



## Abstract

We describe the first high precision real-time analysis of the N<sub>2</sub>O site-specific isotopic composition at ambient mixing ratios. Our technique is based on mid-infrared quantum cascade laser absorption spectroscopy (QCLAS) combined with an automated preconcentration unit. The QCLAS allows for simultaneous and specific analysis of the three main stable N<sub>2</sub>O isotopic species, <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O, <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O, <sup>14</sup>N<sup>14</sup>N<sup>16</sup>O, and the respective site-specific relative isotope ratio differences  $\delta^{15}\text{N}^\alpha$  and  $\delta^{15}\text{N}^\beta$ . Continuous, stand-alone operation is achieved by using liquid nitrogen free N<sub>2</sub>O preconcentration, a quasi-room-temperature quantum cascade laser (QCL), quantitative sample transfer to the QCLAS, and an optimized calibration algorithm. The N<sub>2</sub>O site-specific isotopic composition ( $\delta^{15}\text{N}^\alpha$  and  $\delta^{15}\text{N}^\beta$ ) can be analysed with a long term precision of 0.2‰. The potential of this analytical tool is illustrated by continuous N<sub>2</sub>O isotopomer measurements above a grassland plot over three weeks period, which allowed identification of microbial source and sink processes.

## 1 Introduction

Nitrous oxide (N<sub>2</sub>O) is the most important anthropogenically emitted ozone depleting substance and also a significant greenhouse gas (Ravishankara et al., 2009). N<sub>2</sub>O mixing ratios in the troposphere increased from 270 ppb to the current level of 321.6 ppb (AGAGE 2008) at 0.8 ppb yr<sup>-1</sup> (2005 to 2008) with more than one third of N<sub>2</sub>O emissions being anthropogenic (Montzka et al., 2011; Solomon et al., 2007). For a better understanding of source and sink processes, however, the information obtained from measuring the intramolecular distribution of <sup>15</sup>N on the central ( $\alpha$ ) and the end ( $\beta$ ) position of the linear N<sub>2</sub>O molecule is crucial (Yoshida and Toyoda, 2000).

Abundances of the different isotopic species (<sup>14</sup>N<sup>14</sup>N<sup>16</sup>O, <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O, <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O, etc.) are usually reported in the  $\delta$ -notation, where  $\delta^{15}\text{N}$  denotes the relative difference in the amount of <sup>15</sup>N versus <sup>14</sup>N (abbreviated herein as <sup>15</sup>N/<sup>14</sup>N) in N<sub>2</sub>O in comparison

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5, 813–838, 2012

### Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



to atmospheric N<sub>2</sub> as the reference material (Coplen, 2011). Similarly,  $\delta^{15}\text{N}^\beta$  denotes the relative difference of isotope ratios for  $^{15}\text{N}^{14}\text{N}^{16}\text{O}$  vs.  $^{14}\text{N}^{14}\text{N}^{16}\text{O}$ .

The bulk nitrogen  $\delta$  value ( $\delta^{15}\text{N}^{\text{bulk}} = (\delta^{15}\text{N}^\alpha + \delta^{15}\text{N}^\beta)/2$ ) of tropospheric N<sub>2</sub>O is enriched by  $6.3 \pm 0.3\%$  to  $6.72 \pm 0.12\%$ , depending on the sampling location and time (Kaiser et al., 2003; Park et al., 2004; Röckmann and Levin, 2005; Toyoda et al., 2004), with a strong site preference (SP =  $\delta^{15}\text{N}^\alpha - \delta^{15}\text{N}^\beta$ ) of  $18.7 \pm 2.2\%$  for the central nitrogen atom (Yoshida and Toyoda, 2000). Temporal trends in the N<sub>2</sub>O isotopic composition from firn air, ice core and archived air sample measurements indicate a year to year decrease in  $\delta^{15}\text{N}^{\text{bulk}}$  of  $0.04\%$  yr<sup>-1</sup>, confirming substantial emissions of isotopically depleted N<sub>2</sub>O (Bernard et al., 2006; Ishijima et al., 2007; Röckmann and Levin, 2005). According to isotopic budgetary calculations based on a simple two-box model, this could be due to increased anthropogenic N<sub>2</sub>O emission from agricultural soils, as well as a to a change in their average isotopic signature (Ishijima et al., 2007).

On a local scale, the N<sub>2</sub>O isotopic composition can be applied to disentangle or even quantitatively apportion N<sub>2</sub>O production and destruction pathways. For example, the  $^{15}\text{N}$  depletion in N<sub>2</sub>O produced by autotrophic nitrification was found to be considerably higher as compared to heterotrophic denitrification (Koba et al., 2009; Sutka et al., 2006; Toyoda et al., 2005; Yoshida, 1988). On the other hand, process-specific effects on  $\delta^{15}\text{N}^{\text{bulk}}$  might be masked by shifts in the precursor signature (Well et al., 2008), and theoretical considerations indicate a major impact of the involved bacterial species (Schmidt et al., 2004). In contrast to  $\delta^{15}\text{N}^{\text{bulk}}$ , the site preference is considered to be independent of the isotopic composition of the precursor and thus supplies clear process information even if the isotopic signature of the substrate for N<sub>2</sub>O production is lacking (Frame and Casciotti, 2010; Ostrom et al., 2007; Sutka et al., 2006; Toyoda et al., 2002; Well and Flessa, 2009; Yamagishi et al., 2007).

