

Supporting Information

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Carbon Dioxide as a C₁ Building Block for the Formation of Carboxylic Acids by Formal Catalytic Hydrocarboxylation**

*Thomas G. Ostapowicz, Marc Schmitz, Monika Krystof, Jürgen Klankermayer, and Walter Leitner**

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Supporting Information

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S1 Material and Methods

General. All manipulations involving air-sensitive compounds were carried out under inert atmosphere using schlenk techniques or in a glovebox (*MBraun LabMaster SP*). Argon 4.8 (*Messer*, Germany) was used as inert gas in all cases. Prior to use, all glassware was dried in high vacuum, evacuated and refilled with argon at least three times.

Autoclaves. The catalytic runs were performed in 10 mL stainless steel autoclaves. To avoid blind activity, the steel autoclaves were equipped with glass inlets. The autoclaves were evacuated at high vacuum for at least one hour and then charged with an argon atmosphere.

Solvents and Chemicals. Acetic acid was pre-dried over molecular sieves (4 Å) and then refluxed for 2 h over anhydrous CuSO_4 , distilled, and stored over molecular sieves (4 Å) under argon. Methyl iodide was vacuum distilled at low temperatures prior to use and stored at 4°C under argon. All substrates containing stabilizing agents were distilled prior to use and stored under argon over molecular sieves (4 Å). All other substrates were degassed by three freeze-pump-thaw cycles and stored over molecular sieves (3 or 4 Å) under argon. Deionised water was taken from a reverse-osmotic purification system (*Werner EasyPure II*) and degassed by bubbling argon with a frit for at least 1 h. Water contents of all organic solvents were monitored by Carl-Fischer titration (*Metrohm 756 F Coulometer*) and typically kept on the following levels: Acetic acid < 100 ppm, dichloromethane 5 - 10 ppm, tetrahydrofuran 30 - 50 ppm. Deuterated solvents were degassed by three freeze-pump-thaw cycles and stored over molecular sieves 3 Å or 4 Å under argon. All reagents were commercially supplied and used as received unless stated otherwise.

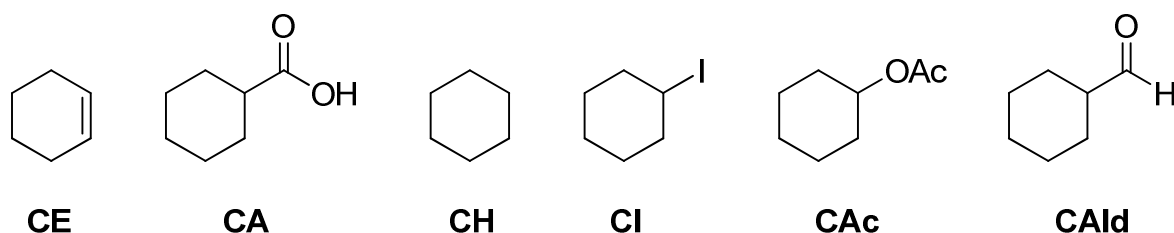
NMR Spectroscopy. NMR spectra were recorded with spectrometers *Bruker AV-600*, *AV-III-400* or *-300* at ambient temperature at the frequency noted. Chemical shifts δ are given in ppm relative to tetramethylsilane (^1H , ^2H and ^{13}C).

Mass Spectrometry. High resolution MS analyses were performed on a *LTQ Orbitrap XL* (*Thermo Fisher Scientific*) by direct ESI from organic solutions without acidification in (+) ionisation. Detected masses are given in m/z and correlated to calculated masses of the respective species.

Gaschromatography. GC analyses were performed on a *Trace GC Ultra* (*Thermo Scientific*) using a packed *CP-WAX-52-CB* column (length = 60 m, diameter = 0.25 mm) isothermally at 70°C for 5 min, then heated to 200°C at 8°C min⁻¹. A constant flow of 2.5 mL min⁻¹ He was applied. The gaschromatograph was equipped with a FID detector.

S2 Catalytic Experiments

The abbreviations for substrates and products are set as follows:



S2.1 Variation of the Metal Source

General procedure: The according metal precursor (93 μmol per metal atom) and cyclohexene (1.87 mmol) were weighed into a Schlenk tube with acetic acid (0.65 mL). In the runs where methyl iodide was applied as a promotor, 925 μmol CH_3I was added. The red brownish solution was transferred via cannula to a stainless steel autoclave with PPh_3 (460 μmol). The autoclave was pressurized with CO_2 (4.0 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting solution the standards 1-phenylethanol (100 mg) and *n*-dodecane (100 mg) were added and the mixture was analyzed by gas chromatography. Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Table S2.1. Carboxylation of cyclohexene with CO_2 and H_2 investigating various metal catalyst precursors. Cf. Table 1 within the manuscript.

Entry	Cat. precursor	Promotor	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	$\text{Fe}_2(\text{CO})_9$	--	16	--	--	--	--	S15
2	$\text{Fe}_2(\text{CO})_9$	CH_3I	20	<1	--	<1	2	S16
3	$\text{Pd}(\text{OAc})_2$	--	8	--	<1	--	--	S17
4	$\text{Pd}(\text{OAc})_2$	CH_3I	22	<1	2	4	10	S18
5	$[\text{RhCl}(\text{CO})_2]_2$	--	20	<1	5	--	--	S19
6	$[\text{RhCl}(\text{CO})_2]_2$	CH_3I	96	69	10	2	1	S20

S2.2 Variation of the Acidic Additive

General procedure: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and the acidic additive (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and *n*-dodecane (100 mg) were added and the mixture was analysed by gas chromatography. Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Table S2.2. Carboxylation of cyclohexene with CO_2 and H_2 investigating the influence of the acidic additive. Cf. Table 1 within the manuscript.

Entry	Acidic additive ^[a]	Amount acidic additive [μmol]	pK_a (DMSO)	Ref. for pK_a	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	HBTA	330	1.7	[1]	97	77	6	5	<1	S21
2	TFA	330	0.5	[2]	85	41	21	<1	4	S22
3	MSA	330	-1.9	[3]	96	65	8	2	<1	S23
4	<i>p</i> -TsOH	330	-2.8	[3]	99	75	4	2	<1	S24
5	<i>p</i> -TsOH·H ₂ O	330	-2.8	[3]	99	88	2	1	<1	S25
6	<i>p</i> -TsOH·H ₂ O	650	-2.8	[3]	99	92	5	2	<1	S27
7	<i>p</i> -TsOH·H ₂ O	1120	-2.8	[3]	99	83	9	2	<1	S28

[a]: HBTA: *bis*(trifluoromethanesulfonyl)imide; TFA: trifluoroacetic acid; MSA: methanesulfonic acid; *p*-TsOH: *para*-toluenesulfonic acid; *p*-TsOH·H₂O: *para*-toluenesulfonic acid monohydrate.

S2.3 Variation of the Solvent

General procedure: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with the according solvent (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and n -dodecane (100 mg) were added and the mixture was analysed by gas chromatography. Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Table S2.3. Carboxylation of cyclohexene with CO_2 and H_2 investigating the influence of the solvent.

Entry	Solvent	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	<i>neat</i>	98	59	12	5	<1	S29
2	propionic acid	98	77	6	3	--	S30
3	tetrahydrofuran	27	<1	1	3	2	S31

S2.4 Variation of the Iodide Source

General procedure: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol) and the according iodide source (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and $n\text{-dodecane}$ (100 mg) were added and the mixture was analysed by gas chromatography. Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Table S2.4. Carboxylation of cyclohexene with CO_2 and H_2 investigating the influence of the iodide source. Cf. Table 1 within the manuscript.

Entry	Iodide source	Amount iodide source [μmol]	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	I_2	925	87	47	9	12	4	S32
2	Lil	925	90	46	15	<1	5	S33
3	NaI	925	36	8	13	<1	2	S34
4	KI	925	26	3	21	<1	2	S35
5	$[\text{CH}_3\text{PPh}_3]\text{I}$	925	26	1	5	--	3	S36
6 ^[a]	CI	925	98	73	<1	4	1	S37
7 ^[a]	CI	184	91	54	22	1	5	S38
8 ^[a]	CI + Lil	184 + 736	95	71	11	2	1	S39

[a]: Conversion and yield calculated for **CE+CI** acting both as substrates.

S2.5 Variation of the Phosphine Ligands

Variation of the phosphine ligand revealed a strong influence on the catalytic performance. Both electronic donating or withdrawing substituents in *para*-position of the phenyl groups showed little effects, and similar yields of **CA** were obtained under otherwise identical conditions (P(*p*-Tol)₃: 66%, P(*p*-CF₃-C₆H₄)₃: 67%). Alkyl phosphines P^{*n*}Oct₃ and PCy₃ showed also very good performance with 69% and 81% yield, respectively. A decrease in selectivity towards **CA** results with sterically more demanding ligands like P^{*t*}Bu₃ (32%) or P(*o*-Tol)₃ (5%). The use of bidentate ligands Ph₂P(CH₂)_nPPh₂ (n=2: dppe, n=3: dppp) or the tridentate ligand H₃CC[(CH₂)PPh₂]₃ (triphos) lead to complete suppression of **CA** formation on the expense of hydrogenation or general loss of activity, respectively. These data strongly suggest dynamic ligand exchange equilibria as important regulators for the system. This is further corroborated by variation of the P/Rh ratio with PPh₃ where maximum **CA** yields of >80% was observed in the range of 5:1 to 8:1, with rapid decay to values below 5% above and below these limits.

General procedure: Under an argon atmosphere, [RhCl(CO)₂]₂ (46 μmol), cyclohexene (1.88 mmol) and CH₃I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The solution was transferred via cannula to a stainless steel autoclave, in which the phosphine ligand (460 μmol á P atom) and *p*-TsOH·H₂O (330 μmol) were already deposited. The autoclave was pressurized with CO₂ (4.1 g) and then additional 10 bar of H₂ were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and *n*-dodecane (100 mg) were added and the mixture was analysed by gas chromatography. Yields were found to be reproducible within ΔY = ±2% in two independent runs for selected experiments.

Table S2.5. Carboxylation of cyclohexene with CO₂ and H₂ investigating different phosphine ligands.

Entry	Ligand ^[a]	Amount ligand [μmol]	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	P ^{<i>t</i>} Bu ₃	460	74	32	10	4	6	S40
2	P(cyclohexyl) ₃	460	>99	81	5	3	1	S41
3	P(<i>n</i> -octyl) ₃	460	99	69	5	2	<1	S42
4	P(<i>p</i> -tolyl) ₃	460	95	66	2	1	<1	S43
5	P(<i>o</i> -tolyl) ₃	460	73	5	<1	10	11	S44
6	P(<i>p</i> -CF ₃ -Ph) ₃	460	98	67	21	1	<1	S45
7	tppms ^[c]	460	98	76	3	3	<1	S46
8	P(OPh) ₃	460	40	<1	<1	4	15	S47
9	O=PPh ₃	460	74	29	<1	13	6	S48
10	dppe	230	32	<1	--	15	6	S49
11	dppp	230	38	<1	<1	14	9	S50
12	triphos	155	33	<1	<1	10	11	S51
13	PPh ₃	46	68	27	3	22	10	S52
14	PPh ₃	690	51	4	22	--	6	S53

[a] dppe: Ph₂P(CH₂)₂PPh₂; dppp: Ph₂P(CH₂)₃PPh₂; triphos: H₃CC[(CH₂)PPh₂]₃.

S2.6 Conversion Time Profile

General procedure: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which the PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After the according time interval, the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and $n\text{-dodecane}$ (100 mg) were added and the mixture was analysed by gas chromatography. Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Table S2.6. Conversion/yield time profile of the carboxylation of cyclohexene with CO_2 and H_2 . Cf. Figure 1 within the manuscript.


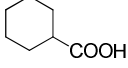

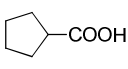

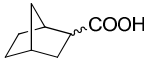


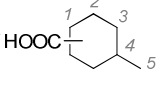

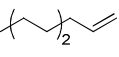
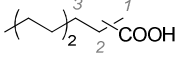
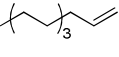
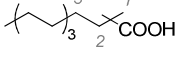
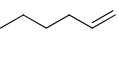
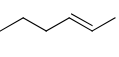
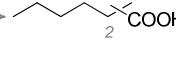
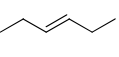

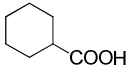
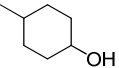
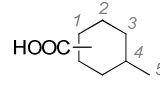
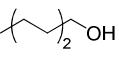

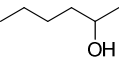
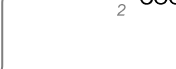

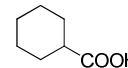

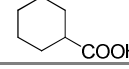
Entry	Reaction time [h]	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	1	40	3	2	15	2	S54
2	2	63	27	3	11	4	S55
3	3	80	47	4	11	5	S56
4	6	90	70	5	9	3	S57
5	9	96	77	5	3	1	S58
6	12	98	81	5	3	1	S59
7	16	>99	85	5	1	<1	S60
8	20	>99	85	5	2	1	S61

S2.7 Substrate Scope

General procedure: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), the according substrate (1.88 mmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C in an aluminium cylinder. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and *n*-dodecane (100 mg) were added and the mixture was analysed by gas chromatography. GC Yields were found to be reproducible within $\Delta Y = \pm 2\%$ in two independent runs for selected experiments.

Aqueous work-up to isolate the carboxylic acid products was performed as follows: After the reaction the autoclave was cooled to 0°C and then carefully vented. The reaction mixture was transferred to a round bottom flask with additional dichloromethane and the solvent was evaporated in vacuo. The residual solid was re-dissolved in dichloromethane (15 mL) and the resulting solution was extracted four times with saturated NaHCO_3 solution (4 x 10 mL). The aqueous phases were combined and concentrated hydrochloric acid was added dropwise until pH 1 was reached. Subsequently, the combined aqueous phases were re-extracted with dichloromethane (5 x 10 mL). The combined dichloromethane phases were dried over Na_2SO_4 and the solvent was removed in vacuo to obtain the carboxylic acid products as slightly yellowish oils or low-melting solids, in agreement with literature melting points. No impurities were detectable by GC chromatography, and only trace amounts of phosphonium ions were detected by ^1H NMR and ^{31}P -NMR spectroscopy. The ^1H NMR spectra are depicted at the according pages for the products along with the GC data (see pages: S26, S63, S68, S72, S76, S79). Completely colorless cyclohexane carboxylic acid was obtained upon recrystallization from pentane at -78°C in 65% yield.

Table S2.7. Carboxylation with CO₂ and H₂ investigating different substrates. Cf. Table 2 within the manuscript.

Entry	Substrate	Conv. [%]	product yield [%]	Isolated yield	GC at page
1		98	 88%	86% yellowish oil, solidifies upon standing (mp _{Lit} = 29°C)	S25
2		98	 91%	81% yellowish oil (mp _{Lit} = 4°C)	S62
3		91	 50% exo 12% endo		S64
4		96	44% COOH at 1 20% COOH at 2/3 9% COOH at 5		S65
5		98	 46% COOH at 1 20% COOH at 2/3 8% COOH at 5		S66
6		99	52% COOH at 1 18% COOH at 2/3 7% COOH at 5	75% yellowish oil	S67
7		99	 42% COOH at 1 22% COOH at 2 10% COOH at 3		S69
8		98	 31% COOH at 1 17% COOH at 2 6% COOH at 3 7% COOH at 4		S70
9		93	53% COOH at 1 26% COOH at 2 11% COOH at 3	76% yellowish oil	S71
10		97	 43% COOH at 1 24% COOH at 2 11% COOH at 3		S73
11		93	45% COOH at 1 23% COOH at 2 10% COOH at 3		S74
12		>99	 74%	73% yellowish oil, solidifies upon standing (mp _{Lit} = 29°C)	S75
13		99	 48% COOH at 1 19% COOH at 2/3 7% COOH at 5		S77
14		> 99	 41% COOH at 1 15% COOH at 2 6% COOH at 3	55% yellowish oil	S78
15		> 99	 37% COOH at 1 19% COOH at 2 8% COOH at 3		S80
16		80	 21%		S81
17		95	 71%		S82

S2.8 Detection of CO and Control Experiments with CO

Procedure for the detection of CO gas: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was pressurized with CO_2 (4.1 g) and then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented into an evacuated gas tube (Figure S2.1 left). To the resulting red solution the standards 1-phenylethanol (100 mg) and n -dodecane (100 mg) were added and both, the gas phase in the gas tube as well as the liquid reaction mixture, were analysed by gas chromatography.

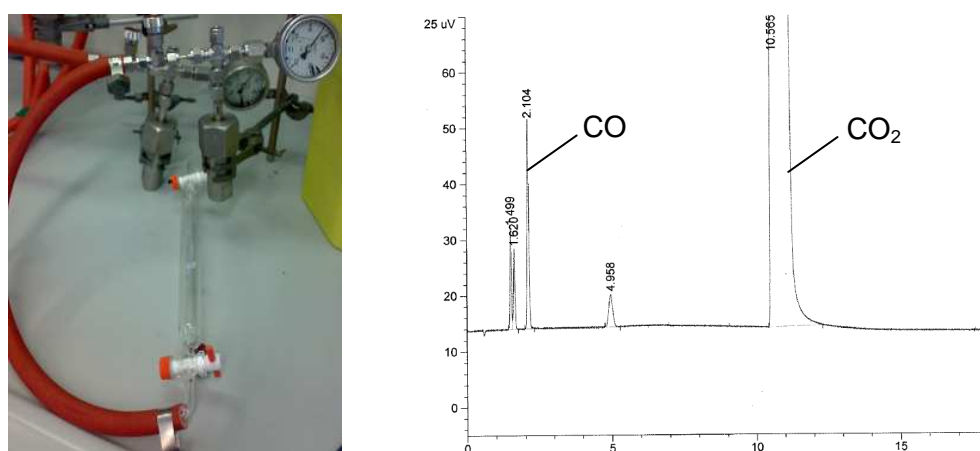


Figure 2.1. Left: Gas tube to trap the gas phase from the autoclave after the reaction. Right: Part of the GC chromatogram taken from the gas phase analysis after the reaction.

Control Experiments applying CO gas: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol), and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were deposited already. The autoclave was pressurized with CO and in some experiments additional H_2 was added at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. To the resulting red solution the standards 1-phenylethanol (100 mg) and n -dodecane (100 mg) were added and the reaction mixture was analysed by gas chromatography.

Table S2.8. Control Experiments applying CO gas instead of CO_2 .

Entry	CO [bar]	H_2 [bar]	Total pressure [bar]	Conv. [%]	Yield of CA [%]	Yield of CH [%]	Yield of CI [%]	Yield of CAc [%]	GC at page
1	30	--	30	91	32	<1	<1	7	S83
2	30	10	40	96	53	<1	<1	5	S84
3	5	10	15	95	79	<1	<1	1	S85

S2.9 Labelling Experiments

Procedure for the ^{13}C labelling experiments: Under an argon atmosphere, $[\text{RhCl}(\text{CO})_2]_2$ (46 μmol), cyclohexene (1.88 mmol) and CH_3I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh_3 (460 μmol) and $p\text{-TsOH}\cdot\text{H}_2\text{O}$ (330 μmol) were already deposited. The autoclave was cooled to different inlet temperatures, then pressurized with $^{13}\text{CO}_2$ and weighed. Afterwards, un-labelled CO_2 was pressurized to reach the total amount of CO_2 between 4.0 and 4.4 g. Then additional 10 bar of H_2 were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C . After 16 h the autoclave was cooled to 0°C and then carefully vented. The resulting red solution was analysed by ^1H and ^{13}C NMR spectroscopy. The analysis of the ^{13}C labelling experiments were conducted by comparing the initial ratios of $^{12}\text{C}:^{13}\text{C}$ in CO_2 with product ^{13}C NMR spectra (Figure S2.2). As internal reference the NMR signal of the ring carbon atom C^b was applied to determine the relative intensity of the signal of C^a . For comparison also Ferrocene was added as external NMR standard (Figure S2.3).

Table S2.9. Labelling experiments using different ratios of $^{13}\text{CO}_2$ and un-labelled CO_2 .

Entry	Inlet Temp. $^{13}\text{CO}_2$ [$^\circ\text{C}$]	Amount $^{13}\text{CO}_2$ [g]	Amount CO_2 [g]	Ratio $^{12}\text{C}:^{13}\text{C}$	Ratio $\text{C}^a:\text{C}^b$ determined. by ^{13}C NMR	Conv. [%]	Yield of CA [%]	^{13}C NMR at page
1	20	--	4.30	89.1:1	1.0:1	99	86	S87
2	20	0.22	3.85	14.5:1	3.6:1	99	84	S87
3	0	0.30	4.00	11.5:1	3.8:1	99	85	S88
4	-40	0.38	4.05	9.4:1	3.9:1	99	81	S88
5	-40	0.39	3.77	8.6:1	4.0:1	98	87	S89
6	-40	0.38	1.83	4.5:1	5.6:1	98	75	S89

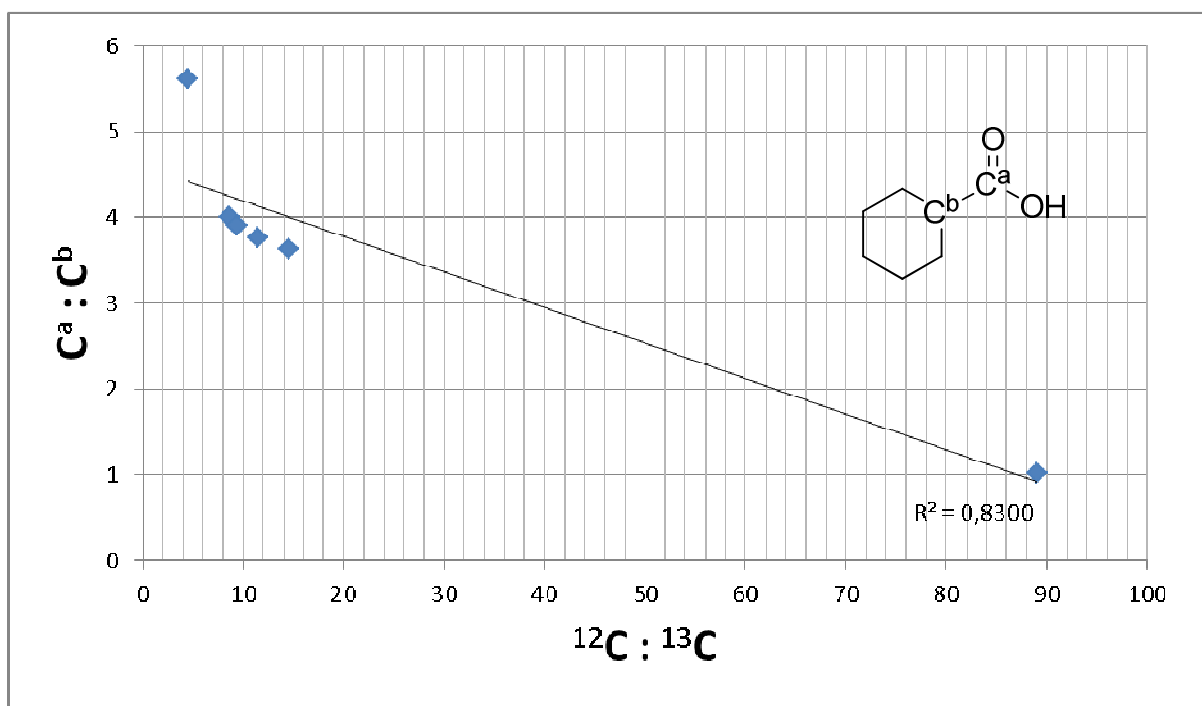


Figure S2.2. Diagram of the ratios of $^{12}\text{C}:^{13}\text{C}$ in CO_2 versus the product data obtained from ^{13}C NMR spectroscopy.

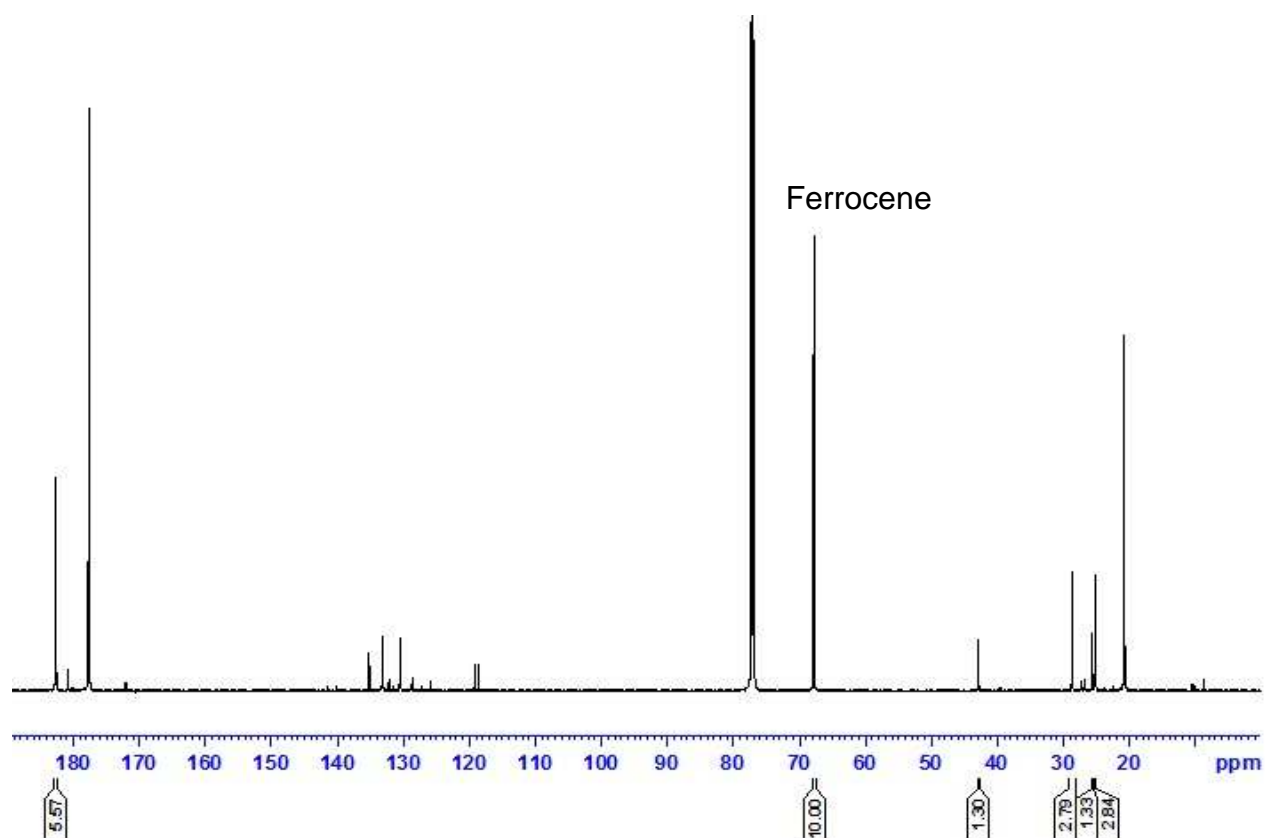
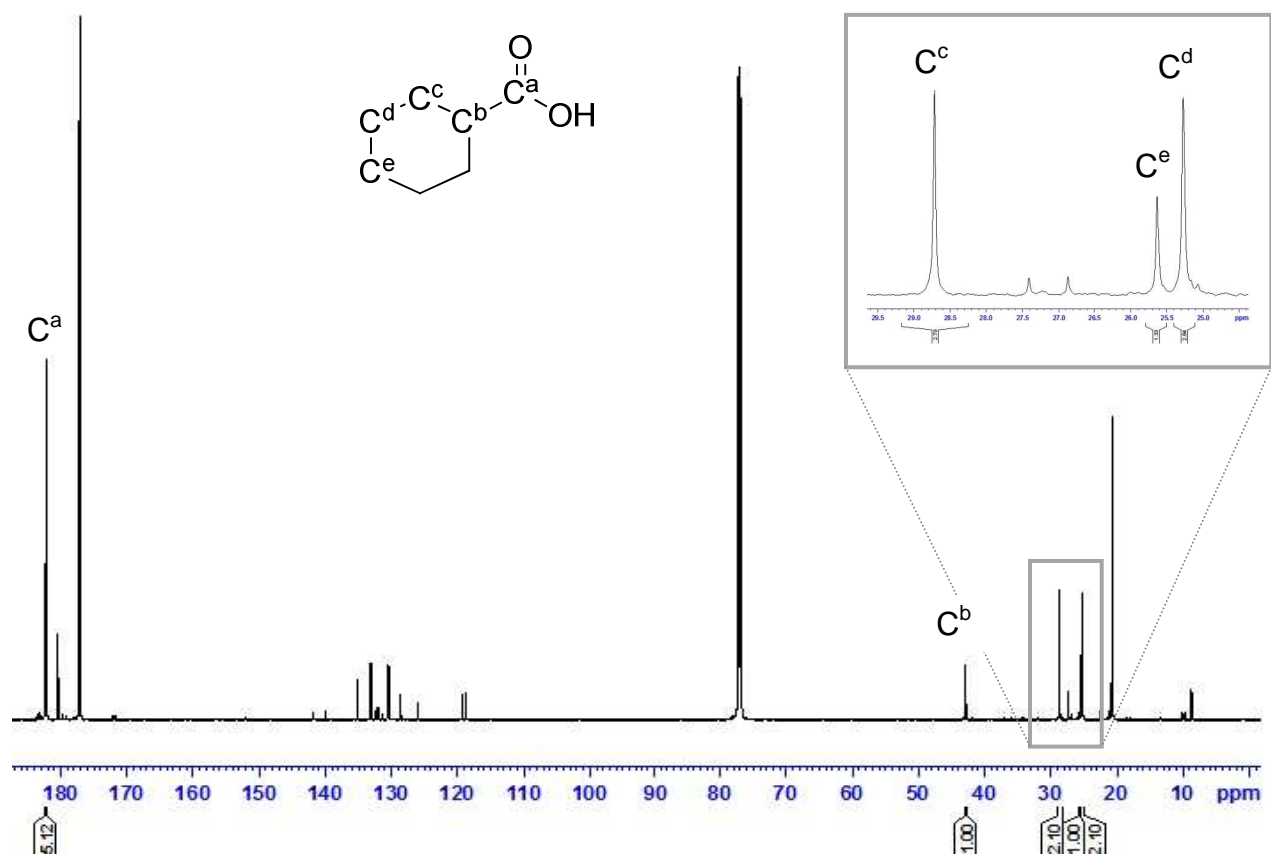


Figure S2.3. ^{13}C NMR spectra of one labeling experiment without (*top*) and with addition of ferrocene (*bottom*) as external NMR standard.

Procedure for the D₂ labelling experiments: Under an argon atmosphere, [RhCl(CO)₂]₂ (46 μmol), cyclohexene (1.88 mmol) and CH₃I (925 μmol) were weighed into a Schlenk tube without addition of acetic acid. The red brownish solution was transferred via cannula to a stainless steel autoclave, in which the PPh₃ (460 μmol) and *p*-TsOH·H₂O (330 μmol) were already deposited. The autoclave was pressurized with 10 bar of D₂ and then CO₂ (4.1 g) was added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C in an aluminium cylinder. After 16 h, the autoclave was cooled to 0°C and then carefully vented. The resulting red solution was analysed by ¹H and ²H NMR spectroscopy. The according spectra are depicted on page 90.

Procedure for the D₂O labelling experiments: Under an argon atmosphere, [RhCl(CO)₂]₂ (46 μmol), cyclohexene (1.88 mmol) and CH₃I (925 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL) and D₂O (0.1 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh₃ (460 μmol) and *p*-TsOH·H₂O (330 μmol) were already deposited. The autoclave was pressurized with CO₂ (4.1 g) and then additional 10 bar of H₂ were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. The resulting red solution was analysed by ¹H and ²H NMR spectroscopy. The according spectra are depicted on page 91.

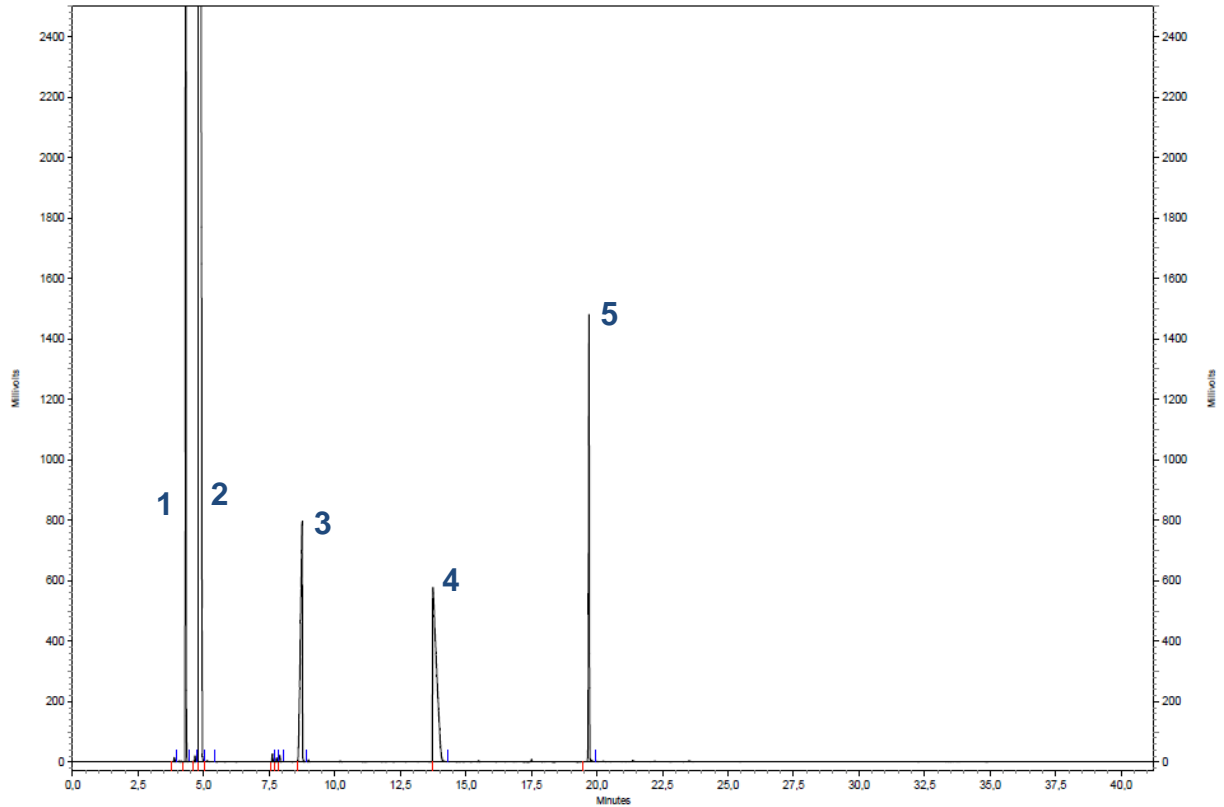
Procedure for the H₂¹⁸O labelling experiments: Under an argon atmosphere, [RhCl(CO)₂]₂ (46 μmol), cyclohexene (1.88 mmol) and **Cl**+LiI (184+ 736 μmol) were weighed into a Schlenk tube along with acetic acid (0.65 mL) and H₂¹⁸O (0.3 mL). The red brownish solution was transferred via cannula to a stainless steel autoclave, in which PPh₃ (460 μmol) and *p*-TsOH·H₂O (330 μmol) were already deposited. The autoclave was pressurized with CO₂ (4.1 g) and then additional 10 bar of H₂ were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. The resulting red solution was analysed by mass spectrometry. Masses were found to be reproducible in accuracy and intensity for the labelled compounds during 3 min of measuring time. The according spectra are depicted on page 92.

