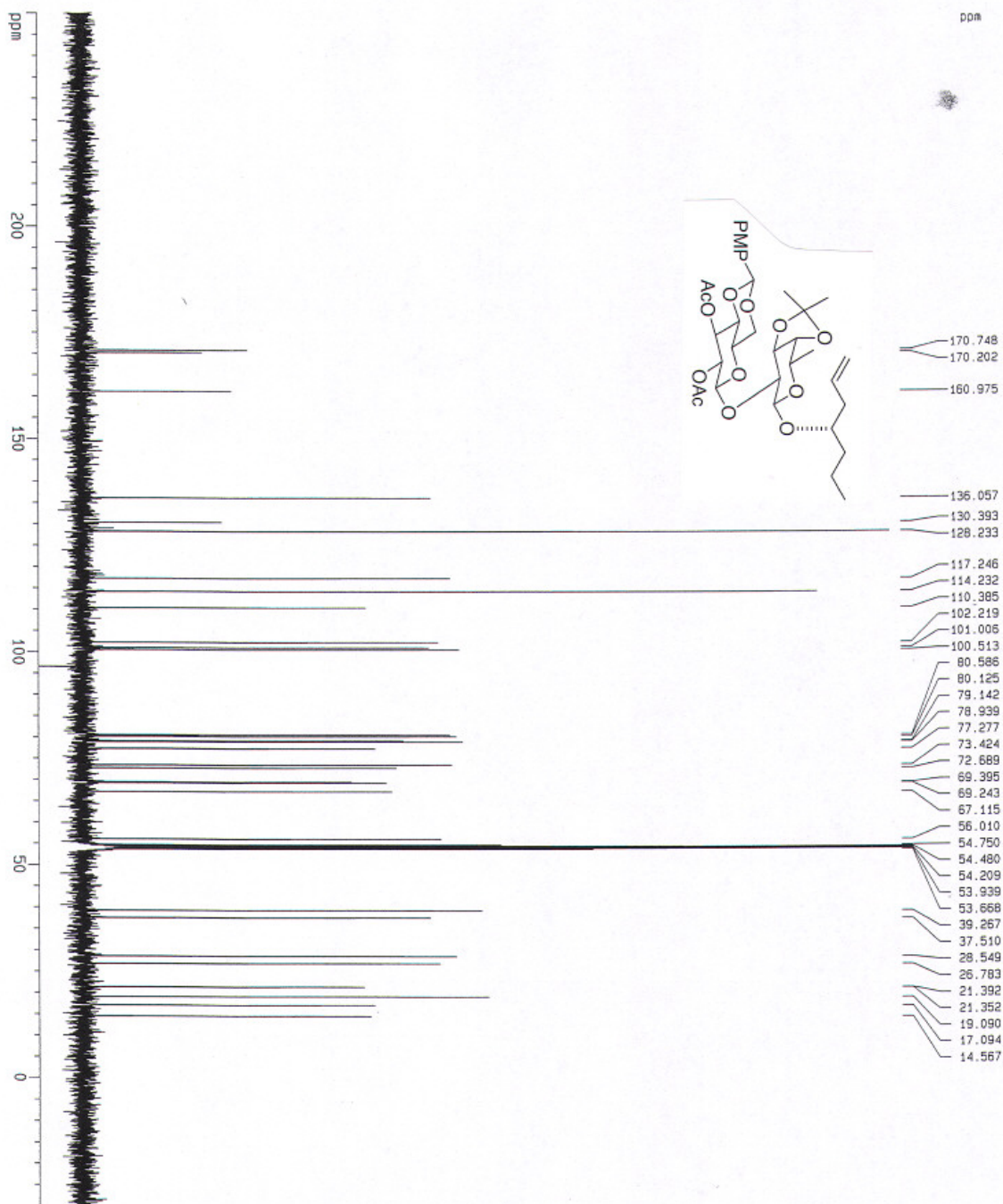
F2 - Acquisition Parameters
 Date_ 20050126
 Time 12.15
 INSTRUM av400
 PROBRD 5 mm BBO BB-4H
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SMH B278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9594243 sec
 RG 114
 DM 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300482 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 DPM
 F1 3601.24 Hz
 F2P -200.07 Hz
 F2 0.45455 DPM/cm
 PPMICK 181.87729 Hz/cm
 HCM

NA6-NA-146-01



Current Data Parameters
 NAME J125039
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060126
 Time 12.43
 INSTRUM av400
 PROBRD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SWH 33112.582 Hz
 FIDRES 0.300258 Hz
 AQ 0.5995439 sec
 RG 16384
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

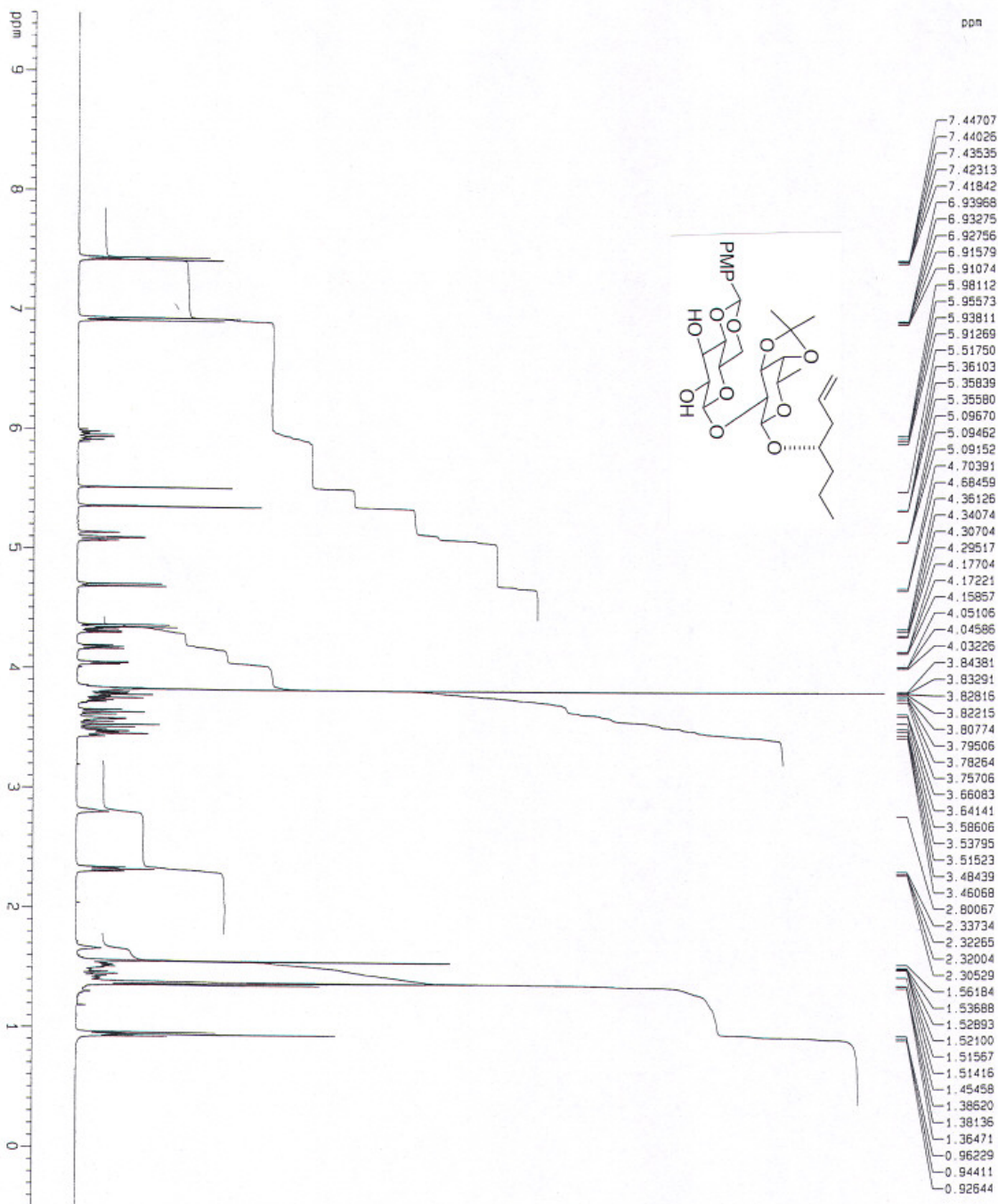
----- CHANNEL f1 -----
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242789 MHz

----- CHANNEL f2 -----
 CPROBR2 waltz16
 NUC2 1H
 POC2 90.00 usec
 PLL2 0.00 dB
 PL12 19.75 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32759
 SF 100.6126889 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 250.000 ppm
 F1 25153.17 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPHCK 12.72227 ppm/cm
 HZCK 1280.52615 Hz/cm

NAG-NA-146-01



- 7.44707
- 7.44026
- 7.43535
- 7.42313
- 7.41842
- 6.93968
- 6.93275
- 6.92756
- 6.91579
- 6.91074
- 5.98112
- 5.95573
- 5.93811
- 5.91269
- 5.51750
- 5.36103
- 5.35839
- 5.35580
- 5.09670
- 5.09462
- 5.09152
- 4.70391
- 4.68459
- 4.36126
- 4.34074
- 4.30704
- 4.29517
- 4.17704
- 4.17221
- 4.15857
- 4.05106
- 4.04586
- 4.03226
- 3.84381
- 3.83291
- 3.82816
- 3.82215
- 3.80774
- 3.79506
- 3.78264
- 3.75706
- 3.66083
- 3.64141
- 3.58606
- 3.53795
- 3.51523
- 3.48439
- 3.46068
- 2.80067
- 2.33734
- 2.32265
- 2.32004
- 2.30529
- 1.56184
- 1.53688
- 1.52693
- 1.52100
- 1.51557
- 1.51416
- 1.45458
- 1.38620
- 1.38136
- 1.36471
- 0.96229
- 0.94411
- 0.92644

Current Data Parameters
 NAME m10101
 EXPNO 20
 PROCNO 1

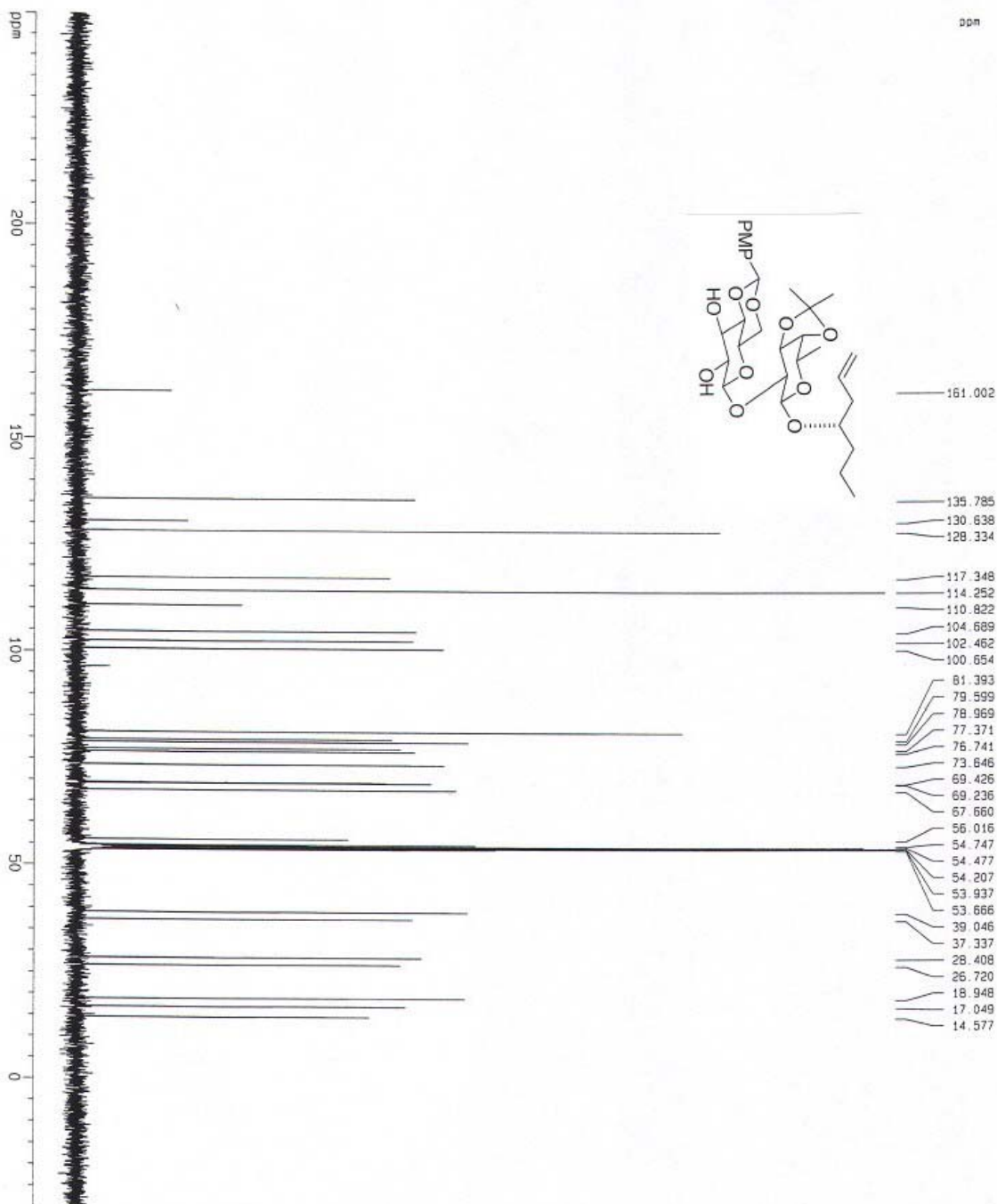
F2 - Acquisition Parameters
 Date_ 20051111
 Time 22.31
 INSTRUM av400
 PROBRD 5 mm BBO BB-HH
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS B
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 114
 DW 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 O1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 KW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 GHz
 F1 3891.24 Hz
 F2P -0.500 MHz
 F2 -200.07 Hz
 PPMCH 0.45455 ppm/cm
 HCM 181.87727 Hz/cm