The standard technique for N<sub>2</sub>O isotopic measurements is laboratory-based isotope-ratio mass-spectrometry (IRMS) in combination with flask-sampling (Brenninkmeijer and Röckmann, 1999; Toyoda and Yoshida, 1999). It is a well-known method with excellent precision of up to  $0.05\%$  for  $\delta^{15}\text{N}^{\text{bulk}}$ ,  $0.1\%$  for  $\delta^{18}\text{O}$ , and  $0.3\%$  for  $\delta^{15}\text{N}^\alpha$

**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



and  $\delta^{15}\text{N}^\beta$  (Bernard et al., 2006; Toyoda et al., 2011a,b). Nevertheless, it also has some disadvantages such as the large size of the instrument, which hinders in-situ field measurements. Laser spectroscopy is a valuable alternative because it is inherently selective, even for molecules with the same mass (Janssen and Tuzson, 2006; Gagliardi et al., 2005; Nakayama et al., 2007; Uehara et al., 2001, 2003; Wächter and Sigrist, 2007), and field-deployable instruments for unattended measurements can be designed. A significant improvement was obtained in recent years by the implementation of quantum cascade laser sources (QCL) to reach a precision of 0.5‰ for  $\delta^{15}\text{N}^\alpha$  and  $\delta^{15}\text{N}^\beta$  at N<sub>2</sub>O mixing ratios of 90 ppm (Wächter et al., 2008).

In the present project we describe the first instrumentation to perform real-time analysis of N<sub>2</sub>O site-specific isotopic composition at atmospheric mixing ratios. This is achieved combining a liquid nitrogen-free fully-automated preconcentration unit optimized and validated for N<sub>2</sub>O isotopomer analysis by Mohn et al. (2010) with an improved version of the QCLAS published by Wächter et al. (2008). The potential of this approach is demonstrated by a three week measurement campaign of atmospheric N<sub>2</sub>O to identify distinct soil microbial N<sub>2</sub>O source and sink processes.

## 2 Materials and methods

A schematic diagram of the measurement setup is shown in Fig. 1. Details on the development, optimization and validation of the N<sub>2</sub>O preconcentration unit and the QC laser spectrometer have been described previously (Mohn et al., 2010; Wächter et al., 2008). Thus, their basic principles and recent modifications are only briefly presented, while analytical improvements, procedures of air sampling, automation and the applied calibration procedure are discussed in more detail.

## 2.1 Sampling site and setup

Field experiments were conducted in Dübendorf at 430 m a.s.l. (47°24'10" N/8°36'43" E). The observation area is located in an industrial and densely populated region near Zurich. A main road passes 100 m south, and a highway around 750 m north of the sampling site. Air was continuously sampled above a grassland plot (5 m × 20 m) at a flow rate of about one standard litre per minute (slpm) through a 15 m long unheated PTFE tubing (ID 4 mm) using a diaphragm vacuum pump (KNF Neuberger, CH). The air intake was first mounted at 1.5 m above ground (8 to 24 September), and then (24 to 31 September) 10 cm above the surface to be more representative for soil N<sub>2</sub>O production. At the pump outlet, the pressure was adjusted to 4 bar by means of a pressure relief valve. Water and CO<sub>2</sub> were quantitatively removed by permeation drying (PD-100T-48, PermaPure Inc., USA) and by chemical trapping with Ascarite (30 g, 10–35 mesh, Fluka, Switzerland) bracketed by Mg(ClO<sub>4</sub>)<sub>2</sub> (2 × 13 g, Fluka, Switzerland). Finally, the sample was passed through a sintered metal filter (SS-6F-MM-2, Swagelok, USA) and directed to the preconcentration unit. An alternative sample input consisted of pressurized air (Messer, Switzerland) employed as target gas (Klausen et al. 2010) which was treated as described above by a second permeation dryer and a chemical trap (20 g Ascarite, 2 × 8 g Mg(ClO<sub>4</sub>)<sub>2</sub>). This setup allows determining the long term stability and precision of the complete analytical procedure, including preconcentration, laser spectroscopic analysis and calibration. The chemical traps were exchanged every 3 to 4 days before reaching their maximal load. To detect any potential breakthrough, the CO<sub>2</sub> concentration was monitored by QCLAS after preconcentration together with the N<sub>2</sub>O isotopomers (CO<sub>2</sub> line at 2188.0 cm<sup>-1</sup>).