Procedure for the H₂¹⁸O control experiment: Under an argon atmosphere, cyclohexane carboxylic acid (1.88 mmol) was weighed into a Schlenk tube along with acetic acid (0.65 mL) and H₂¹⁸O (0.2 mL). The solution was transferred via cannula to a stainless steel autoclave. The autoclave was pressurized with CO₂ (4.5 g) and then additional 10 bar of H₂ were added up to a total pressure of 70 bar at room temperature. The reaction mixture was stirred and heated to 180°C. After 16 h the autoclave was cooled to 0°C and then carefully vented. The resulting solution was analysed by mass spectrometry. Masses were found to be reproducible in accuracy and intensity for the labelled compounds during 3 min of measuring time. The according spectra are depicted on page 93.

S3 Gaschromatographic Data

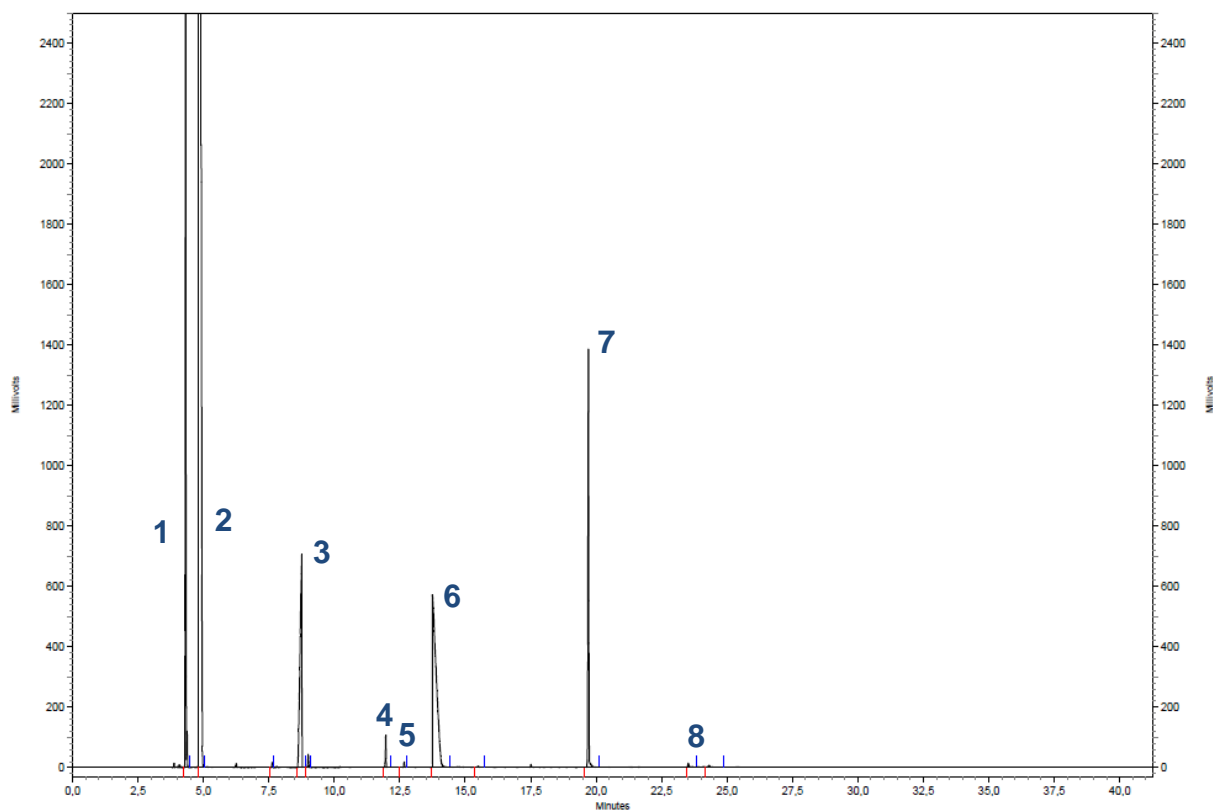
S3.1 Gaschromatograms to Table S2.1

Entry 1



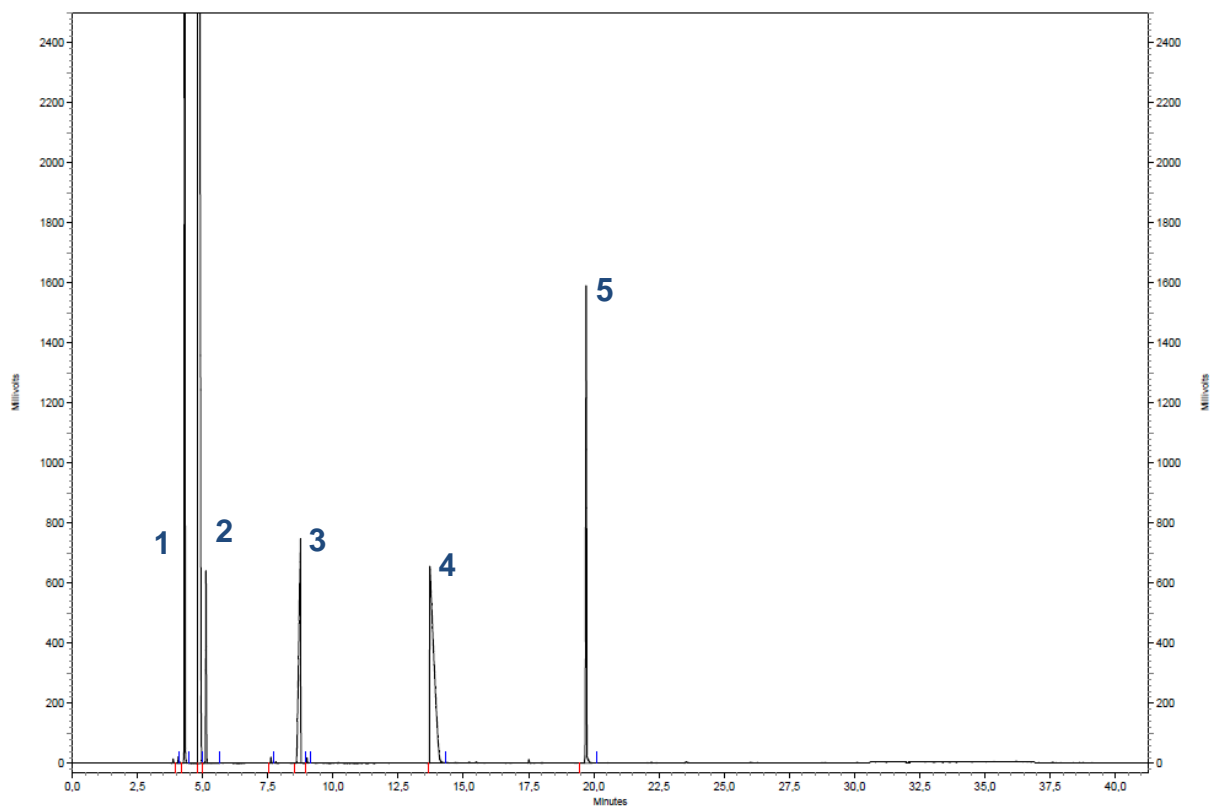
Entry	Retention Time [min]	Substance	Area
1	4.317	CE	58098986
2	4.798	CH ₂ Cl ₂ (Solv.)	538231322
3	8.760	<i>n</i> -Dodecane (Stand.)	38862081
4	13.745	CH ₃ COOH (Solv.)	57959547
5	19.703	1-Phenylethanol (Stand.)	28757687

Entry 2



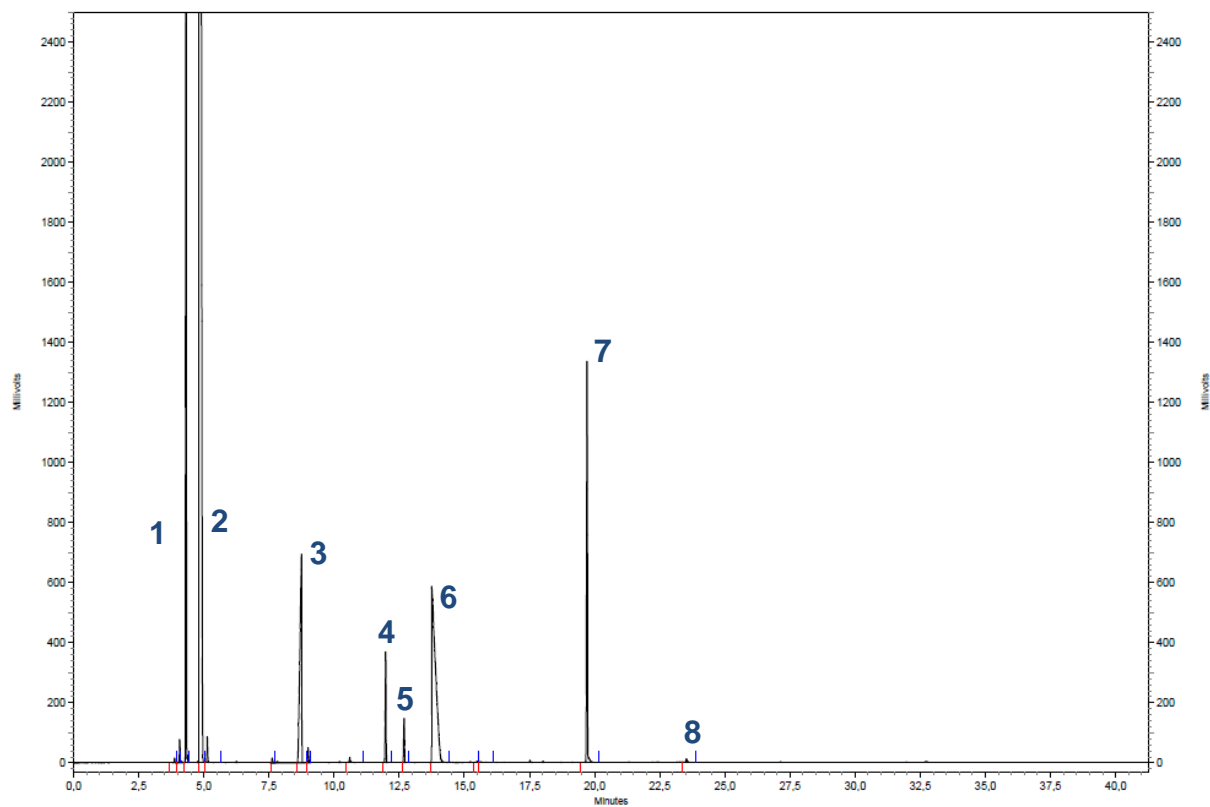
Entry	Retention Time [min]	Substance	Area
1	4.318	CE	50062542
2	4.798	CH ₂ Cl ₂ (Solv.)	542094021
3	8.753	<i>n</i> -Dodecane (Stand.)	32665215
4	11.963	CAc	1747061
5	12.668	CI	310187
6	13.745	CH ₃ COOH (Solv.)	56923094
7	19.703	1-Phenylethanol (Stand.)	27072815
8	23.312	CA	308705

Entry 3



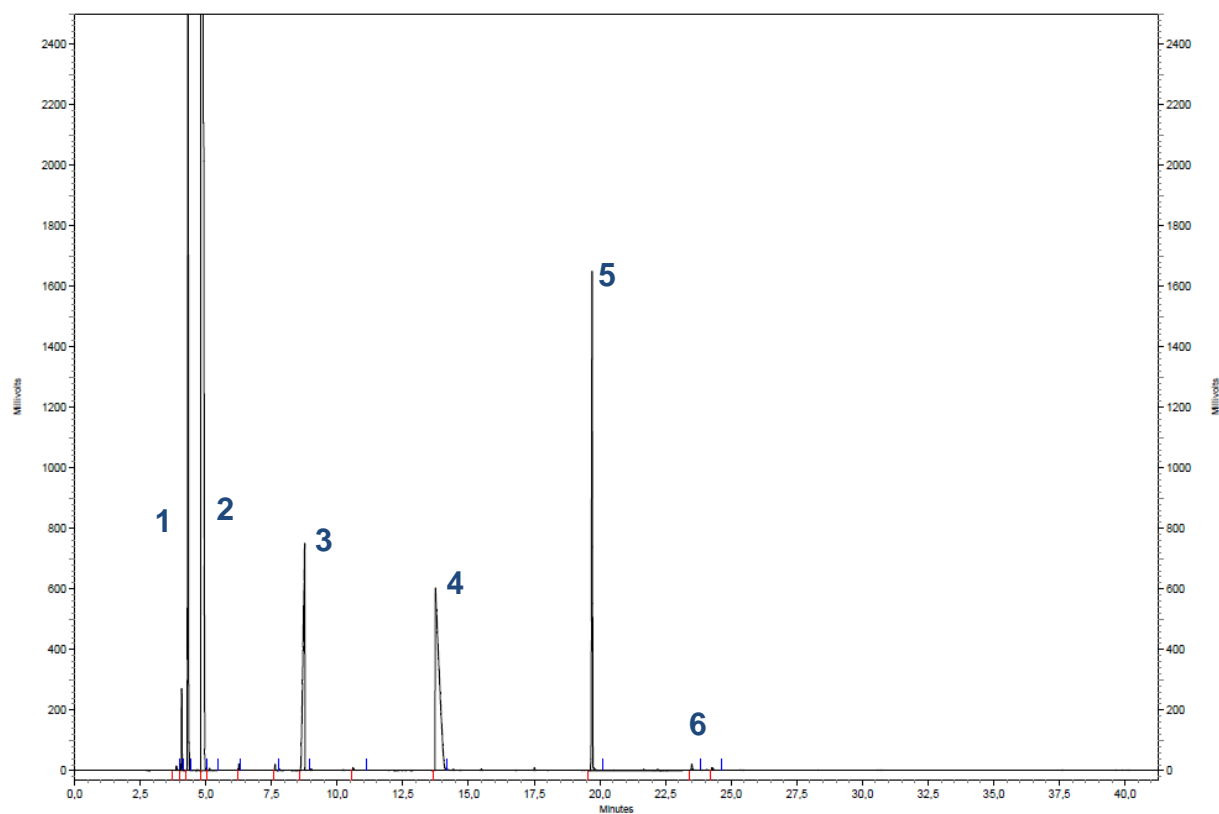
Entry	Retention Time [min]	Substance	Area
1	4.310	CE	66855521
2	4.790	CH ₂ Cl ₂ (Solv.)	535246491
3	8.753	<i>n</i> -Dodecane (Stand.)	34948061
4	13.715	CH ₃ COOH (Solv.)	71609064
5	19.708	1-Phenylethanol (Stand.)	32276946

Entry 4



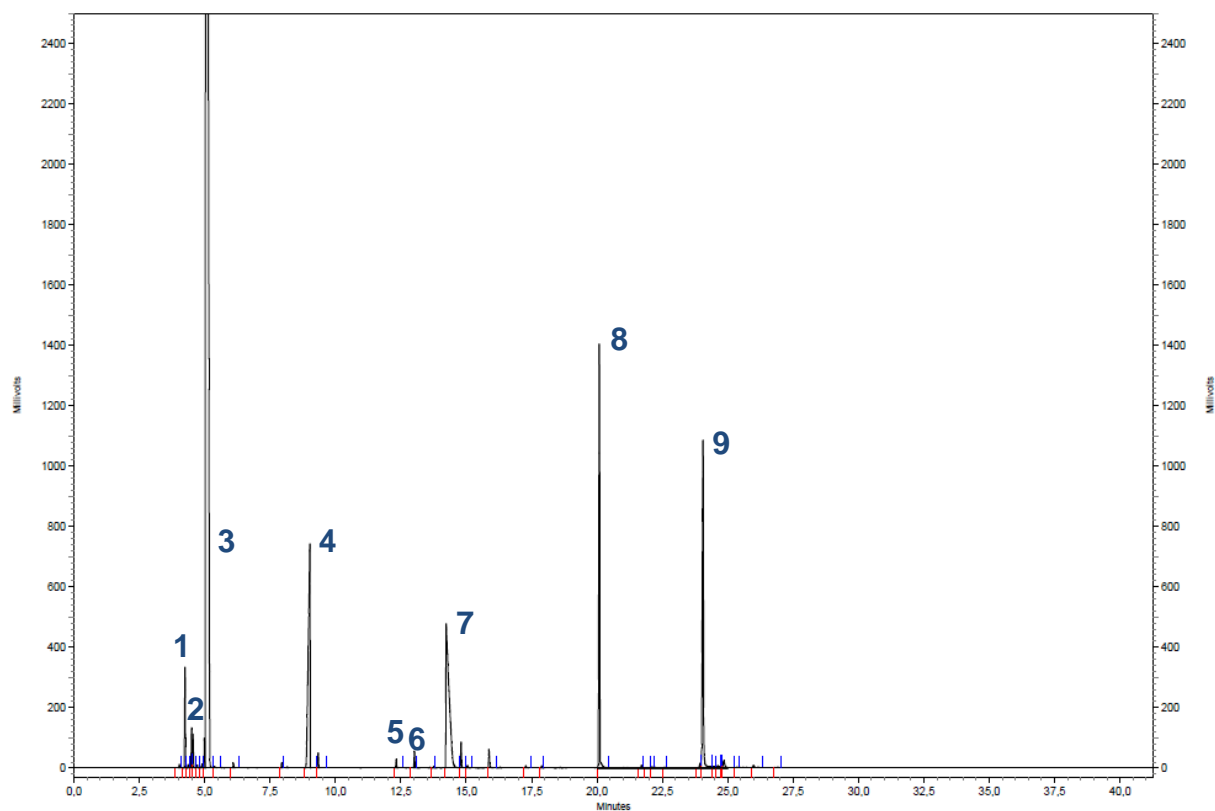
Entry	Retention Time [min]	Substance	Area
1	4.313	CE	48194395
2	4.792	CH ₂ Cl ₂ (Solv.)	538476658
3	8.752	<i>n</i> -Dodecane (Stand.)	32378855
4	11.983	CAC	6877451
5	12.687	CI	2662651
6	13.750	CH ₃ COOH (Solv.)	59668788
7	19.710	1-Phenylethanol (Stand.)	25406361
8	23.525	CA	423929

Entry 5



Entry	Retention Time [min]	Substance	Area
1	4.320	CE	53071059
2	4.798	CH ₂ Cl ₂ (Solv.)	544635844
3	8.765	<i>n</i> -Dodecane (Stand.)	35663246
4	13.740	CH ₃ COOH (Solv.)	62411720
5	19.707	1-Phenylethanol (Stand.)	32192435
6	23.292	CA	350399

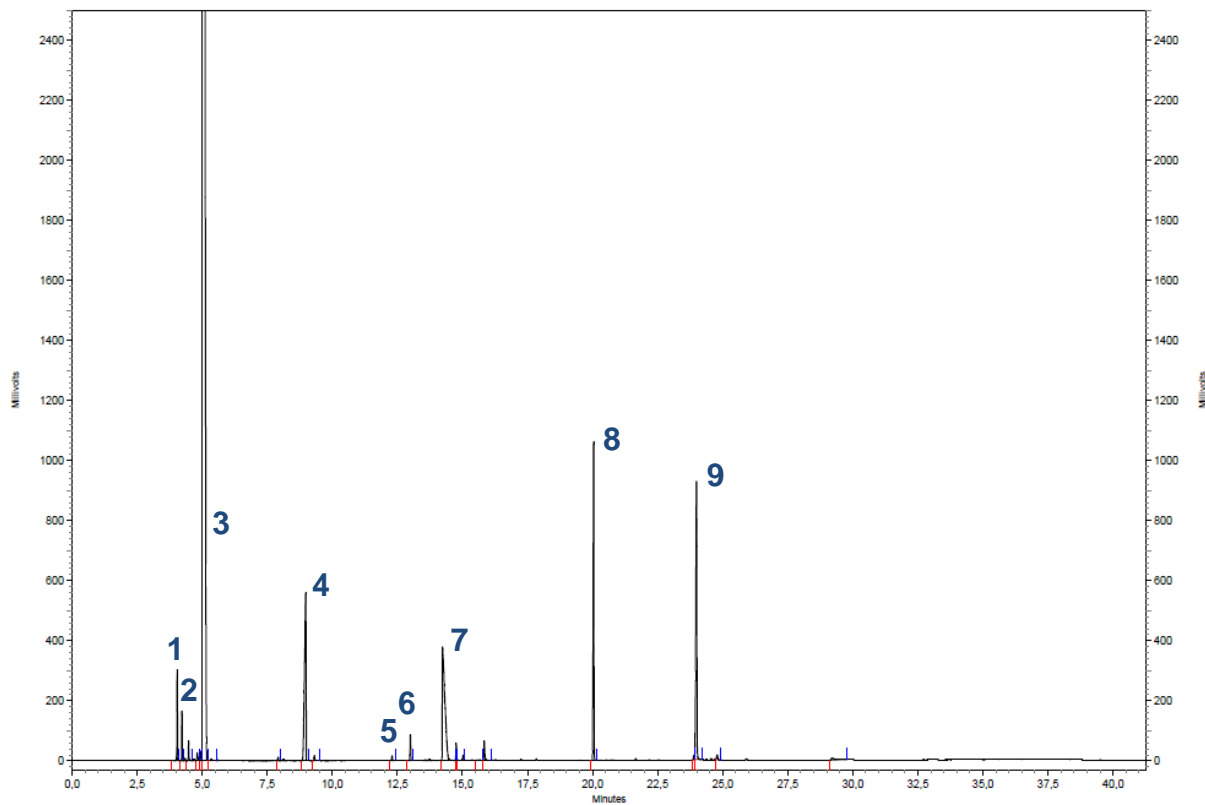
Entry 6



Entry	Retention Time [min]	Substance	Area
1	4.245	CH	4433844
2	4.502	CE	2031212
3	5.028	CH ₂ Cl ₂ (Solv.)	522589517
4	9.023	<i>n</i> -Dodecane (Stand.)	33671343
5	12.323	CAc	529990
6	13.022	CI	1042924
7	14.232	CH ₃ COOH (Solv.)	39395166
8	20.090	1-Phenylethanol (Stand.)	28037869
9	24.053	CA	30891976

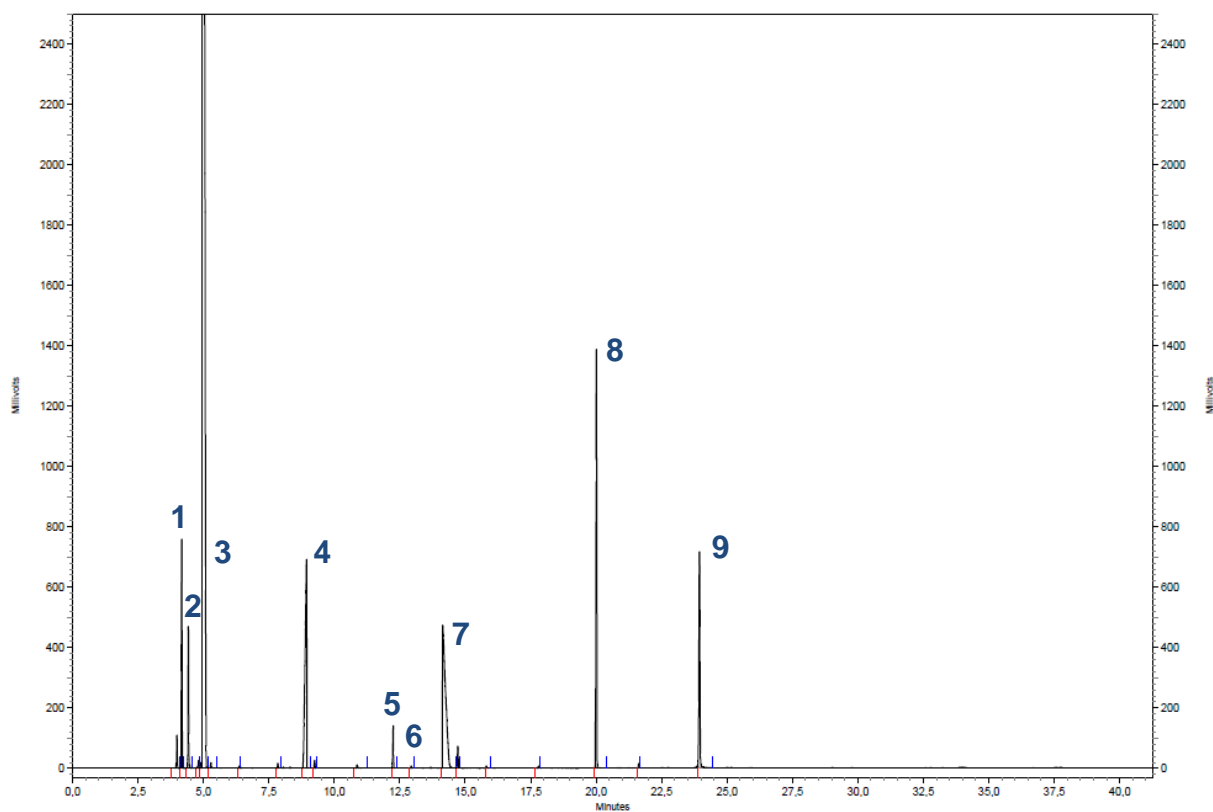
S3.2 Gaschromatograms to Table S2.2

Entry 1



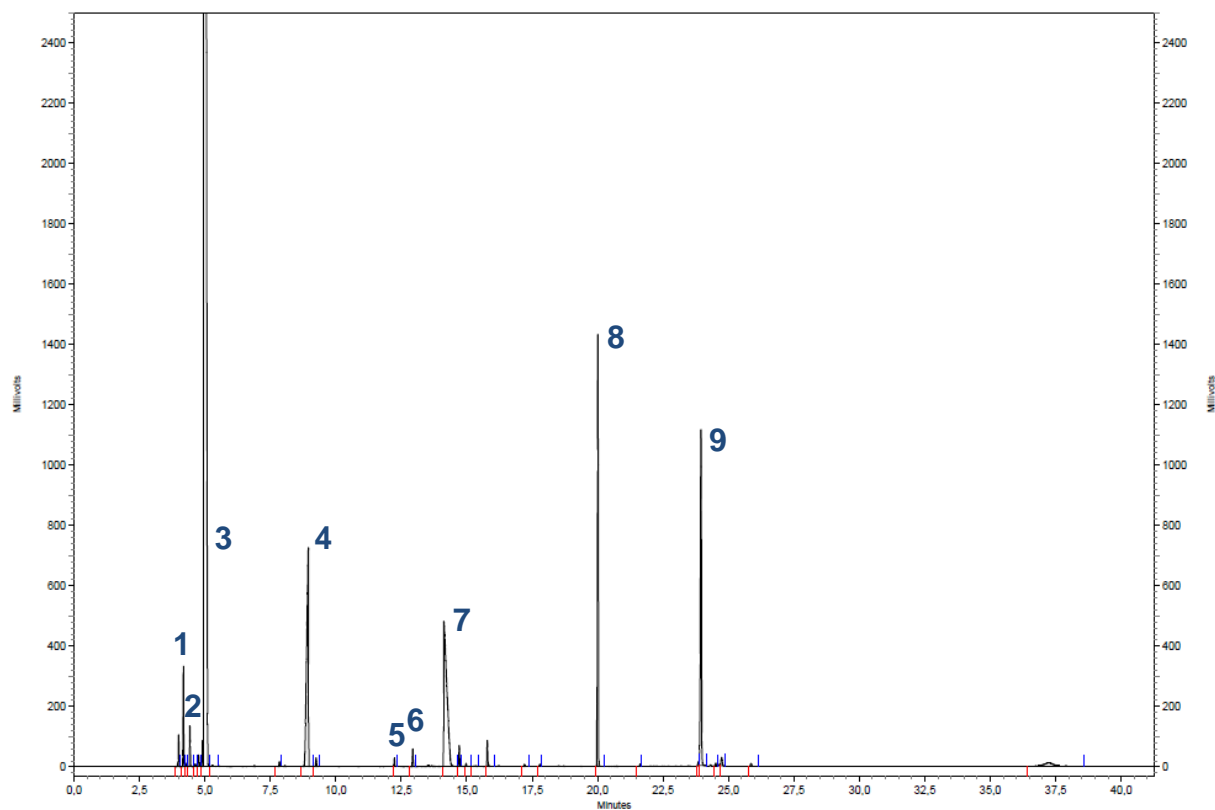
Entry	Retention Time [min]	Substance	Area
1	4.222	CH	1991545
2	4.477	CE	927542
3	5.017	CH ₂ Cl ₂ (Solv.)	392085582
4	8.980	<i>n</i> -Dodecane (Stand.)	20293940
5	12.293	CAc	267475
6	13.000	CI	1517506
7	14.235	CH ₃ COOH (Solv.)	25382742
8	20.038	1-Phenylethanol (Stand.)	18798006
9	23.992	CA	22530242

Entry 2



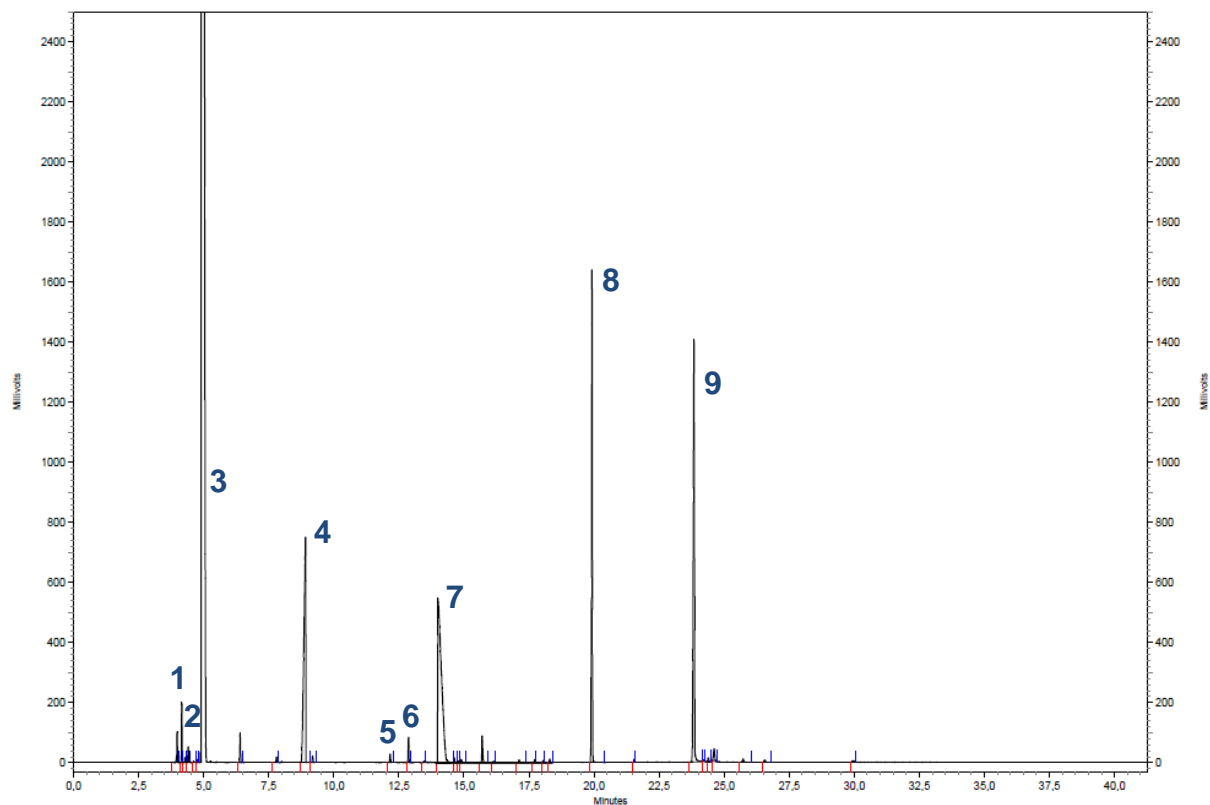
Entry	Retention Time [min]	Substance	Area
1	4.170	CH	10035533
2	4.422	CE	7333320
3	4.930	CH ₂ Cl ₂ (Solv.)	565209382
4	8.943	<i>n</i> -Dodecane (Stand.)	30628600
5	12.245	CAc	2297911
6	12.937	CI	130155
7	14.135	CH ₃ COOH (Solv.)	38394893
8	20.012	1-Phenylethanol (Stand.)	26627134
9	23.948	CA	16921950