NAG-NA-110-01



Current Data Parameters
 NAME m10101
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051111
 Time 23.00
 INSTRUM av400
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SWH 33112.582 Hz
 FIDRES 0.562528 Hz
 AQ 0.9896436 sec
 RG 10390.4
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 D11 0.03000000 sec

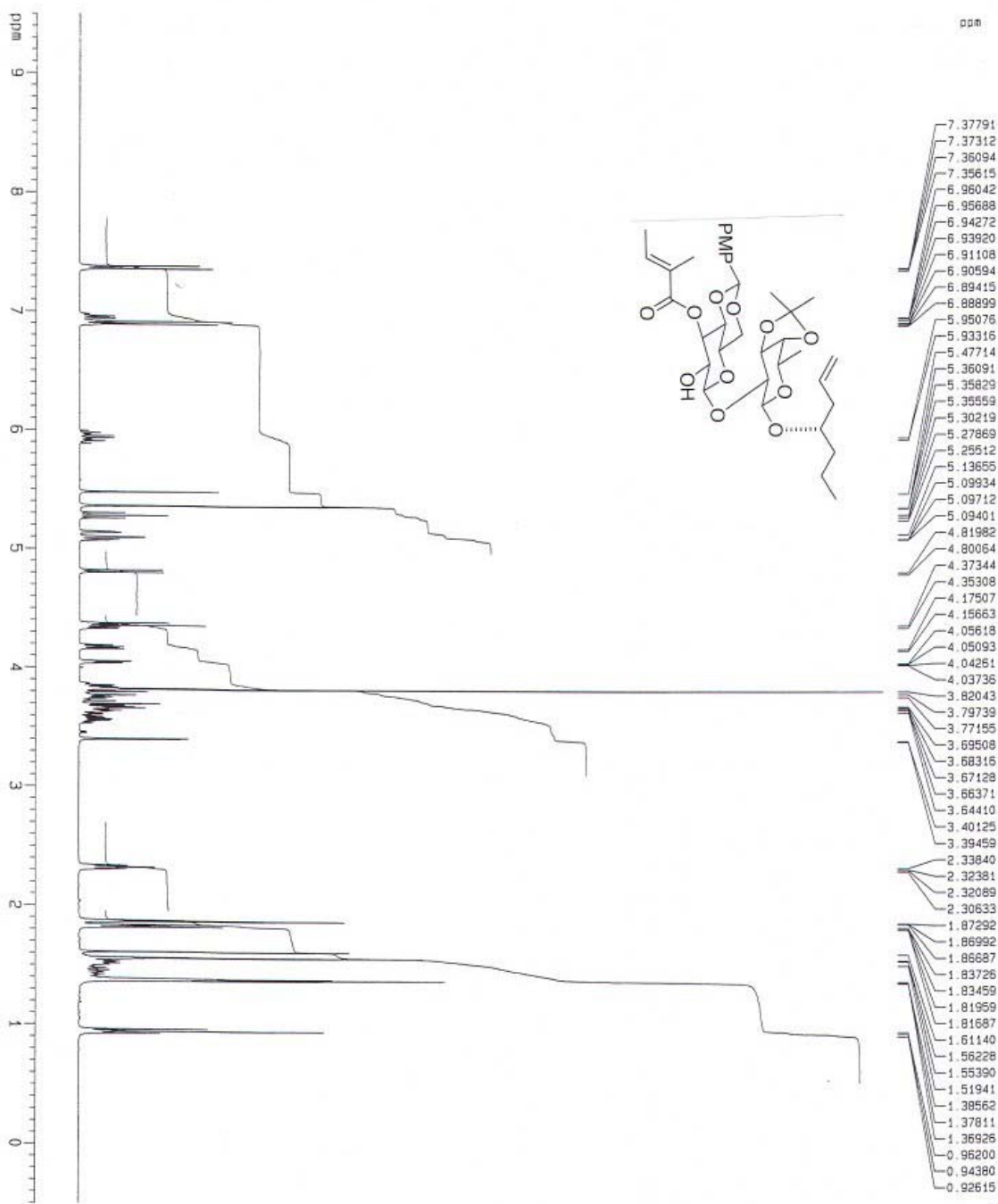
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.04 usec
 PL1 5.00 dB
 SF01 100.6262789 MHz

***** CHANNEL f2 *****
 CROPRG2 waltz16
 NUC2 1H
 PCPG2 90.00 usec
 PL2 0.00 dB
 PL12 19.75 dB
 SF02 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6126888 MHz
 KHZ
 EQ
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.17 Hz
 F2P -30.000 ppm
 F2 -3018.36 Hz
 PRGCM 12.72727 ppm/cm
 HZCM 1280.52515 Hz/cm

MAG-NA-110-01



Current Data Parameters
 NAME: 0707014
 EXNO: 10
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20051207
 Time: 15.13
 INSTRUM: HY400
 PROBNM: 5 mm BBO BB-1H
 PULPROG: zg30
 TO: 69536
 SOLVENT: CDCl2
 NS: 32
 DS: 8
 SMH: 8278.146 Hz
 FIDRES: 0.126314 Hz
 AQ: 3.9964643 sec
 RG: 114
 TM: 60.400 usec
 DE: 0.50 usec
 TE: 303.0 K
 D1: 0.00300000 sec

***** CHANNEL f1 *****
 NUC1: 1H
 P1: 9.25 usec
 PL1: 0.00 dB
 SFO1: 400.1324710 MHz

F2 - Processing parameters
 SI: 32768
 SF: 400.1300000 MHz
 MDW: EM
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 2.00

1D NMR plot parameters
 CX: 22.00 cm
 CY: 15.00 cm
 F1P: 0.500 ppm
 F1: 3801.24 Hz
 F2P: -0.500 ppm
 F2: -200.07 Hz
 PPMCK: 0.46405 ppm/cm
 HZCM: 181.87727 Hz/cm

NAG-NA-127-03
 3-0-acylated?



Current Data Parameters
 NAME dr07036
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051207
 Time 20.26
 INSTRUM av400
 PROBRD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SWH 33112.582 Hz
 FIDRES 0.300239 Hz
 AQ 0.9895435 sec
 RG 16384
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.63000000 sec
 d11 0.63000000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6262789 MHz

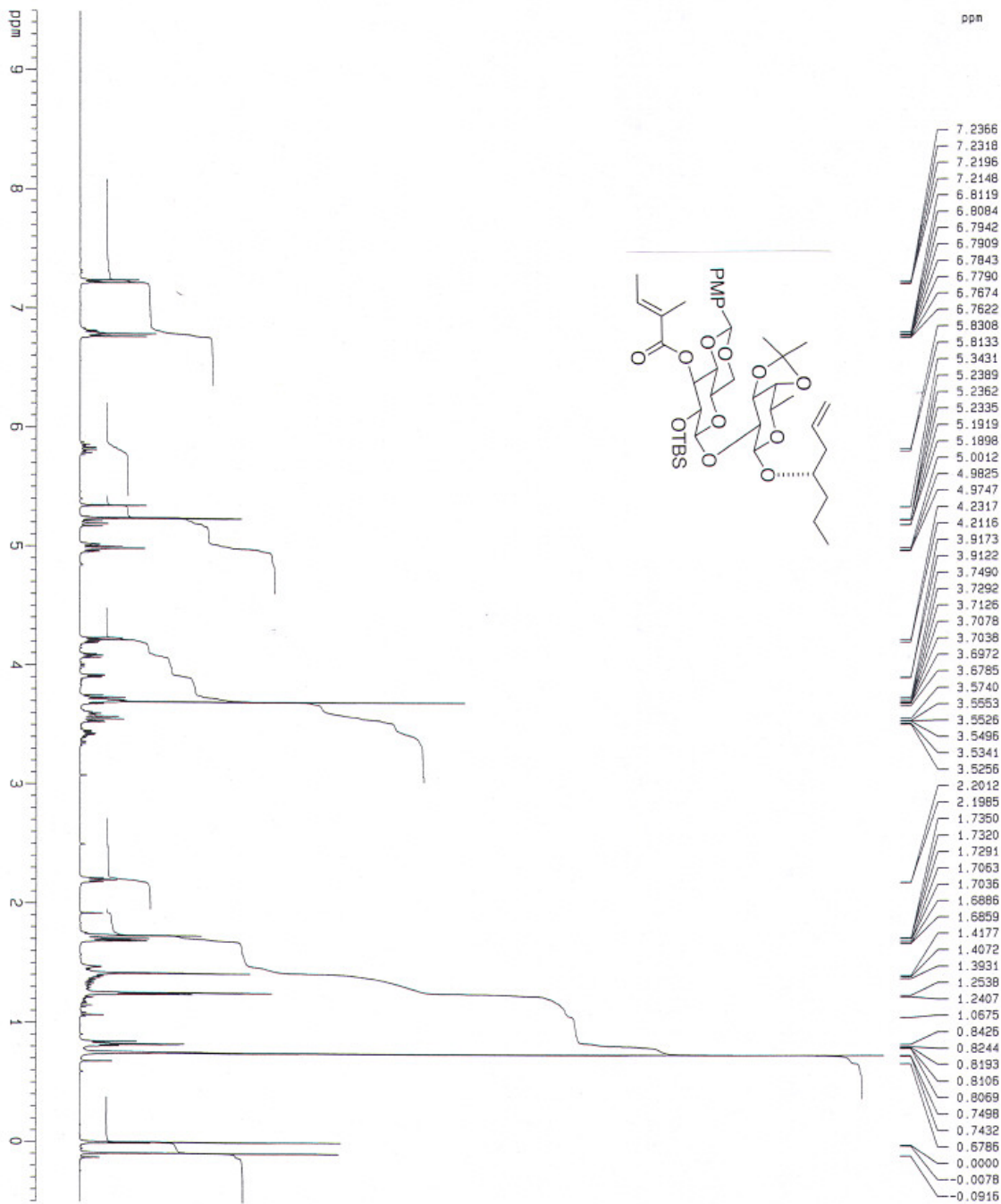
***** CHANNEL f2 *****
 CDPRG2 waltz16
 NUC2 1H
 PCPR2 90.00 usec
 PL2 0.00 dB
 PL12 19.75 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6126988 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 BB 0
 PC 2.00

ID NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.17 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPM0H 12.7272 ppm/cm
 HZ0H 1280.92515 Hz/cm

NAG-NA-127-03

3-O-acetyl?



NAG-NA-133-01

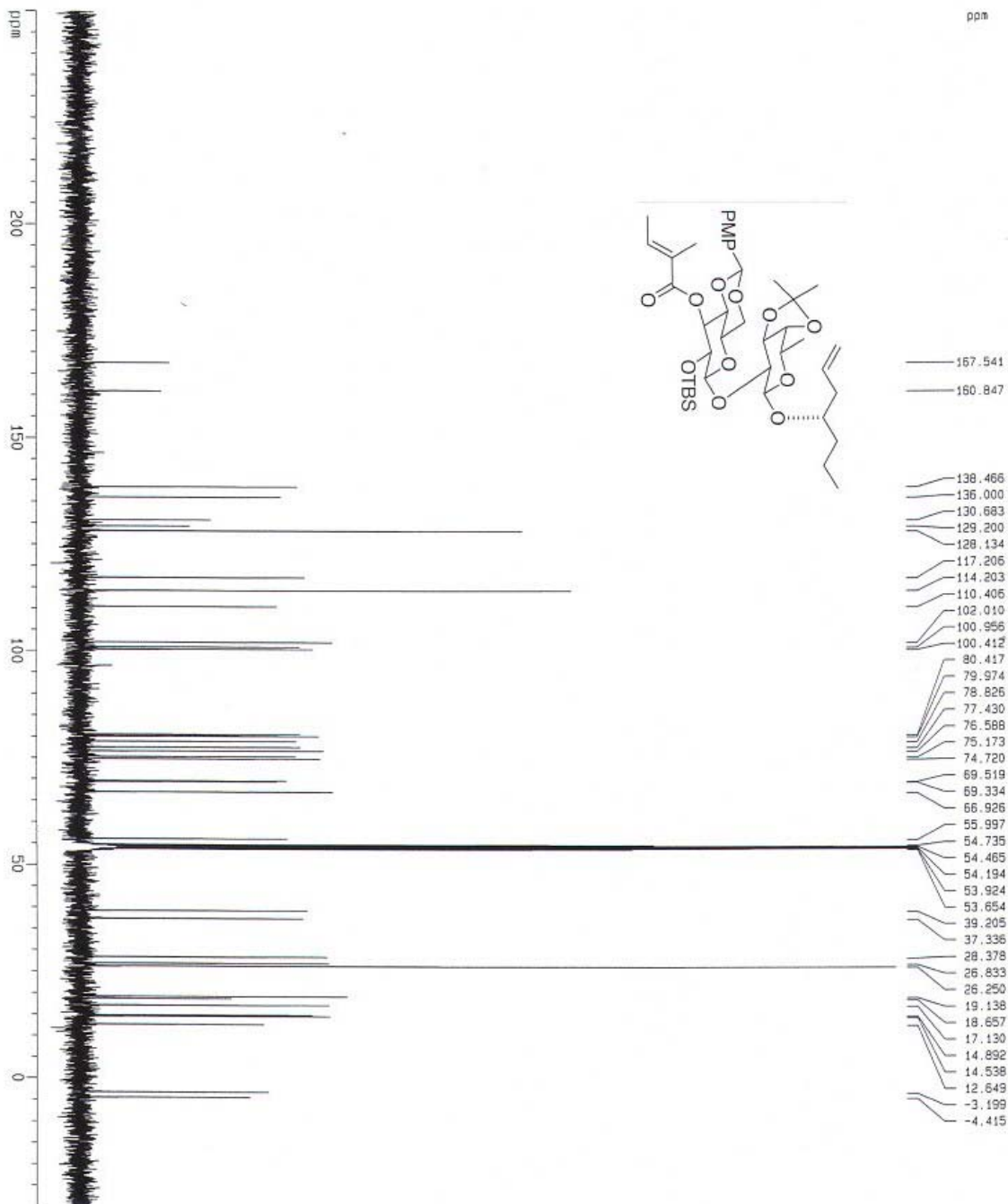
Current Data Parameters
 NAME dr90010
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20031209
 Time 12.03
 INSTRUM av400
 PROBRD 5 mm BBO BB-5H
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9984243 sec
 RG 114
 DW 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 O1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300488 MHz
 KW 4
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.590 ppm
 F1 3801.24 Hz
 F2P -206.07 Hz
 F2 0.4535 ppm/cm
 PPMCK 181.87729 Hz/cm
 HCKM



