

Biogeosciences Discussions is the access reviewed discussion forum of *Biogeosciences*

Does the temperature sensitivity of decomposition vary with soil organic matter quality?

M. Reichstein^{1,2}, T. Kätterer³, O. Andr n³, P. Ciais⁴, E.-D. Schulze⁵, W. Cramer², D. Papale¹, and R. Valentini¹

¹Laboratory of Forest Ecology, Universit  della Tuscia, I-01100 Viterbo, Italy

²Potsdam Inst. for Climate Impact Research, Telegrafenberg C4, D-14473 Potsdam, Germany

³Soil Fertility and Plant Nutrition, SLU, Department of Soil Sciences, P.O.Box 7014, 75007 Uppsala, Sweden

⁴Lab. des Sciences du Climat et de l'Environnement, LSCE, 91191, Gif sur Yvette, France

⁵Max-Planck Institut f r Biogeochemie, PO Box 100164, 07701 Jena, Germany

Received: 3 June 2005 – Accepted: 21 June 2005 – Published: 1 July 2005

Correspondence to: M. Reichstein (reichstein@unitus.it)

  2005 Author(s). This work is licensed under a Creative Commons License.

737

Abstract

Knorr et al. (2005) concluded that soil organic carbon pools with longer turnover times are more sensitive to temperature. We show that this conclusion is equivocal, largely dependent on their specific selection of data and does not persist when the data set of K tterer et al. (1998) is analysed in a more appropriate way. Further, we analyse how statistical properties of the model parameters may interfere with correlative analyses that relate the Q_{10} of soil respiration with the basal rate, where the latter is taken as a proxy for soil organic matter quality. We demonstrate that negative parameter correlations between Q_{10} -values and base respiration rates are statistically expected and not necessarily provide evidence for a higher temperature sensitivity of low quality soil organic matter. Consequently, we reckon it is premature to conclude that stable soil carbon is more sensitive to temperature than labile carbon.

1 Introduction

The temperature sensitivity of soil carbon decomposition is a key factor determining the response of the terrestrial carbon balance to climatic change as most recently shown in coupled global carbon climate-vegetation model studies (e.g., Jones et al., 2003). Consequently, temperature sensitivity of soil respiration and soil organic matter decomposition has received a lot of attention (e.g., Lloyd and Taylor, 1994; Davidson et al., 1998; K tterer et al., 1998; Luo et al., 2001; Reichstein et al., 2003; Sandermann et al., 2003). In particular the questions have arisen, whether the temperature sensitivity differs between labile and stable soil carbon pools (Liski et al., 1999;  gren, 2000; Davidson et al., 2000; Giardina and Ryan, 2000) and whether soil organisms acclimate to higher temperatures (e.g., Luo et al., 2001).

738

2 Analysis and discussion of results found by Knorr et al. (2005)

Recently, Knorr et al. (2005) nicely showed that neither acclimation to higher temperature, nor temperature insensitivity of stable soil organic carbon decomposition is necessary to explain recent results from soil warming and incubation experiments (Giardina and Ryan, 2000; Luo et al., 2001). We agree with Knorr et al. that these findings can be more convincingly explained by the intrinsic dynamics of soil carbon pools with substantially different decomposition rates as implemented in most global carbon cycle models (cf. also Reichstein et al., 2000; Kirschbaum, 2004). Hence, the hypothesis of increased soil organic matter decomposition inducing a positive carbon cycle feedback in response to global warming should not be rejected.