**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## 2.2 Instrumentation

### 2.2.1 N<sub>2</sub>O preconcentration

The technology of our preconcentration unit is based on a previously developed system called “Medusa” (Miller et al., 2008), re-designed and optimized for the preconcentration of N<sub>2</sub>O isotopic species and their subsequent quantification by laser spectroscopy (Mohn et al., 2010). During standard operation, 10 l of ambient air are preconcentrated on a HayeSep D trap and desorbed in approximately 50 ml of synthetic air, yielding a concentration increase from ambient mixing ratios to >71 ppm N<sub>2</sub>O. The system offers quantitative (>99%) N<sub>2</sub>O recovery without any significant isotopic fractionation or relevant spectral interferences from other atmospheric constituents. Modifications to the previous procedure (Mohn et al., 2010) are mainly related to the desorption phase, where the N<sub>2</sub>O concentration profile was further optimized by increasing the trap temperature to 10° C and decreasing the flow rate of high purity synthetic air to 10 sccm (standard cubic centimetre per minute).

### 2.2.2 Laser spectrometer

The employed QCLAS is based on the instrument described by Wächter et al. (2008). It consists of a single-mode, pulsed QCL emitting at 2188 cm<sup>-1</sup>, a multipass absorption cell (optical path length 56 m, volume 0.5 l; Aerodyne Research Inc., USA) and a detection system with pulse normalization. Laser control, data acquisition and simultaneous quantification of the three main N<sub>2</sub>O isotopic species (<sup>14</sup>N<sup>14</sup>N<sup>16</sup>O, <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O, <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O) is accomplished by the TDLWintel software (Aerodyne Research Inc., USA), taking into account path length, gas temperature (~ 305 K), pressure (8 kPa) and laser line width (0.0068 cm<sup>-1</sup>). Employing a new generation thermo-electrically cooled detector (PVI-3TE-5, Vigo System, PL), a new quasi-room temperature QCL (Alpes Lasers SA, Switzerland) and redesigned electronics led to a considerably improved performance of the laser spectrometer, compared to the results of Wächter et al. (2008).

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion





samples (A)–(D), standard I (71 ppm N<sub>2</sub>O) was analysed as a reference point and to correct for drift effects.

## 2.4 Analysis of N<sub>2</sub>O mixing ratios and isotopomer ratios

The N<sub>2</sub>O mixing ratios of ambient air were determined based on the concentration of the main isotopic species <sup>14</sup>N<sup>14</sup>N<sup>16</sup>O and calibrated against laboratory standard I dynamically diluted to different concentration levels (see Fig. 2 step D). The preconcentration step was taken into account via the ratio of the gas volume in the multipass cell ( $V_{\text{cell}}$ ) and the gas volume applied for N<sub>2</sub>O preconcentration ( $V_{\text{precon}}$ ). While  $V_{\text{precon}}$  can be accurately computed based on the adsorption time and flow, for  $V_{\text{cell}}$  this is not possible. Therefore, the exact value for  $V_{\text{cell}}$  under standard conditions was determined analysing preconcentrated N<sub>2</sub>O from a highly accurate standard (319.91 ± 0.12 ppb) provided by the World Meteorological Organization (WMO) Central Calibration Laboratory (CCL) (Hall et al., 2007). N<sub>2</sub>O concentrations of the laboratory standards were quantified by QCLAS against commercial calibration gases (90.5 ± 0.1 ppm N<sub>2</sub>O, Messer, Switzerland) and are indicated in Table 1.

Relative differences of isotopic ratios  $\delta^{15}\text{N}^{\alpha}$  and  $\delta^{15}\text{N}^{\beta}$  were determined employing a set of standard gases produced in our laboratory based on gravimetric and dynamic dilution methods from pure medical N<sub>2</sub>O (Messer, Switzerland) supplemented with distinct amounts of isotopically pure (>98%) <sup>15</sup>N<sup>14</sup>N<sup>16</sup>O and <sup>14</sup>N<sup>15</sup>N<sup>16</sup>O (Cambridge Isotope Laboratories, USA). Primary laboratory standards were analysed for  $\delta^{15}\text{N}^{\alpha}$ ,  $\delta^{15}\text{N}^{\beta}$  and  $\delta^{15}\text{N}^{\text{bulk}}$  by IRMS at the Tokyo Institute of Technology (Toyoda and Yoshida, 1999). Table 1 indicates the isotopic composition of the secondary laboratory standards applied in the current project and analysed against primary standards by QCLAS. The  $\delta^{15}\text{N}^{\text{bulk}}$  of pure medical N<sub>2</sub>O was additionally analysed by mass spectrometry at the IsoLab of the Max-Planck Institute for Biogeochemistry (MPI-BGC, Jena, Germany) using an EA/IRMS setup (Werner et al., 1999). N<sub>2</sub>O was introduced in between the combustion and the reduction tube of the EA using the loop (250 µl) of a manually

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



operated 6-port valve. This setup enabled a direct comparison of N<sub>2</sub> produced from combustion of IAEA-N1 to N<sub>2</sub> obtained from the medical N<sub>2</sub>O by reduction in the 2nd EA reactor. Quantitative N<sub>2</sub>O conversion reaction yield was verified by the absence of any *m/z* 44 ion current response following N<sub>2</sub>O introduction. Using a  $\delta^{15}\text{N}$  value of +0.43‰ for IAEA-N1 as the scale anchor, a  $\delta^{15}\text{N}^{\text{bulk}}$  value of  $1.64 \pm 0.10$ ‰ ( $n = 4$ ) was obtained which was different by 0.39‰ from the Tokyo Tech result. A similar difference of 0.3‰ was observed by Toyoda and Yoshida (1999) for  $\delta^{15}\text{N}^{\text{bulk}}$  of a laboratory standard calculated from  $\delta^{15}\text{N}^{\alpha}$  and  $\delta^{15}\text{N}^{\beta}$  (calibration via NH<sub>4</sub>NO<sub>3</sub> decomposition) and determined after N<sub>2</sub>O to N<sub>2</sub> reduction. Discrepancies were attributed to fractionation during incomplete NH<sub>4</sub>NO<sub>3</sub> decomposition (Toyoda and Yoshida, 1999).