Current Data Parameters
 NAME 0709010
 EXNO 11
 PNOCDNO 1

F2 - Acquisition Parameters
 Date_ 20051209
 Time 12.31
 INSTRM av400
 PROBHD 5 mm BBO BB-H1
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 1900
 DS 2
 SMT 3312.582 HZ
 FIDRES 0.506258 HZ
 AQ 0.9895436 sec
 RG 16384
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

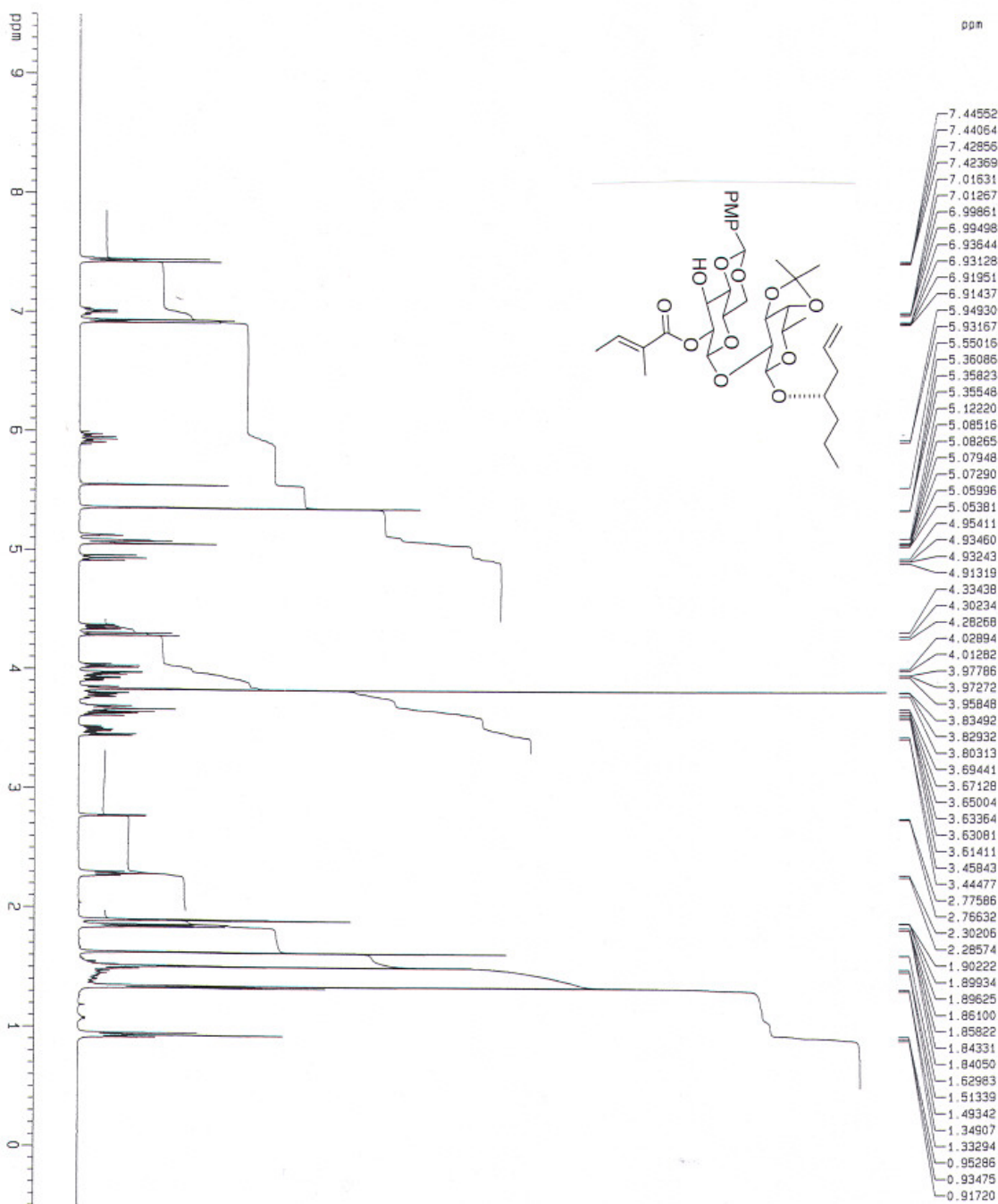
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242789 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 P1P2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6129888 MHz
 MVM EM
 SSB 0
 LB 1.00 HZ
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 250.000 ppm
 F1 25153.17 Hz
 F2 -3018.38 Hz
 PRNCDM 12.72727 ppm/cm
 HZCM 1280.52515 Hz/cm

NAG-NA-133-01



Current Data Parameters
 NAME 0707013
 EXPNO 10
 PROCNO 1

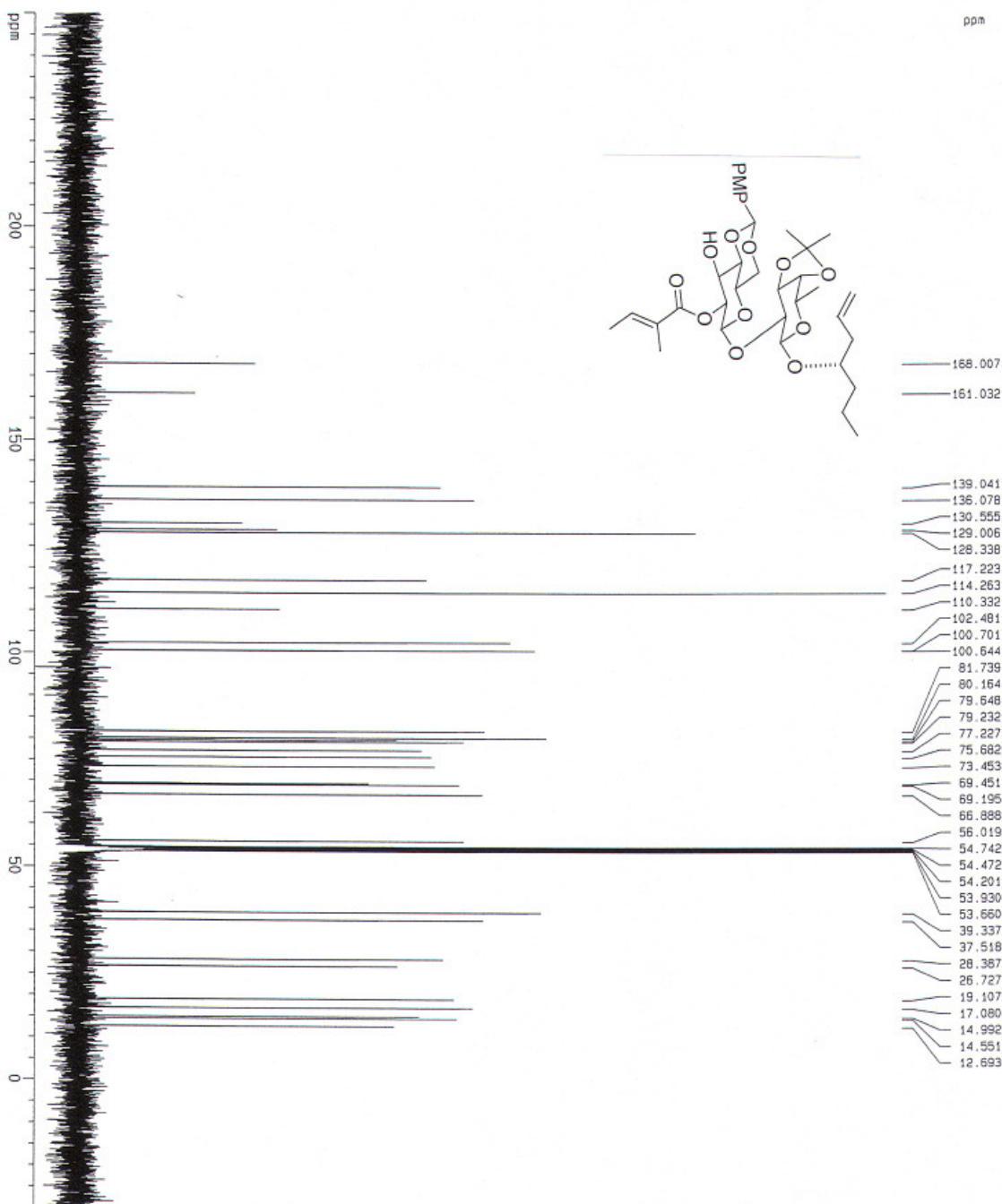
F2 - Acquisition Parameters
 Date_ 20051207
 Time 15.05
 INSTRUM av400
 PPROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 65536
 SFO1 400.124710 MHz
 SOLVENT CDCl3
 NS 32
 DS B
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 114
 DW 60.400 usec
 DE 8.50 usec
 TE 303.0 K
 O1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SFO1 400.124710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 MDW EN
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 0.500 ppm
 F1 3801.24 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPM0X 0.45435 ppm/cm
 PPM0Y 181.87727 Hz/cm

NAG-NA-127-02
 2-0-acylated?



- 168.007
- 161.032
- 139.041
- 136.078
- 130.555
- 129.006
- 128.338
- 117.223
- 114.263
- 110.332
- 102.481
- 100.701
- 100.644
- 81.739
- 80.164
- 79.648
- 79.232
- 77.227
- 75.682
- 73.453
- 69.451
- 69.195
- 66.888
- 56.019
- 54.742
- 54.472
- 54.201
- 53.930
- 53.660
- 39.337
- 37.518
- 28.387
- 26.727
- 19.107
- 17.080
- 14.992
- 14.551
- 12.693

Current Data Parameters
 NAME dr-07035
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051207
 Time 19.54
 INSTRUM av400
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 1900
 DS 2
 SM 33112.582 Hz
 FIDRES 0.5162518 Hz
 AQ 0.9895436 sec
 RG 16394
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

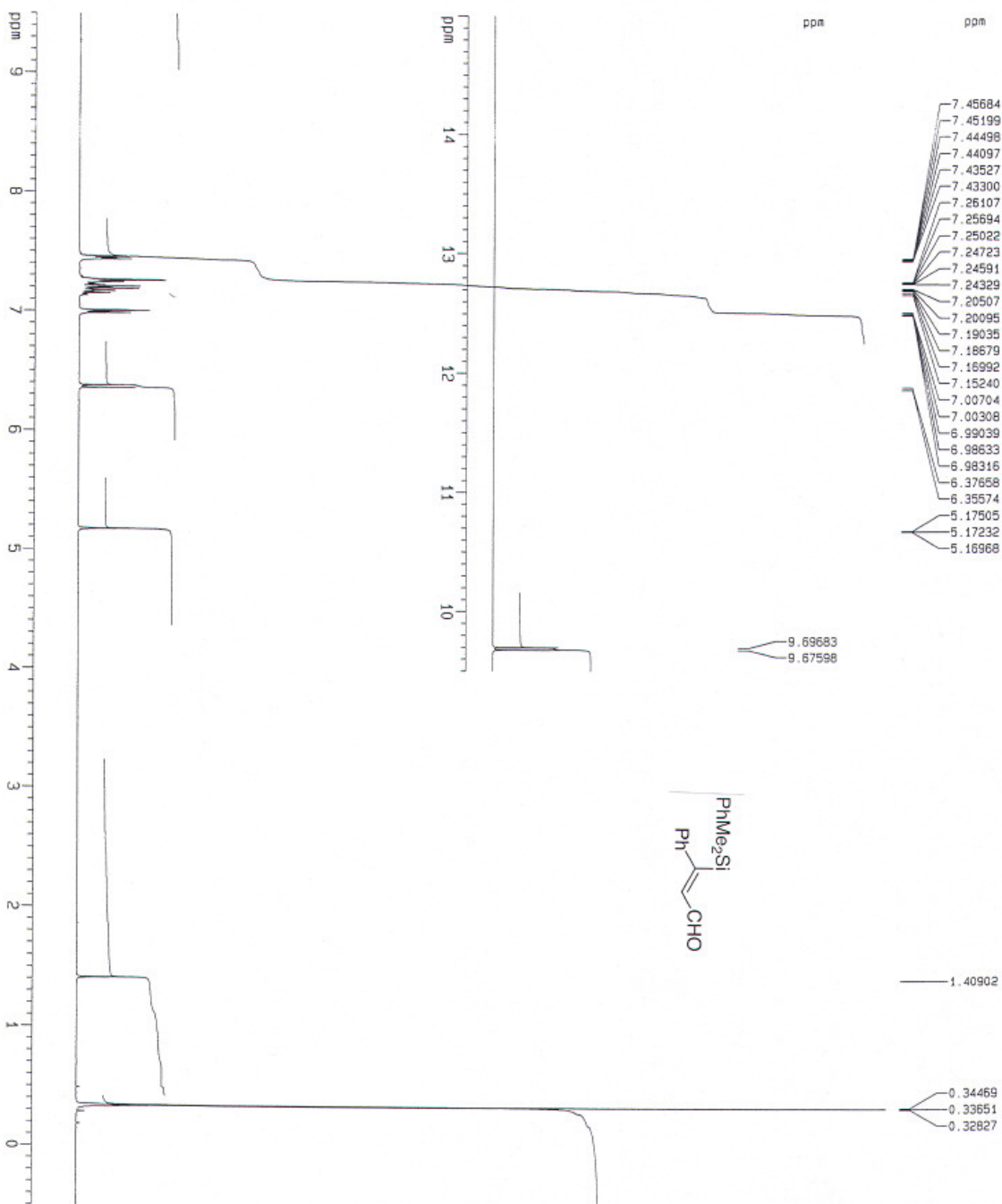
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SF01 100.6242189 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.75 dB
 SF02 400.1324710 MHz

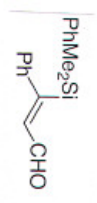
F2 - Processing parameters
 SI 32768
 SF 100.6126888 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.17 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PRNCH 12.7227 ppm/cm
 HZCN 1280.52515 Hz/cm

NA6-NA-127-02
 2-0-acy1?



