Inferential Structure Determination: Overview and new developments

Michael Habeck

Max Planck Institutes for Developmental Biology and for Biological Cybernetics, Tübingen, Germany

michael.habeck@tuebingen.mpg.de

NMR structure determination flowchart



- derivation of conformational restraints
 - NOE classification, calibration, distance bounds
 - parameters of Karplus curve
 - determination of alignment tensor

- derivation of conformational restraints
- choice of restraint potential
 - harmonic
 - flat-bottom harmonic-wall

- derivation of conformational restraints
- choice of restraint potential
- choice of weighting factors
 - "force constants" for NOEs and other data
 - empirical defaults
 - crossvalidation

- derivation of conformational restraints
- choice of restraint potential
- choice of weighting factors
- structure calculation
 - how many?
 - starting structure?

- derivation of conformational restraints
- choice of restraint potential
- choice of weighting factors
- structure calculation
- selection of representative structures
 - selection criterion: energy based?
 - restraint violation?

- derivation of conformational restraints
- choice of restraint potential
- choice of weighting factors
- structure calculation
- selection of representative structures

structures depend on subjective decisions rather than just the data

Goals

- reduce subjective judgment
- quantify the uncertainty that's inherent in NMR structure determination
- calculate "objective" NMR structures
- make statements about reliability

- a single conformational degree of freedom (φ angle)
- data: NOEs for HN-HA distance $d(\phi)$





- a single conformational degree of freedom (ϕ angle)
- data: NOEs for HN-HA distance $d(\phi)$





- a single conformational degree of freedom (φ angle)
- data: NOEs for HN-HA distance $d(\phi)$







- a single conformational degree of freedom (φ angle)
- data: NOEs for HN-HA distance $d(\phi)$







Probability as a measure of uncertainty













Matching $\boldsymbol{\phi}$

overlap between data histogram and model defines $Prob(\phi)$





















Inferential Structure Determination (ISD)

- quantify uncertainties by probabilities
- describe NMR data probabilistically
- express background knowledge by prior probabilities
- consistently combine the probabilities using probability calculus (Bayes' theorem)
- analyse the joint posterior probability of all unknowns (coordinates + model parameters + errors)

Science, 2005; Phys Rev E, 2005

Drawing random samples from probabilities

- in real-world problems probabilities are too complex for visual or analytical analysis
- idea of sampling: pick a set of representatives



Drawing random samples from probabilities

- in real-world problems probabilities are too complex for visual or analytical analysis
- idea of sampling: pick a set of representatives



Drawing random samples from probabilities

- in real-world problems probabilities are too complex for visual or analytical analysis
- idea of sampling: pick a set of representatives



Gibbs sampling

fix coordinates, update additional parameters

fix additional parameters, update coordinates







Generation of structure ensembles

"Temperature"



Optimization algorithms are not for sampling

- optimization algorithms are often used to "sample" the protein conformation space
- but they are designed to locate (global) optima
- multi-start simulated annealing already fails for example



Prior probability of protein conformations

Physical knowledge:

- covalent forces (bond lengths, angles)
- noncovalent forces (van der Waals, electrostatic)
- solvent (hydrophobic forces, entropic effects)

Parametrization in torsion angles



$$\label{eq:prob} \begin{split} & \mathsf{Prob}(\mathsf{structure}) \propto \exp\{-\beta \mathsf{E}_{\mathsf{phys}}(\mathsf{structure})\} \\ & \mathsf{Prior} = \mathsf{Boltzmann} \ \mathsf{ensemble} \end{split}$$

Probabilistic modelling of NMR data

Principle: imagine a process that could have generated your data

This typically comprises

- a forward model (eg. ISPA, Karplus curve)
- an error model (eg. Gaussian distribution)

"nuisance" parameters:

- model parameters (eg. A, B, C in Karplus curve)
- error parameters (eg. width σ of the Gaussian)

Modelling NOEs

forward model: ISPA

• NOE = scale / distance⁶



Modelling NOEs

forward model: ISPA

- NOE = scale / distance⁶
- log(NOE) = log(scale) 6 log(distance)



Modelling NOEs

forward model: ISPA

- NOE = scale / distance⁶
- log(NOE) = log(scale) 6 log(distance)





error model: log-normal

• error = log(NOE) - log(scale) + 6 log(distance)





error model: log-normal

• error = log(NOE) - log(scale) + 6 log(distance)





error model: log-normal

• error = log(NOE) - log(scale) + 6 log(distance)







Idea: calculate average and variance from random samples



Idea: calculate average and variance from random samples



Idea: calculate average and variance from random samples





SH3, 154 distances

Science, 2005



TUDOR, 1875 distances

Phys Rev E, 2005; JACS 2005

Adaptive weighting of data

Standard approach: $E_{hybrid} = W_{data} E_{data} + E_{phys}$

Choice of weight is critical:



Probabilistic interpretation: $w_{data} = 1/\sigma^2$

PNAS, 2006

Adaptive weighting of data

Standard approach: $E_{hybrid} = W_{data} E_{data} + E_{phys}$

Choice of weight is critical:



Probabilistic interpretation: $w_{data} = 1/\sigma^2$

PNAS, 2006

Bayes vs. crossvalidation



2 NMR structures of Josephin





1yzb

2aga



2aga (2960)



1yzb (5525 + 925)

2aga (2960)





 $\sigma = \langle \text{restraint RMS} \rangle_{\text{unbiased}} \propto \sqrt{\mathsf{R}_{\mathsf{free}}}$



Summary

- uncertainties in NMR structure determination must be treated probabilistically
- structure calculation by posterior sampling
- ensemble of sampled structures is statistically meaningful
- model parameters can be estimated (eg. NOE scale, Karplus parameters, alignment tensor)
- error parameters can be estimated (effectively: adaptive weighting of the data)
- estimated errors are useful figures of merit and could replace free R values

Acknowledgement

Andrei Lupas (MPI for Developmental Biology, Tübingen) Bernhard Schölkopf (MPI for Biological Cybernetics, Tübingen)

Annalisa Pastore (NIMR, London)

Michael Nilges (Institut Pasteur, Paris) Wolfgang Rieping (University of Cambridge)