Relative differences of site - selective isotope ratios  $\delta^{15}\text{N}^{\alpha}$  and  $\delta^{15}\text{N}^{\beta}$  of pre-concentrated N<sub>2</sub>O were corrected for dependency of the isotope ratios on the N<sub>2</sub>O mixing ratio (before preconcentration). These corrections were small, about 0.004‰ppb<sup>-1</sup> and 0.016‰ppb<sup>-1</sup> for  $\delta^{15}\text{N}^{\alpha}$  and  $\delta^{15}\text{N}^{\beta}$ , respectively. Moreover, measurements that were significantly influenced by abrupt changes in the laser intensity were discarded. These light intensity changes affected less than 2% of the data, and the laser driver that was identified as the source of instability was recently replaced.

To confirm the accuracy of our measurements, we analysed background air in a cylinder filled in 2006 by the Earth System Research Laboratory (Global Monitoring Division) of the National Oceanic & Atmospheric Administration (NOAA). The trace gas mixing ratios analysed by the WMO CCL are typical for natural air:  $384.40 \pm 0.02$  ppm CO<sub>2</sub>,  $319.91 \pm 0.12$  ppb N<sub>2</sub>O,  $1838.5 \pm 0.4$  ppb CH<sub>4</sub>,  $143.9 \pm 1.0$  ppb CO. The QCLAS analysis of the N<sub>2</sub>O site-selective isotopic composition, with  $\delta^{15}\text{N}^{\alpha} = 15.62 \pm 0.06$ ‰,  $\delta^{15}\text{N}^{\beta} = -2.84 \pm 0.04$ ‰,  $\delta^{15}\text{N}^{\text{bulk}} = 6.39 \pm 0.03$ ‰ and SP =  $18.45 \pm 0.08$ ‰ (the precision indicated is the standard error of the mean), is in perfect agreement with published data for unpolluted tropospheric N<sub>2</sub>O.



capabilities, the statistical uncertainty for repeated measurements (standard error of the mean) is considerably lower.

### 3.2 Source appointment by N<sub>2</sub>O isotopomer analysis

The isotopic signature of a source process can be estimated by the Keeling-plot approach where the variations in the isotopic composition are plotted against the inverse of concentration values. This technique was originally developed for carbon dioxide and its isotopologues and has been employed in numerous studies, recently also in combination with field-deployable instrumentation for continuous CO<sub>2</sub> isotopic analysis (McManus et al., 2010; Mohn et al., 2008; Tuzson et al., 2011). For N<sub>2</sub>O, up to date all process studies on N<sub>2</sub>O isotopic species had to rely on grab sampling followed by IRMS laboratory analysis because real-time analysis was not available with the required precision. Consequently, current research is based on short-term investigations with limited temporal and spatial averaging capabilities (Ostrom et al., 2010; Toyoda et al., 2011a; Yamagishi et al., 2007).

In Fig. 5, data obtained in a 24 h time interval (e.g. from 23 September noon to 24 September noon) was analysed using the Keeling plot approach. Individual data points represent the average N<sub>2</sub>O mixing ratio and isotopic composition over 20 min of N<sub>2</sub>O sampling during preconcentration. Assuming a two source mixing with unpolluted background air, the intercept of the linear regression line corresponds to the isotopic signature of the N<sub>2</sub>O emitting processes for  $\delta^{15}\text{N}_s^{\text{bulk}}$  and SP<sub>s</sub>. This approach implies that the soil microbial N<sub>2</sub>O production pathways and its isotopic signatures are basically constant for one diurnal cycle. As can be observed from the low scatter in the Keeling plots (i.e. Fig. 5), this model is adequate for the studied processes and gives moderate uncertainties in the linear regression parameters (Fig. 6).