- 7.45684
- 7.45199
- 7.44498
- 7.44097
- 7.43527
- 7.43300
- 7.26107
- 7.25694
- 7.25022
- 7.24723
- 7.24591
- 7.24329
- 7.20507
- 7.20095
- 7.19035
- 7.18679
- 7.16992
- 7.15240
- 7.00704
- 7.00308
- 6.99039
- 6.98633
- 6.98316
- 6.37658
- 6.35574
- 5.17505
- 5.17232
- 5.16968



- 9.69683
- 9.67598
- 1.40902
- 0.34469
- 0.33851
- 0.32827

Current Data Parameters
 NAME J11031
 EXPNO 10
 PROCNO 1

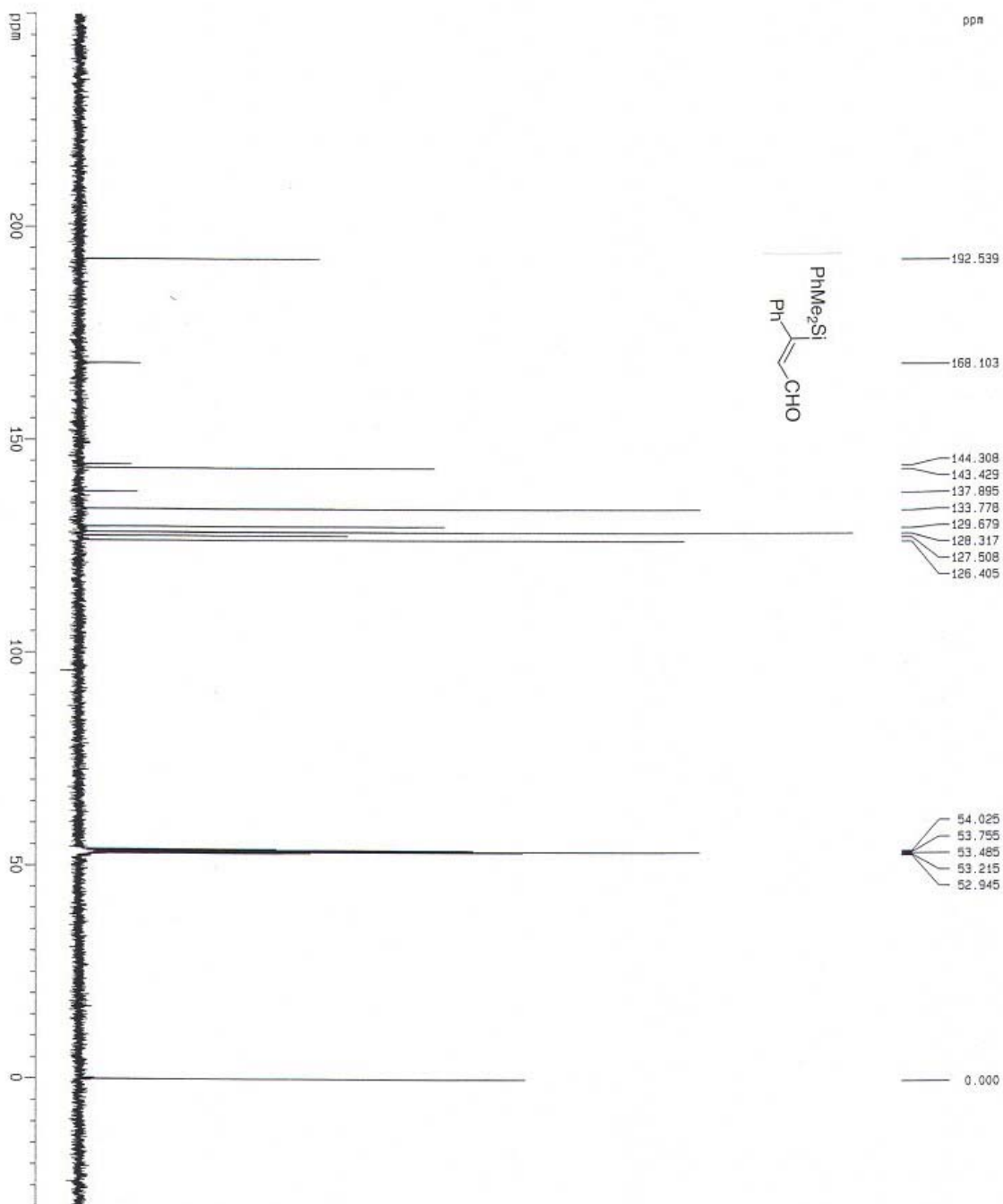
F2 - Acquisition Parameters
 Date_ 20060712
 Time 0.54
 INSTRUM ac400
 PROBNM 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 32
 DS B
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 114
 DW 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300745 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 ppm
 F1 3801.24 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMKCM 0.45455 ppm/cm
 HZCM 181.87730 Hz/cm

NA6-NB-032-01



Current Data Parameters
 NAME JY11031
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060712
 Time 1.23
 INSTRUM sv400
 PROBRD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SMH 33112.582 Hz
 FIDRES 0.505258 Hz
 AQ 0.9896435 sec
 RG 18390.4
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 D11 0.03000000 sec

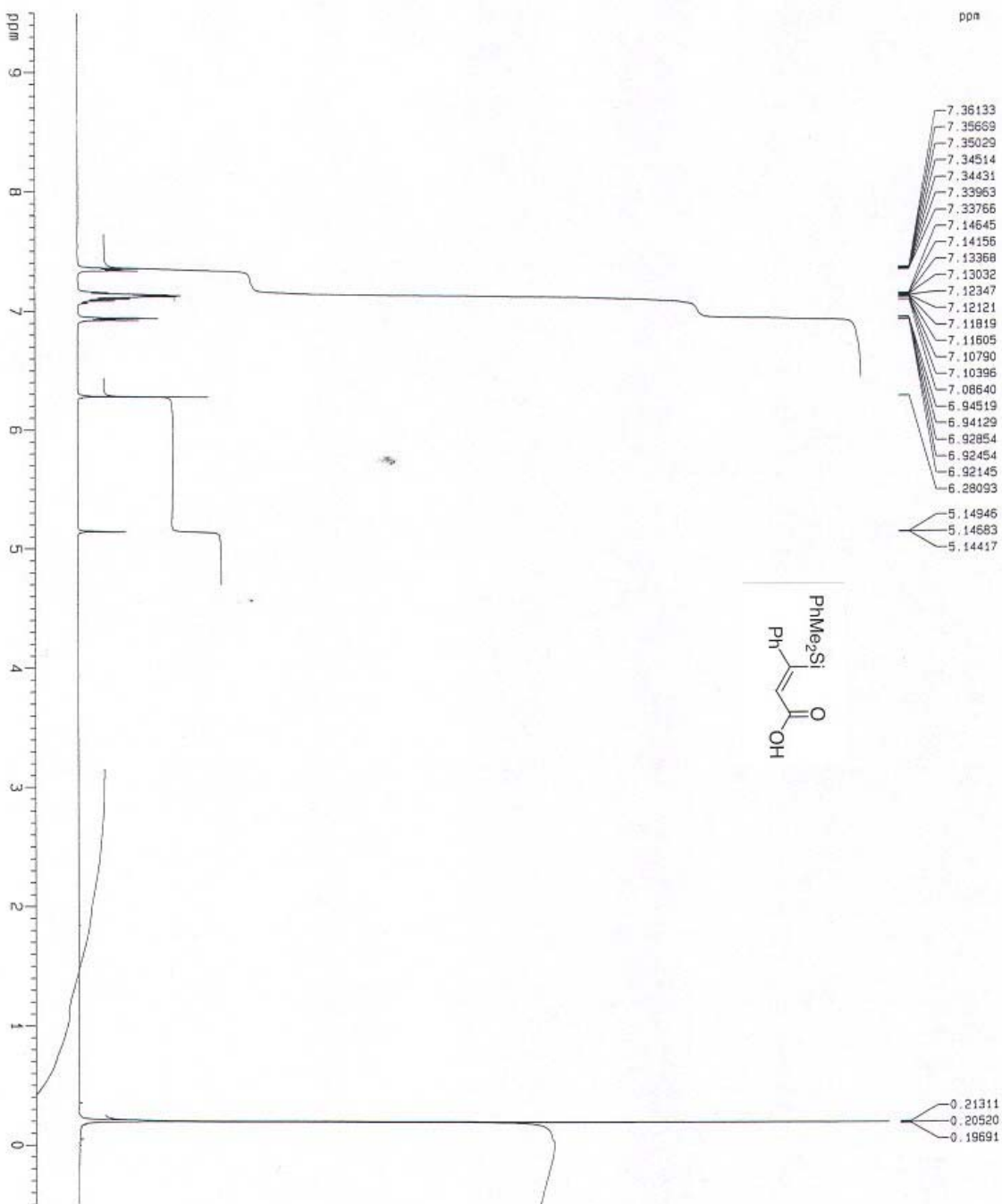
----- CHANNEL f1 -----
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6247789 MHz

----- CHANNEL f2 -----
 CPROG2 hq11z15
 NUC2 1H
 PPOG2 98.00 usec
 PL2 0.00 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127604 MHz
 KDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

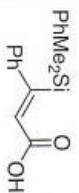
1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 2560.000 DDM
 F1 25553.19 Hz
 F2P -30.000 DDM
 F2 -30416.38 Hz
 PPMCM 12.72727 ppm/cm
 HZCM 1260.55812 Hz/cm

NAG-NB-032-01



- 7.36133
- 7.35659
- 7.35029
- 7.34514
- 7.34431
- 7.33963
- 7.33766
- 7.14645
- 7.14156
- 7.13368
- 7.13032
- 7.12347
- 7.12121
- 7.11819
- 7.11605
- 7.10790
- 7.10396
- 7.08640
- 6.94519
- 6.94129
- 6.92854
- 6.92454
- 6.92145
- 6.28093
- 5.14946
- 5.14683
- 5.14417

- 0.21311
- 0.20520
- 0.19591



```

Current Data Parameters
NAME      J115072
EXPNO     10
PROCNO    1

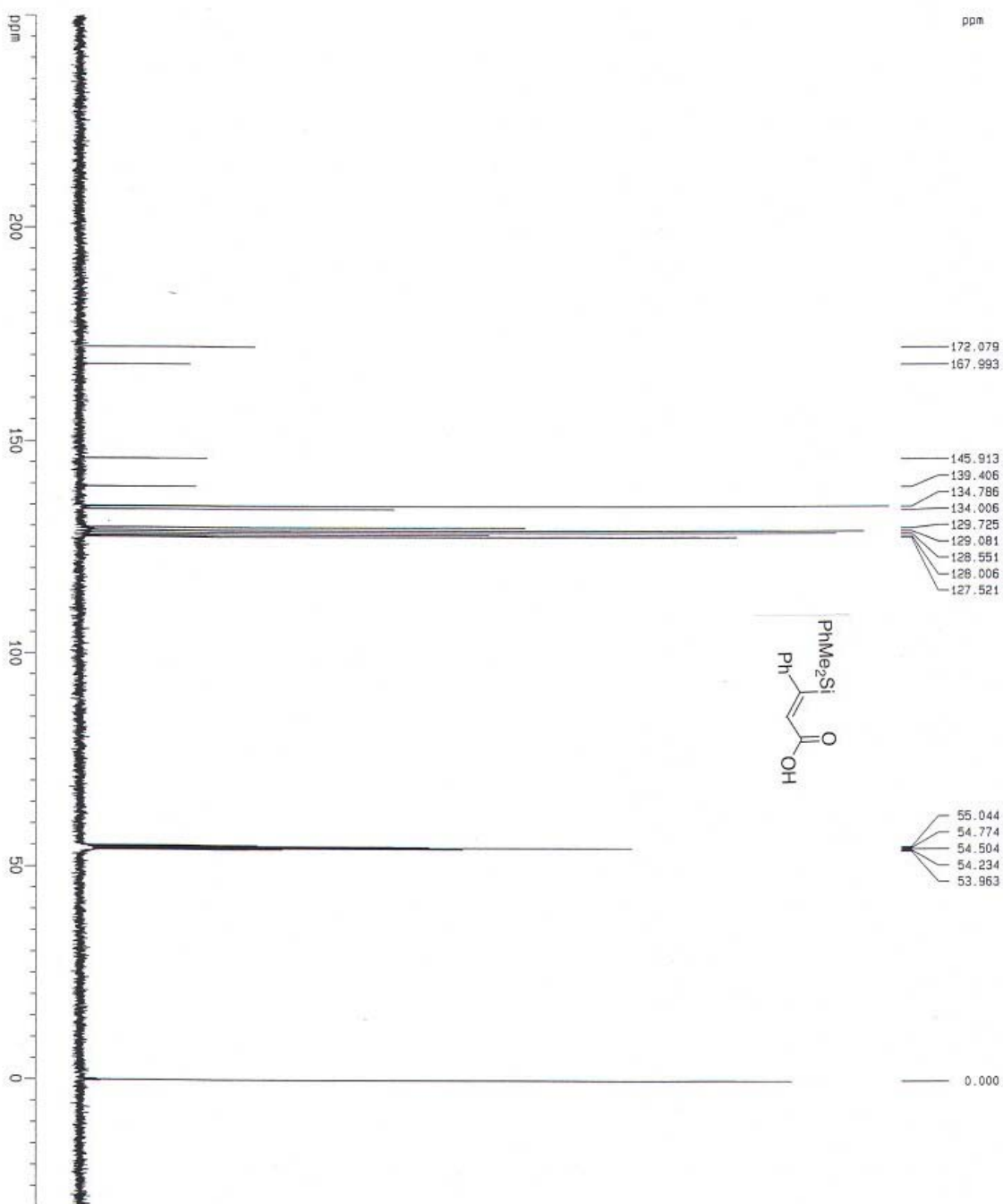
F2 - Acquisition Parameters
Date_     20060713
Time      10.38
INSTRUM   av400
PROBHD    5 mm BBO BB-1H
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         32
DS         2
SWH        8278.146 Hz
FIDRES     0.126314 Hz
AQ         3.9564243 sec
RG         114
DE         60.400 usec
TE         303.0 K
D1         0.00300000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         9.25 usec
PL1        0.00 dB
SFO1       400.1324710 MHz

F2 - Processing parameters
SI         32768
SF         400.1300049 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         2.00

10 MHz plot parameters
CX         22.00 cm
CY         15.00 cm
F1p        9.500 ppm
F1         3801.24 Hz
F2p        -0.500 ppm
F2         -200.07 Hz
PPOCKW    0.45655 ppm/cm
H2CKW     181.87732 Hz/cm
    
```