Entry 3



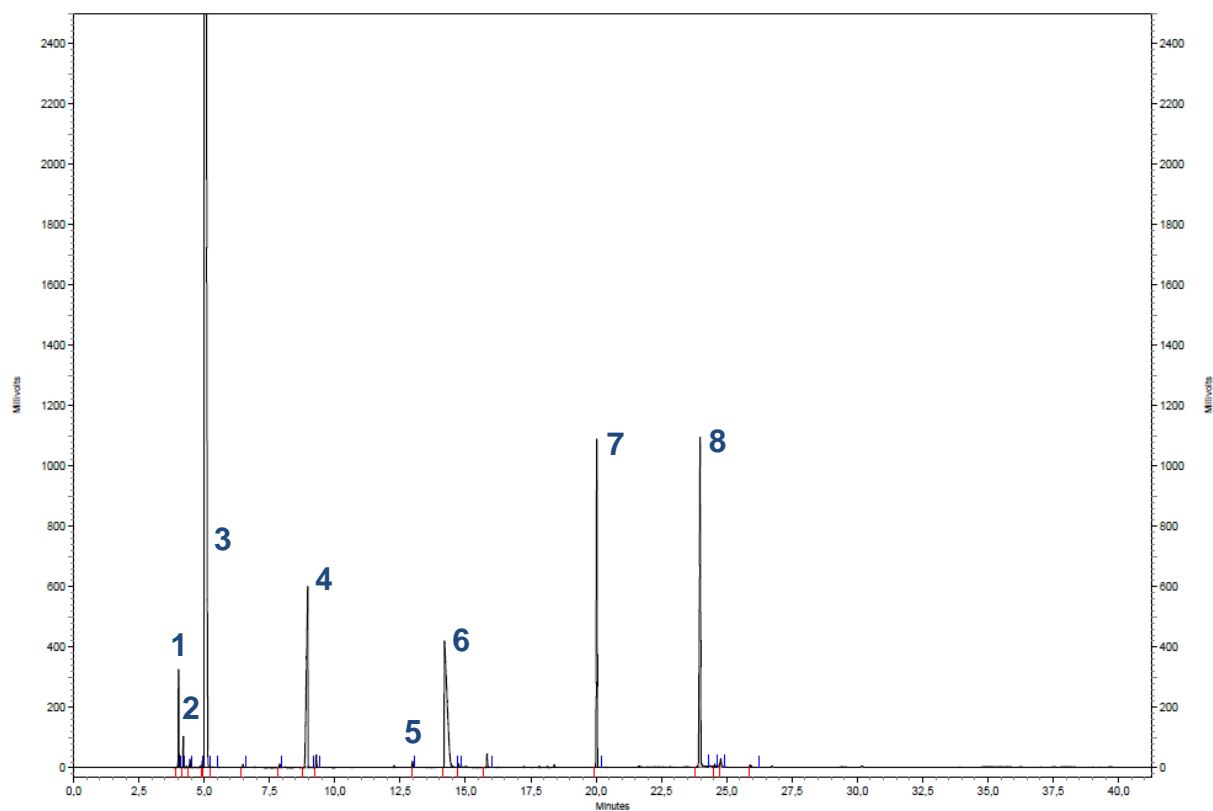
Entry	Retention Time [min]	Substance	Area
1	4.177	CH	4374561
2	4.428	CE	2088590
3	4.938	CH ₂ Cl ₂ (Solv.)	551017028
4	8.952	<i>n</i> -Dodecane (Stand.)	33888780
5	12.238	CAc	444846
6	12.942	CI	1018953
7	14.128	CH ₃ COOH (Solv.)	39703195
8	20.012	1-Phenylethanol (Stand.)	27529373
9	23.953	CA	28584714

Entry 4



Entry	Retention Time [min]	Substance	Area
1	4.158	CH	2679301
2	4.407	CE	848160
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.915	<i>n</i> -Dodecane (Stand.)	35139425
5	12.163	CAc	456082
6	12.873	CI	1462056
7	14.005	CH ₃ COOH (Solv.)	52063194
8	19.925	1-Phenylethanol (Stand.)	32487728
9	23.840	CA	40158028

Entry 5



Entry	Retention Time [min]	Substance	Area
1	4.200	CH	1272338
2	4.453	CE	408694
3	4.988	CH ₂ Cl ₂ (Solv.)	425538821
4	8.963	<i>n</i> -Dodecane (Stand.)	22961031
5	12.972	CI	360050
6	14.195	CH ₃ COOH (Solv.)	31496254
7	20.025	1-Phenylethanol (Stand.)	19652325
8	23.982	CA	28968611

Data of the isolated product to Entry 5

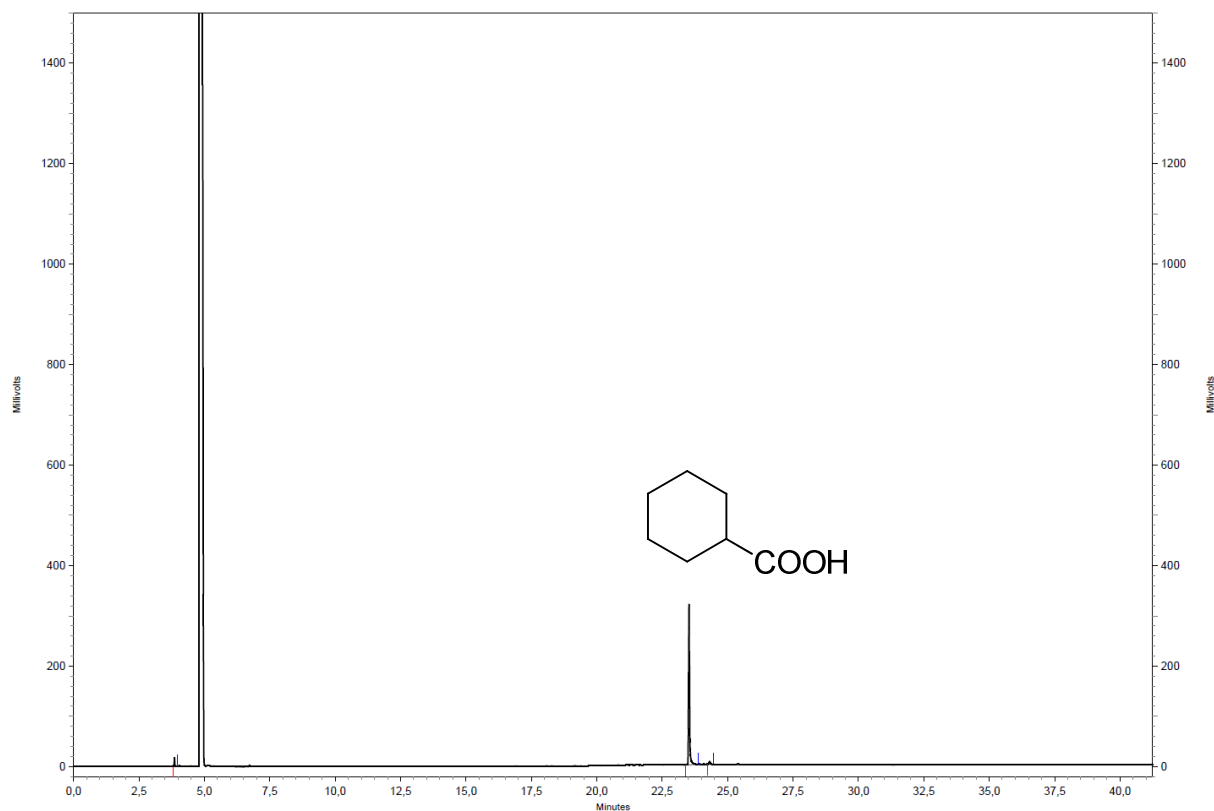


Figure 3.1. GC chromatogram of the isolated cyclohexane carboxylic acid.

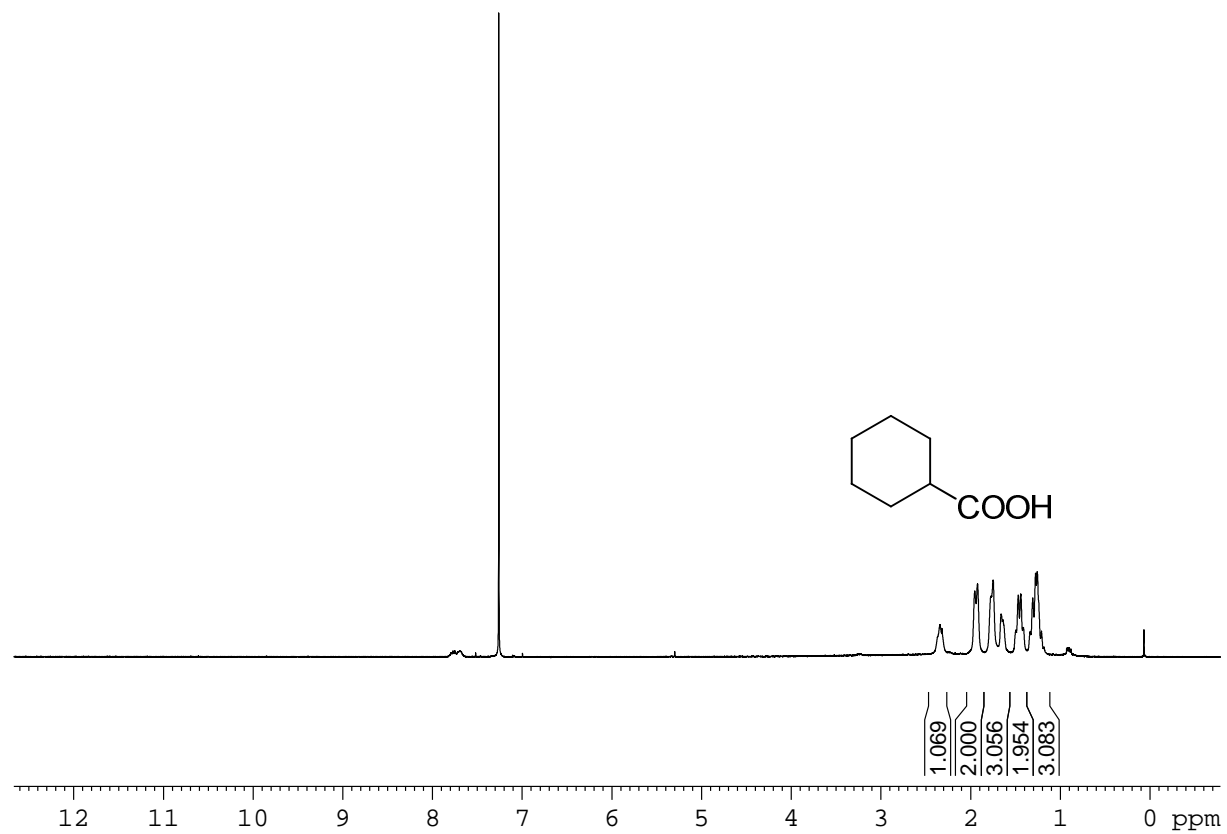
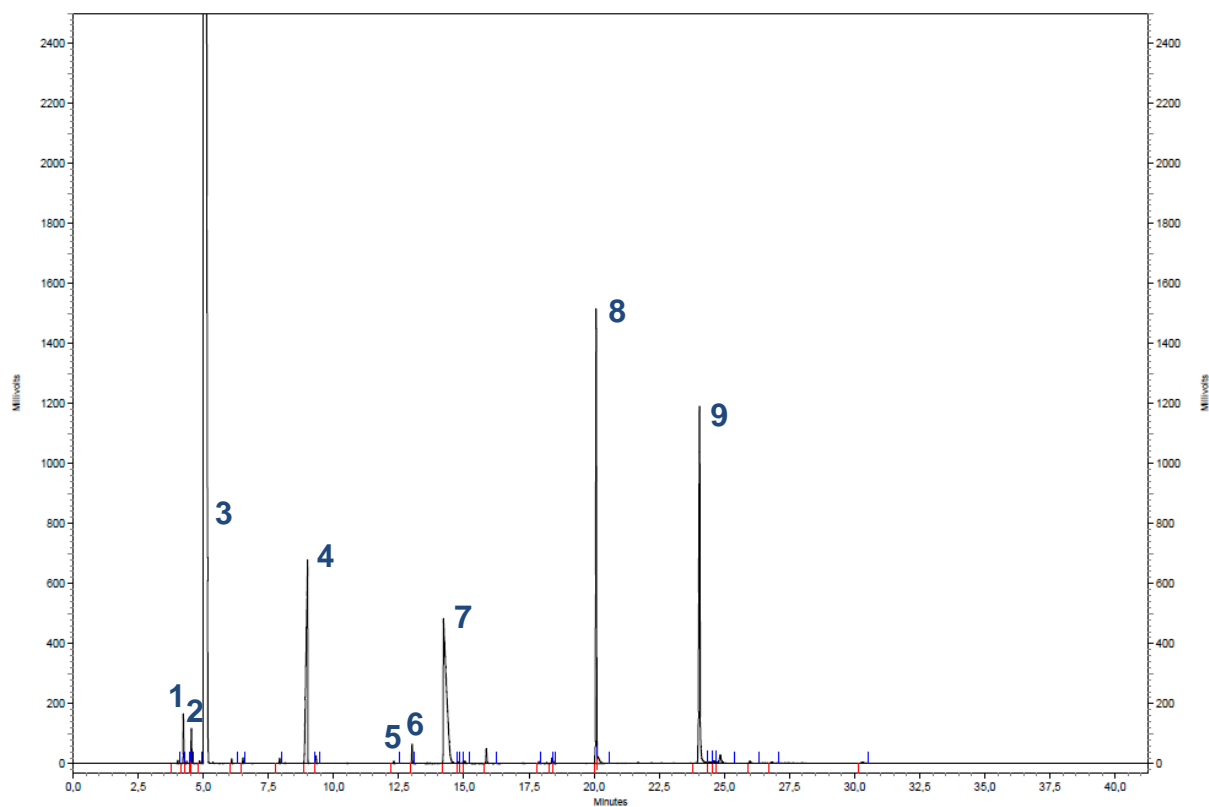


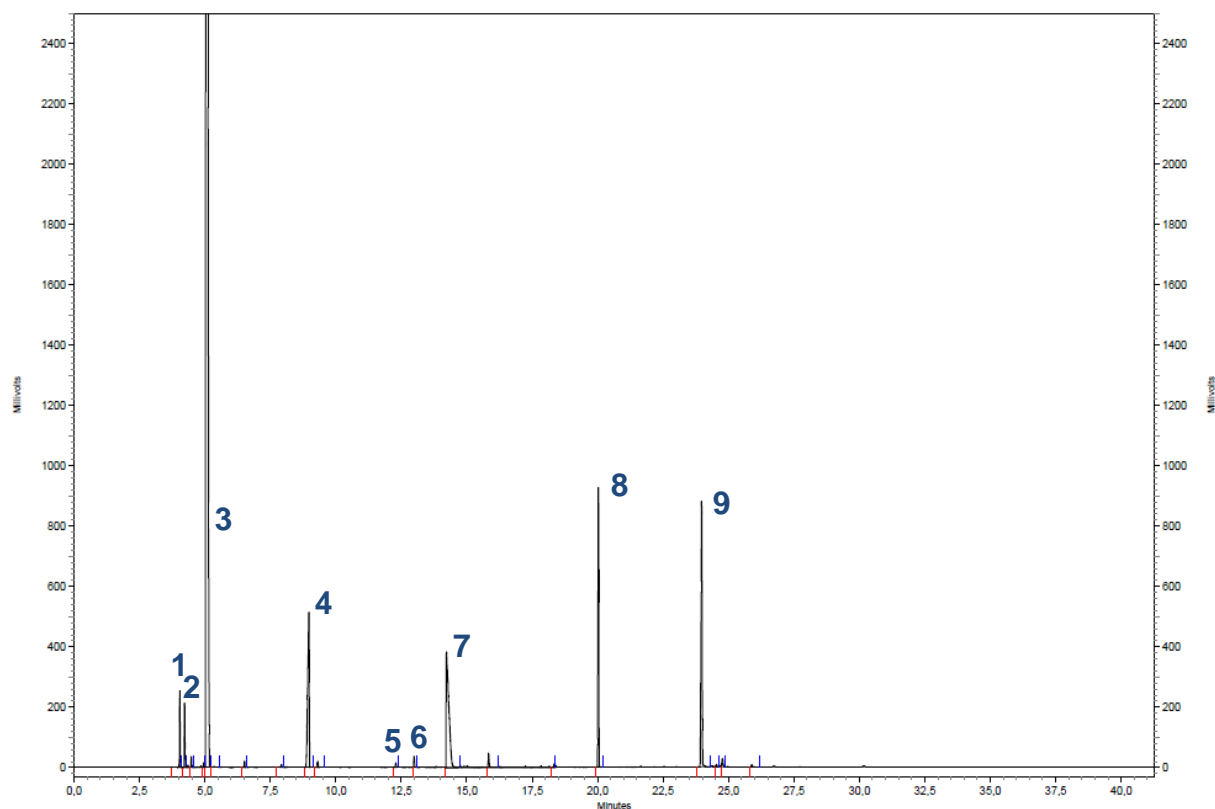
Figure 3.2. ¹H NMR spectrum of the isolated product measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

Entry 6



Entry	Retention Time [min]	Substance	Area
1	4.235	CH	2177669
2	4.542	CE	1783739
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	9.005	<i>n</i> -Dodecane (Stand.)	29490788
5	12.312	CAc	161523
6	13.012	CI	1204964
7	14.213	CH ₃ COOH (Solv.)	41830295
8	20.083	1-Phenylethanol (Stand.)	29884818
9	24.048	CA	35487518

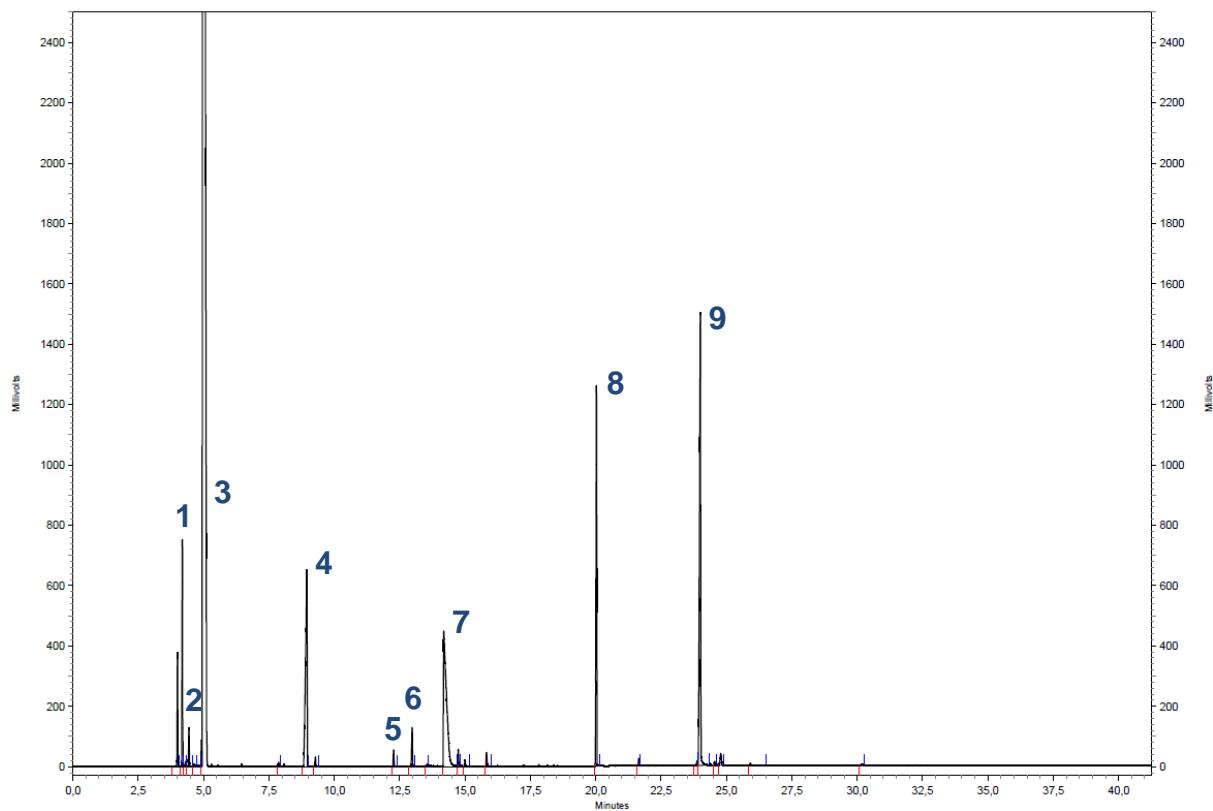
Entry 7



Entry	Retention Time [min]	Substance	Area
1	4.227	CH	2539880
2	4.480	CE	485270
3	5.032	CH ₂ Cl ₂ (Solv.)	329568598
4	8.973	<i>n</i> -Dodecane (Stand.)	17638402
5	12.290	CAc	247847
6	12.993	CI	622962
7	14.225	CH ₃ COOH (Solv.)	26212605
8	20.030	1-Phenylethanol (Stand.)	16186723
9	23.975	CA	21811311

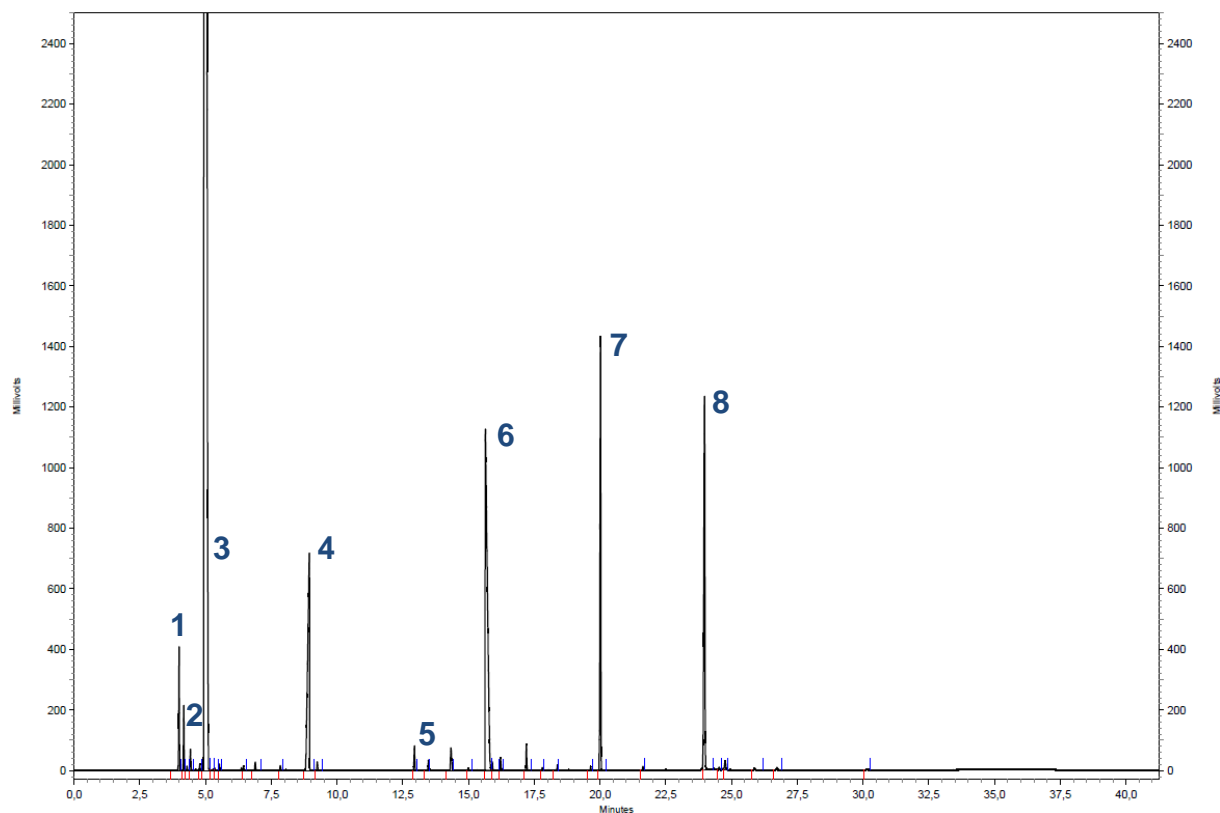
S3.3 Gaschromatograms to Table S2.3

Entry 1



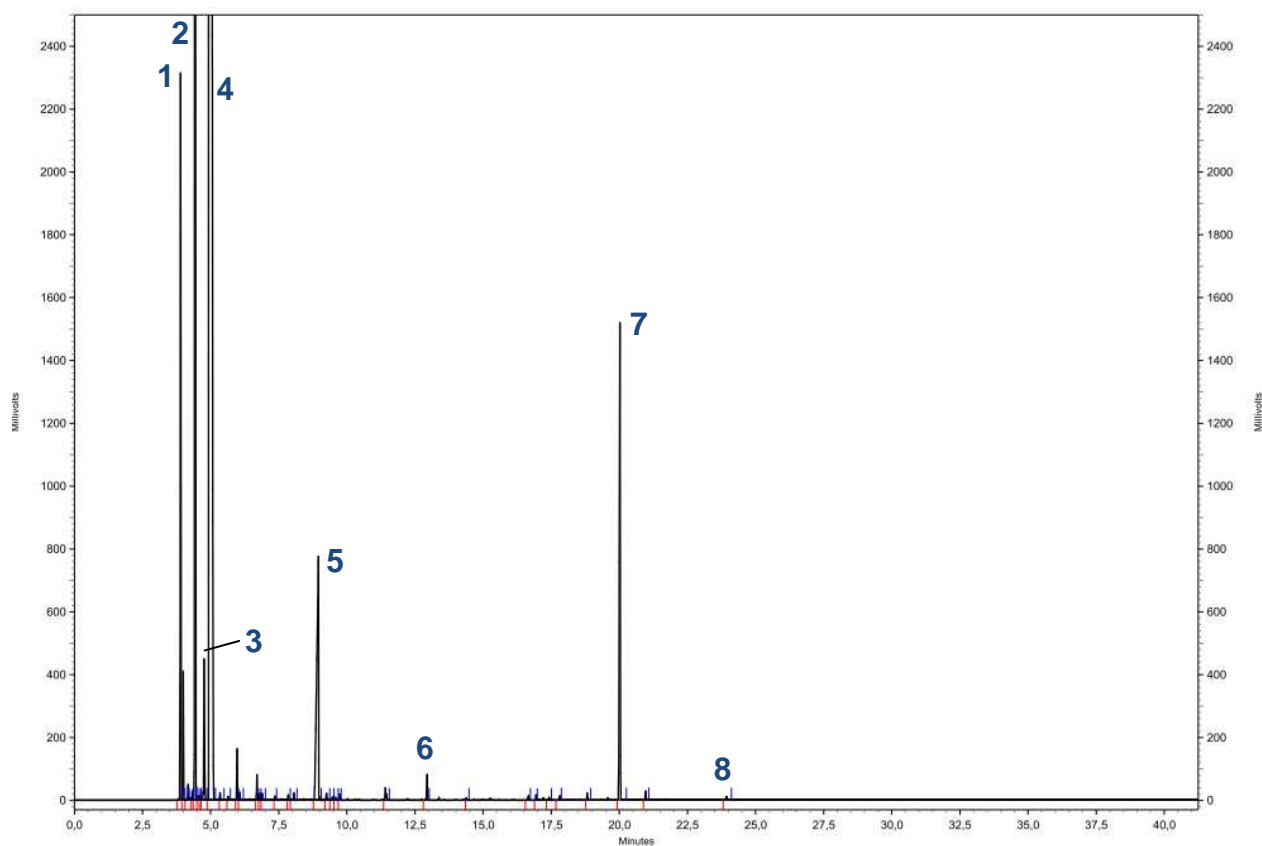
Entry	Retention Time [min]	Substance	Area
1	4.183	CH	9605788
2	4.438	CE	1889030
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.955	<i>n</i> -Dodecane (Stand.)	26190459
5	12.268	CAc	890076
6	12.973	CI	2287680
7	14.185	CH ₃ COOH (Solv.)	35089734
8	20.027	1-Phenylethanol (Stand.)	23278985
9	23.998	CA	43927273

Entry 2



Entry	Retention Time [min]	Substance	Area
1	4.172	CH	2823840
2	4.425	CE	1093745
3	4.935	CH ₂ Cl ₂ (Solv.)	554829493
4	8.947	<i>n</i> -Dodecane (Stand.)	32640783
5	12.945	CI	1468257
6	15.624	Propionic acid (Solv.)	63238146
7	20.018	1-Phenylethanol (Stand.)	27675391
8	23.973	CA	33090337

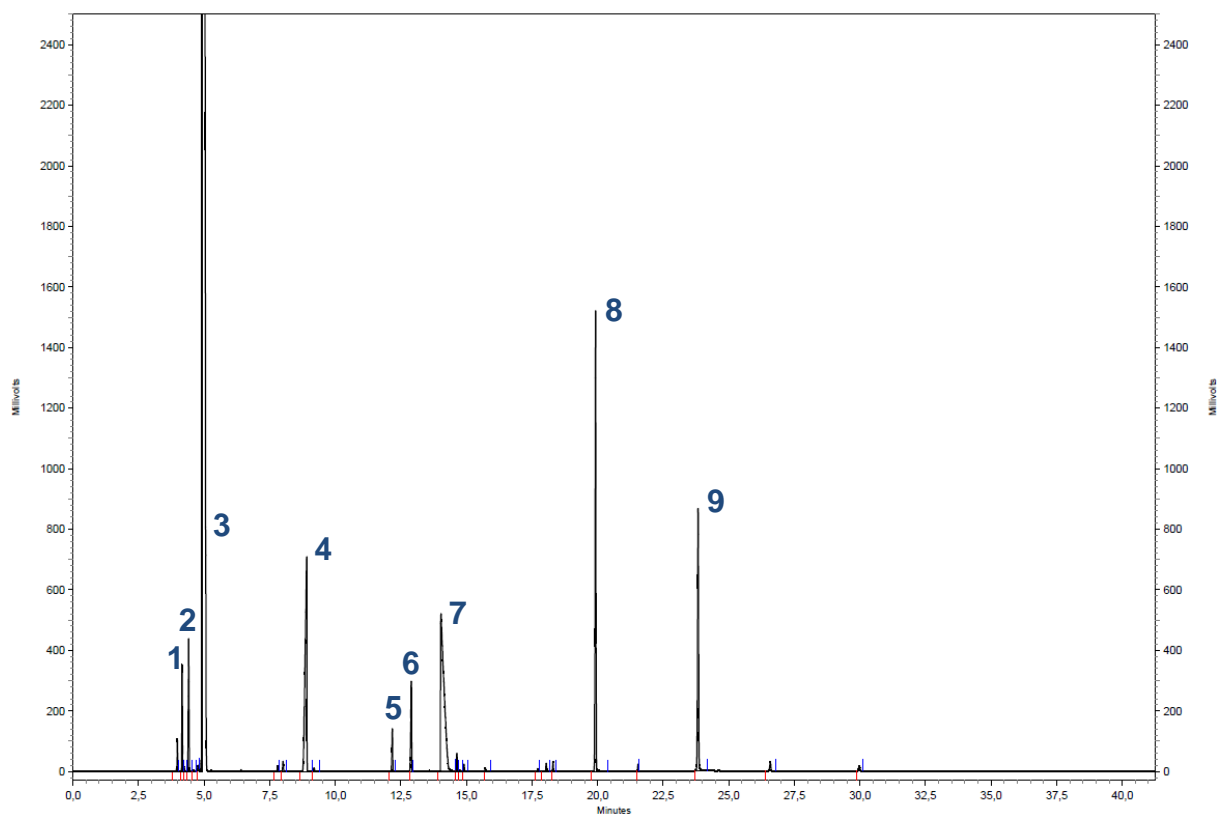
Entry 3



Entry	Retention Time [min]	Substance	Area
1	4.175	CH	689970
2	4.430	THF (Solv.)	102123861
3	4.520	CE	473567
4	4.938	CH ₂ Cl ₂ (Solv.)	555651727
5	8.960	<i>n</i> -Dodecane (Stand.)	36062420
6	12.945	CI	1450316
7	20.020	1-Phenylethanol (Stand.)	29402117
8	23.932	CA	239182