Before fertilizer addition the diurnal variability in N<sub>2</sub>O mixing ratios was small, in the range of 3.0 to 10.5 ppb, accompanied by only a slight shift to lower relative isotope ratio differences at higher N<sub>2</sub>O mixing ratios. It was, nevertheless, possible to resolve these small changes and calculate daily (24 h time intervals, noon to noon) N<sub>2</sub>O source

**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures



Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

signatures ( $\delta^{15}\text{N}_s^{\text{bulk}}$ ,  $\text{SP}_s$ ). These varied between  $-5 \pm 7\text{‰}$  to  $-26 \pm 6\text{‰}$  for  $\delta^{15}\text{N}_s^{\text{bulk}}$  and  $3 \pm 13\text{‰}$  to  $29 \pm 5\text{‰}$  for  $\text{SP}_s$  with a temporal trend (9 to 22 September) from low to high  $\delta^{15}\text{N}_s^{\text{bulk}}$  and high to low  $\text{SP}_s$  values (Fig. 6). Periods with changes in N<sub>2</sub>O mixing ratios below 6.5 ppb (2%) were not considered. To estimate the net isotope effect ( $\Delta\delta^{15}\text{N}_s^{\text{bulk}} = \delta^{15}\text{N}(\text{substrate}) - \delta^{15}\text{N}_s^{\text{bulk}}$ ) of the microbial source process, the <sup>15</sup>N content of the substrate for N<sub>2</sub>O production needs to be known. As the focus of the present study was on method development for ambient air monitoring and not on soil science, no supplementary soil parameters were determined. However, a  $\delta^{15}\text{N}$  (substrate) of 5‰ may be assumed for the inorganic soil nitrogen pool (Makarov et al., 2010). This results in a net isotope effect ( $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$ ) between 10 and 31‰, which is characteristic for N<sub>2</sub>O produced by heterotrophic denitrification. In pure culture studies with denitrifying bacteria, Sutka et al. (2006) and Toyoda et al. (2005) reported up to 39‰ for  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  of N<sub>2</sub>O, and Koba et al. (2009) assigned the range 0 to 39‰ for N<sub>2</sub>O derived from heterotrophic denitrification. In contrast, N<sub>2</sub>O produced by nitrifying bacteria leads to a significantly higher <sup>15</sup>N depletion with a net isotope effect ( $\Delta\delta^{15}\text{N}_s^{\text{bulk}} = \delta^{15}\text{N}(\text{NH}_4^+) - \delta^{15}\text{N}_s^{\text{bulk}}$ ) between 46.9‰ (Sutka et al., 2006) and 68‰ (Yoshida, 1988).

The observed co-variation of  $\text{SP}_s$  and  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  (Fig. 6) for N<sub>2</sub>O emitted before fertilizer application with a slope of  $1.18 \pm 0.32$  can be attributed to a partial consumption by N<sub>2</sub>O reductase activity of denitrifying bacteria (Koba et al., 2009; Yamagishi et al., 2007). Similar values between 1.0 and 1.2 for  $\text{SP}_s / \Delta\delta^{15}\text{N}_s^{\text{bulk}}$  were reported by Ostrom et al. (2007) for N<sub>2</sub>O reduction by two denitrifier species. The temporal trend from high to low  $\text{SP}_s$  and low to high  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  values (Fig. 6) can thus be interpreted as a decreasing share of N<sub>2</sub>O reduction vs. N<sub>2</sub>O production. The N<sub>2</sub>O vs. (N<sub>2</sub>O + N<sub>2</sub>) ratio increase correlates with a night-time air temperature decrease from 13°C to 6°C at the nearby NABEL station (data not shown), which is consistent with the temperature dependence observed in laboratory studies on soil samples (Avalakki et al., 1995; Bailey and Beauchamp, 1973).

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



After fertilizer application the isotopic source signatures are better constrained because of larger diurnal changes in N<sub>2</sub>O mixing ratios (Fig. 6). Furthermore, much less variation was observed in the isotopic source signatures with values between  $-0.1 \pm 0.8\text{‰}$  to  $-5.1 \pm 1.3\text{‰}$  for SP<sub>s</sub> and  $41.8 \pm 0.9\text{‰}$  to  $23.2 \pm 0.7\text{‰}$  for  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$ . To calculate the net isotope effect ( $\Delta\delta^{15}\text{N}_s^{\text{bulk}} = (\delta^{15}\text{N}(\text{substrate}) - \delta^{15}\text{N}_s^{\text{bulk}})$ ), the <sup>15</sup>N content of the fertilizer N analysed by IRMS ( $\delta^{15}\text{N}(\text{NH}_4\text{NO}_3) = 1.3 \pm 0.3\text{‰}$ ) was applied. The resulting SP<sub>s</sub> and  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  values are indicative for N<sub>2</sub>O production by heterotrophic denitrification without or with only minor N<sub>2</sub>O to N<sub>2</sub> reduction (Sutka et al., 2006; Toyoda et al., 2005). The observed low N<sub>2</sub>O consumption agrees with a recent publication assuming that the N<sub>2</sub>O/(N<sub>2</sub>O + N<sub>2</sub>) product ratio of denitrification is positively correlated with the NO<sub>3</sub><sup>-</sup> availability in soils (Senbayram et al., 2011). The successive increase in  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  suggests a shift in the isotope composition of the soil nitrate pool due to fractionation during denitrification.