NAG-NB-033-01



Current Data Parameters
 NAME 1Y12072
 EXNO 11
 PRUNO 1

F2 - Acquisition Parameters
 Date_ 20060713
 Time 11.06
 INSTRUM av400
 PROBRD 5 mm BBO BB-H1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1900
 DS 2
 SWH 33112.582 Hz
 FIDRES 0.505258 Hz
 AQ 0.3896426 sec
 RG 18398.4
 DW 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 O1 0.03000000 sec
 d11 0.03000000 sec

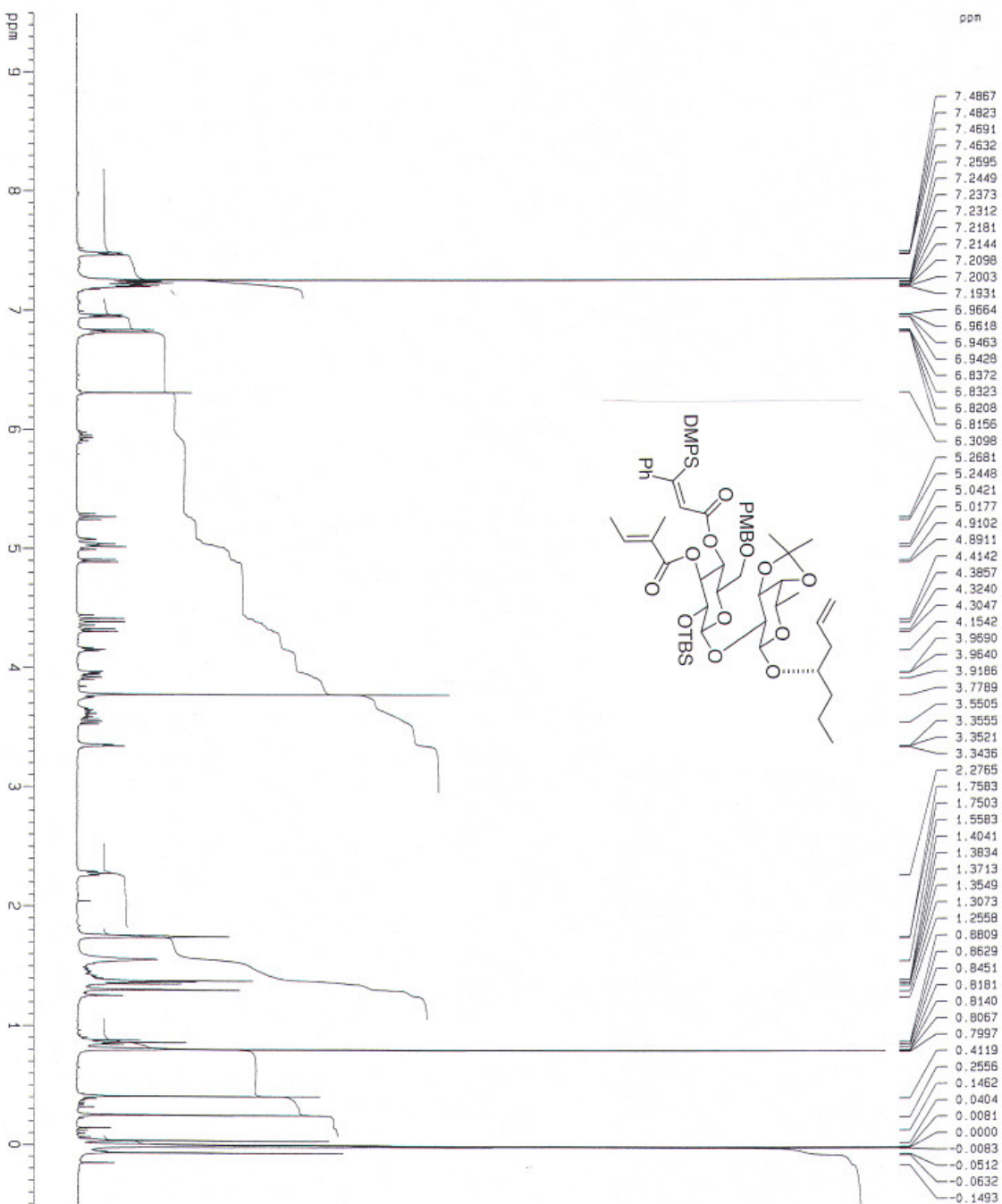
----- CHANNEL f1 -----
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242789 MHz

----- CHANNEL f2 -----
 CPDPRG2 MALT216
 NUC2 1H
 P2 90.00 usec
 PCPR2 0.00 dB
 PL2 19.75 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.612590 MHz
 MEW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.17 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PRNOM 12.72127 ppm/cm
 HZCM 1280.52466 Hz/cm

NAG-NB-033-01



Current Data Parameters
 NAME mp08205
 EXPNO 10
 PROCNO 1

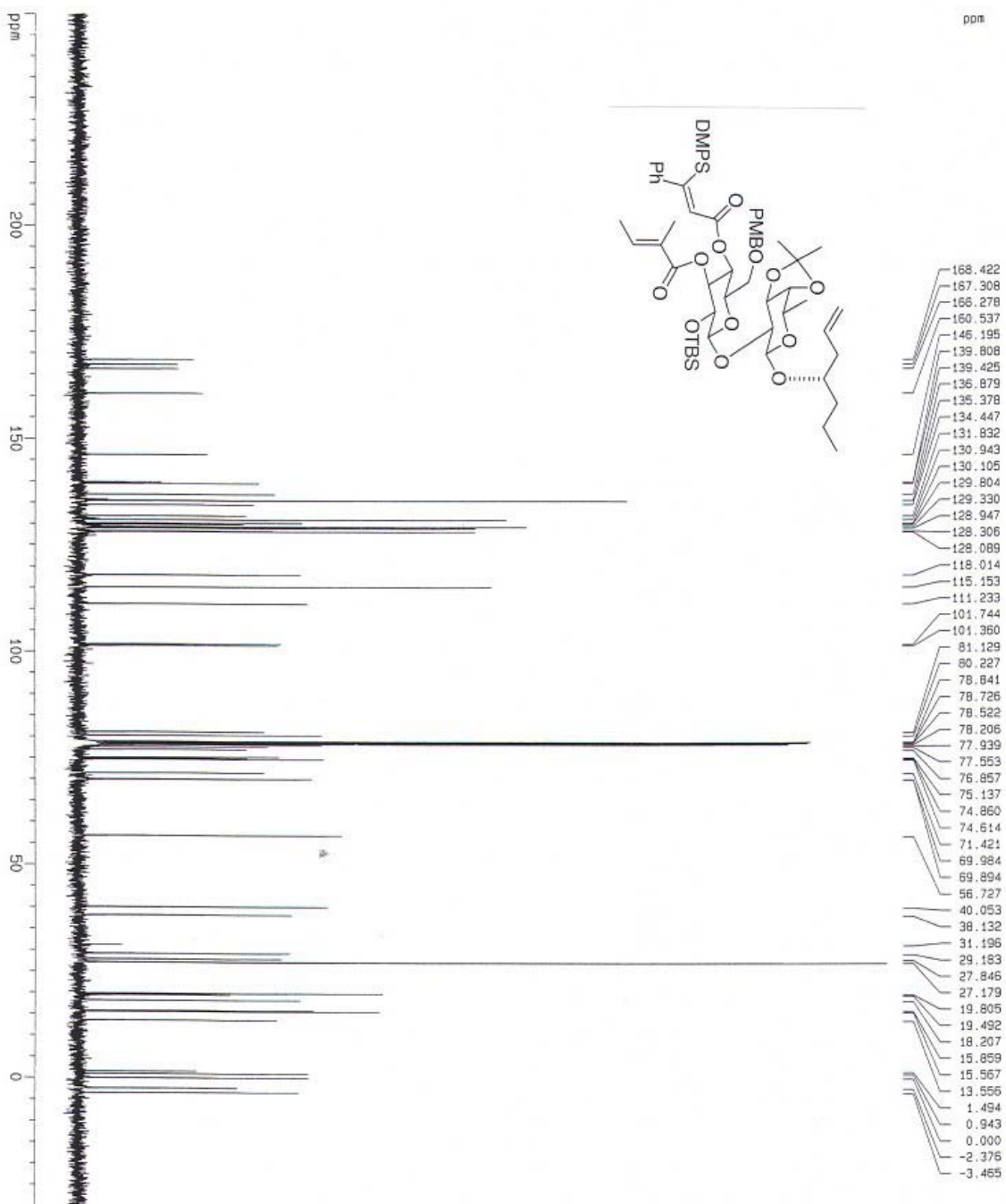
F2 - Acquisition Parameters
 Date_ 20060508
 Time 14:16
 INSTRUM spect
 PROBHD 5 mm BBO BB-3H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 322.5
 OW 60.400 usec
 OE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SFO1 400.1264710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300099 MHz
 NQW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 ppm
 F1 3901.24 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPGM h1
 0.48455 ppm/cm
 181.8727 Hz/cm

NA6-NA-229-02



Current Data Parameters
 NAME ey09031
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060509
 Time 16.21
 INSTRUM av400
 PROBR0 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SMH 33112.982 Hz
 FIDRES 0.5002568 Hz
 AQ 0.9896435 sec
 RG 18390.4
 DM 15.100 USEC
 DE 8.00 USEC
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

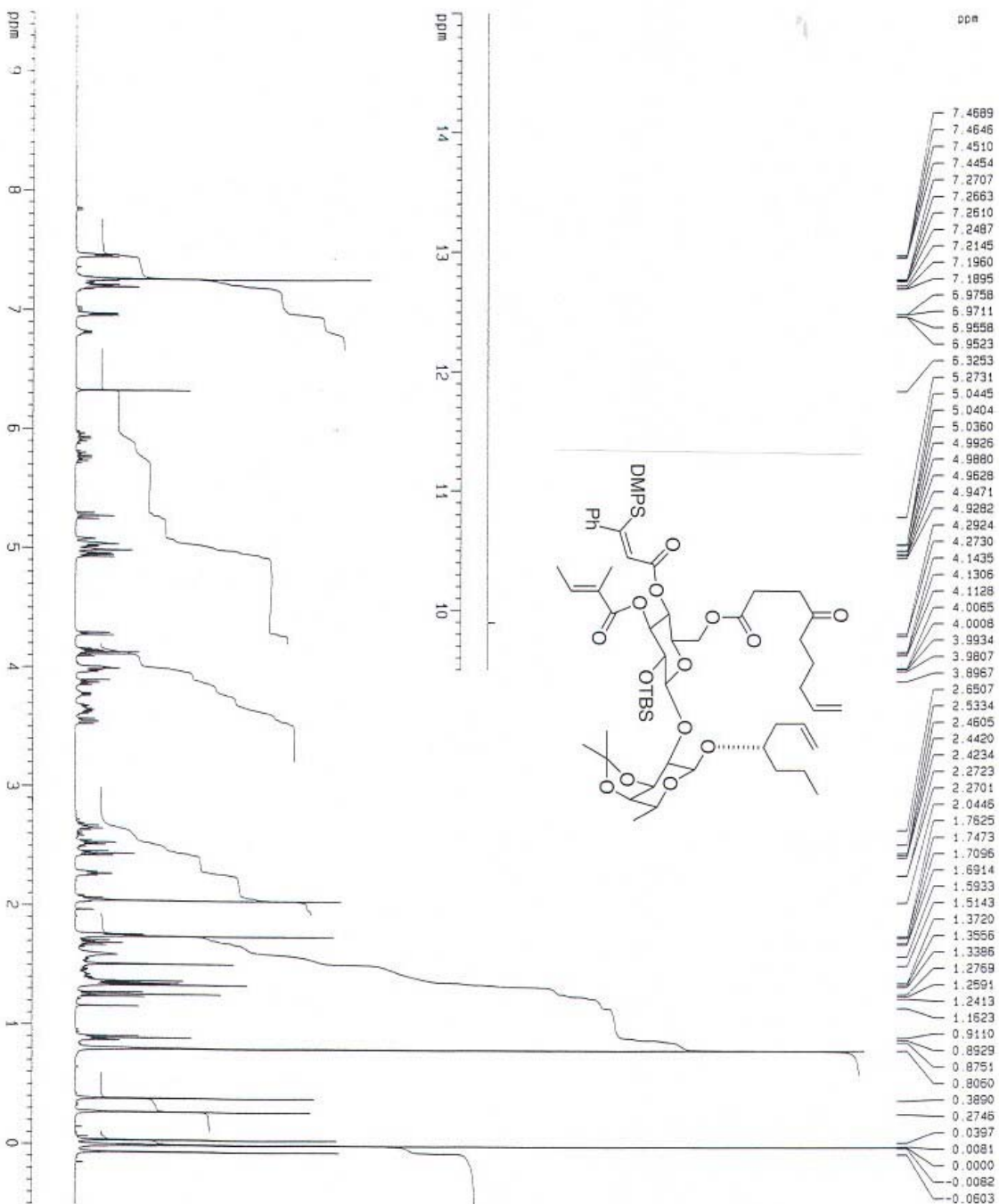
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242789 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 P2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters:
 SI 32768
 SF 100.6126177 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