Knorr et al. (2005) also conclude from their study that the decomposition of stable soil organic matter is even more sensitive to increases in temperature than that of labile pools, thereby exerting an even stronger positive feedback to global warming than assumed by current carbon cycle models. However, we propose that this conclusion is largely contingent on the subset of the incubation review data from Kätterer et al. (1998) chosen for the analysis. A significant positive correlation between the activation energy (E_a) in the Arrhenius-equation (which determines the temperature sensitivity) and the log-transformed apparent turnover time of the sample appears when studies with incubation times of less than 100 days are excluded (Fig. 1, red crosses and red line). However, if all data from Kätterer et al. (1998) are retained, no significant correlation remains (Fig. 1, diamonds and black line). Short-term soil incubation studies are less reliable for parameter estimations when turnover is slow. Hence, in our view it is more appropriate to exclude incubation studies according to a criterion relating incubation time to apparent turnover time of the sample. Excluding all soil incubation studies that lasted less than 69% of the apparent turnover time, i.e. less than the apparent half-life, leads to no correlation at all (Fig. 1, blue squares and line).

Moreover, analysing the original data with a two-compartment decomposition model, where the rate constants of the labile and stable pool are allowed to vary independently

739

with temperature, yields a small reverse difference between the temperature sensitivity of the stable and labile pools, the activation energy being 47.3 ± 0.9 kJ/mol for the labile and 44.5 ± 0.6 kJ/mol for the stable pools, respectively.

In general, studies where parameters are estimated from experimental data and a posteriori correlated against each other (e.g., Fierer et al., 2005), should be interpreted with care while taking into account the expected statistical correlation of model parameters (e.g., Draper and Smith, 1981) that does not necessarily have an ecological meaning. We demonstrate this here using a Monte-Carlo approach, where we create simulated data sets by adding normal random errors (representing experimental noise) to a deterministic temperature (T_{soil}) response of soil respiration (R_{soil}) described by $R_{\text{soil}} = B \cdot Q_{10}^{0.1 \cdot T_{\text{soil}}}$ ($B = 0.1 \mu\text{g C g}^{-1} \text{ soil h}^{-1}$; $Q_{10} = 2.5$; T_{soil} range 10–30°C). These settings are similar to the data recently presented by Fierer et al. (2005). In this way, we produced each 100 data set realizations with random error standard deviations (σ_ϵ) of 0.125, 0.25 and $0.5 \mu\text{g C g}^{-1} \text{ soil h}^{-1}$, respectively (cf. Fig. 2a). Subsequently, for each data set the parameters B and Q_{10} were estimated by a non-linear least squares algorithm (nlinlsq in PV-WAVE 7.0; Visual Numerics Inc., 2003). Figure 2b shows the resulting parameter distribution for $\sigma_\epsilon = 0.25 \mu\text{g C g}^{-1} \text{ soil h}^{-1}$, yielding Q_{10} -values between 2.3 and 3.2, which are strongly correlated with the basal respiration rate parameter B . This correlation is merely an effect of the statistical model properties (Draper and Smith, 1981), since the constructed data set originates from a simple Q_{10} response with simulated noise. While the negative correlation between the parameters is high, with all magnitudes of the experimental error (σ_ϵ), it is evident that with decreasing experimental error the range of Q_{10} -values declines (Table 1), which is in agreement with statistical theory, i.e. lower data uncertainties result in lower standard errors of parameters. Hence, it should be checked to which extent the results by Fierer et al. (2005) are affected by such statistical correlation, or – stated differently – if the correlations found by them are significantly higher than those expected from the statistical effect shown here.