## 4 Conclusions

This study presents to our knowledge the first real-time analysis of N<sub>2</sub>O site-selective isotopic composition at atmospheric mixing ratios. Our approach is based on a cryogenic free instrumentation which comprises a mid-IR QCL absorption spectrometer and a fully automated N<sub>2</sub>O preconcentration unit. During three weeks of continuous field measurements nearly 550 air samples were analysed for N<sub>2</sub>O mixing ratios and site-specific isotopic composition. Long term precision for  $\delta^{15}\text{N}^\alpha$  and  $\delta^{15}\text{N}^\beta$  was found to be superior to state-of-the-art IRMS. Additionally, precision for N<sub>2</sub>O mixing ratios determined by QCLAS was comparable to the standard technique applied in global monitoring networks (GC-ECD).

The excellent analytical precision allowed resolving even small changes in N<sub>2</sub>O mixing ratios and isotope composition,  $\delta^{15}\text{N}_s^{\text{bulk}}$  and SP and calculating daily (24 h time intervals, noon to noon) source signatures ( $\delta^{15}\text{N}_s^{\text{bulk}}$ , SP<sub>s</sub>) for N<sub>2</sub>O emitted from



**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

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**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion



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## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

Full Screen / Esc

Printer-friendly Version

Interactive Discussion

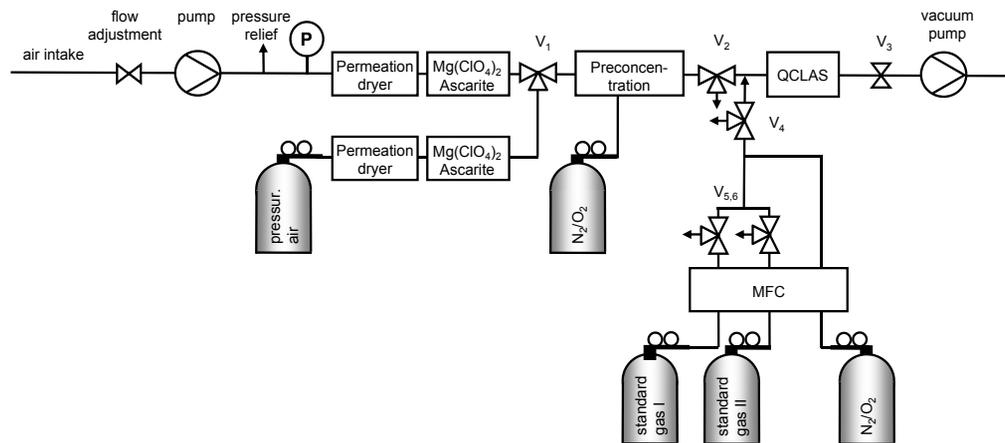


**Table 1.** N<sub>2</sub>O mixing ratios and relative differences of isotopic ratios  $\delta^{15}\text{N}^\alpha$  and  $\delta^{15}\text{N}^\beta$  of secondary laboratory standards applied in the current project (the precision indicated is the standard error of the mean). Standard Ia was replaced by standard Ib 22 September.

	N <sub>2</sub> O [ppm]	$\delta^{15}\text{N}^\alpha$ [‰]	$\delta^{15}\text{N}^\beta$ [‰]
Standard Ia	246.9±0.1	2.1±0.1	2.0±0.2
Standard Ib	250.1±0.05	15.2±0.1	2.0±0.1
Standard II	249.1±0.1	25.0±0.1	24.8±0.2

**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.



**Fig. 1.** Experimental setup for on-line N<sub>2</sub>O isotopomer analysis in ambient air. V<sub>i</sub> are solenoid valves and MFC mass flow controllers.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

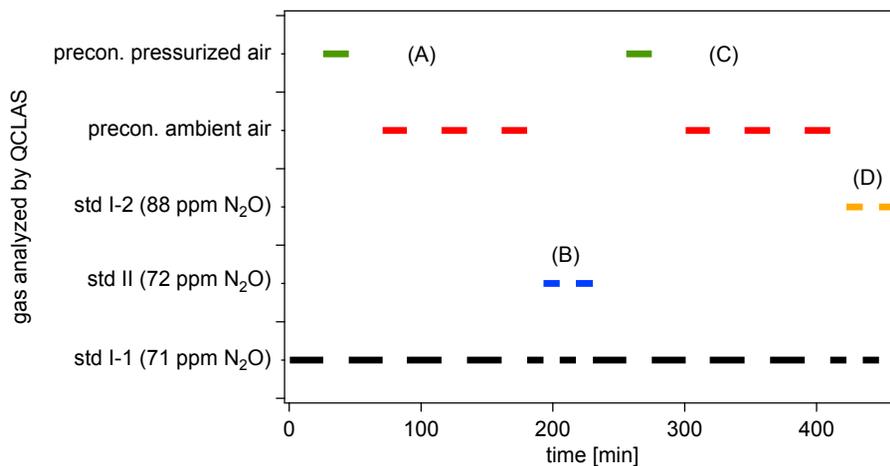
Full Screen / Esc

Printer-friendly Version

Interactive Discussion

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.