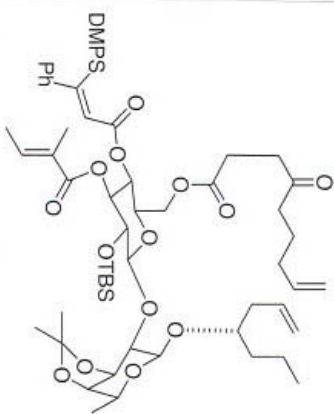
10 NMR plot parameters:
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.16 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPM/CH 12.72727 ppm/cm
 HZ/CM 1380.52417 Hz/cm

NAG-NA-229-02



ppm

| |
|--------|
| 7.4589 |
| 7.4646 |
| 7.4510 |
| 7.4454 |
| 7.2707 |
| 7.2663 |
| 7.2610 |
| 7.2487 |
| 7.2145 |
| 7.1960 |
| 7.1895 |
| 6.9758 |
| 6.9711 |
| 6.9558 |
| 6.9523 |
| 6.3253 |
| 5.2731 |
| 5.0445 |
| 5.0404 |
| 5.0360 |
| 4.9926 |
| 4.9880 |
| 4.9628 |
| 4.9471 |
| 4.9282 |
| 4.2924 |
| 4.2730 |
| 4.1435 |
| 4.1305 |
| 4.1128 |
| 4.0065 |
| 4.0008 |
| 3.9934 |
| 3.9807 |
| 3.8967 |
| 2.6507 |
| 2.5334 |
| 2.4605 |
| 2.4420 |
| 2.4234 |
| 2.2723 |
| 2.2701 |
| 2.0446 |
| 1.7625 |
| 1.7473 |
| 1.7096 |
| 1.6914 |
| 1.5933 |
| 1.5143 |
| 1.3720 |
| 1.3556 |
| 1.3386 |
| 1.2769 |
| 1.2591 |
| 1.2413 |
| 1.1623 |
| 0.9110 |
| 0.8929 |
| 0.8751 |
| 0.8060 |
| 0.3890 |
| 0.2746 |
| 0.0397 |
| 0.0081 |
| 0.0000 |
| 0.0082 |
| 0.0503 |



Current Data Parameters
 NAME 1Y19027
 EXPNO 10
 PROCNO 1

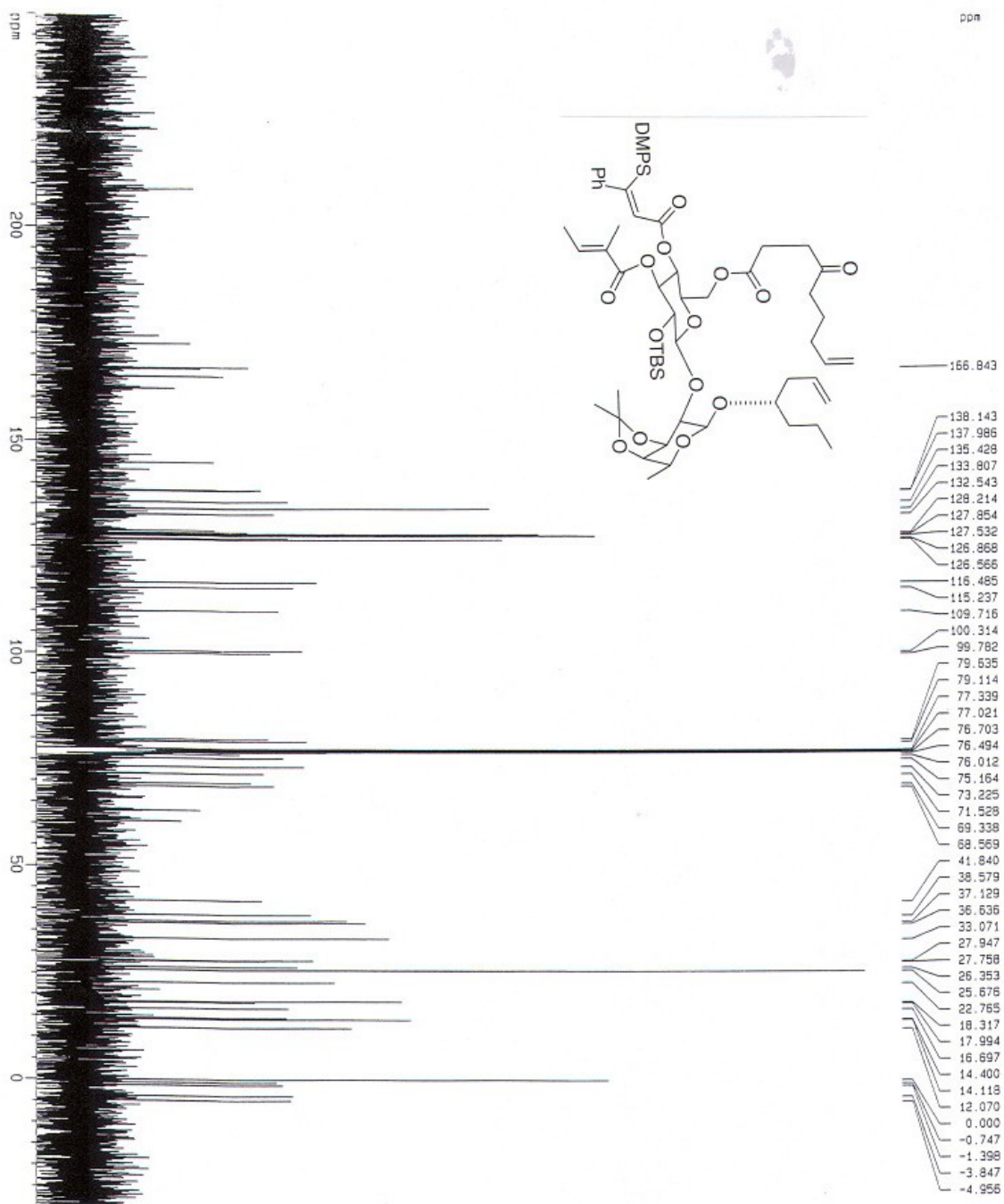
F2 - Acquisition Parameters
 Date_ 20060719
 Time 23.29
 INSTRUM 84400
 PROBHD 5 mm 880 86-1H
 PULPROG zg30
 TO 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SH 8278 146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 181
 DM 60.400 usec
 DE 6.50 usec
 TE 303.2 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300933 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters:
 CX 22.00 cm
 CY 15.00 cm
 FIP 9.500 ppm
 FL 8801.24 Hz
 F2P -0.500 ppm
 FZ -200.07 Hz
 PRNOM 0.45455 ppm/cm
 NCH 181.8777 Hz/cm

NAG-NB-040-01



| Chemical Shift (ppm) |
|----------------------|
| 166.843 |
| 138.143 |
| 137.986 |
| 135.428 |
| 133.807 |
| 132.543 |
| 128.214 |
| 127.854 |
| 127.532 |
| 126.868 |
| 126.566 |
| 116.485 |
| 115.237 |
| 109.716 |
| 100.314 |
| 99.782 |
| 79.635 |
| 79.114 |
| 77.339 |
| 77.021 |
| 76.703 |
| 76.494 |
| 76.012 |
| 75.164 |
| 73.225 |
| 71.528 |
| 69.338 |
| 68.569 |
| 41.840 |
| 38.579 |
| 37.129 |
| 36.636 |
| 33.071 |
| 27.947 |
| 27.758 |
| 26.353 |
| 25.676 |
| 22.765 |
| 18.317 |
| 17.994 |
| 16.697 |
| 14.400 |
| 14.118 |
| 12.070 |
| 0.000 |
| -0.747 |
| -1.398 |
| -3.847 |
| -4.956 |

Current Data Parameters
 NAME 1Y19027
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060719
 Time 22:57
 INSTRUM sv400
 PULPROG zgpg30
 F1 65536
 F2 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SMI 33112.582 Hz
 FIDRES 0.505258 Hz
 AQ 0.9806436 sec
 RG 16394
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

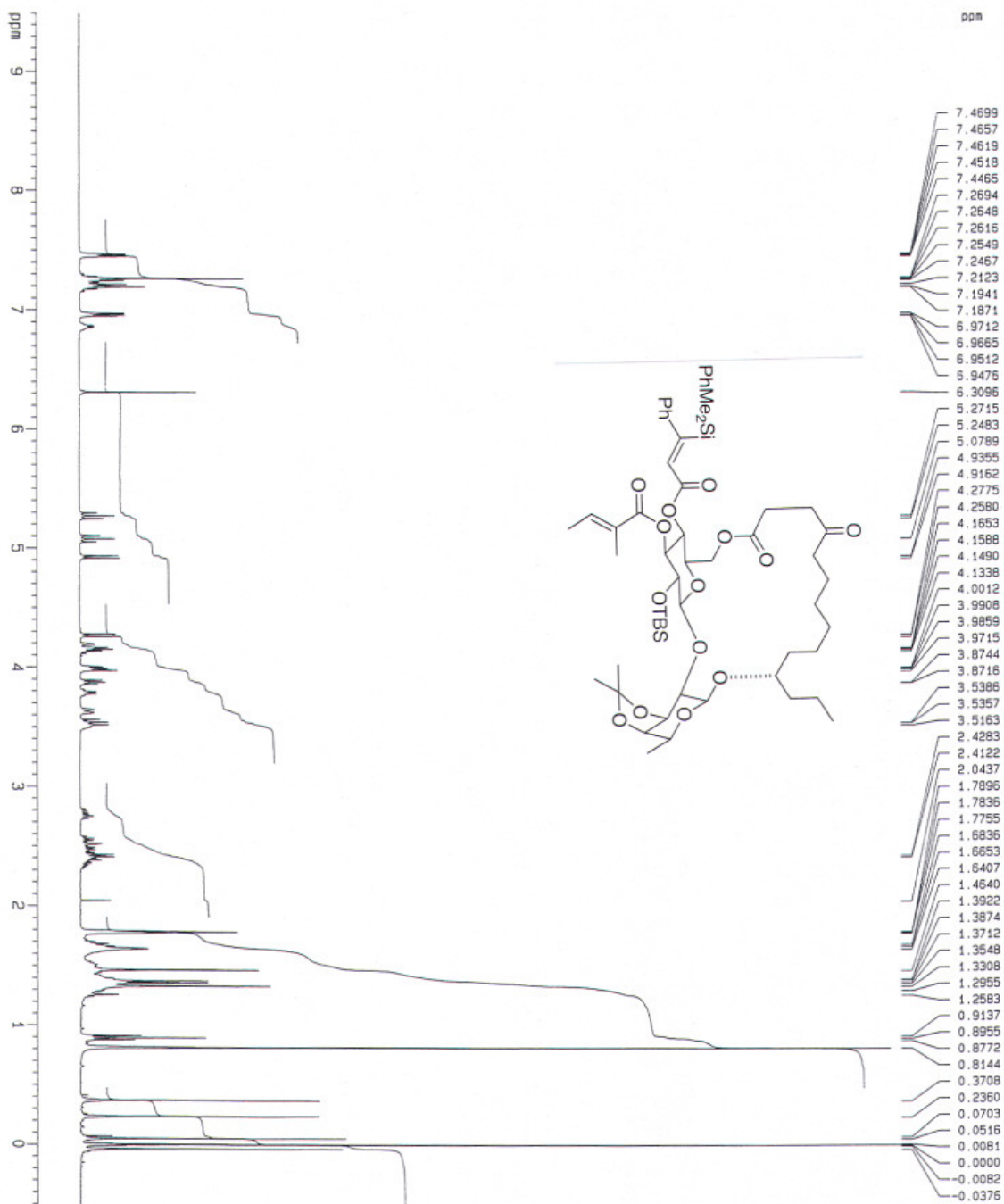
***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.624789 MHz

***** CHANNEL f2 *****
 PULPROG waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 S1 32768
 SF 100.6127664 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.19 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPM/CM 12.72727 ppm/cm
 HZ/CM 1280.58512 Hz/cm