740

Well-designed studies are necessary to test hypotheses regarding how soil carbon pool properties and their temperature sensitivities are related. Recent laboratory experiments indicate very similar responses of labile and more stable soil organic matter to temperature (Fang et al., 2005; Reichstein et al., 2005). However, due to their short duration, these studies can only differentiate between pools with very short and moderate turnover times, while no information on the behaviour of very old, resistant and or physically protected soil organic matter fractions can be obtained. Moreover, by definition, short-term studies, where high-frequency temperature oscillations are imposed (Fang et al., 2005; Reichstein et al., 2005) are not designed for detecting any possible acclimation effects. On the other hand, longer-term studies might create very unnatural conditions in the incubated soil samples since no fresh carbon input enters the soil (unlike in the ecosystem). This factor may induce microbial starvation and associated modifications of microbial assimilation efficiency, and may affect the respiratory quotient (ratio of CO₂ production and O₂ consumption) (Persson et al., 2000), cause accumulation of metabolic by-products (e.g., Kirschbaum, 1995), and change the microbial community structure. The latter has been clearly shown in situ in a girdling experiment, where the soil organismic community changed severely during one year of no supply of photosynthates (Schulze et al., 2004). Moreover, while the apparent acclimation of respiration observed during warming experiments can be favourably explained just by the dynamics of soil organic matter pools without real acclimation (Reichstein et al., 2000; Kirschbaum, 2004; Knorr et al., 2005), it cannot be doubted that acclimation does occur in biological systems as e.g. recently reviewed for plants (Atkin et al., 2005) and studied for fungi (Lange and Green, 2005). However the importance of these effects at the ecosystem level in the context of global warming remains unclear. Until these issues are more completely resolved, we have to live with more uncertainty with respect to the temperature sensitivity of soil organic matter decomposition than the Knorr et al. (2005) study seems to convey.

741

3 Conclusions

Based on our analysis, we believe that it is premature to conclude that stable soil organic matter is more sensitive to temperature than labile organic matter. Although it is reasonable from a theoretical point of view that low quality substrates with long turnover times may have stronger temperature dependence than high quality substrates (Bosatta and Ågren, 1999), the implementation of this assumption in global carbon models is currently not justified by empirical evidence. The temperature sensitivity of different soil carbon pools merits further high-priority scientific experimental and theoretical analysis. Recent advances in stable isotope and soil organic matter fractionation techniques offer promising perspectives (Leifeld and Fuhrer, 2005).

Acknowledgements. We would like to thank V. Brovkin, M. Heimann and W. Lucht for motivation and insightful comments during the preparation of this manuscript. The analysis was facilitated by a European Marie-Curie fellowship to MR at the University of Tuscia, Viterbo (INTERMODE project; MEIF-CT-2003-500696).

References

- Ågren, G. I.: Temperature dependence of old soil organic matter, *Ambio*, 29, 55–55, 2000.
- Atkin, O. K., Bruhn, D., Hurry, V. M., and Tjoelker, M. G. The hot and the cold: unravelling the variable response of plant respiration to temperature, *Functional Plant Biology*, 32, 87–105, 2005.
- Bosatta, E. and Ågren, G. I.: Soil organic matter quality interpreted thermodynamically, *Soil Biology and Biochemistry*, 31, 1889–1891, 1999.
- Davidson, E. A., Belk, E., and Boone, R. D.: Soil water content and temperature as independent or confounded factors controlling soil respiration in temperate mixed hardwood forest, *Global Change Biology*, 4, 217–227, 1998.
- Davidson, E. A., Trumbore, S. E., and Amundson, R.: Soil warming and organic carbon content, *Nature*, 408, 789–790, 2000.
- Draper, N. and Smith, H.: *Applied Regression Analysis*, Wiley, New York, 1981.