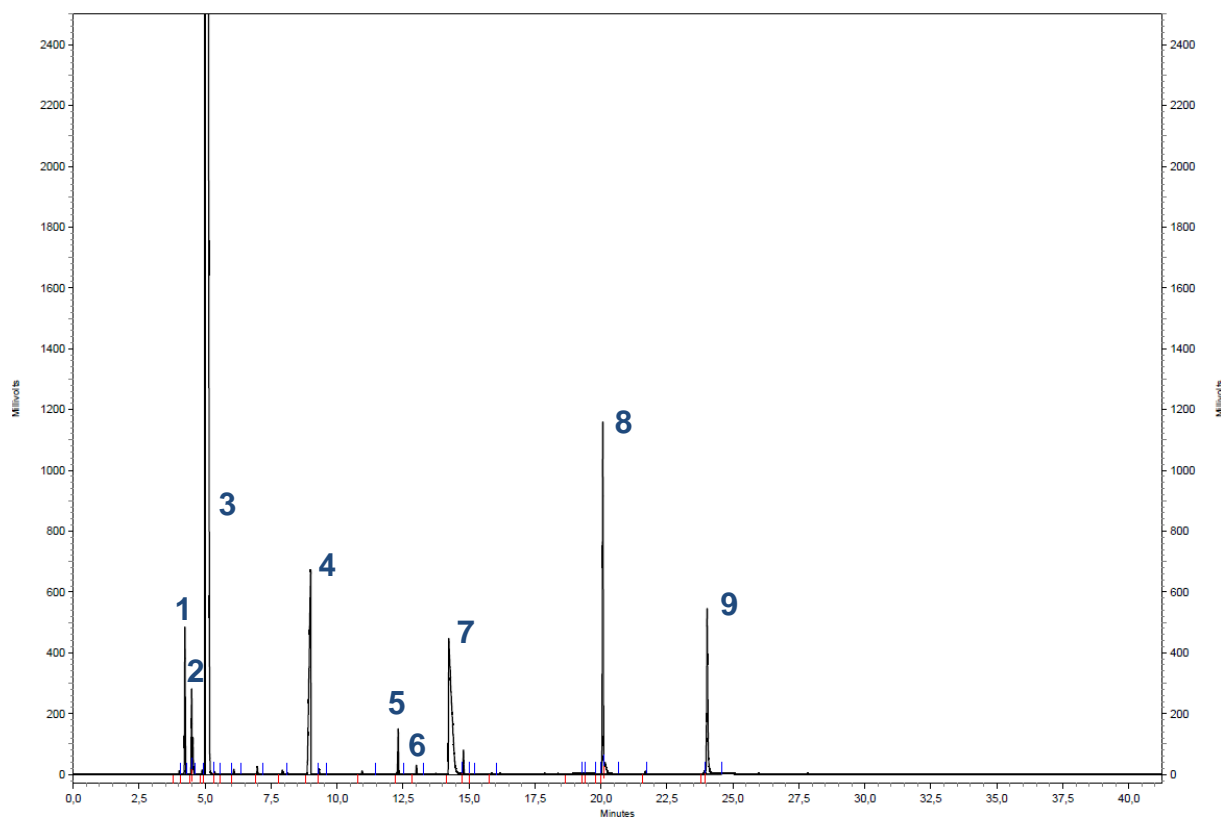
S3.4 Gaschromatograms to Table S2.4

Entry 1



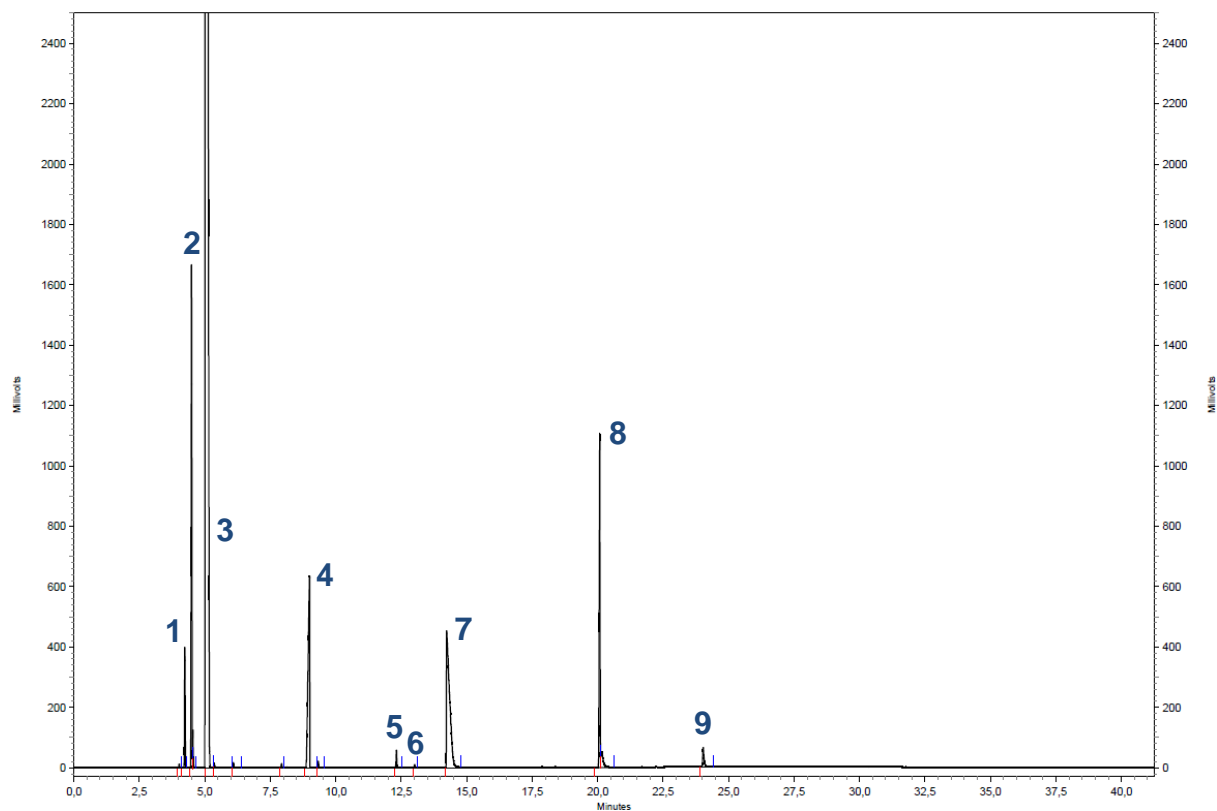
Entry	Retention Time [min]	Substance	Area
1	4.162	CH	4623719
2	4.412	CE	6724551
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.910	<i>n</i> -Dodecane (Stand.)	30930081
5	12.173	CAC	2314710
6	12.898	CI	6311692
7	14.028	CH ₃ COOH (Solv.)	46117037
8	19.927	1-Phenylethanol (Stand.)	29236760
9	23.833	CA	21649809

Entry 2



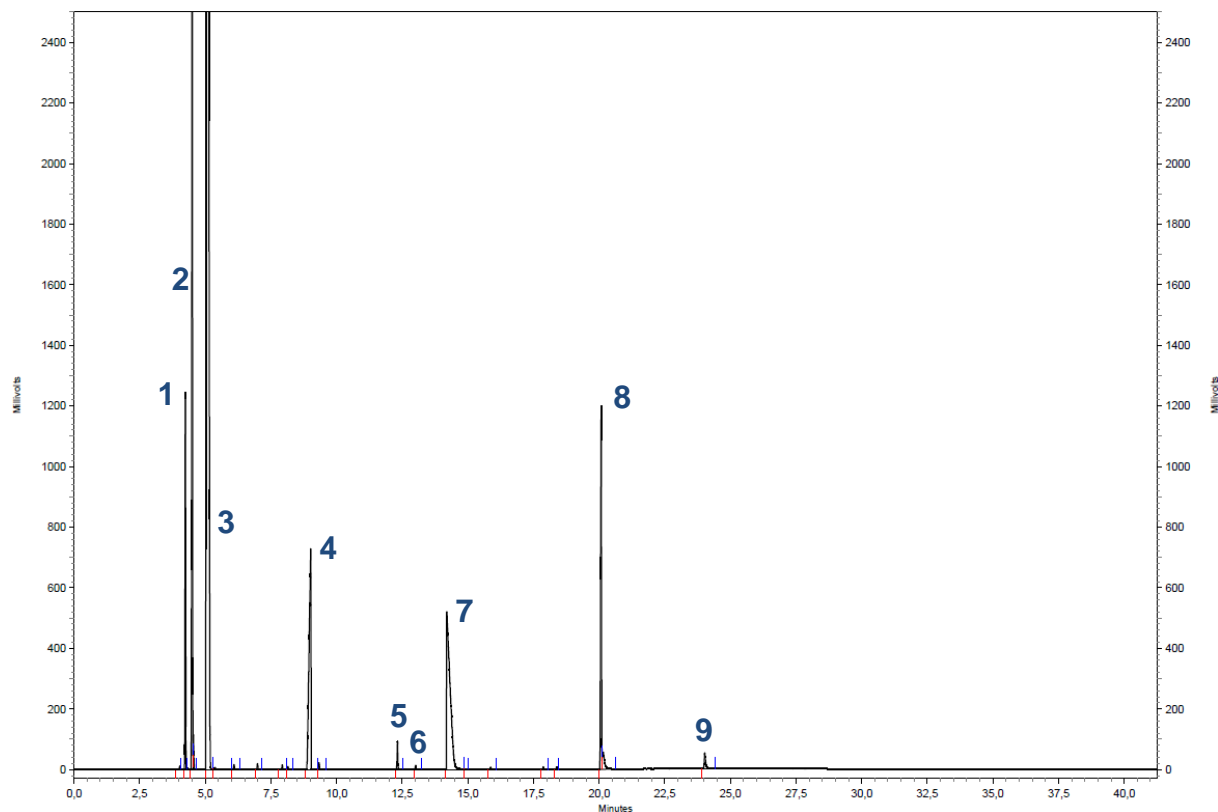
Entry	Retention Time [min]	Substance	Area
1	4.233	CH	6407863
2	4.490	CE	4261870
3	5.012	CH ₂ Cl ₂ (Solv.)	538587171
4	9.002	<i>n</i> -Dodecane (Stand.)	29123450
5	12.315	CAc	2617854
6	13.007	CI	553475
7	14.233	CH ₃ COOH (Solv.)	35607530
8	20.073	1-Phenylethanol (Stand.)	21793779
9	24.027	CA	16019251

Entry 3



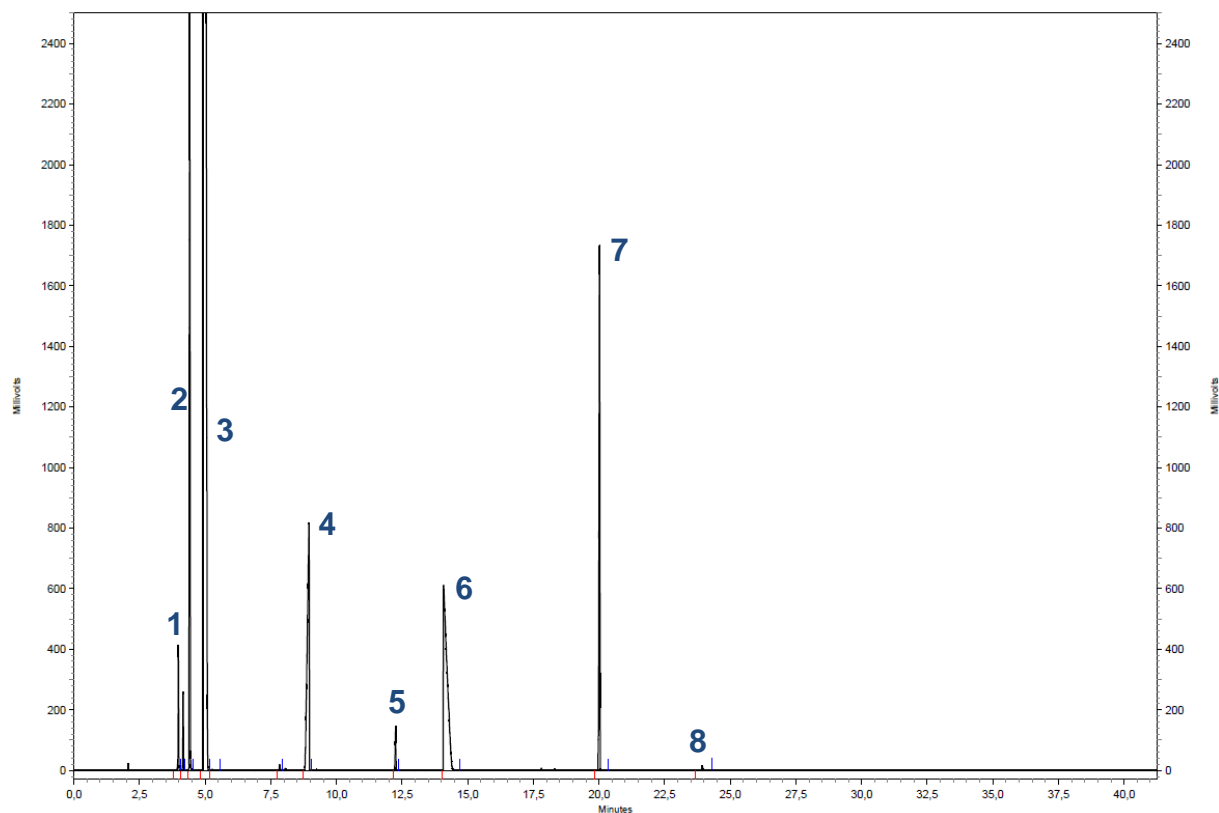
Entry	Retention Time [min]	Substance	Area
1	4.235	CH	5144918
2	4.492	CE	25230166
3	5.013	CH ₂ Cl ₂ (Solv.)	544970234
4	8.998	<i>n</i> -Dodecane (Stand.)	26142032
5	12.317	CAc	991745
6	13.010	CI	195450
7	14.233	CH ₃ COOH (Solv.)	37599694
8	20.078	1-Phenylethanol (Stand.)	20853886
9	24.028	CA	2439710

Entry 4



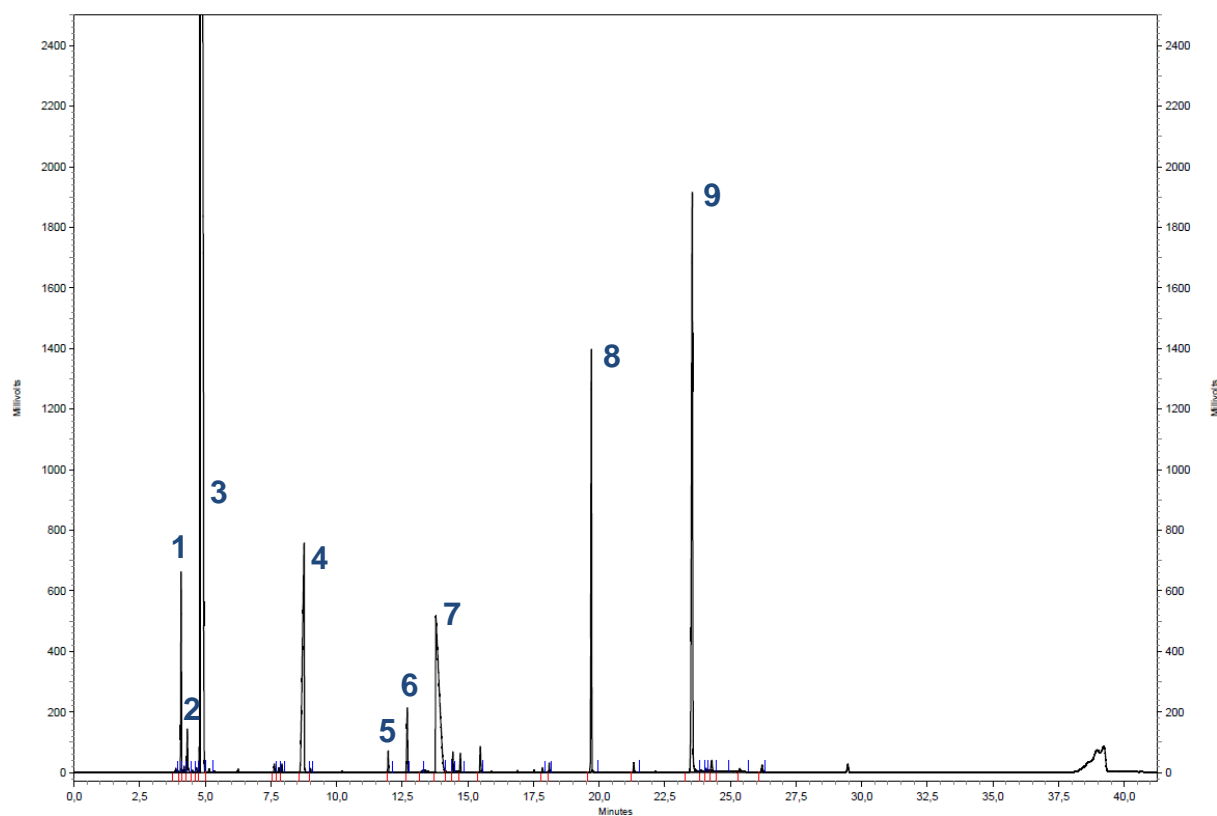
Entry	Retention Time [min]	Substance	Area
1	4.240	CH	16346362
2	4.498	CE	55861908
3	5.022	CH ₂ Cl ₂ (Solv.)	522642505
4	9.015	<i>n</i> -Dodecane (Stand.)	32957009
5	12.320	CAc	1666006
6	13.010	CI	265242
7	14.200	CH ₃ COOH (Solv.)	47136194
8	20.075	1-Phenylethanol (Stand.)	23201249
9	24.025	CA	1839809

Entry 5



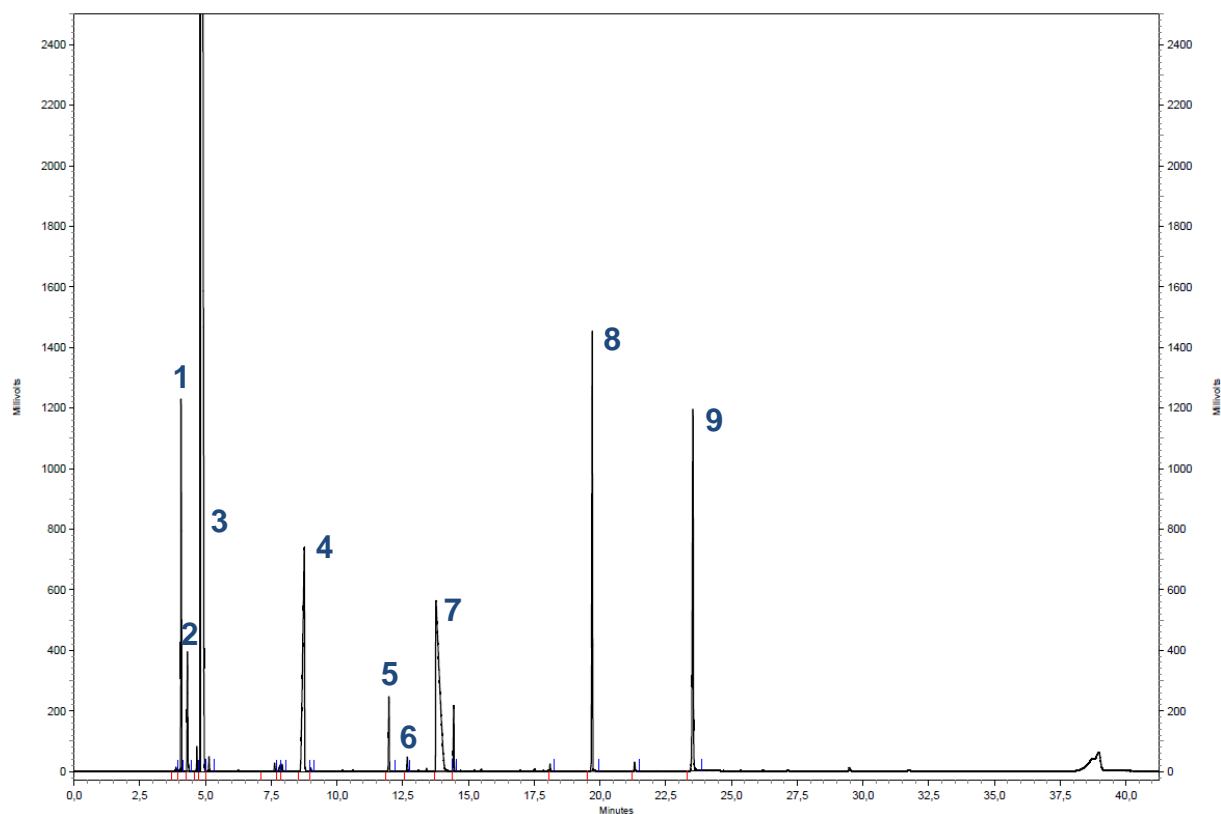
Entry	Retention Time [min]	Substance	Area
1	4.158	CH	3365695
2	4.412	CE	46005793
3	4.922	CH ₂ Cl ₂ (Solv.)	544941656
4	8.960	<i>n</i> -Dodecane (Stand.)	40741507
5	12.252	CAC	2408441
6	14.080	CH ₃ COOH (Solv.)	58967864
7	20.022	1-Phenylethanol (Stand.)	34575327
8	23.937	CA	356243

Entry 6



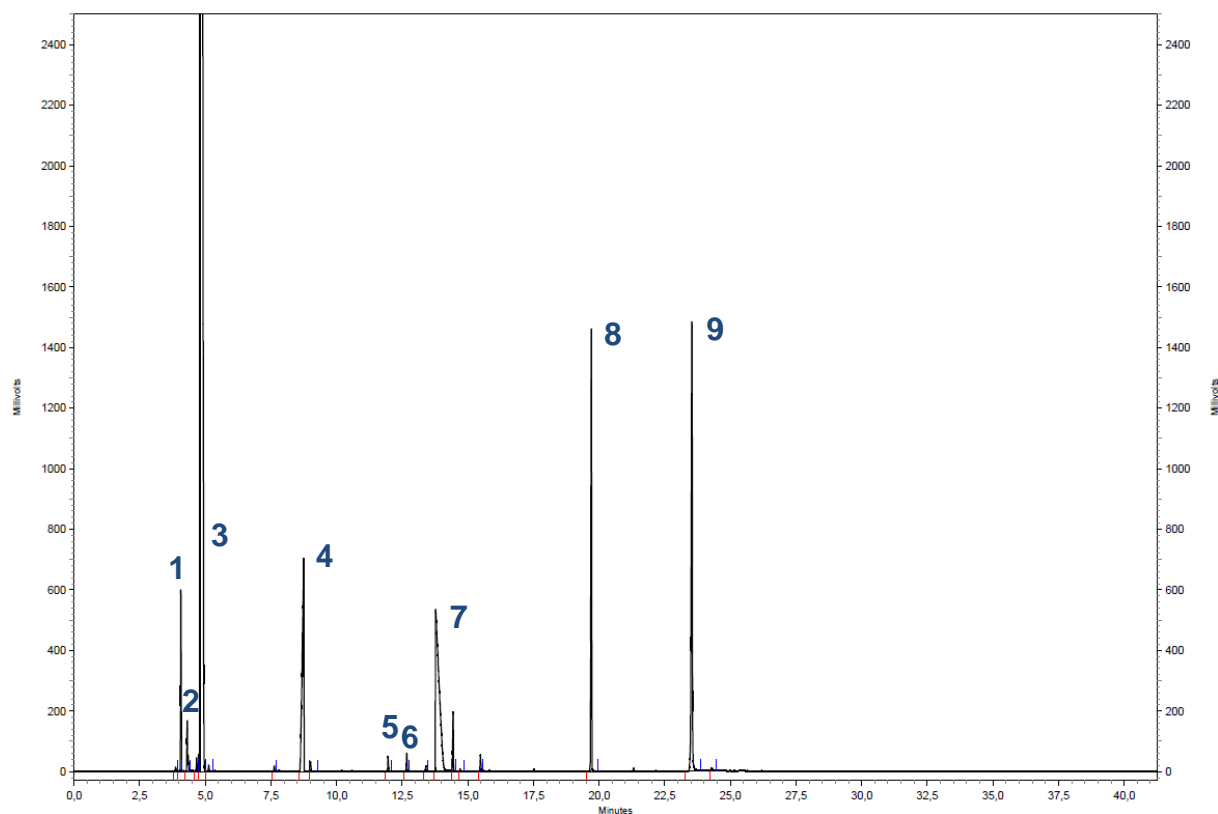
Entry	Retention Time [min]	Substance	Area
1	4.073	CH	8443705
2	4.313	CE	2321799
3	4.795	CH ₂ Cl ₂ (Solv.)	53904642
4	8.762	<i>n</i> -Dodecane (Stand.)	35617834
5	11.963	CAc	1153530
6	12.692	CI	4076805
7	13.777	CH ₃ COOH (Solv.)	47859651
8	19.705	1-Phenylethanol (Stand.)	25810587
9	23.548	CA	57425083

Entry 7



Entry	Retention Time [min]	Substance	Area
1	4.072	CH	15599781
2	4.312	CE	6099375
3	4.792	CH ₂ Cl ₂ (Solv.)	544760547
4	8.758	<i>n</i> -Dodecane (Stand.)	34454803
5	11.977	CAc	4301103
6	12.675	CI	813872
7	13.757	CH ₃ COOH (Solv.)	56141915
8	19.705	1-Phenylethanol (Stand.)	27244280
9	23.535	CA	30588932

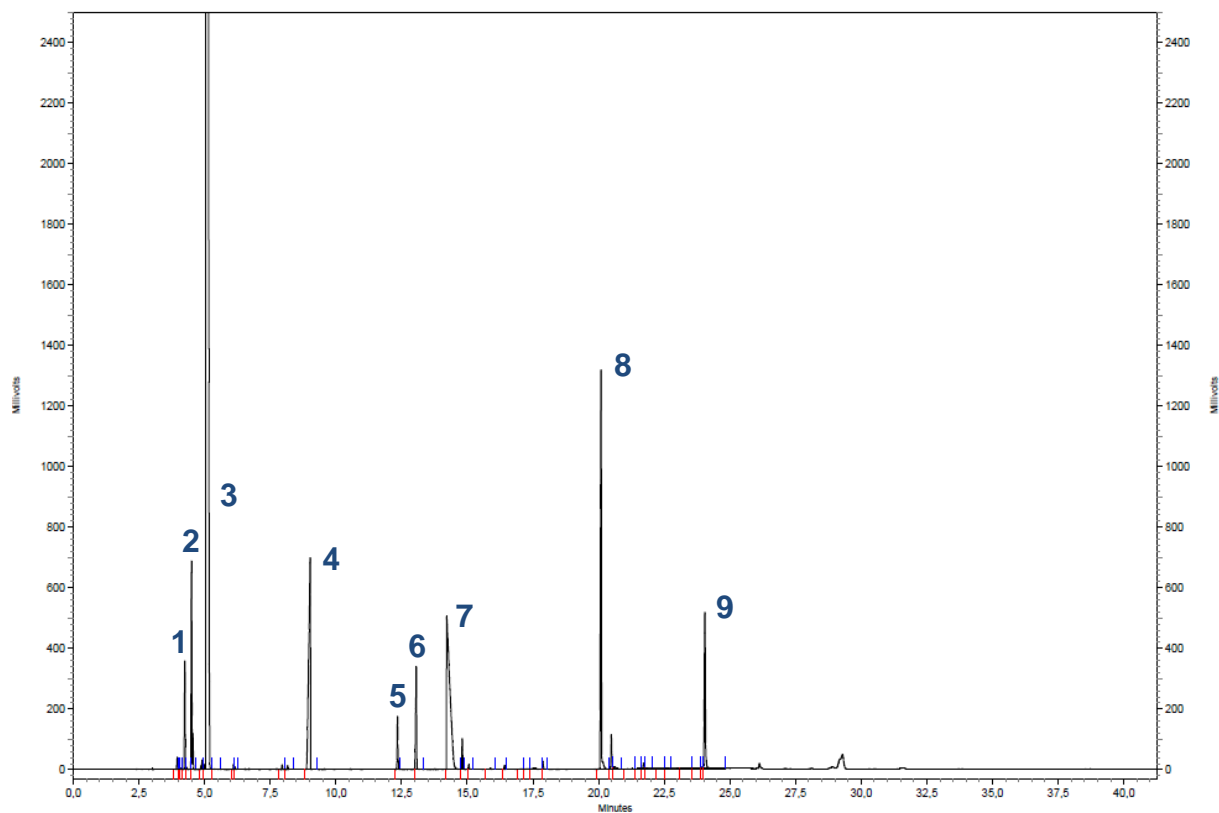
Entry 8



Entry	Retention Time [min]	Substance	Area
1	4.070	CH	7688845
2	4.310	CE	3263295
3	4.788	CH ₂ Cl ₂ (Solv.)	552348130
4	8.753	<i>n</i> -Dodecane (Stand.)	31910631
5	11.962	CAC	787350
6	12.675	CI	1050554
7	13.773	CH ₃ COOH (Solv.)	50790928
8	19.705	1-Phenylethanol (Stand.)	27606546
9	23.537	CA	40727922

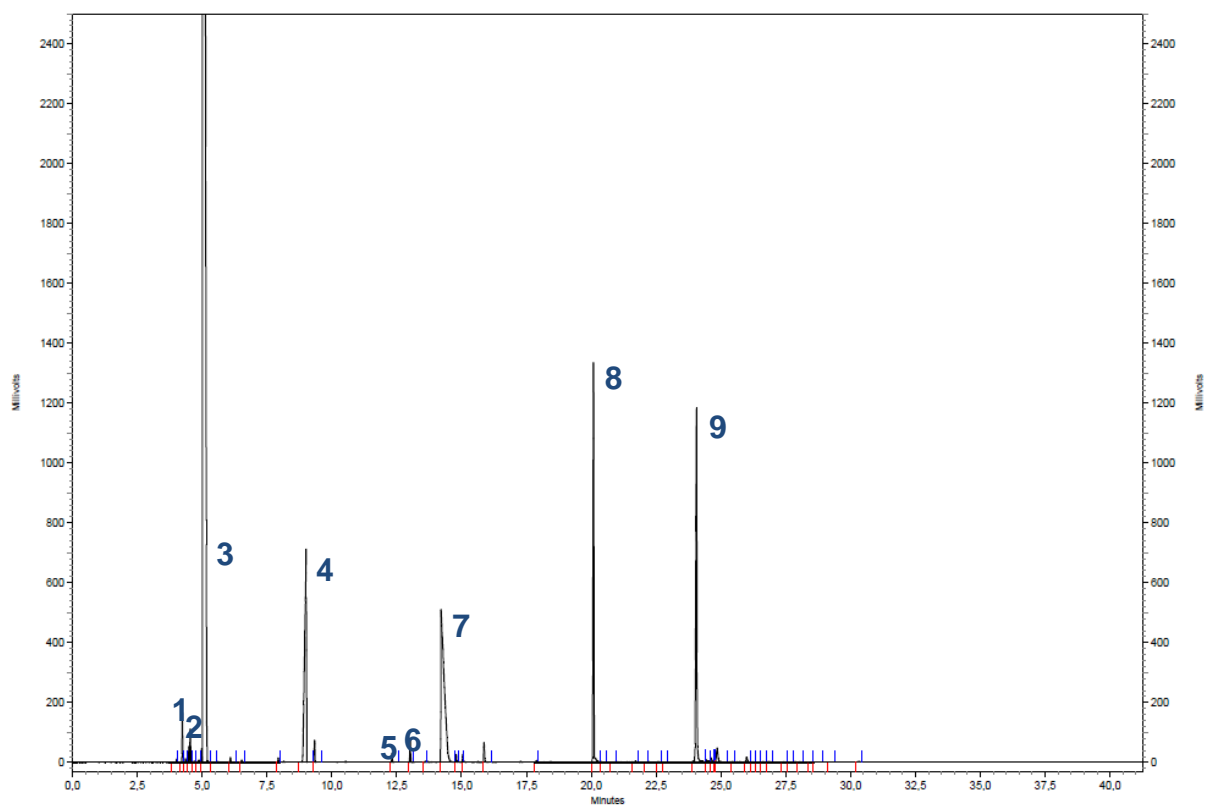
S3.5 Gaschromatograms to Table S2.5

Entry 1



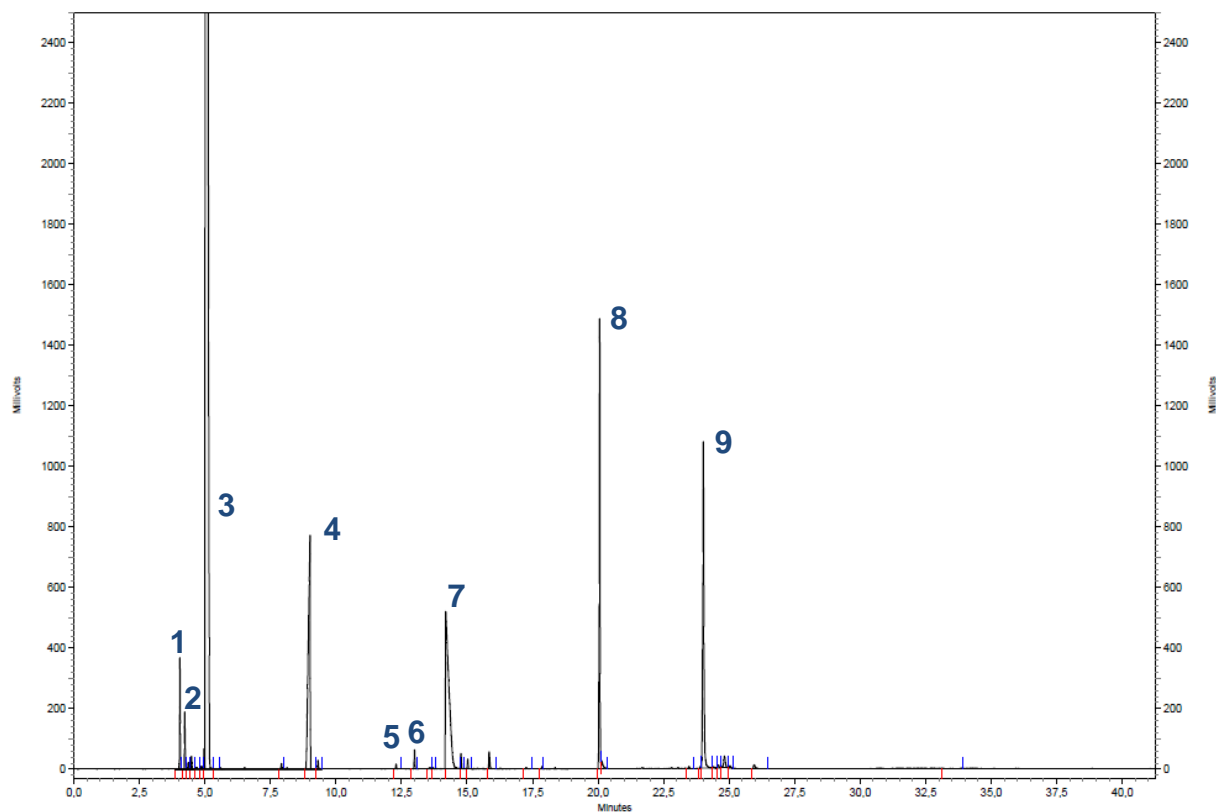
Entry	Retention Time [min]	Substance	Area
1	4.245	CH	4755632
2	4.502	CE	12208263
3	5.027	CH ₂ Cl ₂ (Solv.)	538971362
4	9.023	<i>n</i> -Dodecane (Stand.)	30517966
5	12.347	CAc	3354729
6	13.058	CI	7869117
7	14.218	CH ₃ COOH (Solv.)	44047706
8	20.092	1-Phenylethanol (Stand.)	26530706
9	24.047	CA	13544121