**Fig. 2.** Measurement cycle: **(A) + (C)** analysis of ambient air or pressurized air (target gas), **(B)** determination of calibration factors for  $\delta^{15}\text{N}^{\alpha}$ ,  $\delta^{15}\text{N}^{\beta}$ , and **(D)** the N<sub>2</sub>O mixing ratio as well as its influence on  $\delta$  values.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

Full Screen / Esc

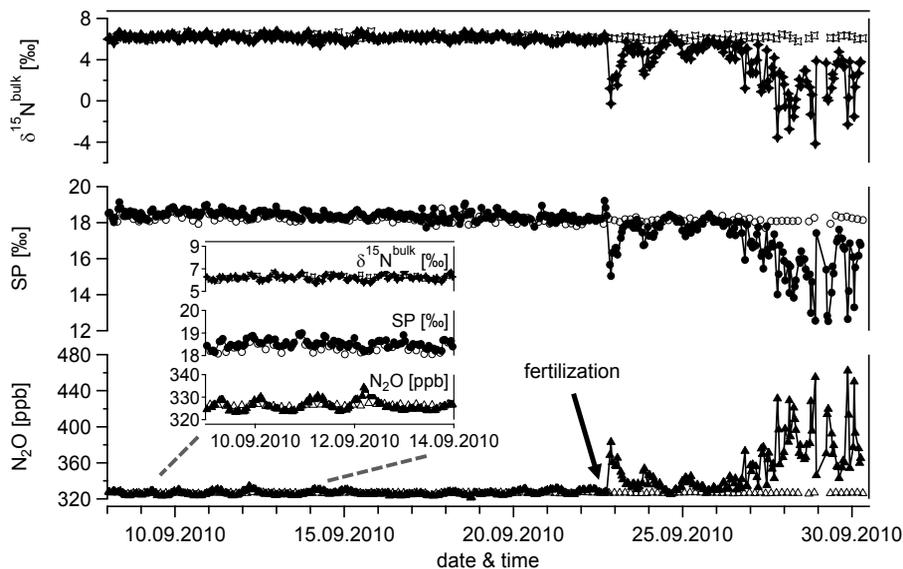
Printer-friendly Version

Interactive Discussion



## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.



**Fig. 3.** Time series of N<sub>2</sub>O mixing ratio and the corresponding  $\delta^{15}\text{N}^{\text{bulk}}$  and SP values of ambient air (closed symbols) and pressurized air (target gas, open symbols) analysed by QCLAS after preconcentration. Relative isotope ratio differences are based on the Tokyo Tech calibration of the primary laboratory standards. Strong emissions of  $^{15}\text{N}$  depleted N<sub>2</sub>O were observed after fertilizer addition (200 kg N ha<sup>-1</sup> NH<sub>4</sub>NO<sub>3</sub>, 400 kg C ha<sup>-1</sup> sucrose) indicated by an arrow.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