742

- Fang, C., Smith, P., Moncrieff, J., and Smith, J. U.: Similar response of labile and resistant soil organic matter pools to changes in temperature, *Nature*, 433, 57–59, 2005.
- Fierer, N., Craine, J. M., McLauchlan, K., and Schimel, J. P.: Litter quality and the temperature sensitivity of decomposition, *Ecology*, 86, 320–326, 2005.
- 5 Giardina, C. P. and Ryan, M. G.: Evidence that decomposition rates of organic carbon in mineral soil do not vary with temperature, *Nature*, 404, 858–861, 2000.
- Jones, C., Cox, P., and Huntingford, C.: Uncertainty in climate–carbon-cycle projections associated with the sensitivity of soil respiration to temperature, *Tellus*, 55B, 642–648, 2003.
- Kätterer, T., Reichstein, M., Andrén, O., and Lomander, A.: Temperature dependence of organic matter decomposition: a critical review using literature data analysed with different models, *Biology and Fertility of Soils*, 27, 258–262, 1998.
- 10 Kirschbaum, M. U. F.: The temperature dependence of soil organic matter decomposition, and the effect of global warming on soil organic carbon storage, *Soil Biology and Biochemistry*, 27, 753–760, 1995.
- 15 Kirschbaum, M. U. F.: Soil respiration under prolonged soil warming: are rate reductions caused by acclimation or substrate loss? *Global Change Biology*, 10, 1870–1877, 2004.
- Knorr, W., Prentice, I. C., House, J. I., and Holland, E. A.: Long-term sensitivity of soil carbon turnover to warming, *Nature*, 433, 298–301, 2005.
- Lange, O. L. and Green, T. G. A.: Lichens show that fungi can acclimate their respiration to seasonal changes in temperature, *Oecologia*, 142, 20–27, 2005.
- 20 Leifeld, J. and Fuhrer J.: The temperature response of CO₂ production from bulk soils and soil fractions is related to soil organic matter quality, *Biogeochemistry*, in press, 2005.
- Liski, J., Ilvesniemi, H., Mäkelä, A., and Westman, K. J.: CO₂ emissions from soil in response to climatic warming are overestimated – the decomposition of old soil organic matter is tolerant of temperature, *Ambio*, 28, 171–174, 1999.
- 25 Lloyd, J. and Taylor, J. A.: On the temperature dependence of soil respiration, *Functional Ecology*, 8, 315–323, 1994.
- Luo, Y., Wan, S., Hui, D., and Wallace, L. L. Acclimatization of soil respiration to warming in a tall grass prairie, *Nature*, 413, 622–625, 2001.
- 30 Persson, T., Karlsson, P. S., Seyferth, U., et al.: Carbon mineralization in European Forest soils, *Ecological Studies*, 142, 257–275, 2000.
- Reichstein, M., Bednorz, F., Broll, G., and Kätterer, T.: Temperature dependence of carbon mineralization: conclusions from a long-term incubation of subalpine soil samples, *Soil Biology*

743

- and *Biochemistry*, 32, 947–958, 2000.
- Reichstein, M., Rey, A., Freibauer, A., et al.: Modelling temporal and large-scale spatial variability of soil respiration from soil water availability, temperature and vegetation productivity indices, *Global Biogeochemical Cycles*, 17, 1104, doi:10.1029/2003GB002035, 2003.
- 5 Reichstein, M., Subke, J.-A., Angeli, A. C., and Tenhunen, J. D.: Does the temperature sensitivity of decomposition of soil organic matter depend upon water content, soil horizon, or incubation time? *Global Change Biology*, in press, 2005.
- Sandermann, J., Amundson, R., and Baldocchi, D.: Application of eddy covariance measurements to the temperature dependence of soil organic matter mean residence time, *Global*
- 10 *Biogeochemical Cycles*, 17, 1061, doi:10.1029/2001GB001833, 2003.
- Schulze, X. W., Gleixner, G., Kaiser, K., et al.: A proteomic fingerprint of dissolved organic carbon and of soil particles, *Oecologia*, 142, 335–343, 2004.
- Visual Numerics Inc.: PV-Wave Advantage Reference, VNI Press, Houston, 2003.

744

Table 1. The effect of the random error standard deviation (σ_ε) on the statistical properties of the parameter estimates of the Q_{10} model as found by the Monte-Carlo analysis. See text for details.