Entry 2



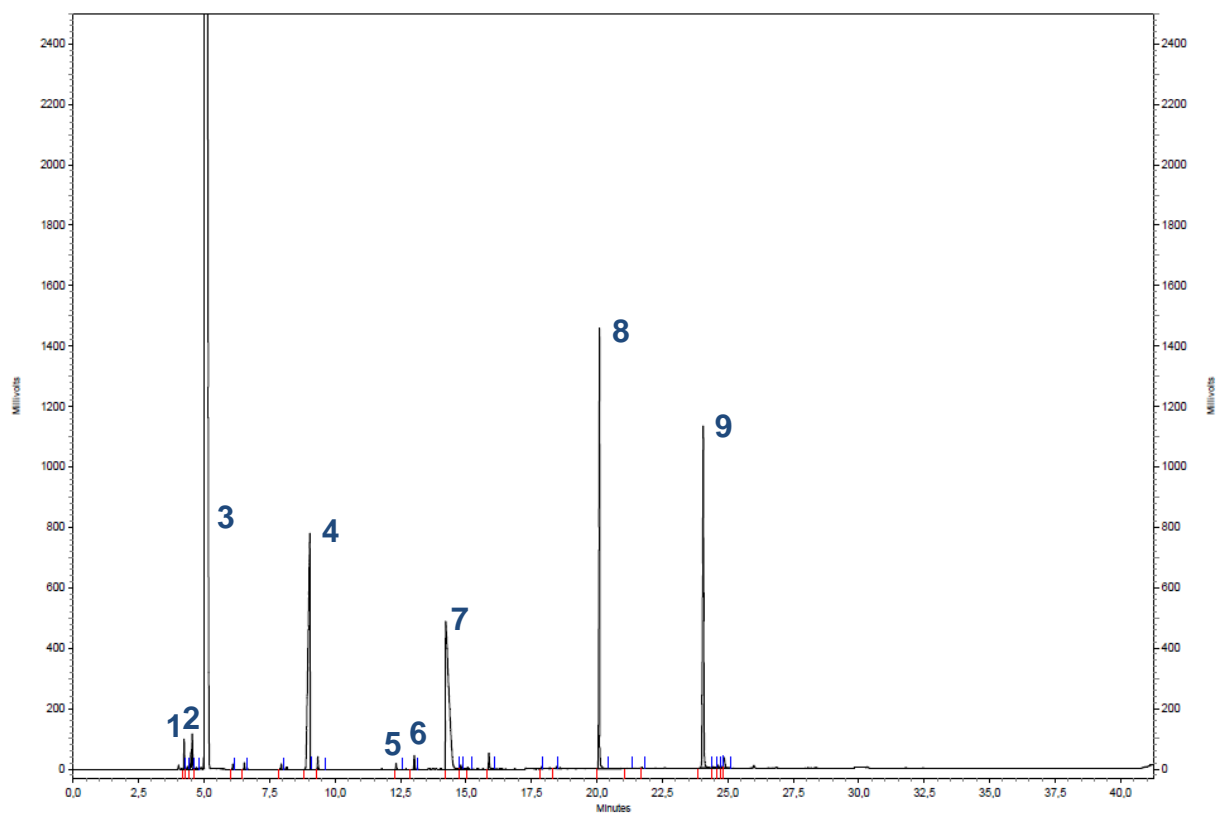
Entry	Retention Time [min]	Substance	Area
1	4.237	CH	2212263
2	4.543	CE	2549632
3	5.018	CH ₂ Cl ₂ (Solv.)	523505336
4	9.013	<i>n</i> -Dodecane (Stand.)	31575593
5	12.322	CAc	543484
6	13.022	CI	1491064
7	14.212	CH ₃ COOH (Solv.)	44837575
8	20.087	1-Phenylethanol (Stand.)	26305575
9	24.058	CA	34121033

Entry 3



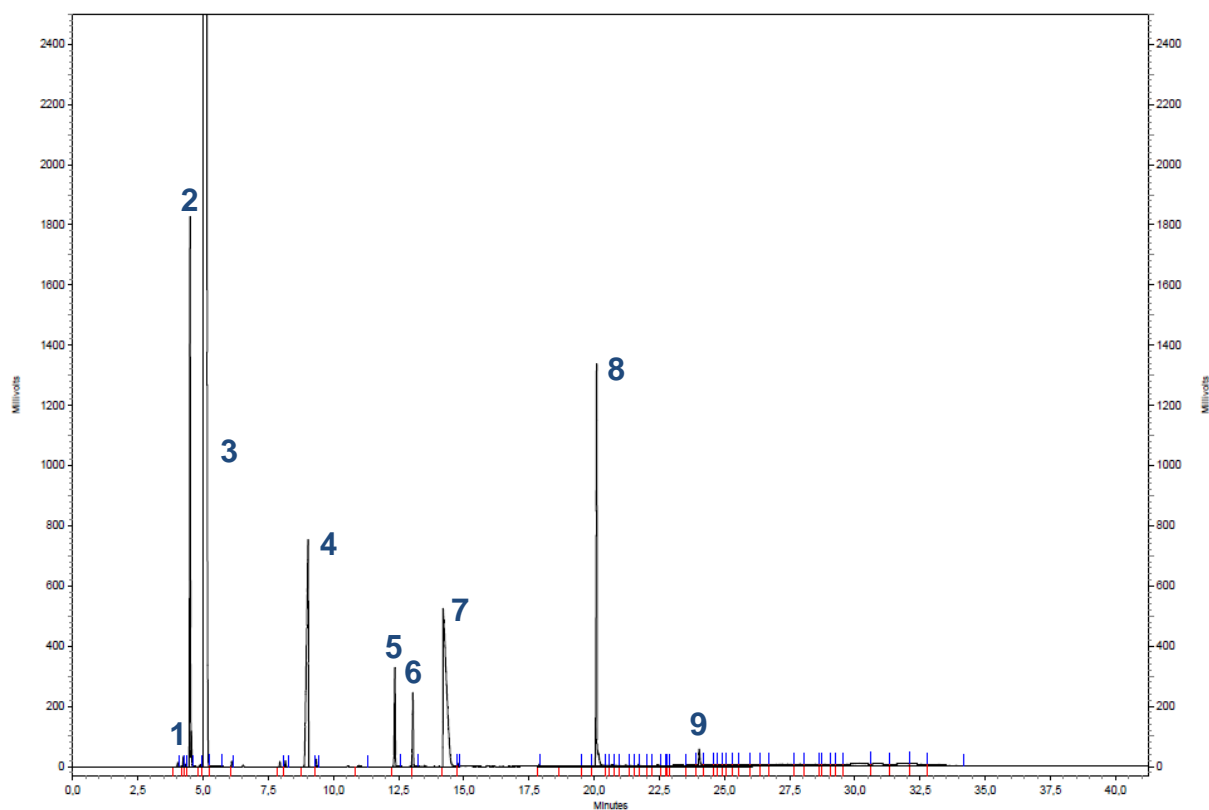
Entry	Retention Time [min]	Substance	Area
1	4.230	CH	2536833
2	4.485	CE	677684
3	5.012	CH ₂ Cl ₂ (Solv.)	516660834
4	9.012	<i>n</i> -Dodecane (Stand.)	36570073
5	12.295	CAc	275615
6	12.998	CI	1168625
7	14.185	CH ₃ COOH (Solv.)	45849429
8	20.062	1-Phenylethanol (Stand.)	29227806
9	24.018	CA	33012551

Entry 4



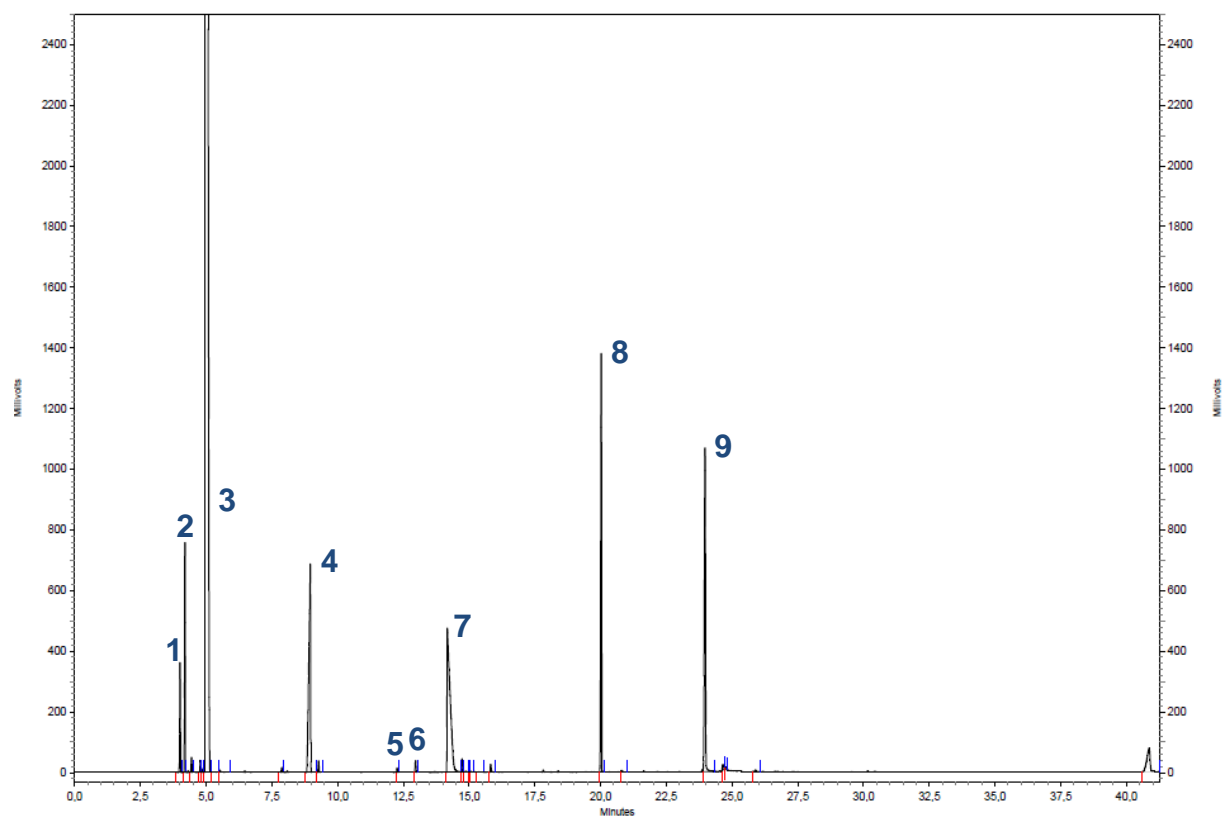
Entry	Retention Time [min]	Substance	Area
1	4.242	CH	1300713
2	4.550	CE	2590549
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	9.037	<i>n</i> -Dodecane (Stand.)	37018450
5	12.333	CAc	326345
6	13.030	CI	821294
7	14.228	CH ₃ COOH (Solv.)	41566839
8	20.098	1-Phenylethanol (Stand.)	29190086
9	24.063	CA	32288328

Entry 5



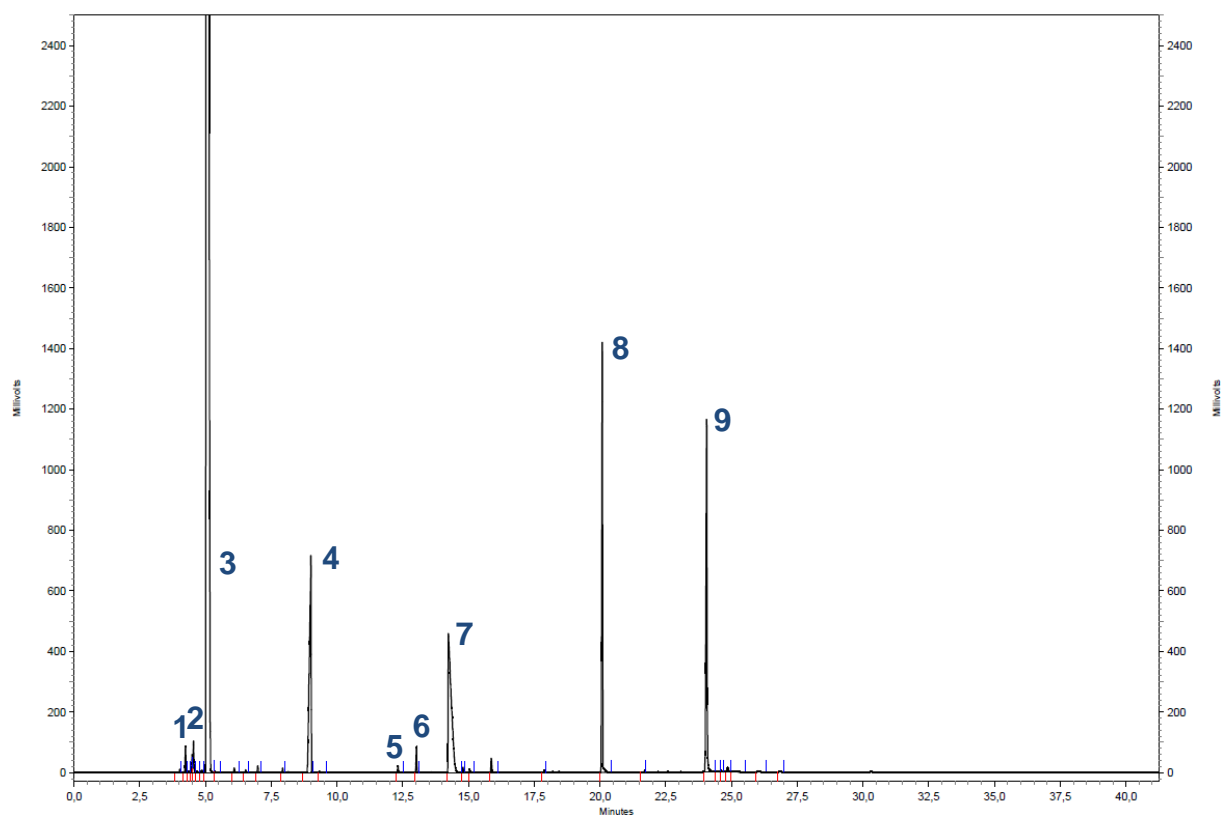
Entry	Retention Time [min]	Substance	Area
1	4.243	CH	463012
2	4.502	CE	29574068
3	5.027	CH ₂ Cl ₂ (Solv.)	534155611
4	9.033	<i>n</i> -Dodecane (Stand.)	35038431
5	12.358	CAc	6954729
6	13.052	CI	5116405
7	14.210	CH ₃ COOH (Solv.)	47295142
8	20.093	1-Phenylethanol (Stand.)	28523371
9	24.033	CA	2112362

Entry 6



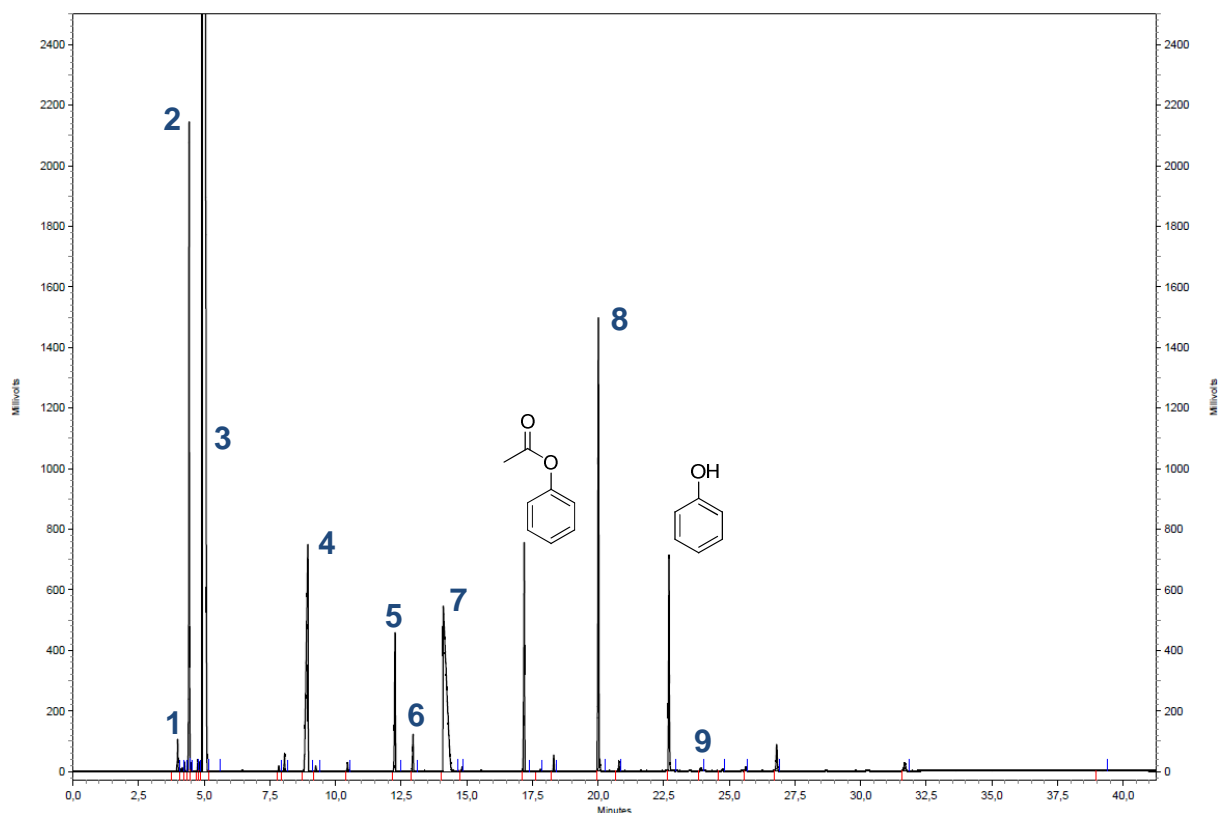
Entry	Retention Time [min]	Substance	Area
1	4.192	CH	9516376
2	4.445	CE	736403
3	4.970	CH ₂ Cl ₂ (Solv.)	481626659
4	8.968	<i>n</i> -Dodecane (Stand.)	29571531
5	12.267	CAc	227325
6	12.967	CI	669337
7	14.170	CH ₃ COOH (Solv.)	38218067
8	20.027	1-Phenylethanol (Stand.)	25971566
9	23.975	CA	27930573

Entry 7



Entry	Retention Time [min]	Substance	Area
1	4.238	CH	1331223
2	4.543	CE	1804530
3	5.018	CH ₂ Cl ₂ (Solv.)	545775329
4	9.012	<i>n</i> -Dodecane (Stand.)	31634188
5	12.318	CAc	376806
6	13.020	CI	1531813
7	14.233	CH ₃ COOH (Solv.)	37546431
8	20.085	1-Phenylethanol (Stand.)	27903054
9	24.057	CA	32949156

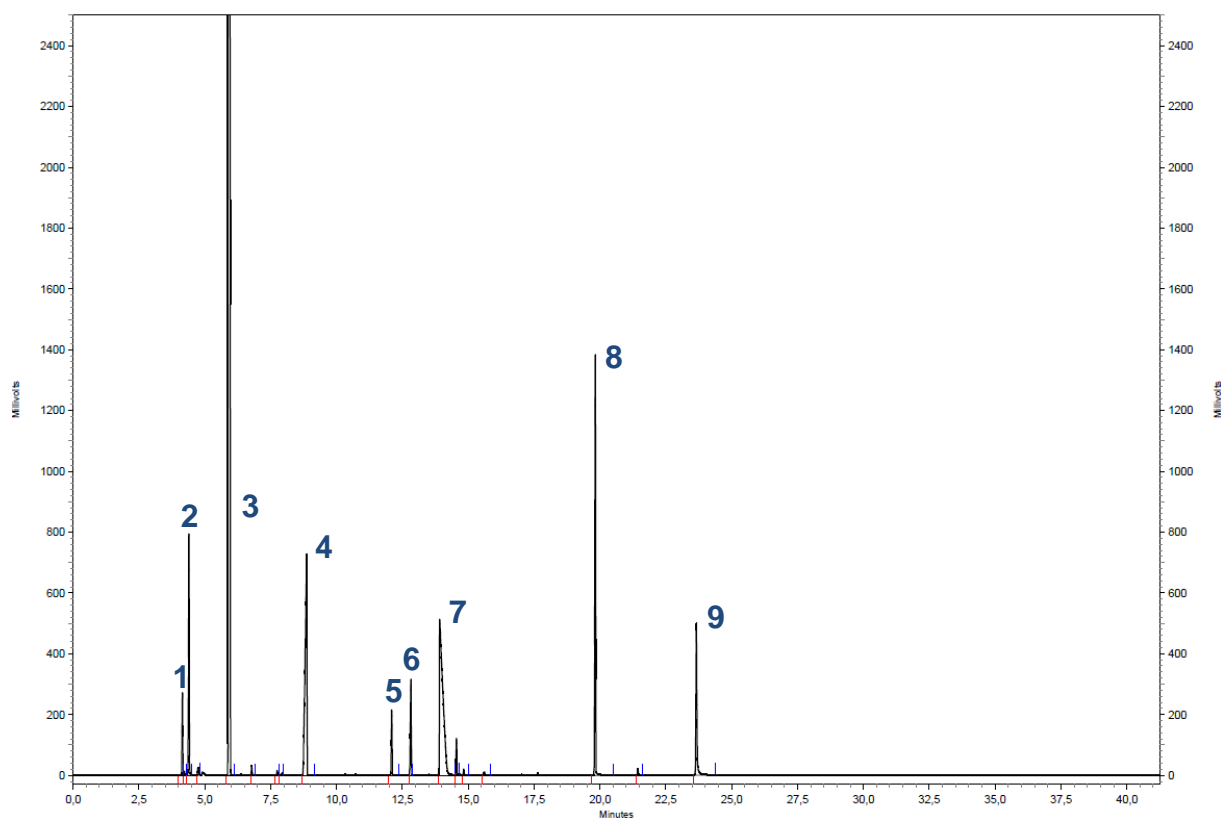
Entry 8



Phenol and acetic acid phenyl ester derive from ligand decomposition under reaction conditions.

Entry	Retention Time [min]	Substance	Area
1	4.168	CH	262557
2	4.422	CE	32763520
3	4.932	CH ₂ Cl ₂ (Solv.)	553455134
4	8.952	<i>n</i> -Dodecane (Stand.)	34844775
5	12.270	CAc	9424417
6	12.952	CI	2149744
7	14.107	CH ₃ COOH (Solv.)	48125055
8	20.013	1-Phenylethanol (Stand.)	29333737
9	23.935	CA	356222

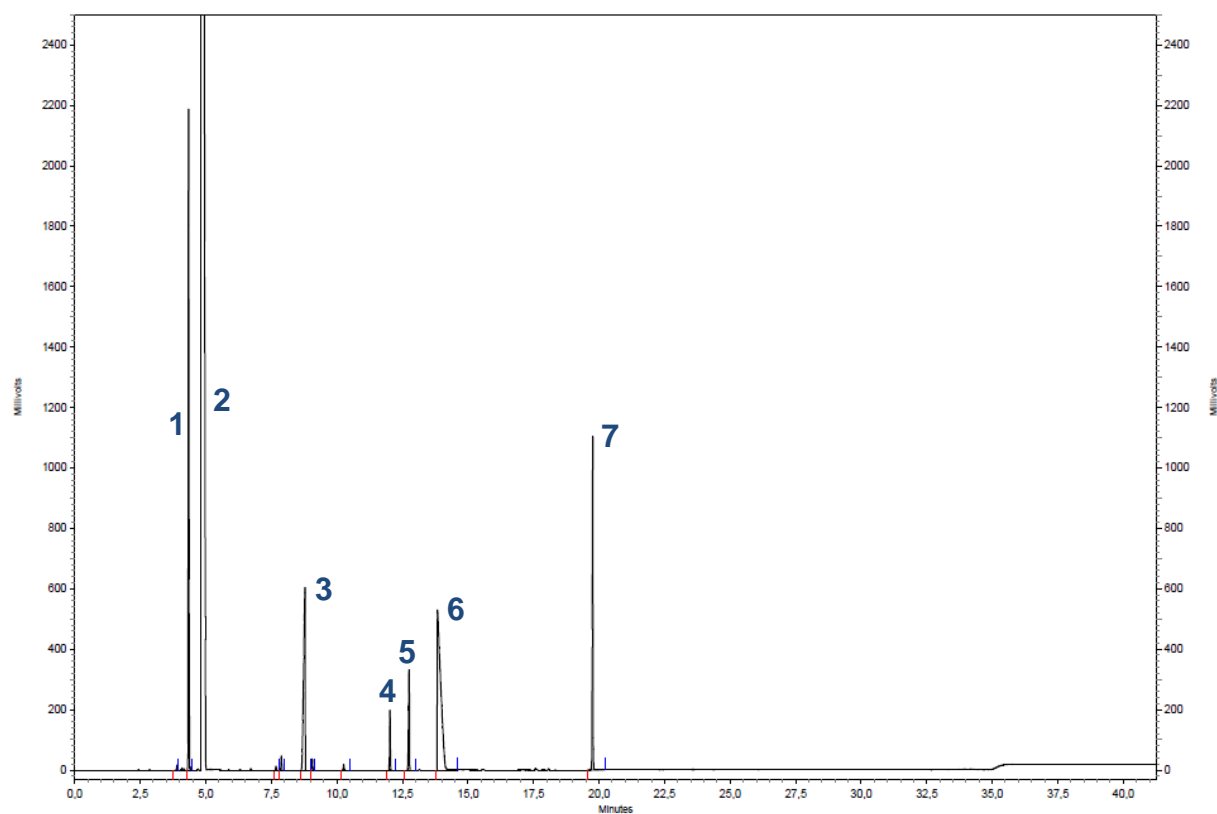
Entry 9



Entry	Retention Time [min]	Substance	Area
1	4.148	CH	3958223
2	4.393	CE	13722225
3 ^[a]	5.892	CDCl ₃ (Solv.)	386368933
4	8.872	<i>n</i> -Dodecane (Stand.)	33464320
5	12.095	CAc	3811090
6	12.825	CI	6818725
7	13.913	CH ₃ COOH (Solv.)	46197680
8	19.825	1-Phenylethanol (Stand.)	26247824
9	23.663	CA	13232421

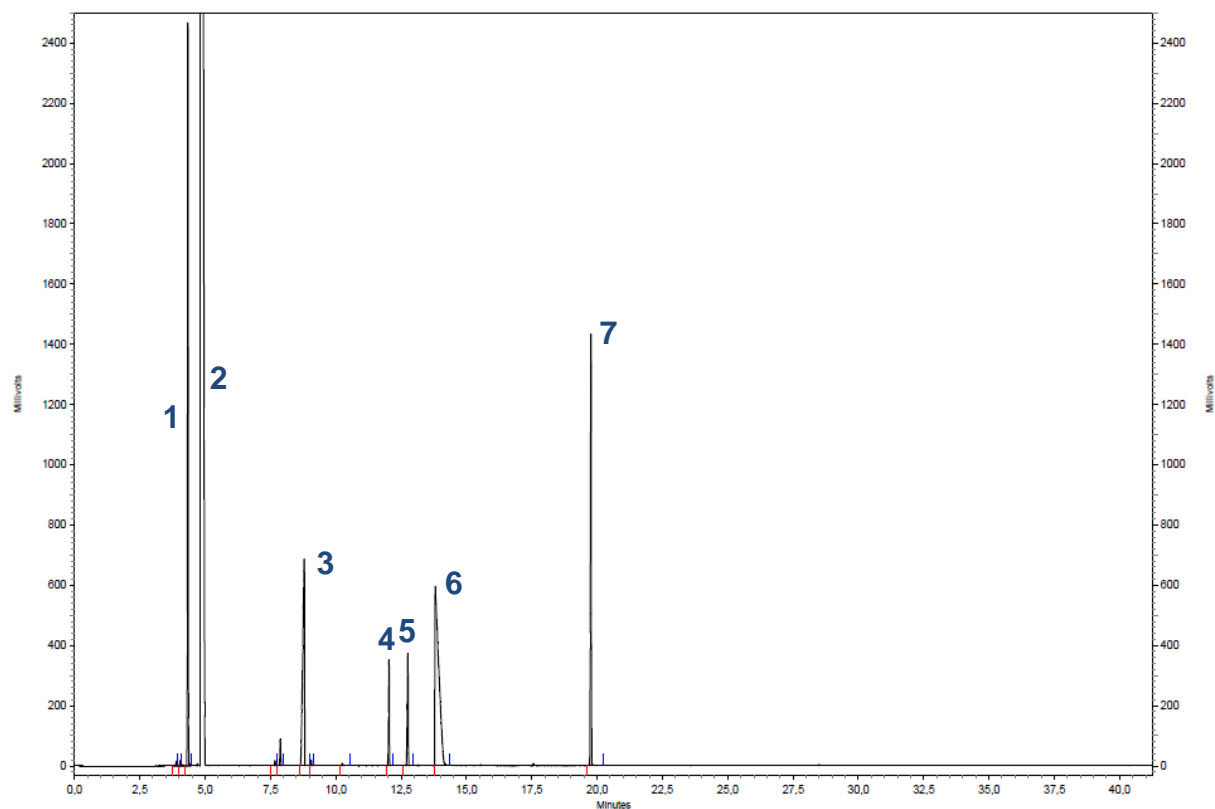
[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Entry 10



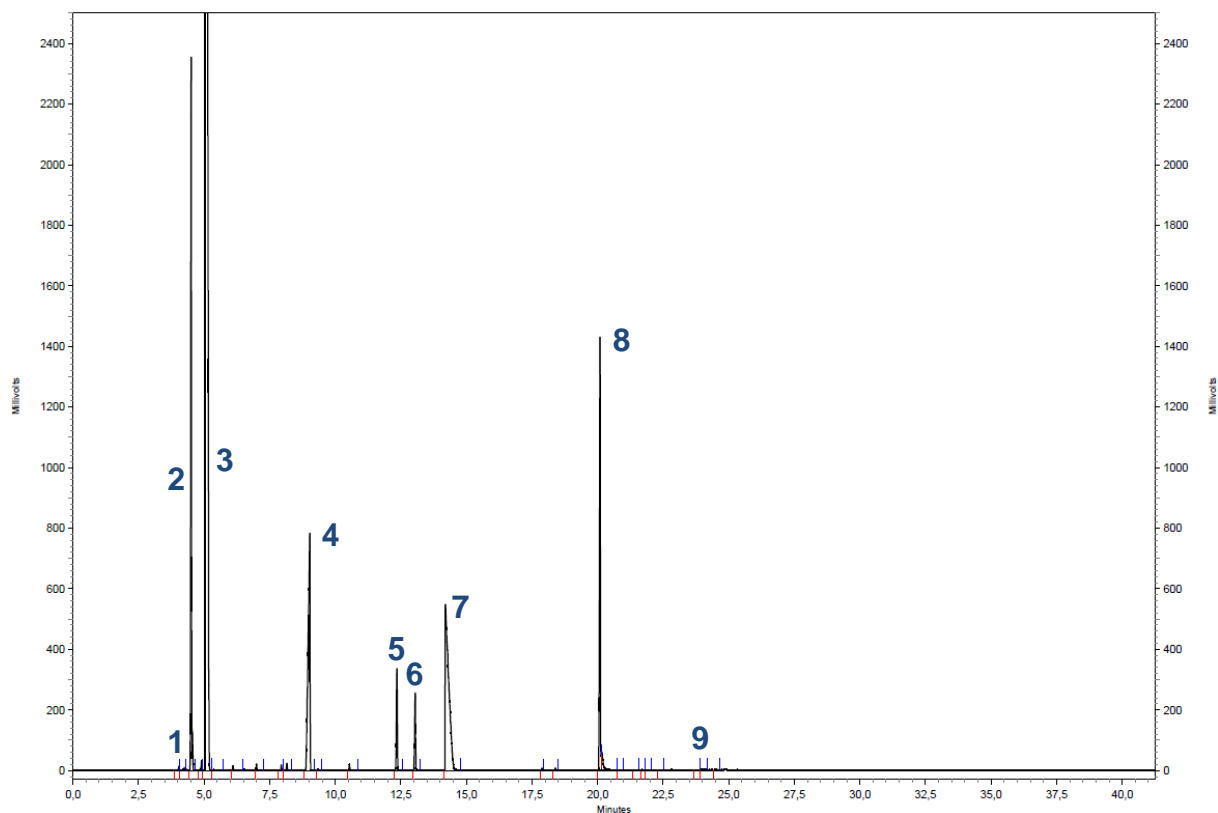
Entry	Retention Time [min]	Substance	Area
1	4.345	CE	32798302
2	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
3	8.783	<i>n</i> -Dodecane (Stand.)	23821426
4	12.027	CAc	3374657
5	12.757	CI	7129707
6	13.835	CH ₃ COOH (Solv.)	48159978
7	19.758	1-Phenylethanol (Stand.)	19839278