◀

▶

◀

▶

Back

Close

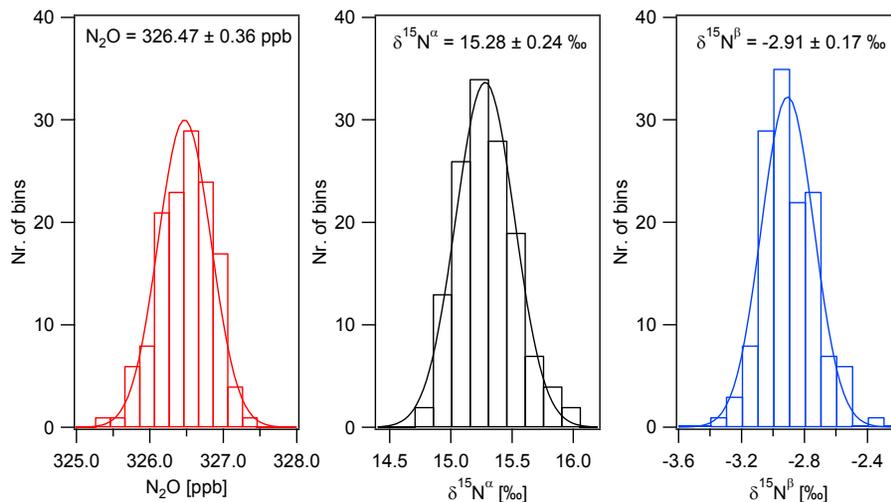
Full Screen / Esc

Printer-friendly Version

Interactive Discussion

**Site selective  
real-time  
measurements of  
atmospheric N<sub>2</sub>O**

J. Mohn et al.

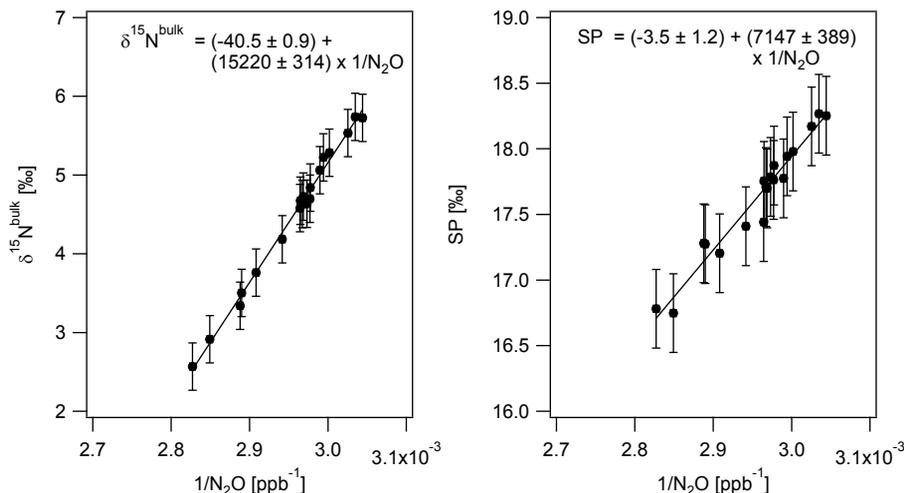


**Fig. 4.** Repeated measurements of pressurized air (target gas) during the field experiment. N<sub>2</sub>O mixing ratios and relative differences of isotope ratios were plotted as a histogram with bin widths of 0.25 ppb (N<sub>2</sub>O), 0.15 ‰ (δ<sup>15</sup>N<sup>α</sup>) and 0.1 ‰ (δ<sup>15</sup>N<sup>β</sup>), respectively (the precision indicated is the standard deviation).

[Title Page](#)[Abstract](#)[Introduction](#)[Conclusions](#)[References](#)[Tables](#)[Figures](#)[⏪](#)[⏩](#)[◀](#)[▶](#)[Back](#)[Close](#)[Full Screen / Esc](#)[Printer-friendly Version](#)[Interactive Discussion](#)

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.



**Fig. 5.** Exemplary 24 h Keeling plot (23 September noon–24 September 2010 noon) after fertilizer addition. Site preference and  $\delta^{15}\text{N}^{\text{bulk}}$  are plotted vs. the inverse of the N<sub>2</sub>O concentration. The intercept of the ordinary least square linear regression corresponds to the isotopic signature of the main N<sub>2</sub>O emitting process ( $\delta^{15}\text{N}_s^{\text{bulk}}$ ,  $\text{SP}_s$ ) and is given together with its 1 $\sigma$  uncertainty.

Title Page

Abstract

Introduction

Conclusions

References

Tables

Figures

⏪

⏩

◀

▶

Back

Close

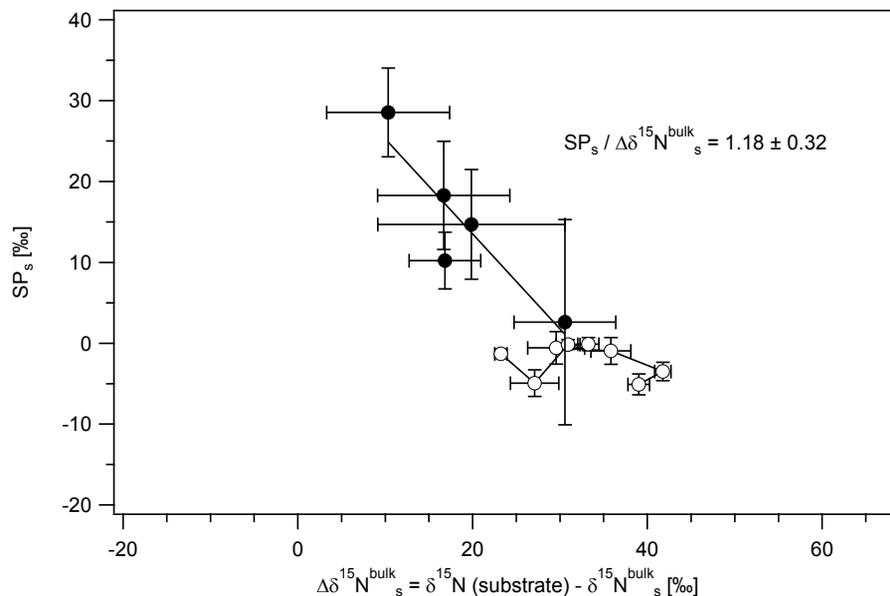
Full Screen / Esc

Printer-friendly Version

Interactive Discussion

## Site selective real-time measurements of atmospheric N<sub>2</sub>O

J. Mohn et al.



**Fig. 6.**  $\text{SP}_s$  vs.  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  plot to interpret the biogeochemistry of soil emitted N<sub>2</sub>O. Isotopic source signatures indicate heterotrophic denitrification as the main N<sub>2</sub>O production process, with  $\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  values between 10 and 42‰. Before fertilizer application (closed symbols, 9 to 22 September, periods with N<sub>2</sub>O concentration changes >6.5 ppb) a  $\text{SP}_s/\Delta\delta^{15}\text{N}_s^{\text{bulk}}$  ratio of  $1.18 \pm 0.32$  indicates N<sub>2</sub>O reductase activity which ceased after fertilizer addition (open symbols, 22 to 30 September) identified by low  $\text{SP}_s$  values between 0 and –5‰. For  $\delta^{15}\text{N}$  (substrate) 5‰ were assumed before fertilizer addition (before 22 September) according to Makarov et al. (2010), afterwards the <sup>15</sup>N content of the applied NH<sub>4</sub>NO<sub>3</sub> fertilizer ( $1.3 \pm 0.3\%$ ) was used.