σ_ε [$\mu\text{g C g}^{-1} \text{ soil h}^{-1}$]	Percentiles for Q_{10}		Percentiles for base respiration		Linear correlation Q_{10} and base respiration
	5%	95%	5%	95%	
0.125	2.43	2.72	0.080	0.108	-0.99
0.25	2.41	2.98	0.068	0.112	-0.96
0.5	2.03	3.30	0.049	0.176	-0.94

745

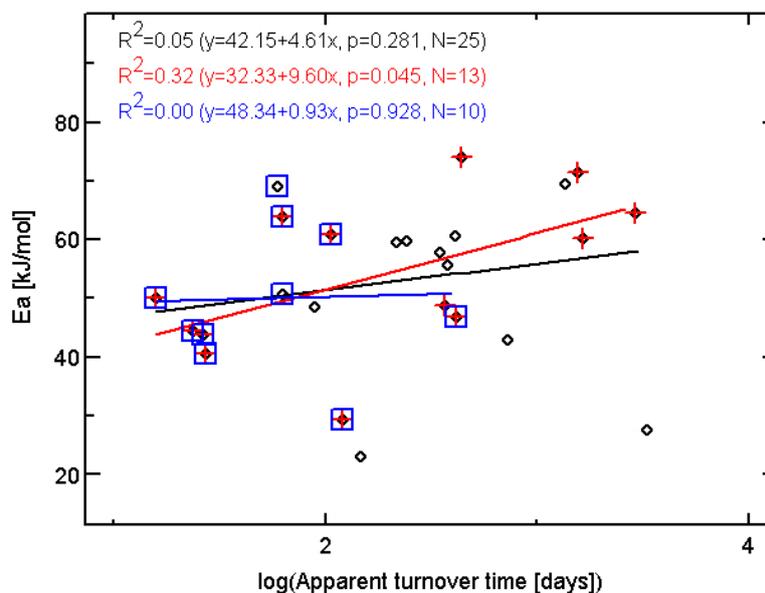


Fig. 1. Correlations between activation energy (E_a) and turnover time using data from Kätterer et al. (1998). Black diamonds and black regression line reflect all data in Kätterer et al. (1998). Red crosses and red regression line represent studies with incubation time over 100 days (as in Knorr et al., 2005). Blue squares and regression lines represent all studies where incubation time was at least as long as the apparent half-life of the sample. Coefficients of determination (R^2), the regression equation, the significance level (p) and the number of the studies included (N) are given at the top in the respective colour.

746

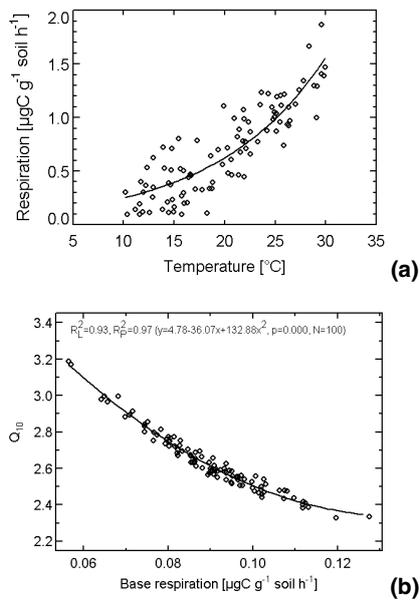


Fig. 2. (a) One single realisation of the data set in the Monte-Carlo analysis. This data set (symbols) was created by adding a normally distributed random error with a standard deviation of $0.25 \mu\text{C g}^{-1} \text{soil h}^{-1}$ to the function $R_{\text{soil}}=B \cdot Q_{10}^{0.1 \cdot T_{\text{soil}}}$ ($B=0.1 \mu\text{C g}^{-1} \text{soil h}^{-1}$; $Q_{10}=2.5$). One hundred of those realizations were constructed, and for each realization the parameters B and Q_{10} estimated by non-linear regression. Panel **(b)** shows the distribution of the parameters after this procedure. A 2nd order polynomial regression line is added, and the linear (R_L^2) and the polynomial (R_P^2) coefficients of determination, the polynomial regression equation, the significance (p) and the number of data points (N) are indicated at the top.