Entry 11



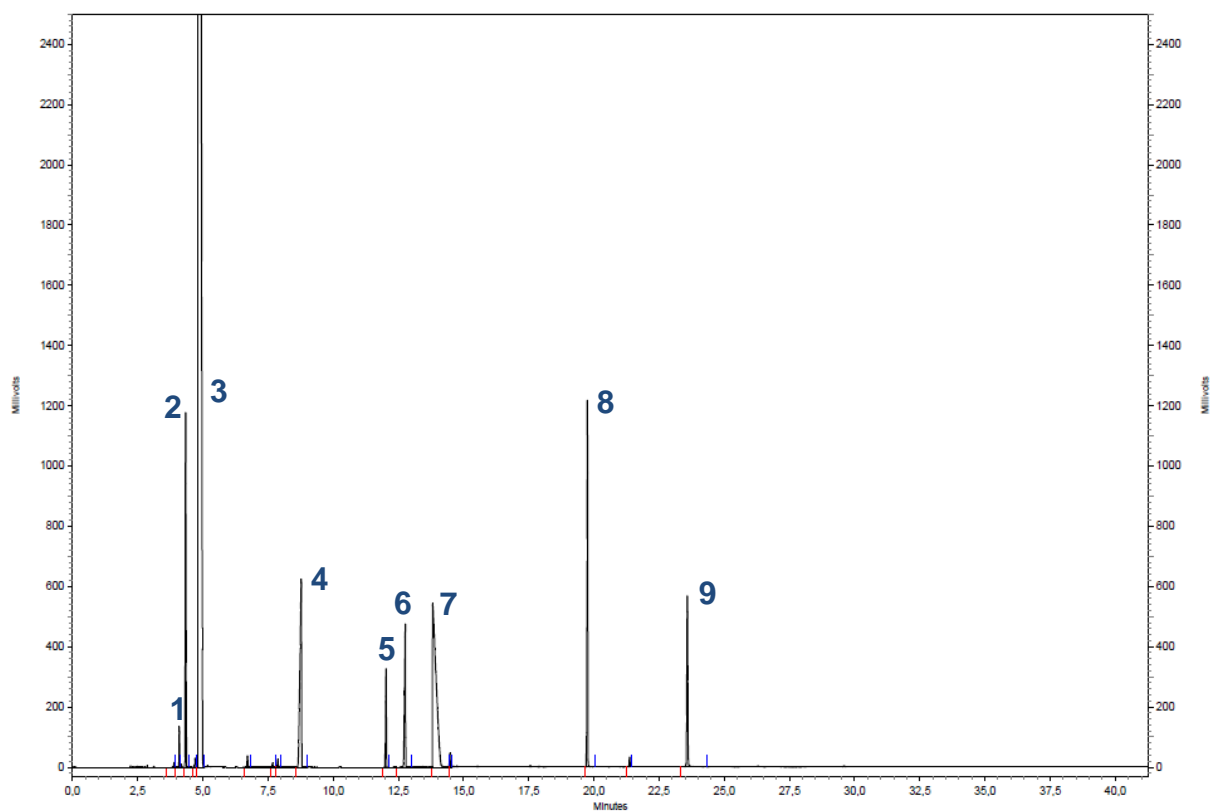
Entry	Retention Time [min]	Substance	Area
1	4.338	CE	36838685
2	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
3	8.795	<i>n</i> -Dodecane (Stand.)	30001100
4	12.038	CAc	6531078
5	12.762	CI	8435655
6	13.803	CH ₃ COOH (Solv.)	59775994
7	19.765	1-Phenylethanol (Stand.)	27215247

Entry 12



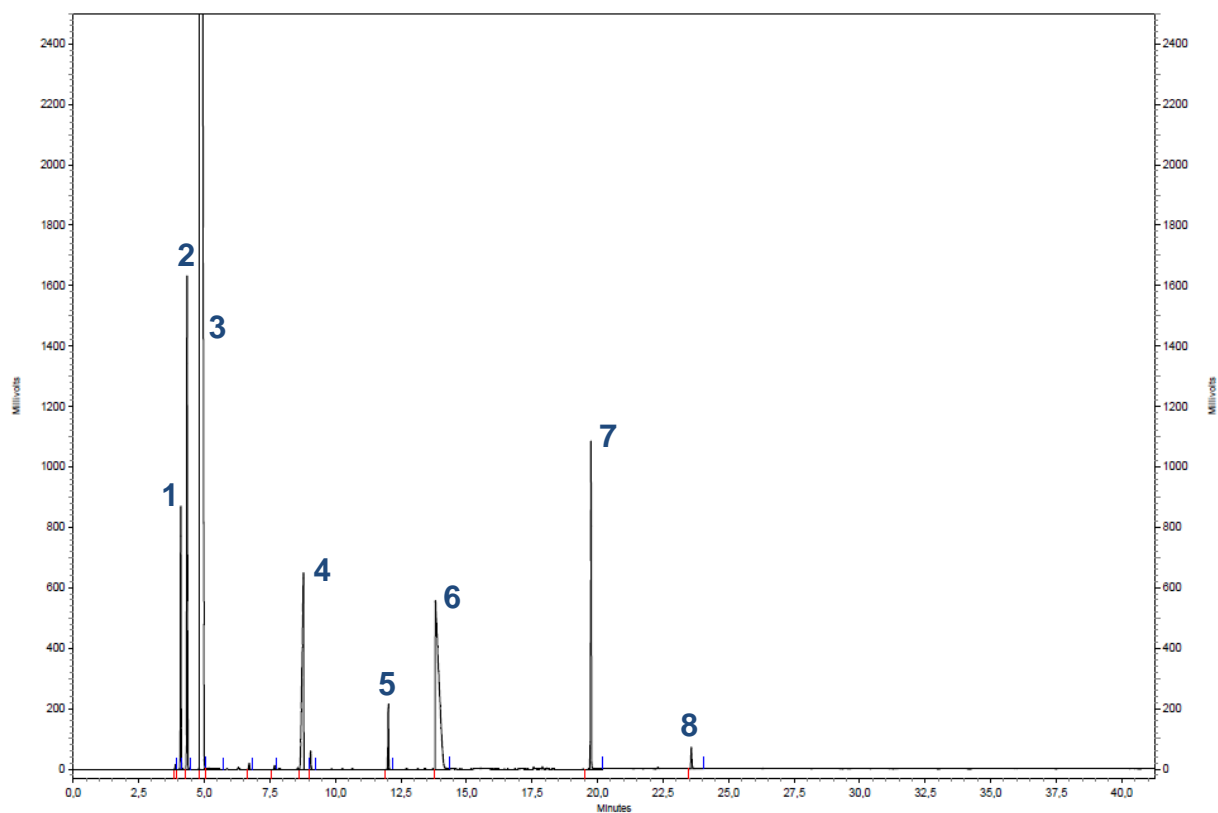
Entry	Retention Time [min]	Substance	Area
1	4.245	CH	182931
2	4.503	CE	38260389
3	5.028	CH ₂ Cl ₂ (Solv.)	527915450
4	9.032	<i>n</i> -Dodecane (Stand.)	37414196
5	12.352	CAc	7412775
6	13.048	CI	5515266
7	14.200	CH ₃ COOH (Solv.)	50794739
8	20.093	1-Phenylethanol (Stand.)	28896099
9	24.042	CA	213664

Entry 13



Entry	Retention Time [min]	Substance	Area
1	4.102	CH	1704673
2	4.345	CE	17177792
3	4.828	CH ₂ Cl ₂ (Solv.)	564983185
4	8.782	<i>n</i> -Dodecane (Stand.)	25360706
5	12.030	CAc	6017472
6	12.765	CI	11665187
7	13.820	CH ₃ COOH (Solv.)	50357720
8	19.753	1-Phenylethanol (Stand.)	21901588
9	23.585	CA	12788526

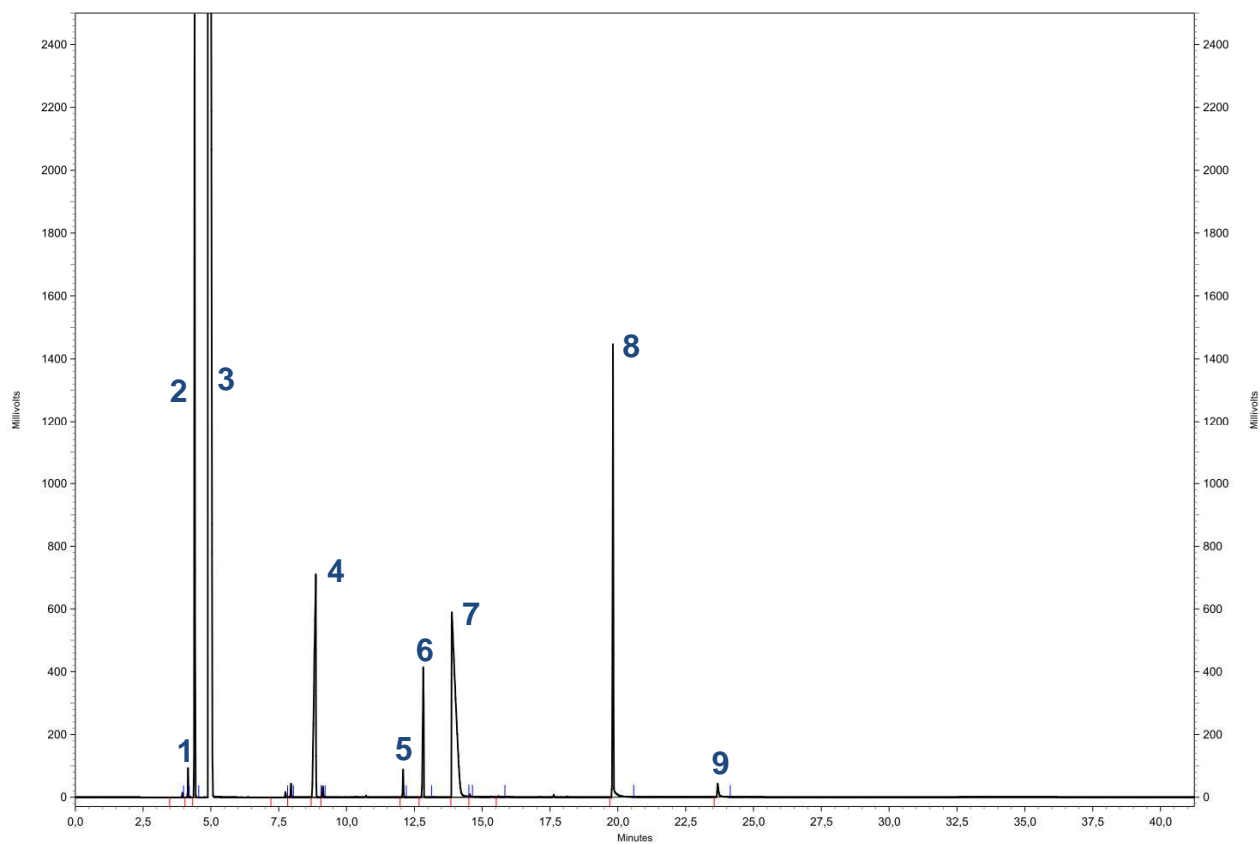
Entry 14



Entry	Retention Time [min]	Substance	Area
1	4.103	CH	12078281
2	4.345	CE	26052188
3	4.833	CH ₂ Cl ₂ (Solv.)	543849939
4	8.783	<i>n</i> -Dodecane (Stand.)	26618126
5	12.022	CAC	3729234
6	13.822	CH ₃ COOH (Solv.)	52073206
7	19.752	1-Phenylethanol (Stand.)	19140448
8	23.578	CA	1678163

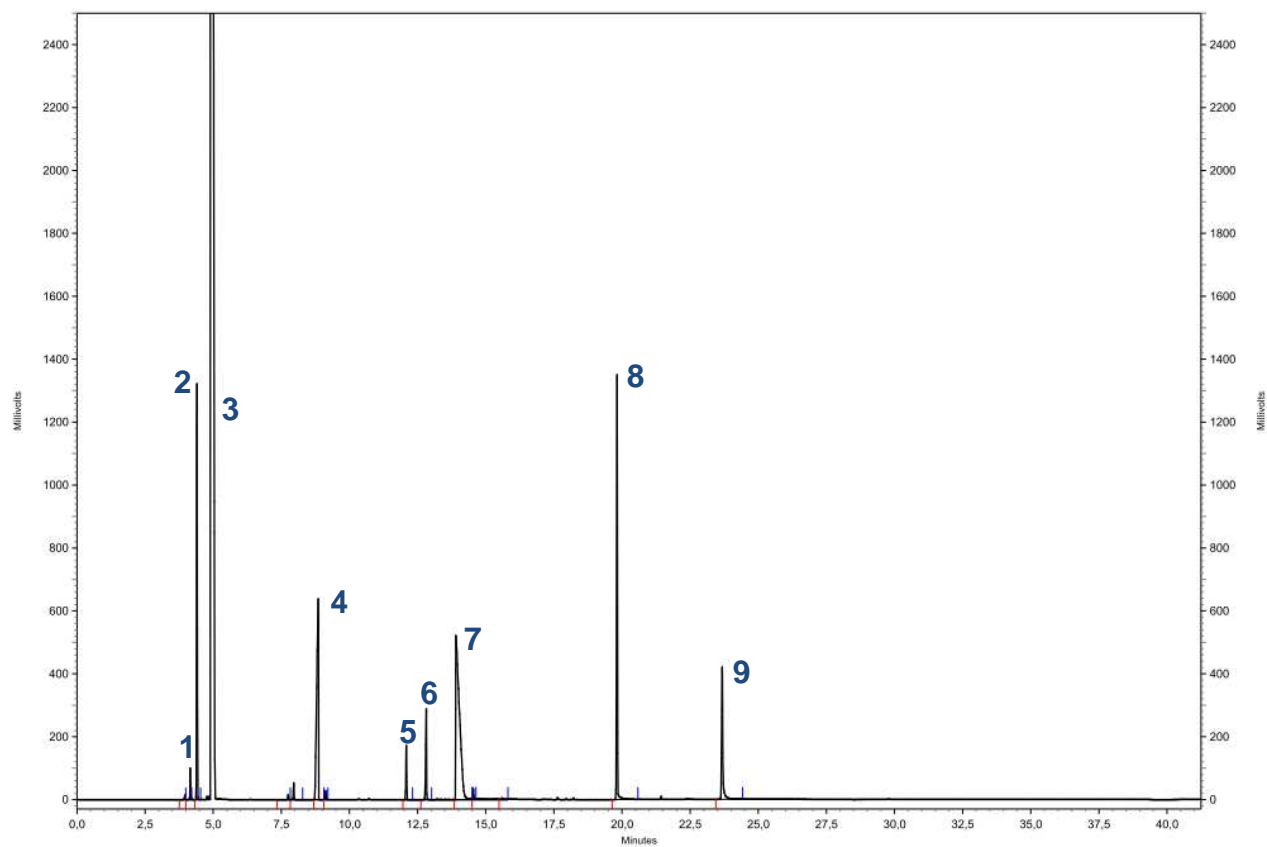
S3.6 Gaschromatograms to Table S2.6

Entry 1



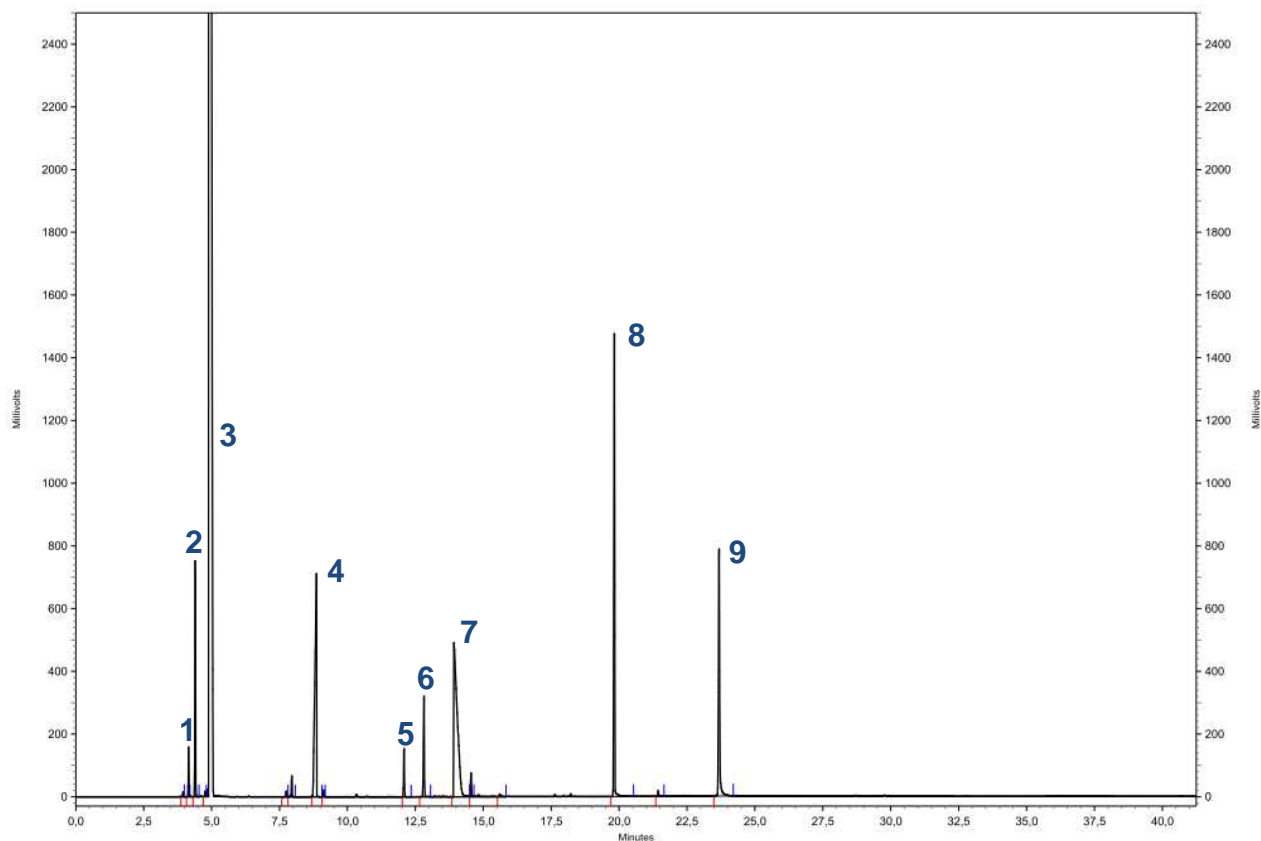
Entry	Retention Time [min]	Substance	Area
1	4.157	CH	1253709
2	4.403	CE	38041907
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.868	<i>n</i> -Dodecane (Stand.)	32513740
5	12.085	CAC	1519053
6	12.832	CI	9855061
7	13.880	CH ₃ COOH (Solv.)	59349994
8	19.823	1-Phenylethanol (Stand.)	29204886
9	23.680	CA	1587142

Entry 2



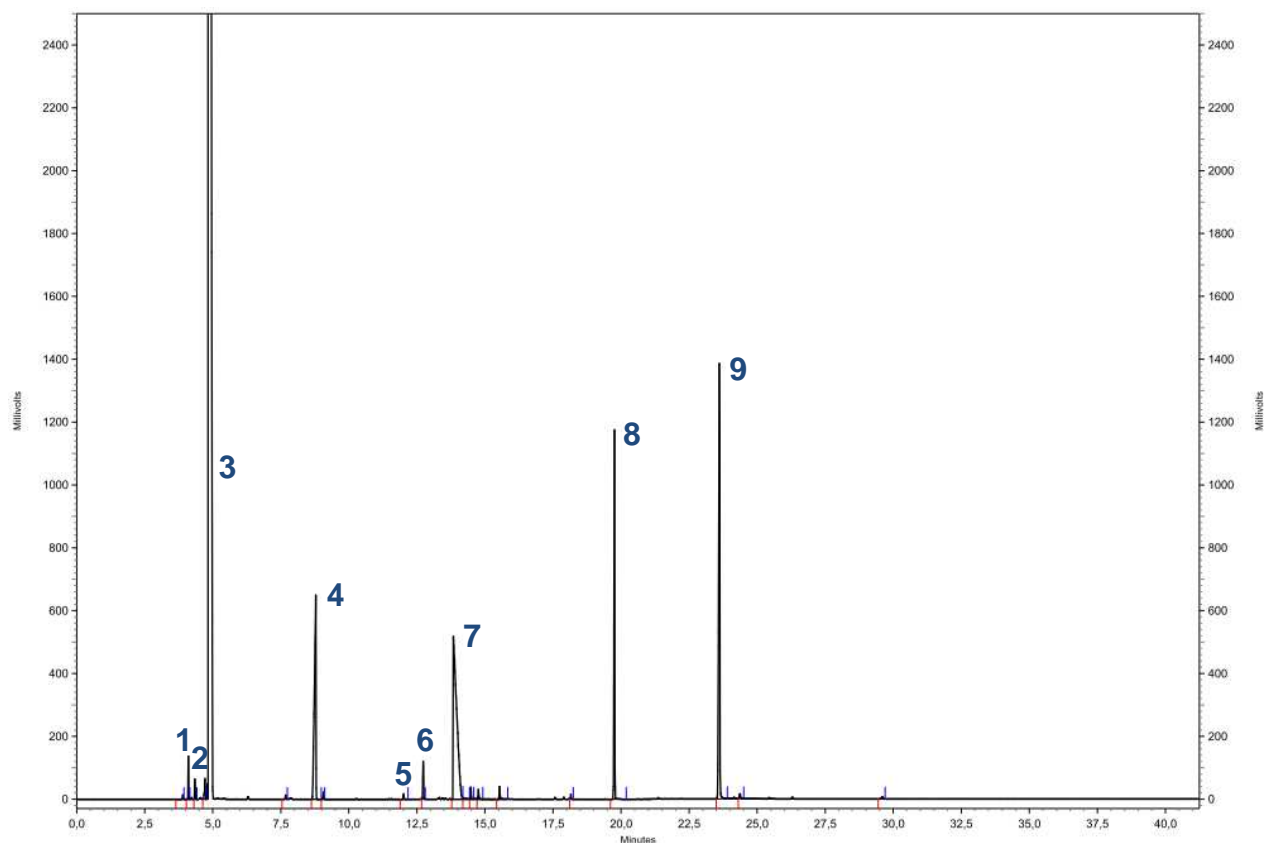
Entry	Retention Time [min]	Substance	Area
1	4.158	CH	1360010
2	4.403	CE	20203678
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.857	<i>n</i> -Dodecane (Stand.)	27430014
5	12.085	CAc	3046430
6	12.818	CI	6202747
7	13.907	CH ₃ COOH (Solv.)	48552998
8	19.820	1-Phenylethanol (Stand.)	26169017
9	23.675	CA	12613377

Entry 3



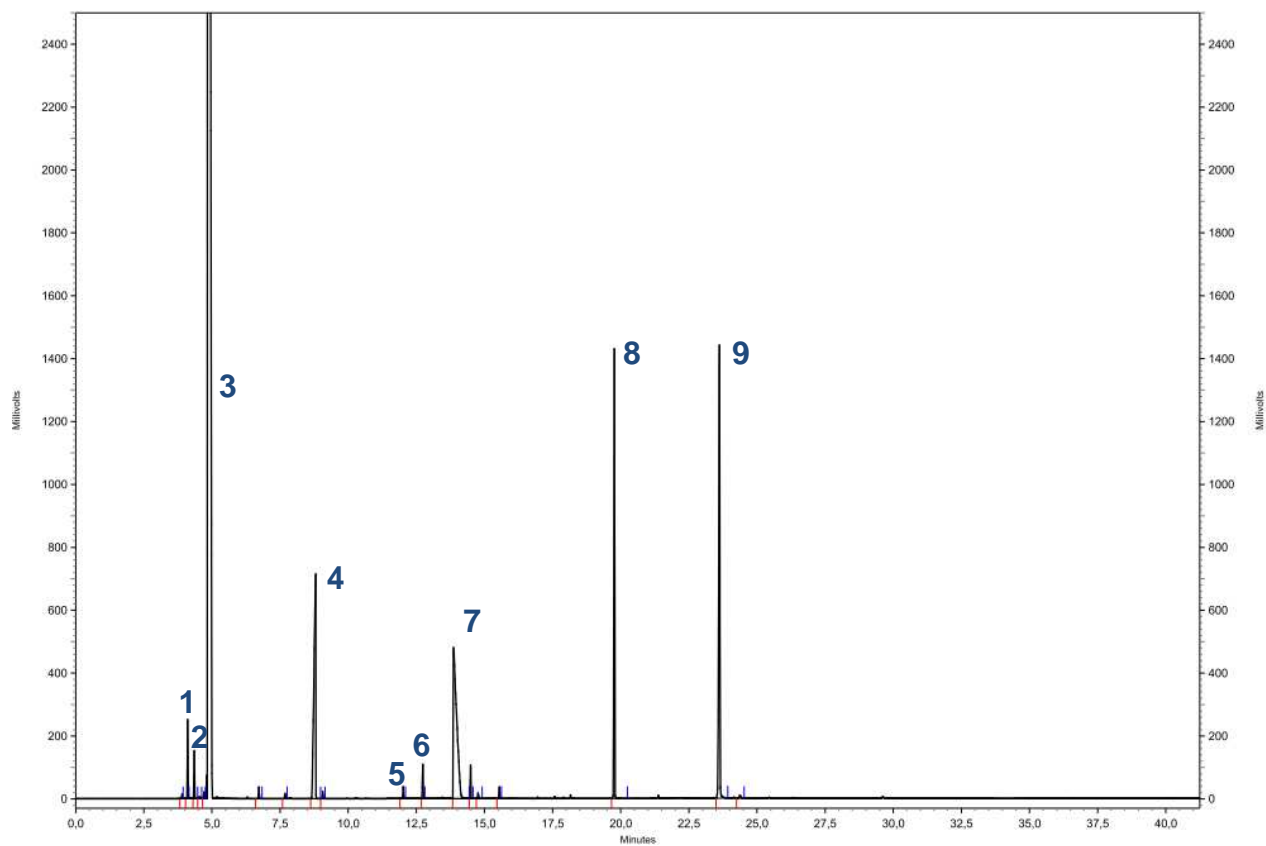
Entry	Retention Time [min]	Substance	Area
1	4.152	CH	2035683
2	4.397	CE	11078406
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.862	<i>n</i> -Dodecane (Stand.)	30954072
5	12.082	CAc	2552132
6	12.822	CI	6986287
7	13.922	CH ₃ COOH (Solv.)	43032339
8	19.827	1-Phenylethanol (Stand.)	29246822
9	23.683	CA	21673896

Entry 4



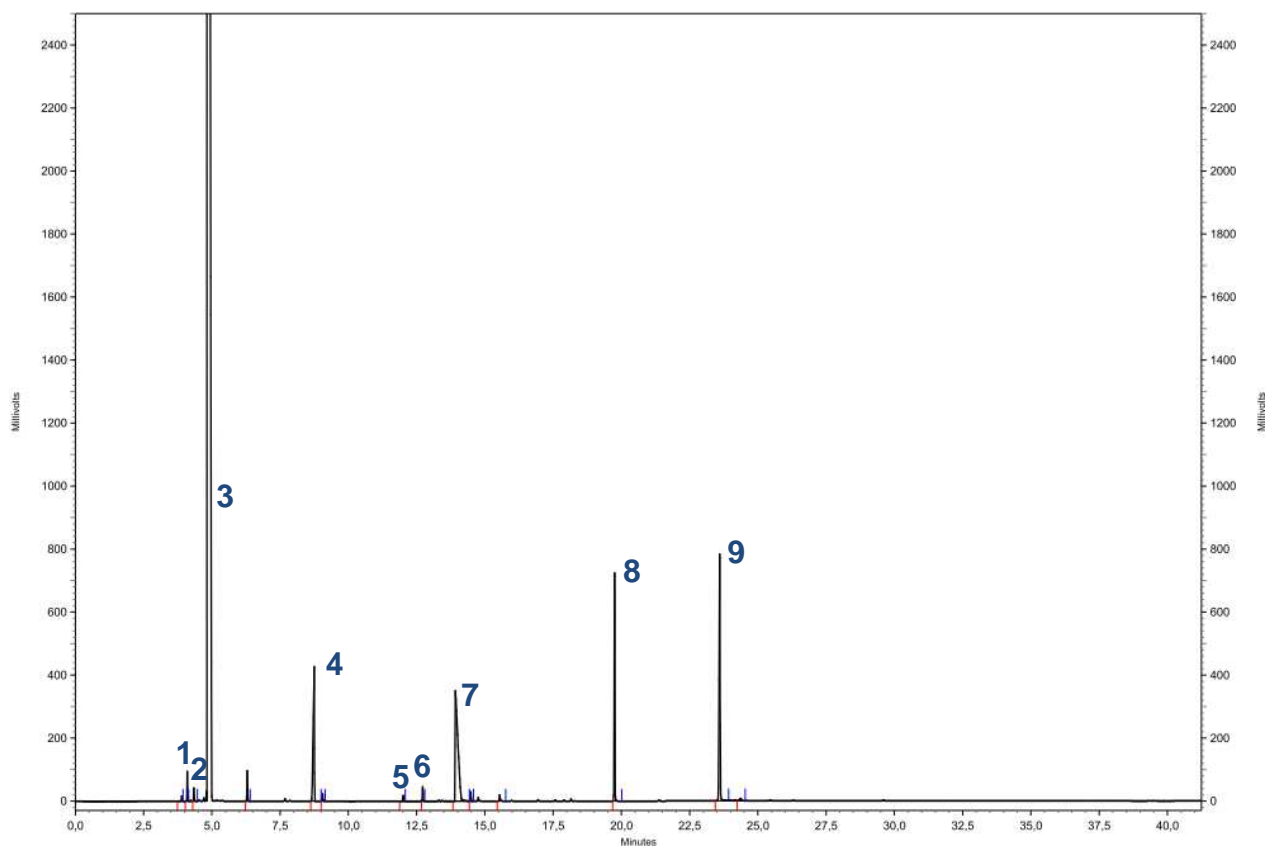
Entry	Retention Time [min]	Substance	Area
1	4.105	CH	1755261
2	4.347	CE	954342
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.788	<i>n</i> -Dodecane (Stand.)	26385371
5	12.012	CAc	280721
6	12.735	CI	2112889
7	13.838	CH ₃ COOH (Solv.)	46449859
8	19.758	1-Phenylethanol (Stand.)	20877781
9	23.612	CA	37072381

Entry 5



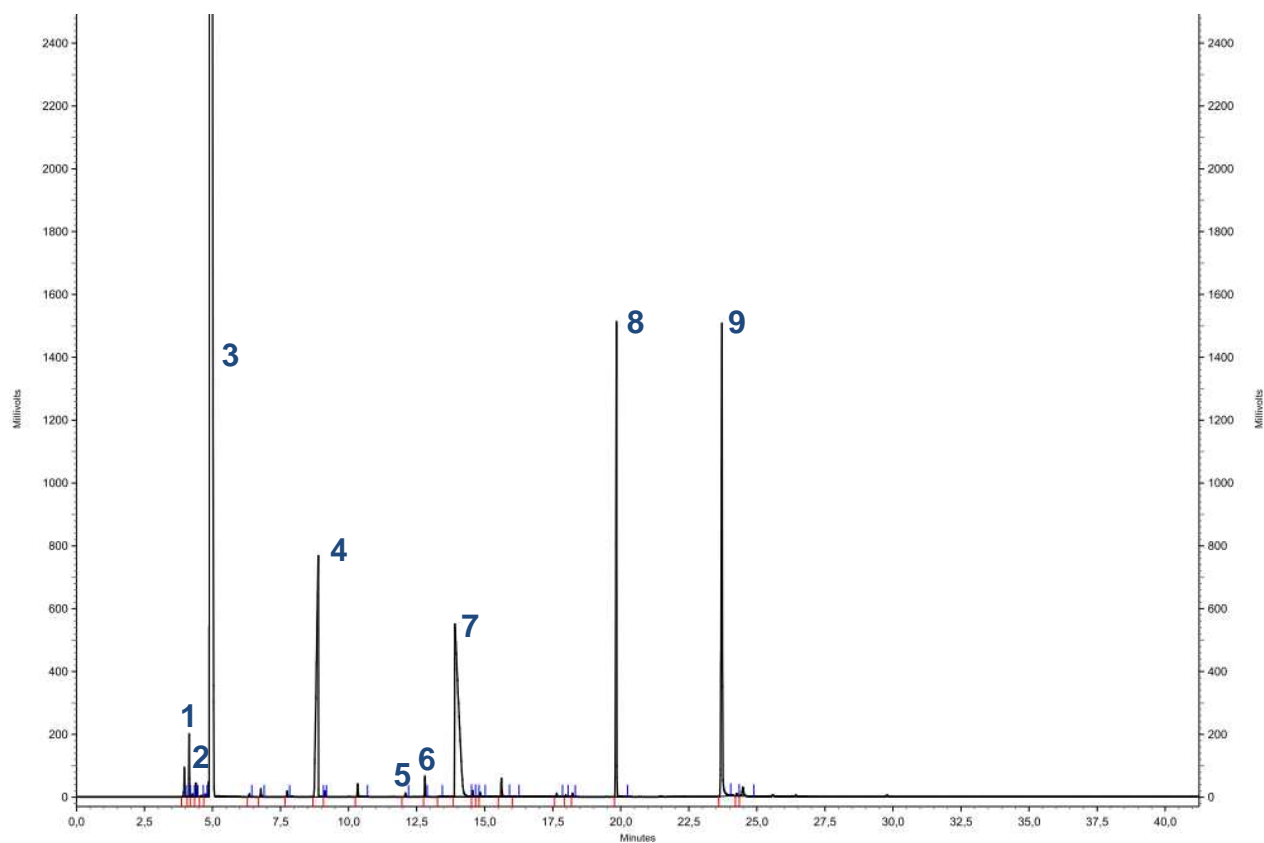
Entry	Retention Time [min]	Substance	Area
1	4.108	CH	3238297
2	4.352	CE	2280269
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.807	<i>n</i> -Dodecane (Stand.)	32192057
5	12.018	CAc	629956
6	12.738	CI	1930831
7	13.865	CH ₃ COOH (Solv.)	40637845
8	19.768	1-Phenylethanol (Stand.)	26679121
9	23.623	CA	38750777