NAG-NB-040-01



ppm

- 7.4699
- 7.4657
- 7.4619
- 7.4518
- 7.4465
- 7.2694
- 7.2648
- 7.2616
- 7.2549
- 7.2467
- 7.2123
- 7.1941
- 7.1871
- 6.9712
- 6.9665
- 6.9512
- 5.9476
- 6.3096
- 5.2715
- 5.2483
- 5.0789
- 4.9355
- 4.9162
- 4.2775
- 4.2580
- 4.1653
- 4.1588
- 4.1490
- 4.1338
- 4.0012
- 3.9908
- 3.9859
- 3.9715
- 3.8744
- 3.8716
- 3.5386
- 3.5357
- 3.5163
- 2.4283
- 2.4122
- 2.0437
- 1.7896
- 1.7836
- 1.7755
- 1.6836
- 1.6653
- 1.6407
- 1.4640
- 1.3922
- 1.3874
- 1.3712
- 1.3548
- 1.3308
- 1.2955
- 1.2583
- 0.9137
- 0.8955
- 0.8772
- 0.8144
- 0.3708
- 0.2360
- 0.0703
- 0.0516
- 0.0081
- 0.0000
- 0.0082
- 0.0376

Current Data Parameters
 NAME 1Y29524
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080723
 Time 15.09

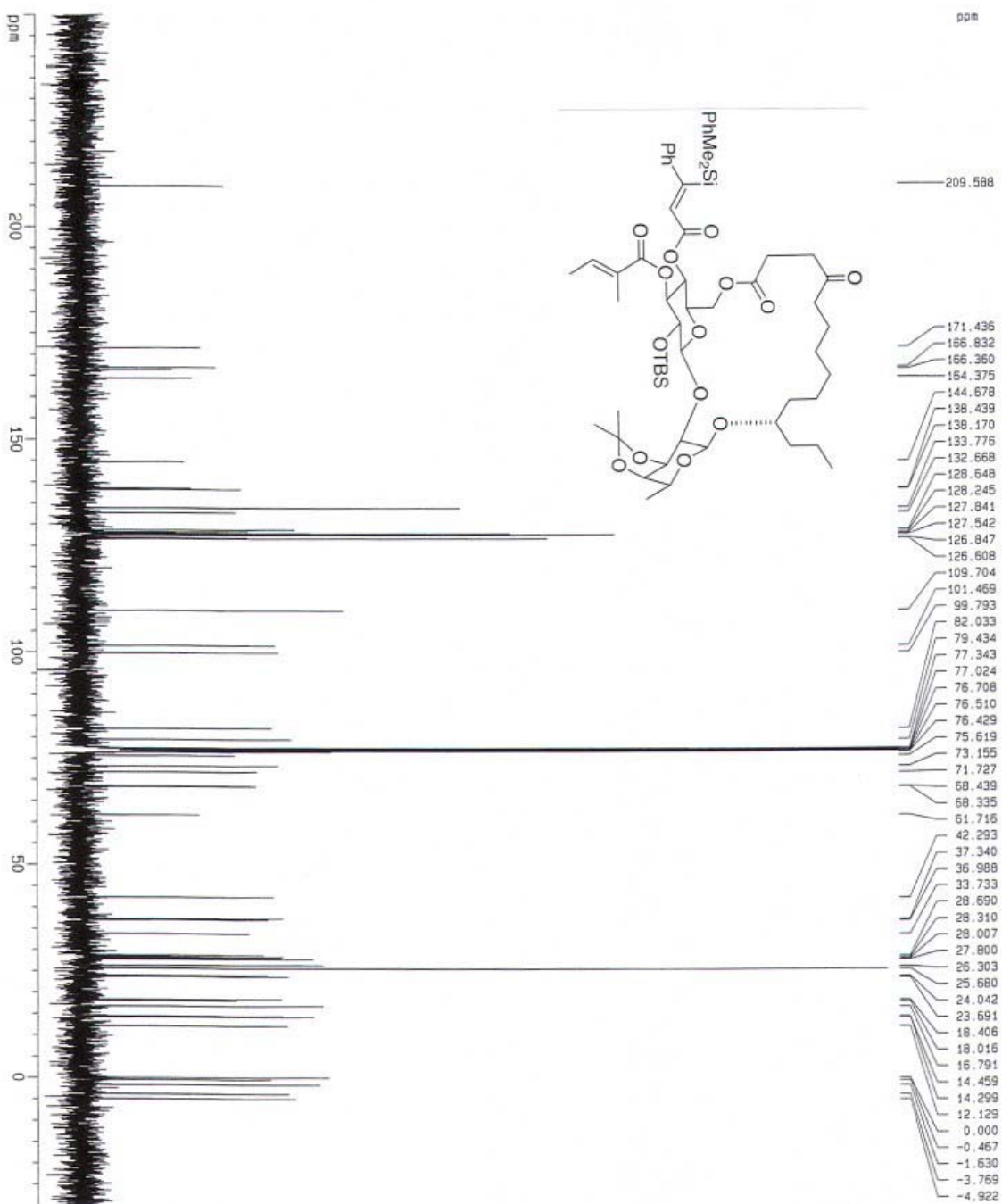
INSTRUM av400
 PROBRD 5 mm BBO BB-1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SMH 8278.146 Hz
 FTRES 0.126314 Hz
 AQ 3.9594243 sec
 RG 114
 DM 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00200000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.23 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300091 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.500 ppm
 F1 3801.24 Hz
 F2P -200.07 Hz
 F2 0.46555 ppm/cm
 PPRCM 181.87727 Hz/cm
 HZCM

NAG-NB-049-01
 IPOB H2 column



Current Data Parameters
 NAME 1J25024
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060725
 Time 15:37
 INSTRUM av400
 PROBRD 5 mm BBO BB-H1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 1900
 DS 2
 SMH 33112.592 Hz
 FIDRES 0.505259 Hz
 AQ 0.9896436 sec
 RG 18390.4
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 O1 0.03000000 sec
 D11 0.03000000 sec

***** CHANNEL f1 *****
 NUC1 ¹³C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.6242189 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 ¹H
 PCP02 90.00 usec
 PL2 0.00 dB
 PL12 19.75 dB
 SFO2 400.1324710 MHz

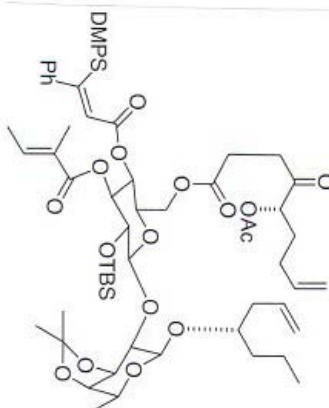
F2 - Processing parameters
 SI 32768
 SF 100.6127654 MHz
 KW EN
 LB 1.00 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 25153.19 Hz
 F2P -30.000 ppm
 F2 -3016.38 Hz
 PRHCK 12.72727 ppm/cm
 HZCM 1280.52612 Hz/cm

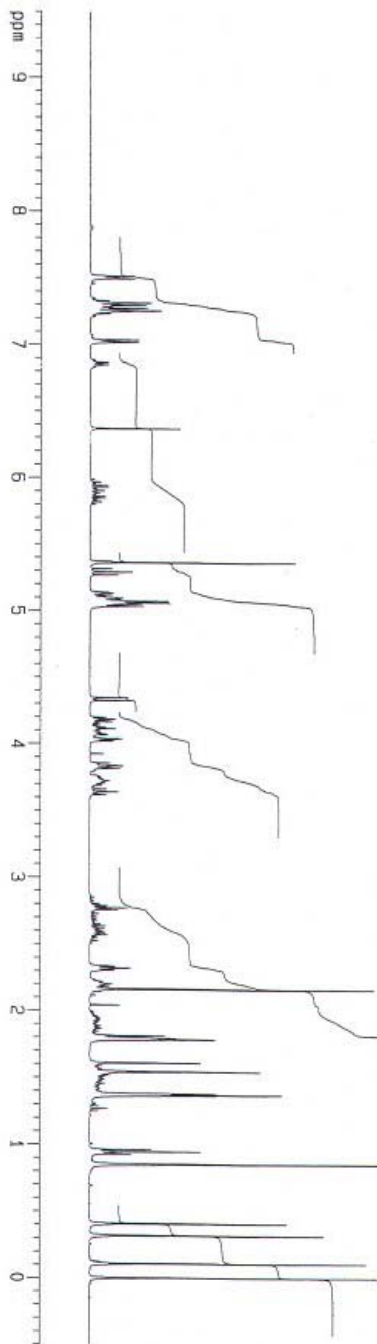
NAG-NB-049-01
 IpOB H2 column

ppm

- 7.51274
- 7.50888
- 7.49418
- 7.48997
- 7.31548
- 7.31059
- 7.30860
- 7.28988
- 7.26790
- 7.24995
- 7.24574
- 7.24162
- 7.03591
- 7.03137
- 7.01579
- 7.01222
- 6.36844
- 6.36492
- 5.36163
- 5.35900
- 5.35623
- 5.28936
- 5.26608
- 5.09853
- 5.09363
- 5.08857
- 5.08088
- 5.07003
- 5.05614
- 5.05081
- 5.04396
- 5.03282
- 4.34426
- 4.32402
- 4.11181
- 4.03813
- 4.03335
- 3.83472
- 3.83307
- 3.81625
- 3.63980
- 2.77053
- 2.75442
- 2.32260
- 2.31992
- 2.16288
- 2.15817
- 2.04254
- 1.81329
- 1.81072
- 1.79295
- 1.78495
- 1.78197
- 1.61190
- 1.60702
- 1.54289
- 1.37992
- 1.37225
- 1.36366
- 0.96105
- 0.94294
- 0.92543
- 0.84866
- 0.40637
- 0.40417
- 0.31707
- 0.10695
- 0.00000



ppm



Current Data Parameters
NAME 1J17020
EXNO 10
PROBHD 1

F2 - Acquisition Parameters
Date_ 20060717
Time 16.01

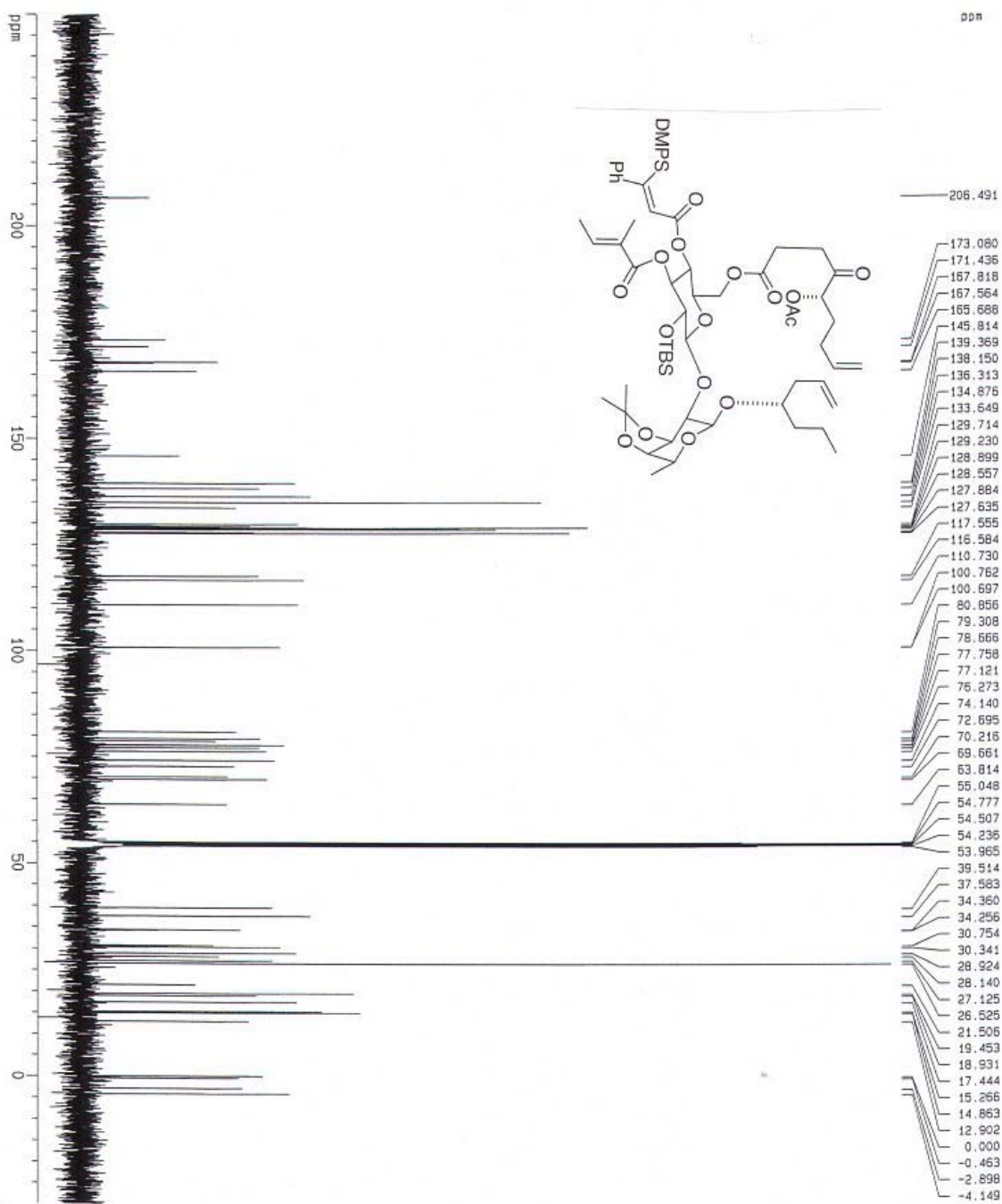
INSTRUM 9V400
PROBHD 5 mm BBO BB-H1
PULPROG zg30
TO 65536
SOLVENT CDCl3
NS 32
DS 8
SMH 8278.146 Hz
FLORES 0.126314 Hz
AQ 3.9894243 sec
RG 66
DM 114
DE 60.400 usec
TE 5.50 usec
TD 30310 K
D1 0.00300000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 9.25 usec
PL1 0.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300000 MHz
MDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 2.00