Entry 6



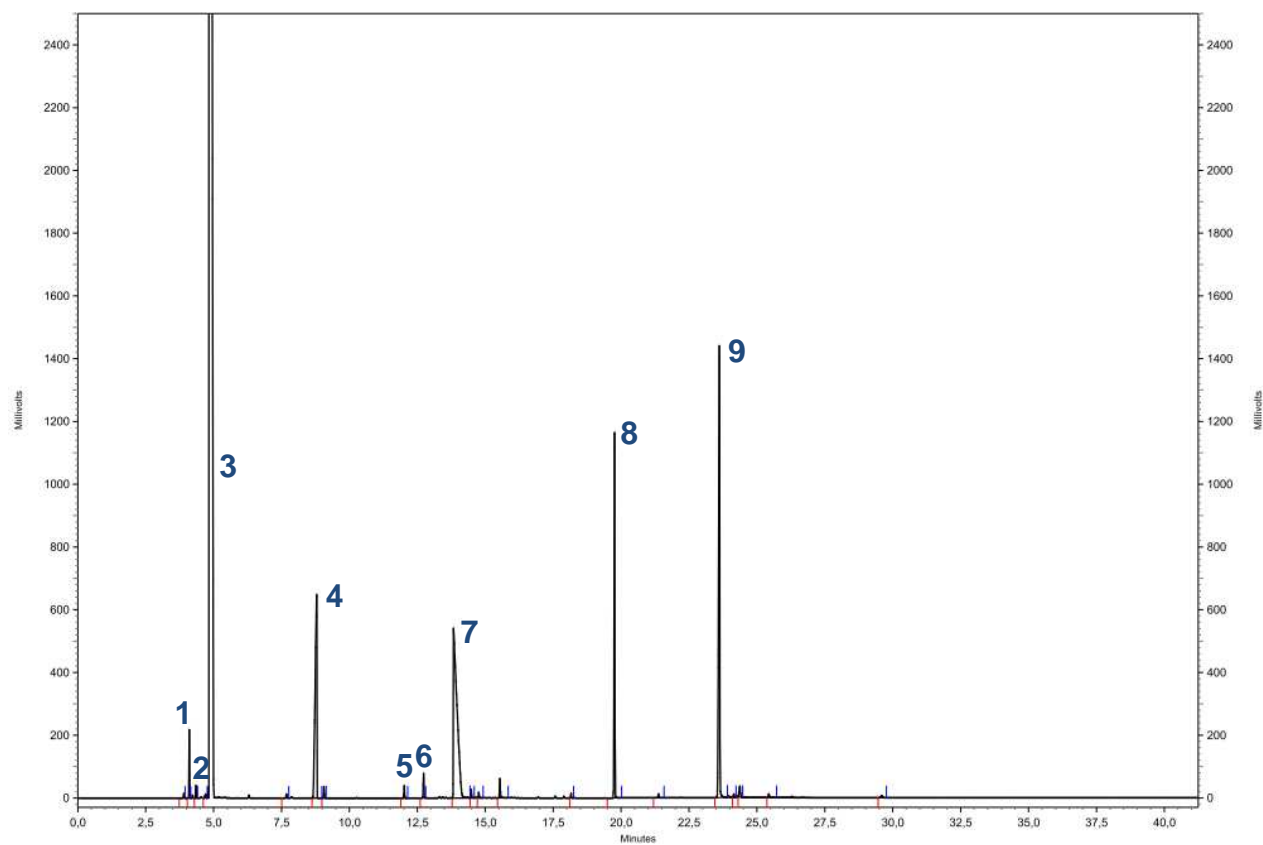
Entry	Retention Time [min]	Substance	Area
1	4.100	CH	1214717
2	4.343	CE	641059
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.750	<i>n</i> -Dodecane (Stand.)	12656494
5	12.010	CAc	298821
6	12.727	CI	793040
7	13.922	CH ₃ COOH (Solv.)	23583787
8	19.753	1-Phenylethanol (Stand.)	12066681
9	23.602	CA	18100772

Entry 7



Entry	Retention Time [min]	Substance	Area
1	4.145	CH	2645996
2	4.390	CE	690070
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.887	<i>n</i> -Dodecane (Stand.)	38410164
5	12.088	CAc	206428
6	12.805	CI	1165824
7	13.907	CH ₃ COOH (Solv.)	52427679
8	19.838	1-Phenylethanol (Stand.)	29284032
9	23.720	CA	43379657

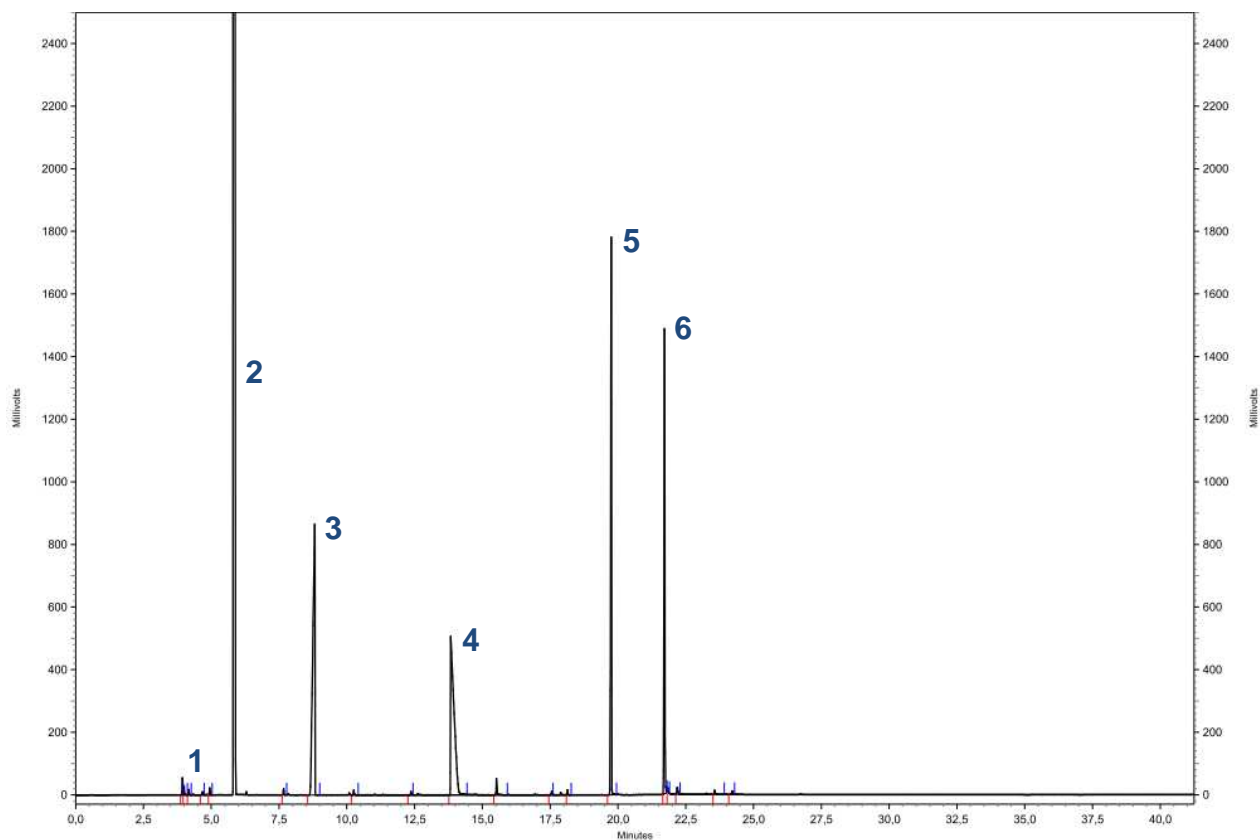
Entry 8



Entry	Retention Time [min]	Substance	Area
1	4.107	CH	2784368
2	4.348	CE	601856
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.797	<i>n</i> -Dodecane (Stand.)	27777498
5	12.017	CAc	634682
6	12.733	CI	1348814
7	13.832	CH ₃ COOH (Solv.)	50006752
8	19.762	1-Phenylethanol (Stand.)	20877959
9	23.617	CA	38595273

S3.7 Gaschromatograms to Table S2.7

Entry 2



Entry	Retention Time [min]	Substance	Area
1	3.997	Cyclopentene	497410
2 ^[a]	<i>cutted</i>	CD ₂ Cl ₃ (Solv.)	--
3	8.817	<i>n</i> -Dodecane (Stand.)	44779679
4	13.832	CH ₃ COOH (Solv.)	44799715
5	19.752	1-Phenylethanol (Stand.)	35436584
6	21.715	Cyclopentane carboxylic acid	28834332

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Data of the isolated product to Entry 2

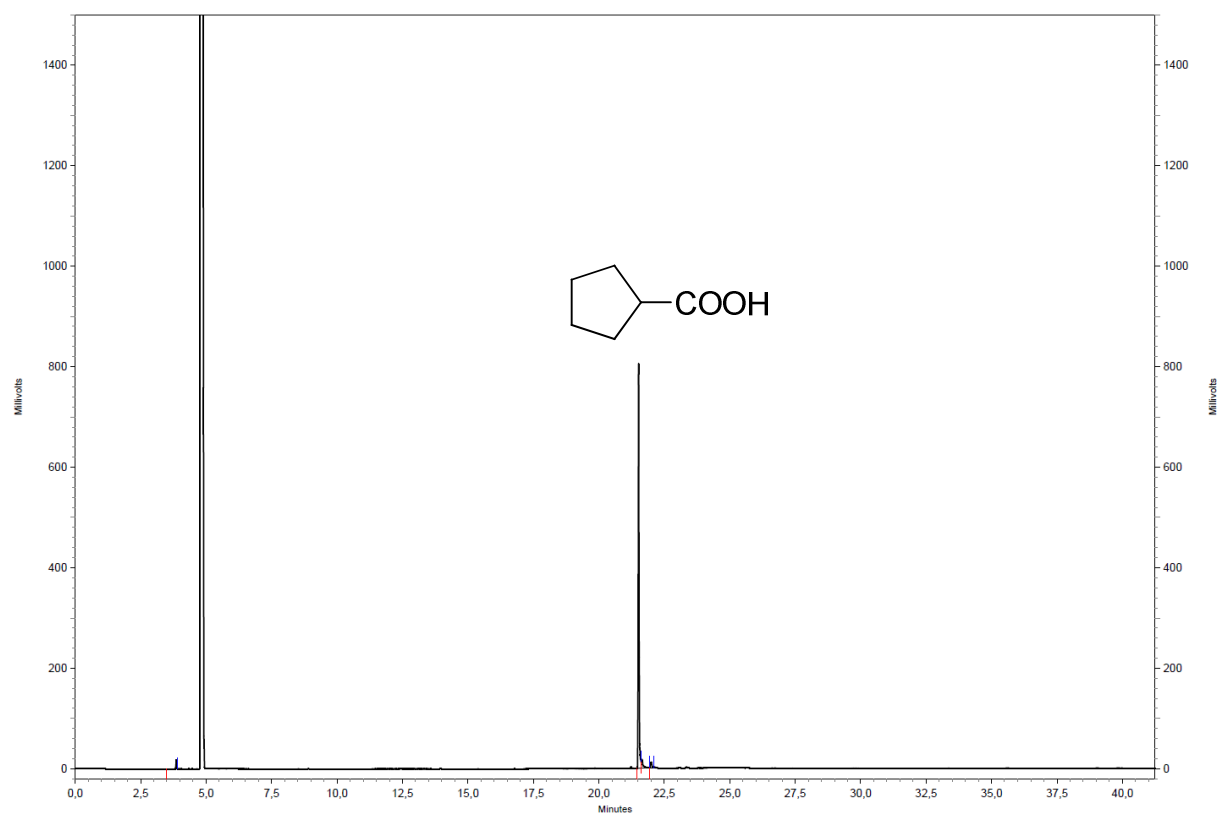


Figure 3.3 GC Chromatogram of the isolated cyclopentane carboxylic acid.

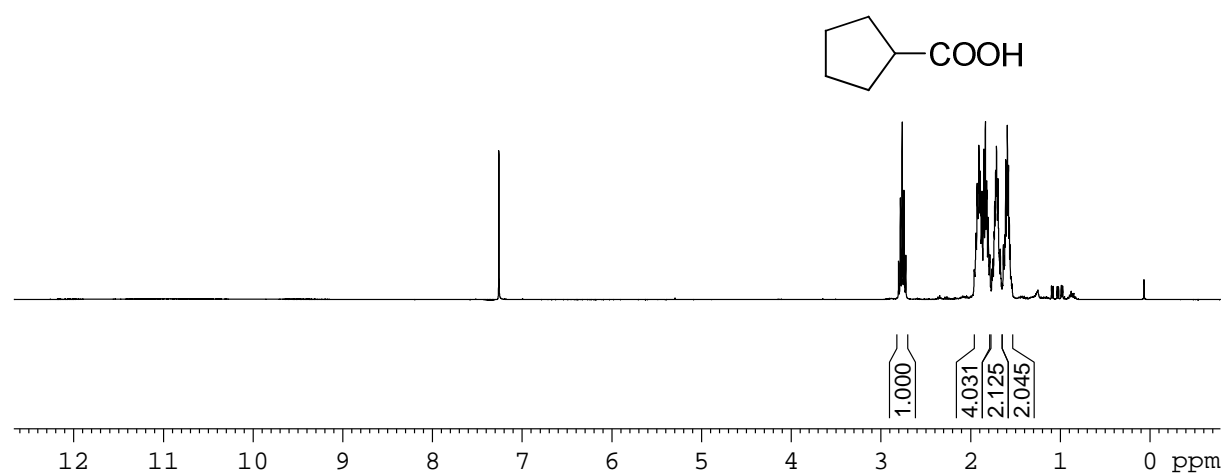
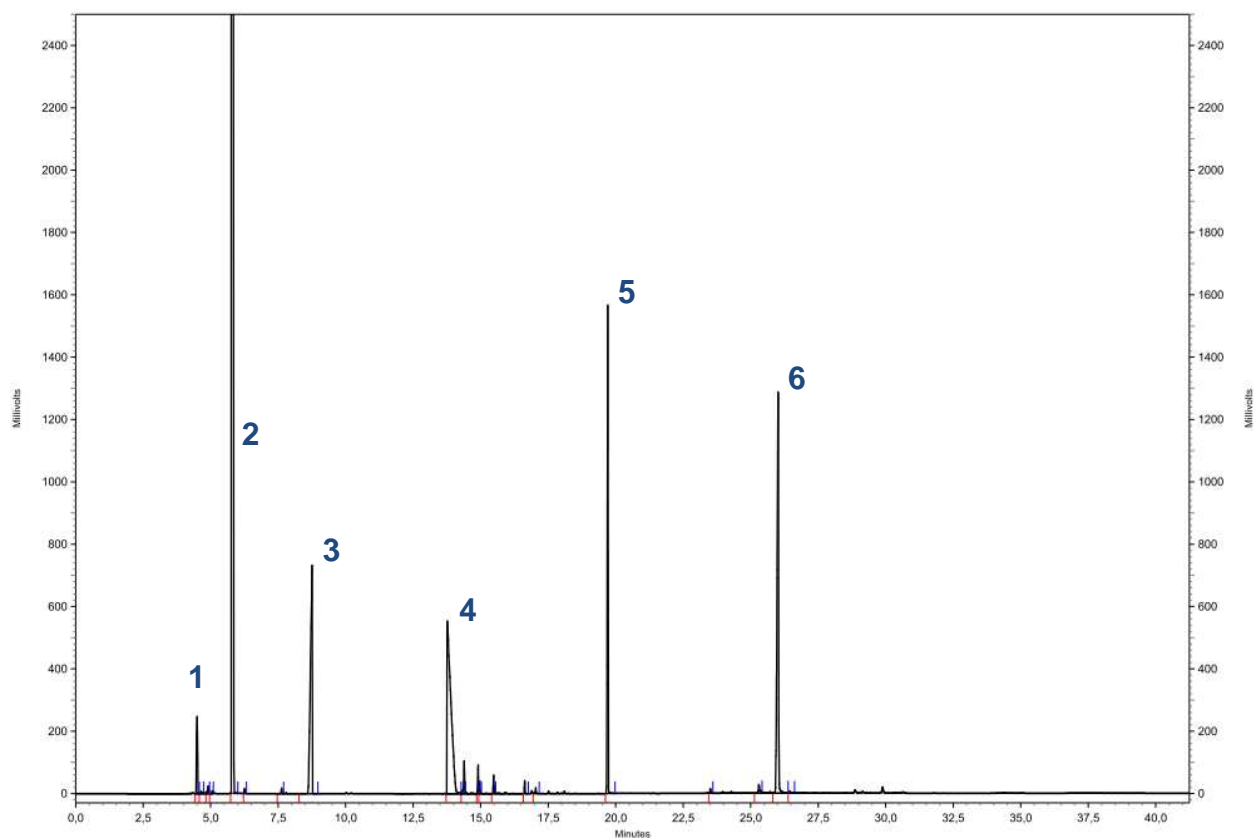


Figure 3.4. ¹H NMR spectrum of the isolated product measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

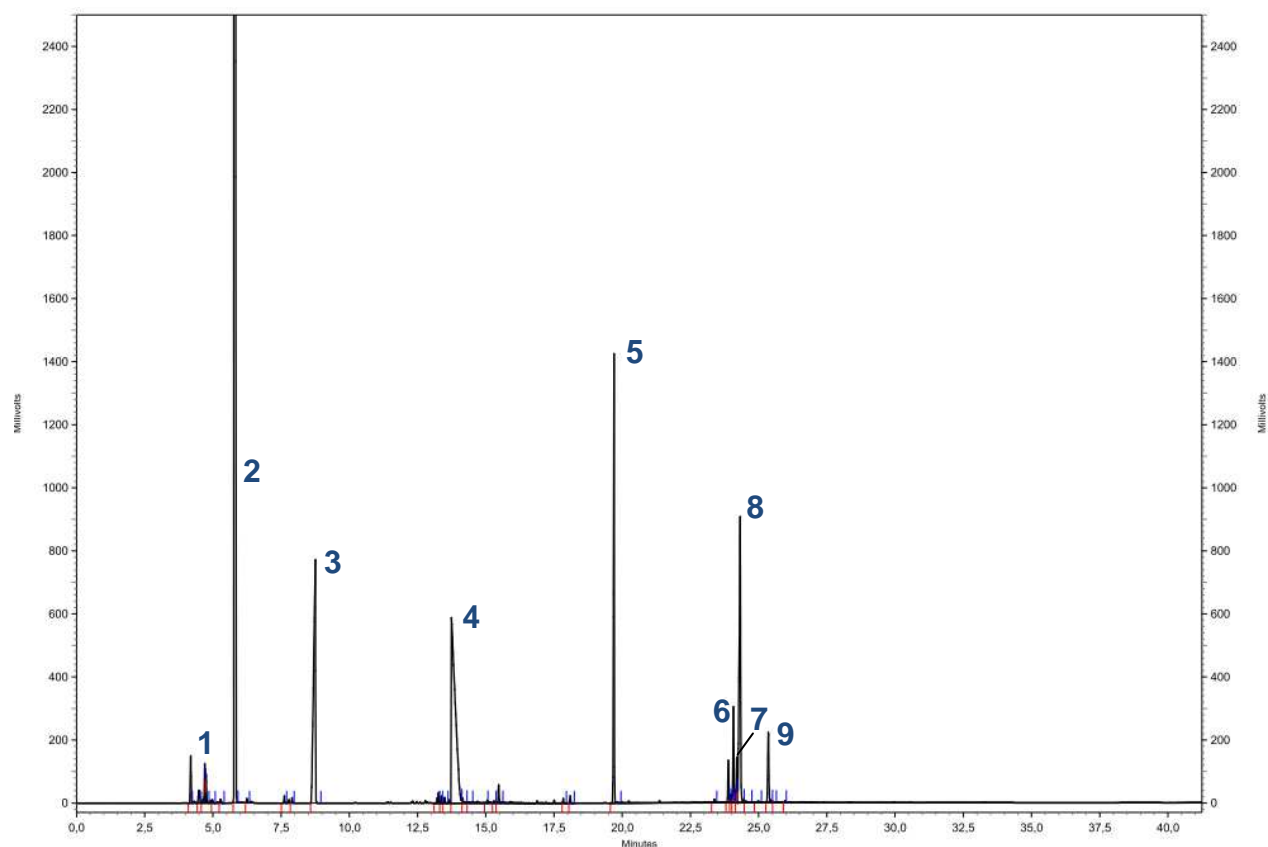
Entry 3



Entry	Retention Time [min]	Substance	Area
1	4.497	Norbornene	6705878
2 ^[a]	5.780	CDCl ₃ (Solv.)	371538304
3	8.757	<i>n</i> -Dodecane (Stand.)	33373713
4	13.765	CH ₃ COOH (Solv.)	53248514
5	19.708	1-Phenylethanol (Stand.)	30539476
6	26.017	<i>exo/endo</i>-Norbornane carboxylic acid	47434972

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

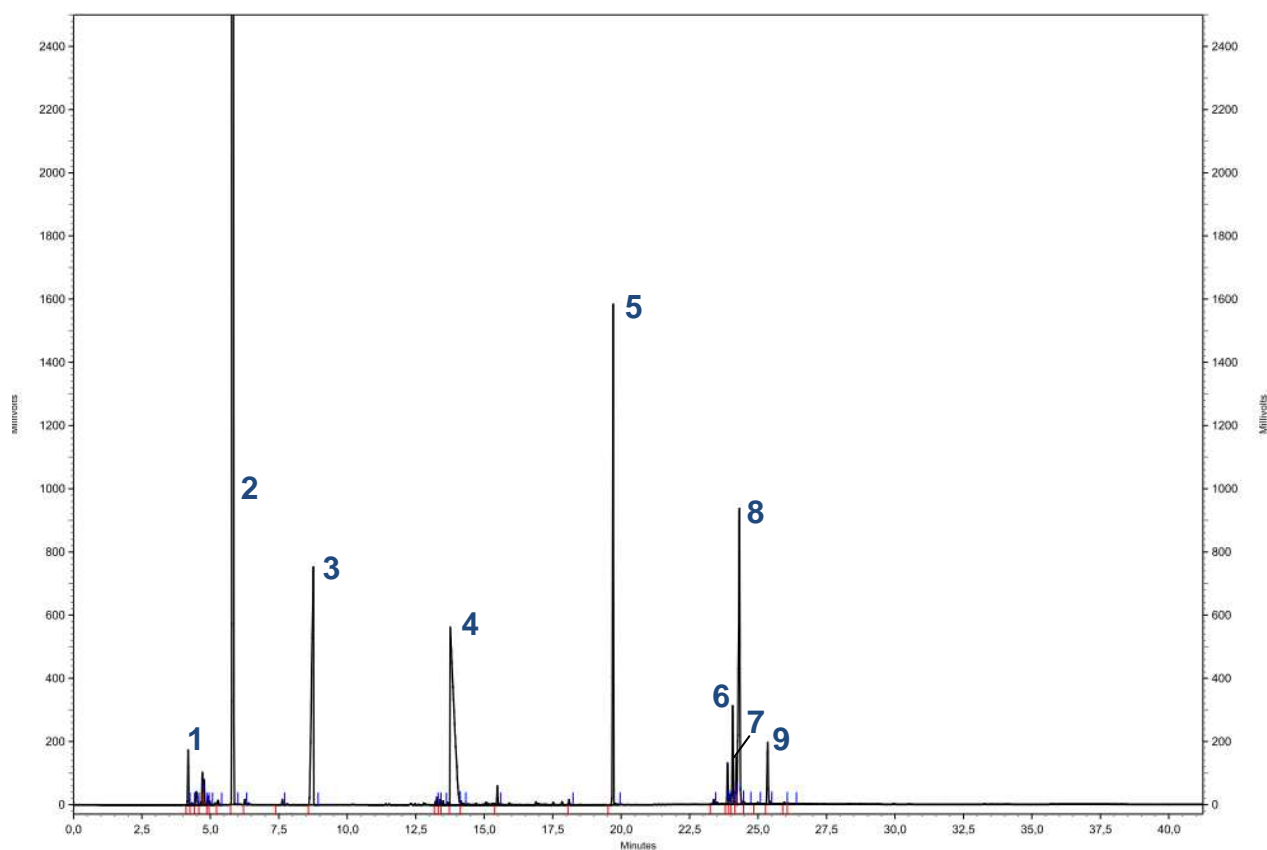
Entry 4



Entry	Retention Time [min]	Substance	Area
1	4.702	1-Methyl cyclohexene	3189899
2 ^[a]	5.777	CDCl ₃ (Solv.)	374078246
3	8.753	<i>n</i> -Dodecane (Stand.)	36190743
4	13.740	CH ₃ COOH (Solv.)	59138955
5	19.698	1-Phenylethanol (Stand.)	26539261
6	24.077	2-Methyl cyclohexane carboxylic acid	6957721
7	24.202	3-Methyl cyclohexane carboxylic acid	3302187
8	24.325	4-Methyl cyclohexane carboxylic acid	31599858
9	25.357	Carboxymethyl cyclohexane carboxylic acid	6345452

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

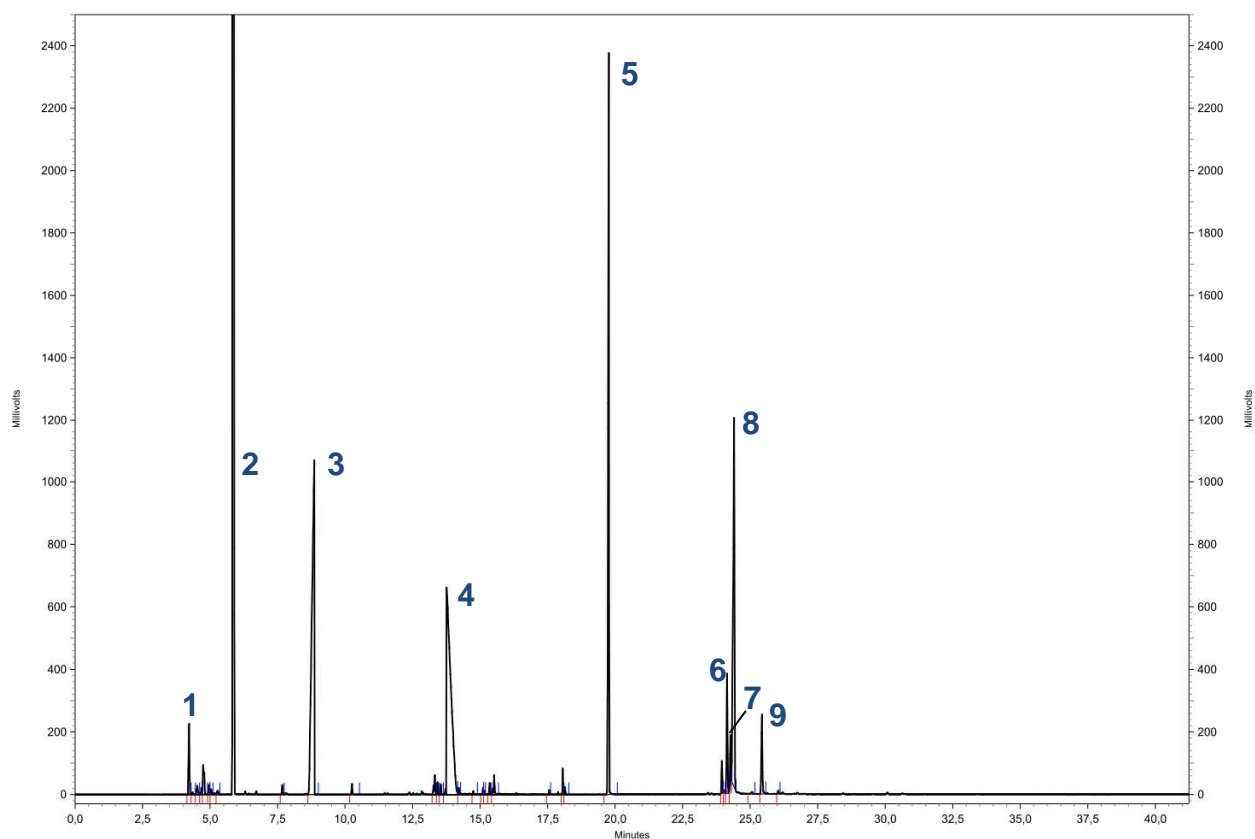
Entry 5



Entry	Retention Time [min]	Substance	Area
1	4.492	3-Methyl cyclohexene	1548035
2 ^[a]	5.783	CDCl ₃ (Solv.)	371868926
3	8.762	<i>n</i> -Dodecane (Stand.)	35212762
4	13.757	CH ₃ COOH (Solv.)	54646215
5	19.708	1-Phenylethanol (Stand.)	29990772
6	24.073	2-Methyl cyclohexane carboxylic acid	7105626
7	24.200	3-Methyl cyclohexane carboxylic acid	3443924
8	24.323	4-Methyl cyclohexane carboxylic acid	32876073
9	25.352	Carboxymethyl cyclohexane carboxylic acid	5608579

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Entry 6



Entry	Retention Time [min]	Substance	Area
1	4.223	4-Methyl cyclohexene	4856166
2 ^[a]	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
3	8.862	<i>n</i> -Dodecane (Stand.)	68225275
4	13.758	CH ₃ COOH (Solv.)	73330100
5	19.765	1-Phenylethanol (Stand.)	54515566
6	24.145	2-Methyl cyclohexane carboxylic acid	8691609
7	24.272	3-Methyl cyclohexane carboxylic acid	2237887
8	24.407	4-Methyl cyclohexane carboxylic acid	41916337
9	25.433	Carboxymethyl cyclohexane carboxylic acid	6620257

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Data of the isolated product to Entry 6

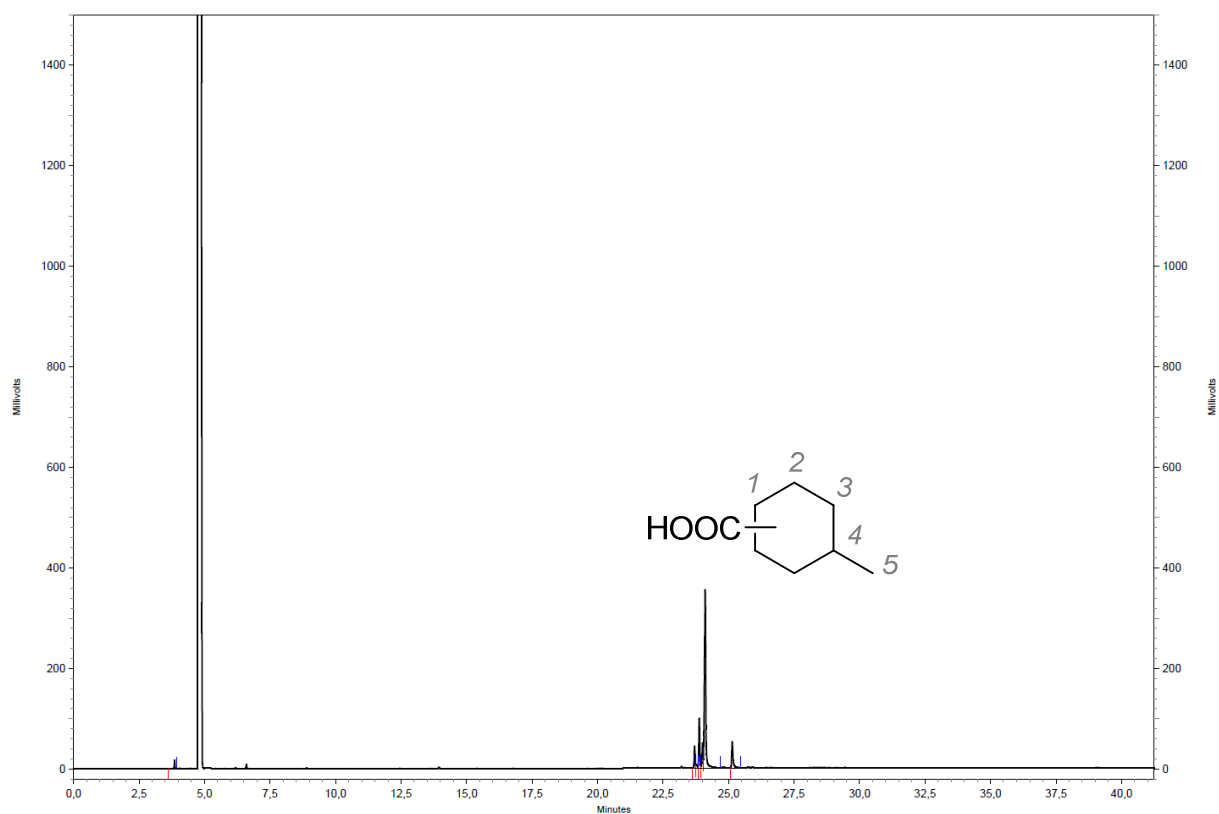


Figure 3.5 GC Chromatogram of the isolated product mixture.