10 NMR plot parameters:
CX 23.00 cm
CY 15.00 cm
F1P 9.500 B0M
F1 3801.24 Hz
F2P -0.500 B0M
F2 -200.07 Hz
FREQM 0.43455 B0V/cm
H2CM 181.87277 Hz/cm

NAG-NB-038-02



Current Data Parameters

| | |
|--------|---------|
| NAME | 11/1020 |
| EXPNO | 11 |
| PROCNO | 1 |

F2 - Acquisition Parameters

| | |
|---------|----------------|
| Date_ | 20060717 |
| Time | 16.29 |
| INSTRUM | AV400 |
| PROBHD | 5 mm BBO BB-4H |
| PULPROG | ZPGC30 |
| TD | 65536 |
| SOLVENT | CDCl3 |
| NS | 1500 |
| DS | 2 |
| SWH | 33112.582 Hz |
| FIDRES | 0.1505258 Hz |
| AQ | 0.5895436 sec |
| RG | 16384 |
| DM | 15.100 usec |
| DE | 8.00 usec |
| TE | 303.0 K |
| D1 | 0.03000000 sec |
| d11 | 0.03000000 sec |

----- CHANNEL f1 -----

| | |
|------|-----------------|
| NUC1 | 13C |
| PL1 | 10.94 usec |
| PL12 | 3.00 dB |
| SFO1 | 100.6242789 MHz |

----- CHANNEL f2 -----

| | |
|---------|-----------------|
| CPDPRG2 | waltz16 |
| NUC2 | 1H |
| PCPD2 | 90.00 usec |
| PL2 | 0.00 dB |
| PL12 | 19.76 dB |
| SFO2 | 400.1324710 MHz |

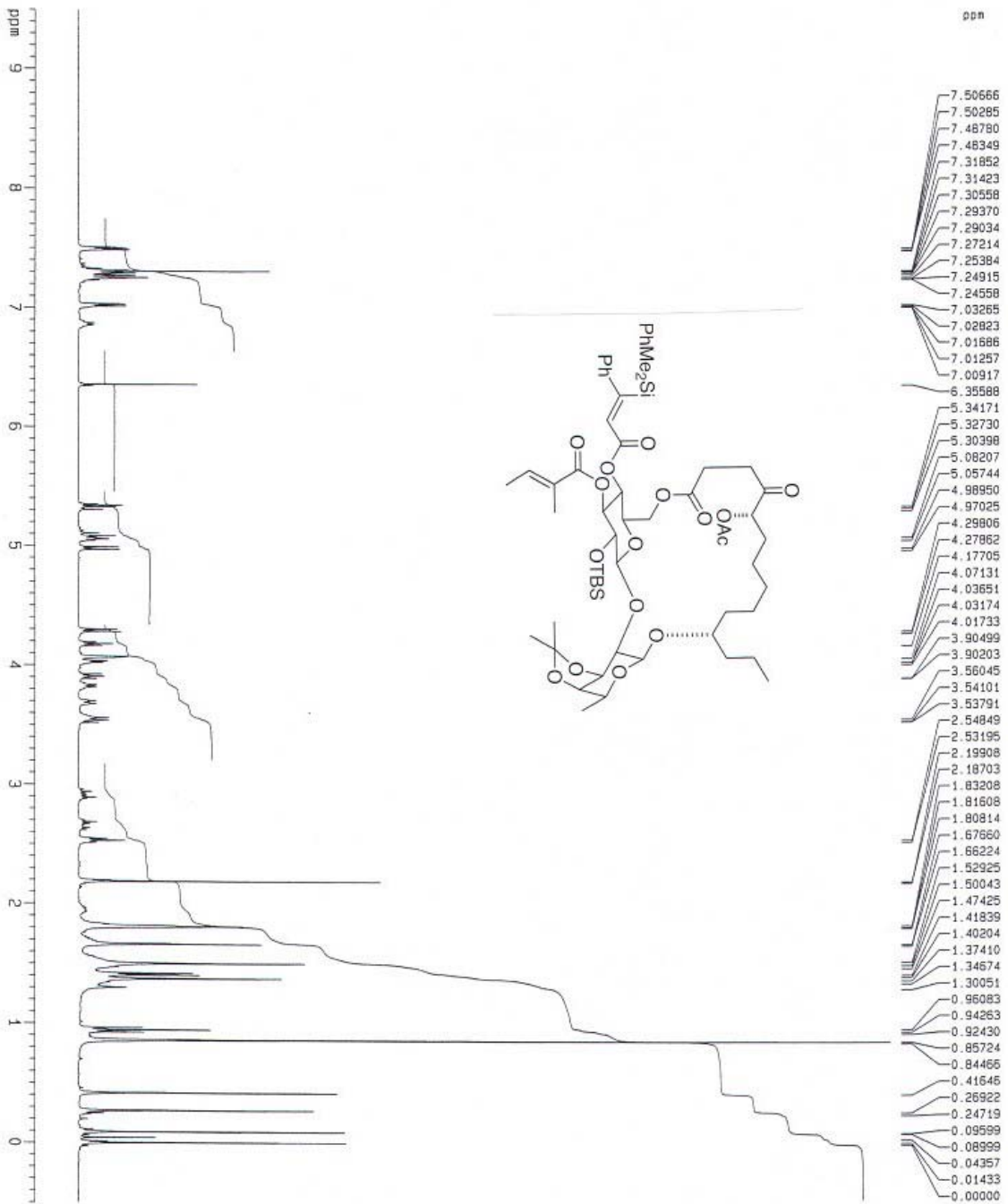
F2 - Processing parameters

| | |
|-----|-----------------|
| SI | 32758 |
| SF | 100.6126571 MHz |
| MVM | EM |
| SSB | 0 |
| LB | 1.00 Hz |
| GB | 0 |
| PC | 2.00 |

10 NMR plot parameters

| | |
|-------|------------------|
| CX | 22.00 cm |
| CV | 15.00 cm |
| FIP | 250.000 ppm |
| F1 | 25153.17 Hz |
| F2P | -30.000 ppm |
| F2 | -3018.38 Hz |
| PRMCH | 12.72727 ppm/cm |
| HZCM | 1280.52466 Hz/cm |

NA6-NB-038-02



ppm

| |
|---------|
| 7.50666 |
| 7.50285 |
| 7.48780 |
| 7.48349 |
| 7.31852 |
| 7.31423 |
| 7.30558 |
| 7.29370 |
| 7.29034 |
| 7.27214 |
| 7.25384 |
| 7.24915 |
| 7.24558 |
| 7.03265 |
| 7.02823 |
| 7.01686 |
| 7.01257 |
| 7.00917 |
| 6.35588 |
| 5.34171 |
| 5.32730 |
| 5.30398 |
| 5.08207 |
| 5.05744 |
| 4.98950 |
| 4.97025 |
| 4.29806 |
| 4.27862 |
| 4.17705 |
| 4.07131 |
| 4.03651 |
| 4.03174 |
| 4.01733 |
| 3.90499 |
| 3.90203 |
| 3.56045 |
| 3.54101 |
| 3.53791 |
| 2.54849 |
| 2.53195 |
| 2.19908 |
| 2.18703 |
| 1.83208 |
| 1.81608 |
| 1.80814 |
| 1.67660 |
| 1.66224 |
| 1.52925 |
| 1.50043 |
| 1.47425 |
| 1.41839 |
| 1.40204 |
| 1.37410 |
| 1.34674 |
| 1.30051 |
| 0.96083 |
| 0.94263 |
| 0.92430 |
| 0.85724 |
| 0.84665 |
| 0.41646 |
| 0.26922 |
| 0.24719 |
| 0.09599 |
| 0.08999 |
| 0.04357 |
| 0.01433 |
| 0.00000 |

Current Data Parameters
 NAME Jn28097
 EXPNO 10
 PROCNO 1

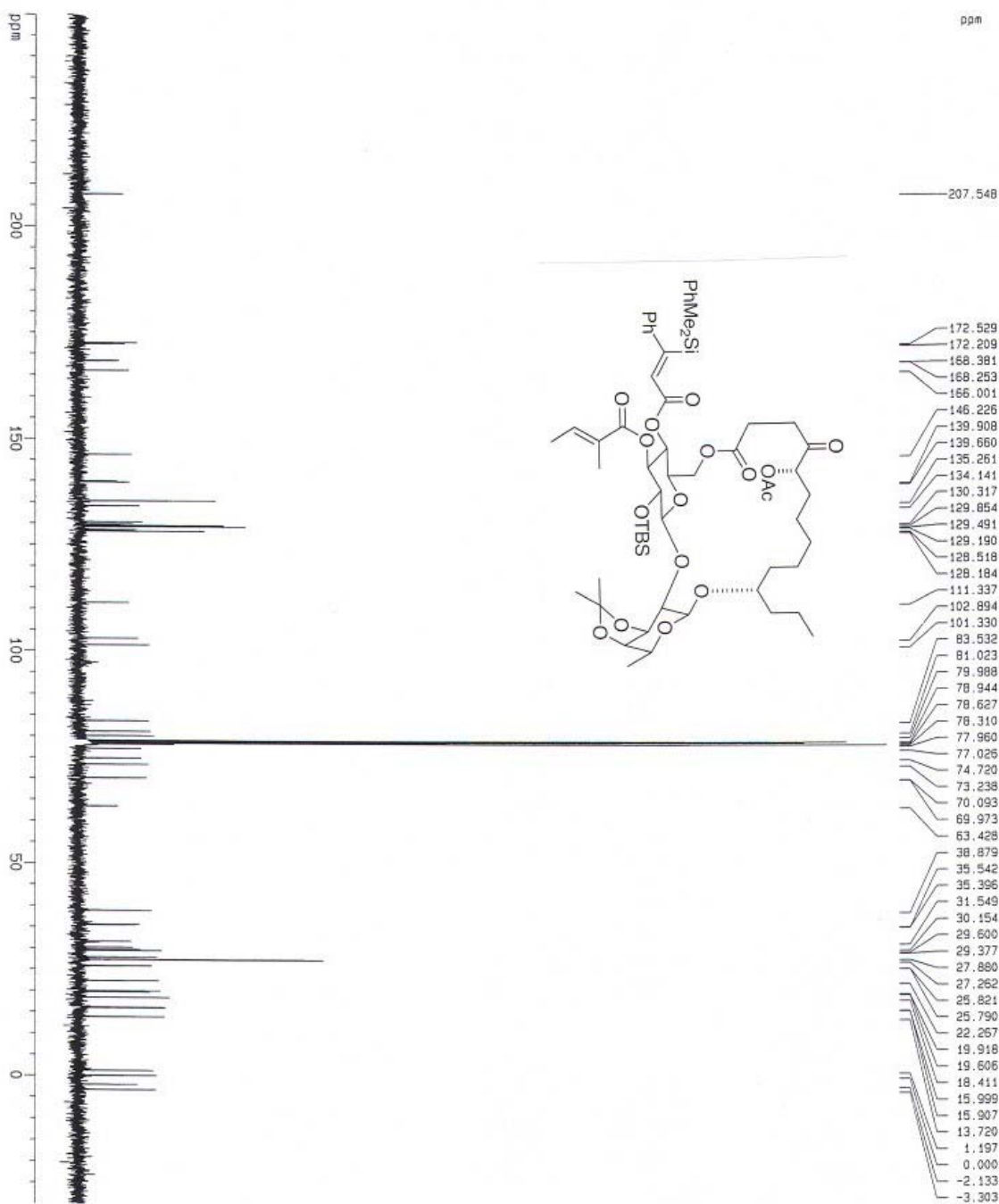
F2 - Acquisition Parameters
 Date_ 20060628
 Time 0.07
 INSTRUM av400
 PROBRD 5 mm BBO BB-5H
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 8
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9984243 sec
 RG 114
 DW 60.400 usec
 DE 6.50 usec
 TE 303.0 K
 D1 0.00300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.25 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1299915 MHz
 KW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

10 NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 FIP 9.500 ppm
 F1 3801.24 Hz
 F2 -200.07 Hz
 FREQH 0.45453 ppm/cm
 FREQV 181.8727 Hz/cm

NAG-NB-015-02



- 172.529
- 172.209
- 168.381
- 168.253
- 166.001
- 146.226
- 139.908
- 139.660
- 135.261
- 134.141
- 130.317
- 129.854
- 129.491
- 129.190
- 128.518
- 128.184
- 111.337
- 102.894
- 101.330
- 83.532
- 81.023
- 79.988
- 78.944
- 78.627
- 78.310
- 77.960
- 77.026
- 74.720
- 73.238
- 70.093
- 69.973
- 63.428
- 38.879
- 35.542
- 35.386
- 31.549
- 30.154
- 29.600
- 29.377
- 27.880
- 27.262
- 25.821
- 25.790
- 22.267
- 19.918
- 19.606
- 18.411
- 15.999
- 15.907
- 13.720
- 1.197
- 0.000
- 2.133
- 3.303

Current Data Parameters
 NAME Jn2697
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060528
 Time 0.50
 INSTRUM av400
 PROBRD 5 mm BBO BB-H1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1500
 DS 2
 SMH 33112.582 Hz
 FIDRES 0.506239 Hz
 AQ 0.3895438 sec
 RG 16384
 DM 15.100 usec
 DE 8.00 usec
 TE 303.0 K
 D1 0.03000000 sec
 d11 0.03000000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 10.94 usec
 PL1 5.00 dB
 SFO1 100.626789 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 90.00 usec
 PL2 0.00 dB
 PL12 19.76 dB
 SFO2 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6126055 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

10 MHz plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 250.000 ppm
 F1 28153.15 Hz
 F2P -30.000 ppm
 F2 -3018.38 Hz
 PPHCVM 12.72727 ppm/cm
 HZCM 1280.52405 Hz/cm

NAG-NB-015-02