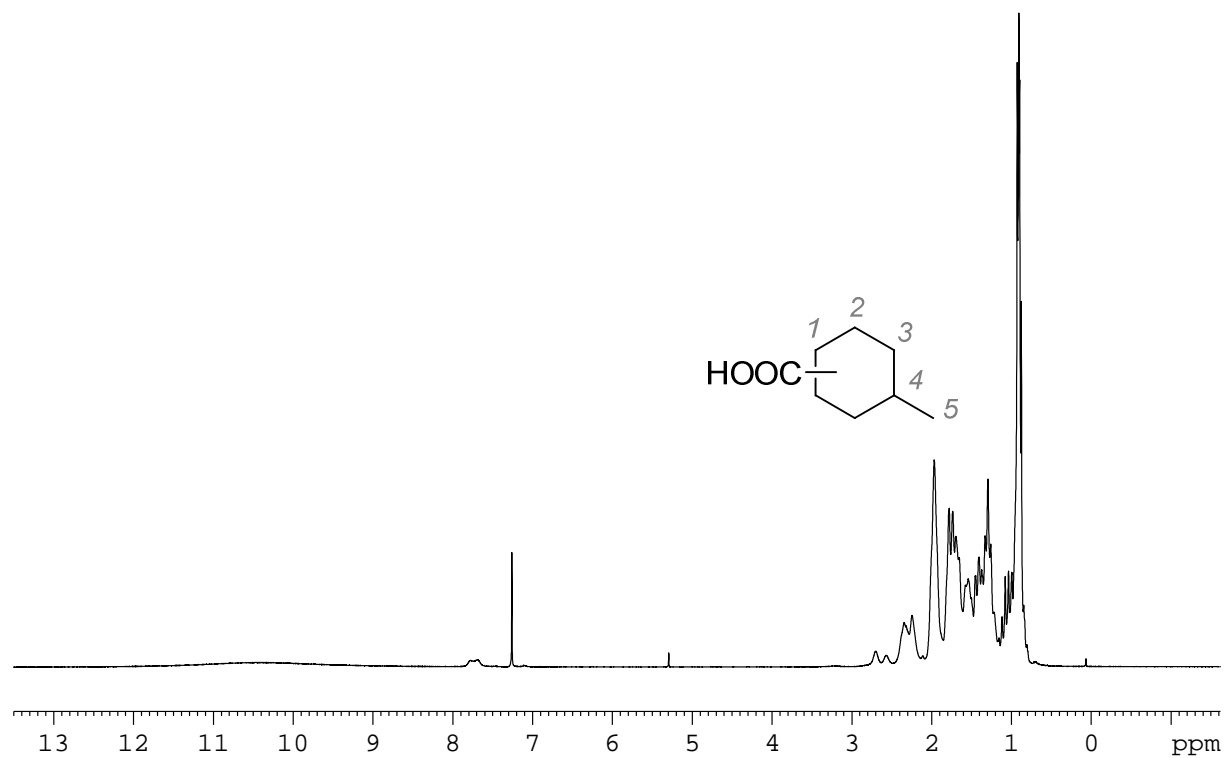
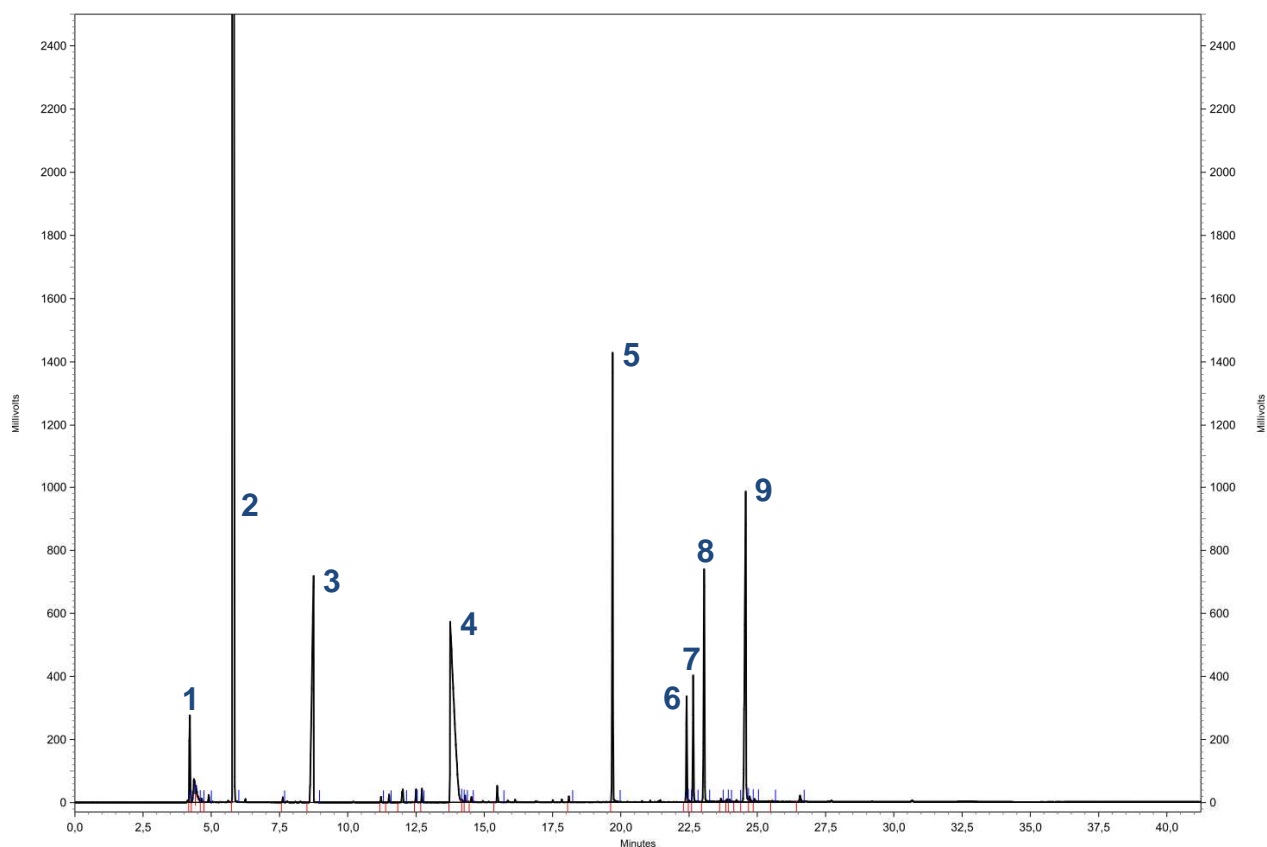


Figure 3.6. ¹H NMR spectrum of the isolated product mixture measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

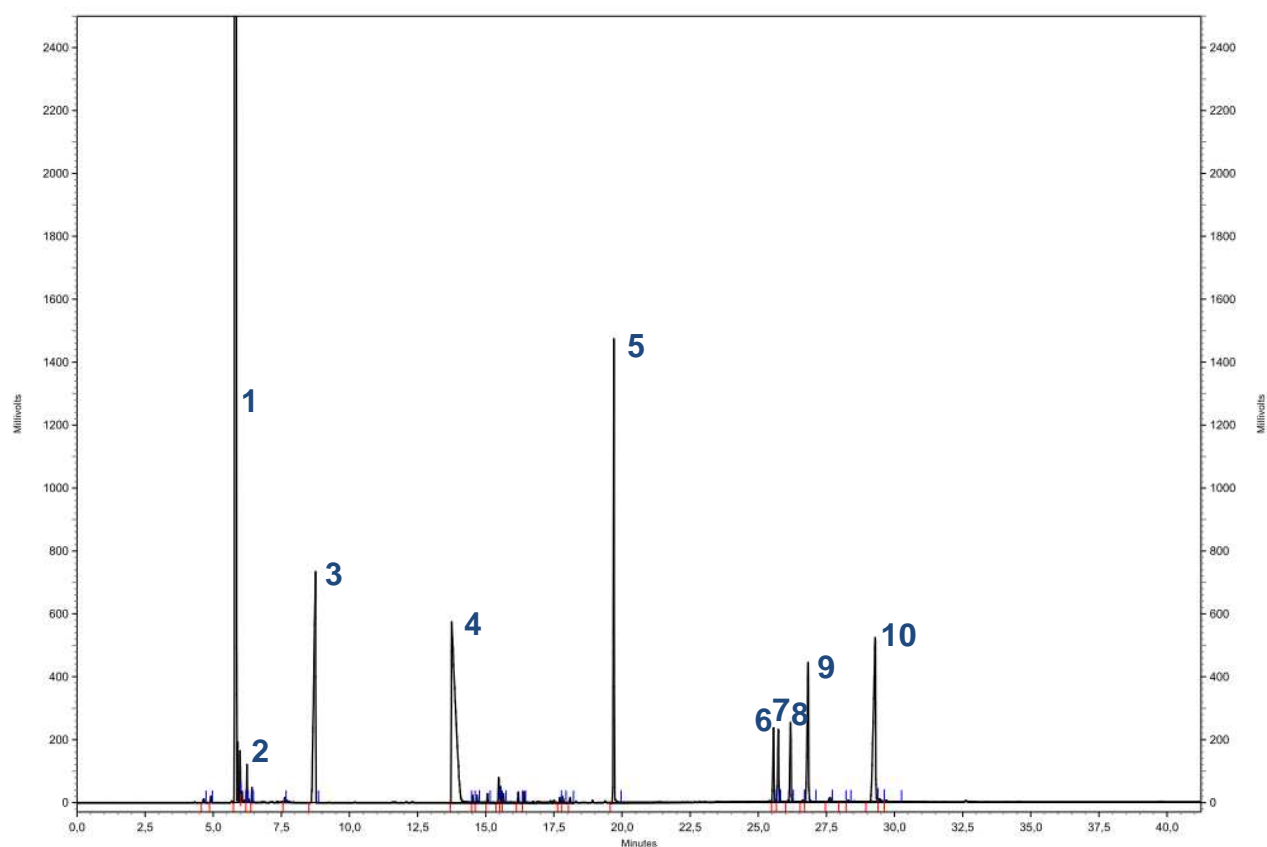
Entry 7



Entry	Retention Time [min]	Substance	Area
1	4.908	1-Octene	553025
2 ^[a]	5.780	CDCl ₃ (Solv.)	359078923
3	8.745	<i>n</i> -Dodecane (Stand.)	32088523
4	13.752	CH ₃ COOH (Solv.)	57547470
5	19.703	1-Phenylethanol (Stand.)	27356427
6	22.410	2-Propyl hexanoic acid	6277807
7	22.652	2-Ethyl heptanoic acid	7866978
8	23.055	2-Methyl octanoic acid	17459174
9	24.573	<i>n</i>-Nonanoic acid	32665607

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

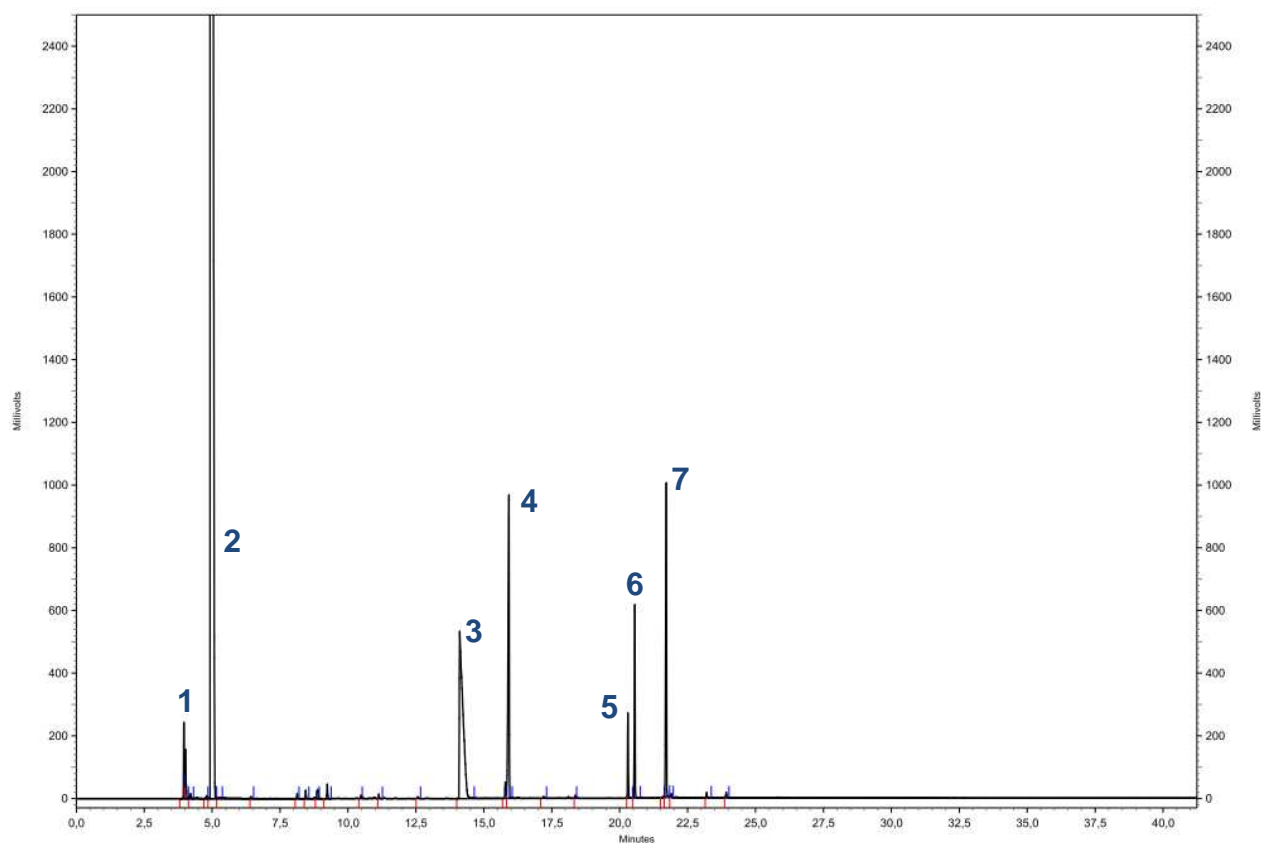
Entry 8



Entry	Retention Time [min]	Substance	Area
1 ^[a]	5.787	CDCl ₃ (Solv.)	374207983
2	6.235	1-Decene	1708970
3	8.753	<i>n</i> -Dodecane (Stand.)	33940046
4	13.745	CH ₃ COOH (Solv.)	58204489
5	19.703	1-Phenylethanol (Stand.)	28592481
6	25.562	2-Butyl heptanoic acid	6474445
7	25.737	2-Propyl octanoic acid	6161028
8	26.187	2-Ethyl nonane carboxylic acid	7543975
9	26.830	2-Methyl decanoic acid	16369660
10	29.290	<i>n</i>-Undecanoic acid	29869254

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Entry 9



Entry	Retention Time [min]	Substance	Area
1	4.022	1-Hexene	2844752
2	4.928	CH ₂ Cl ₂ (Solv.)	549670979
3	14.110	CH ₃ COOH (Solv.)	46299377
4 ^[a]	15.927	<i>n</i> -Octanol (Stand.)	26466472
5	20.305	2-Ethyl pentanoic acid	4229028
6	20.555	2-Methyl hexanoic acid	9939627
7	21.712	<i>n</i>-Heptanoic acid	19681225

[a]: *n*-Octanol was used in this experiment to avoid signal overlap with the substrate peak.

Data of the isolated product to Entry 9

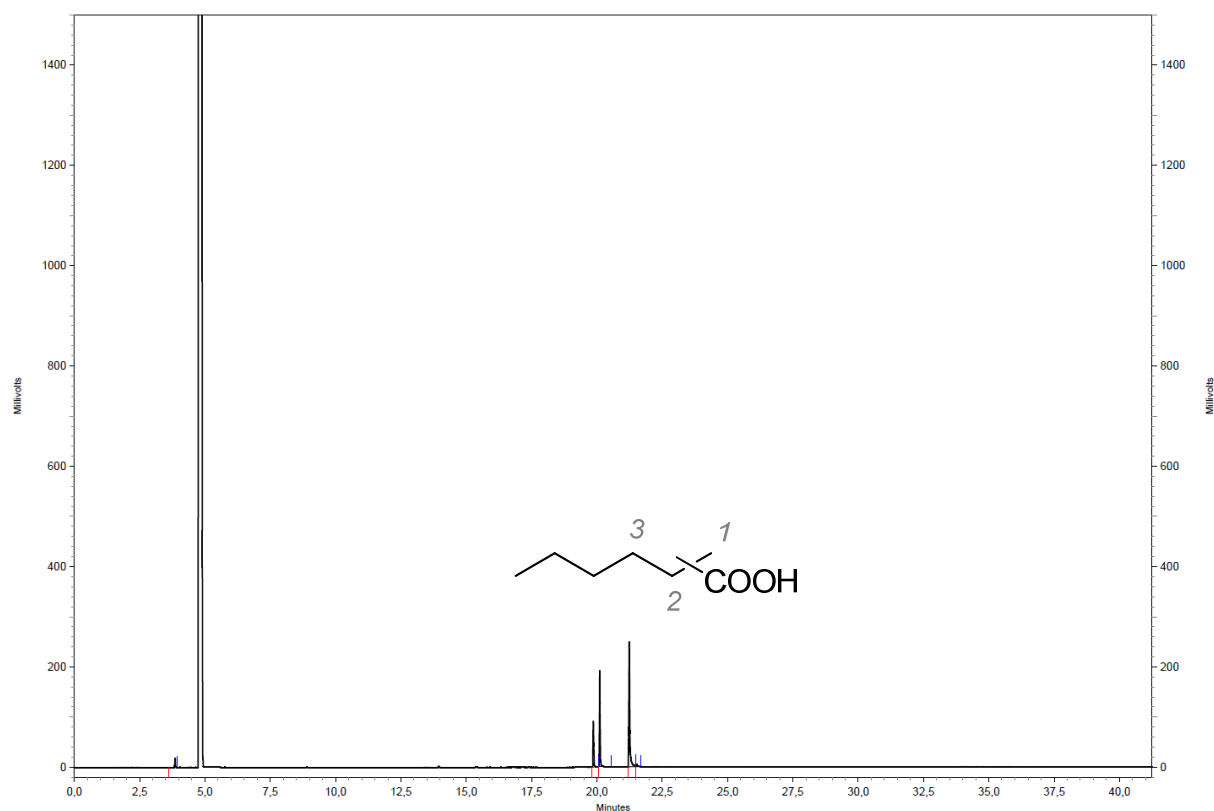


Figure 3.7 GC chromatogram of the isolated product mixture.

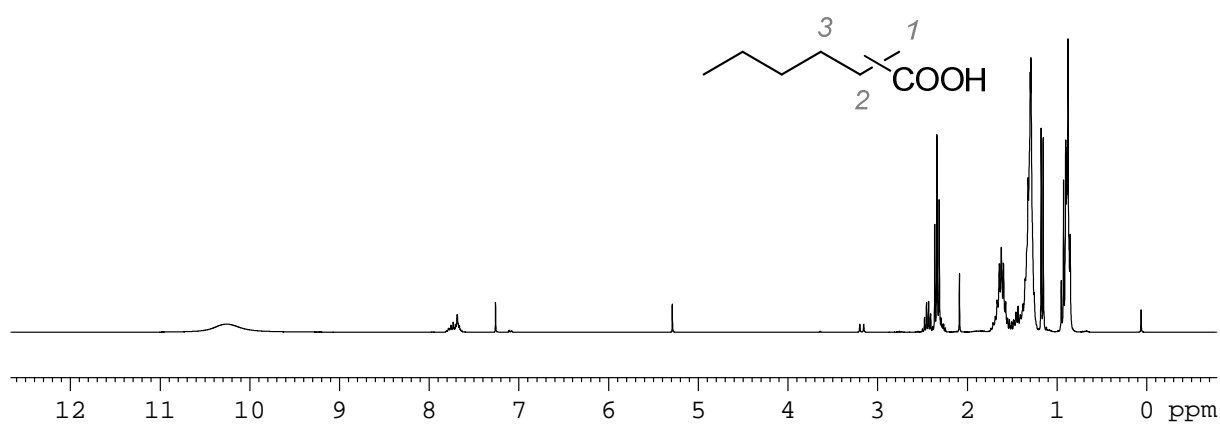
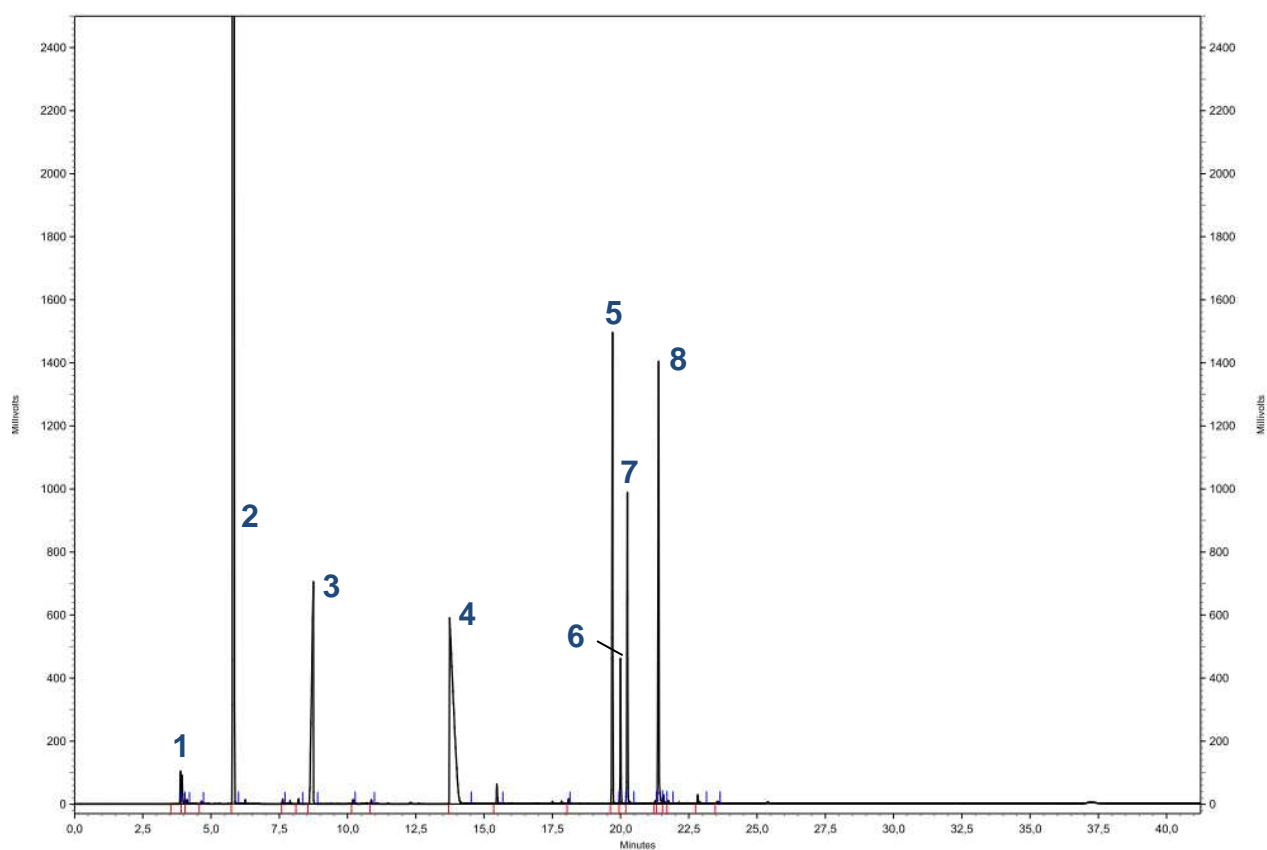


Figure 3.8. ¹H NMR spectrum of the isolated product mixture measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

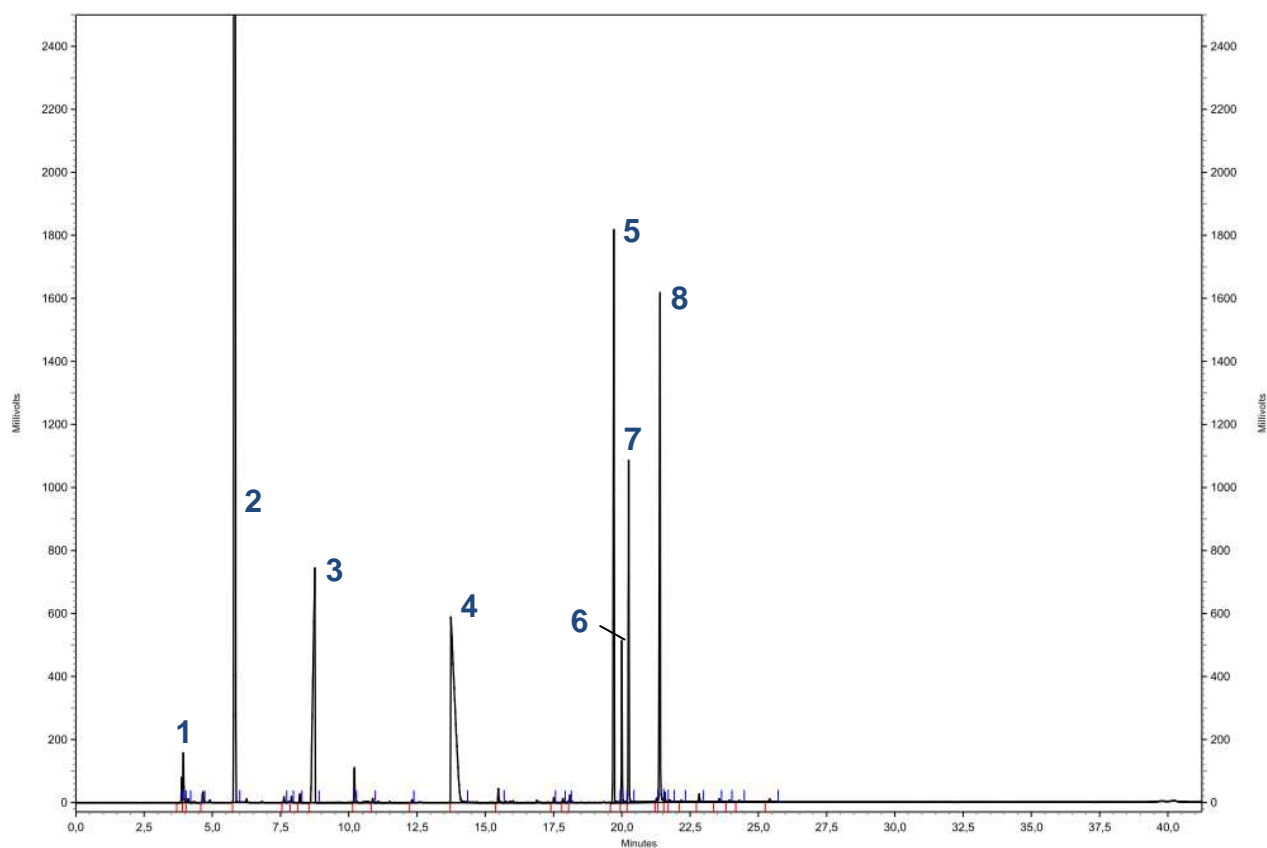
Entry 10



Entry	Retention Time [min]	Substance	Area
1	3.940	2-Hexene	1987613
2 ^[a]	5.785	CDCl ₃ (Solv.)	369417755
3	8.747	<i>n</i> -Dodecane (Stand.)	32575311
4	13.738	CH ₃ COOH (Solv.)	60717693
5	19.705	1-Phenylethanol (Stand.)	28888805
6	19.998	2-Ethyl pentanoic acid	7441529
7	20.250	2-Methyl hexanoic acid	16837208
8	21.393	<i>n</i>-Heptanoic acid	30026305

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

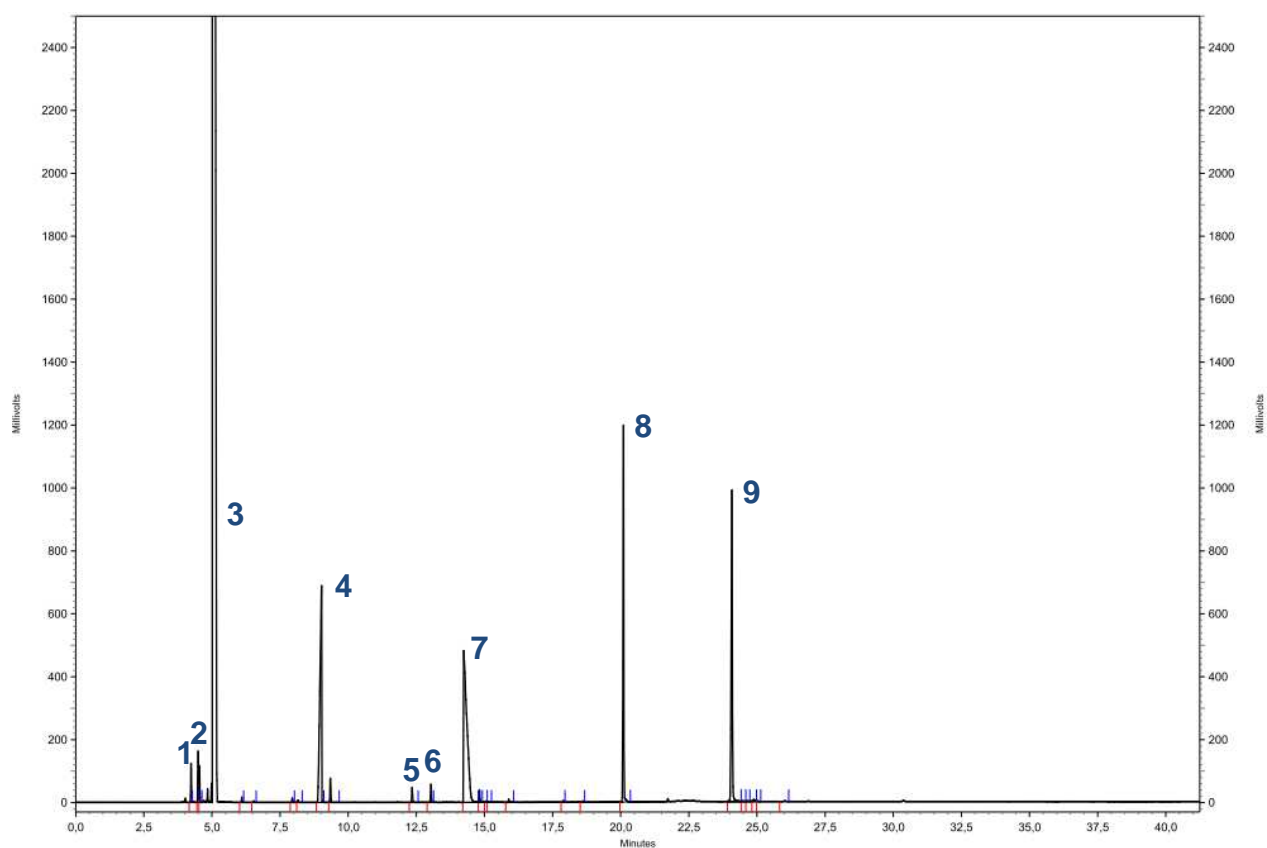
Entry 11



Entry	Retention Time [min]	Substance	Area
1	3.938	3-Hexene	3238228
2 ^[a]	5.787	CDCl ₃ (Solv.)	367788054
3	8.755	<i>n</i> -Dodecane (Stand.)	36634777
4	13.740	CH ₃ COOH (Solv.)	59999654
5	19.708	1-Phenylethanol (Stand.)	36647525
6	20.000	2-Ethyl pentanoic acid	8206306
7	20.252	2-Methyl hexanoic acid	18575822
8	21.398	<i>n</i>-Heptanoic acid	36340221

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Entry 12



Entry	Retention Time [min]	Substance	Area
1	4.235	CH	1595466
2	4.545	CE	1736854
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	9.023	<i>n</i> -Dodecane (Stand.)	30367527
5	12.347	CAc	845689
6	13.038	CI	1050040
7	14.240	CH ₃ COOH (Solv.)	40342430
8	20.102	1-Phenylethanol (Stand.)	22697158
9	24.080	CA	27697583

Data of the isolated product to Entry 12

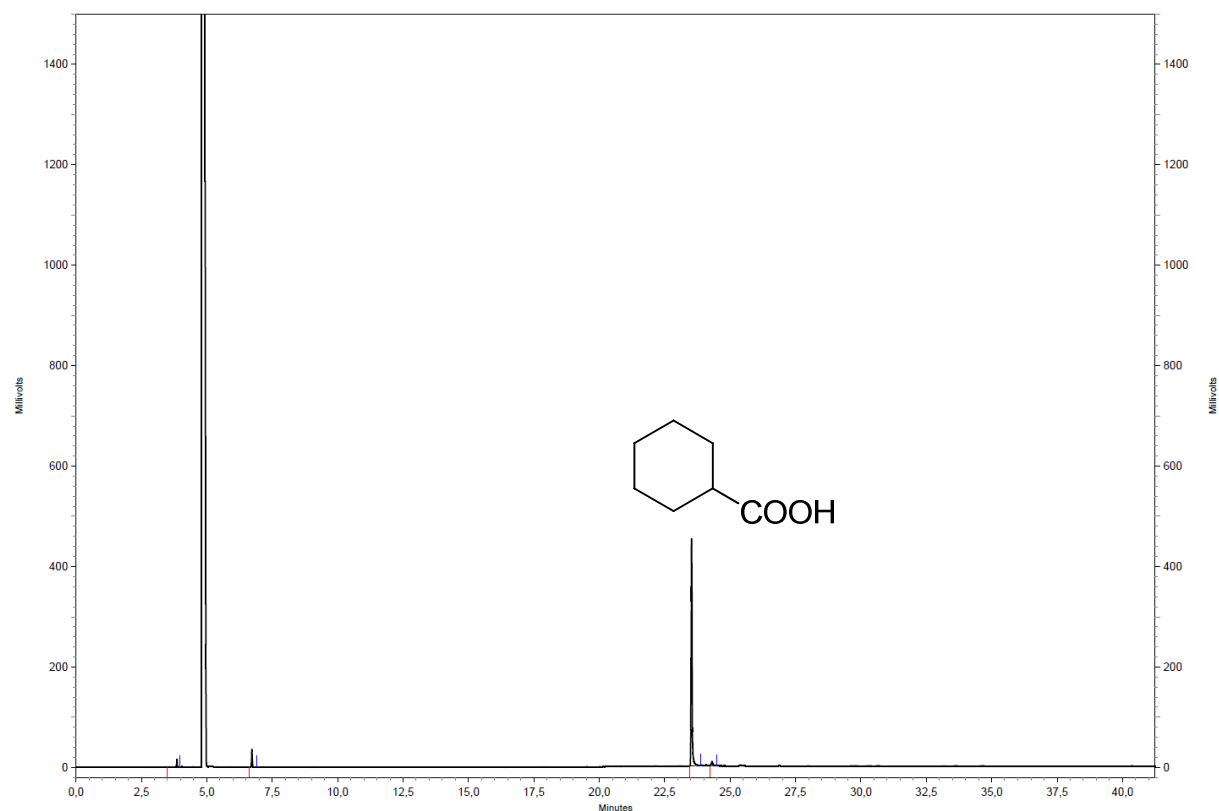


Figure 3.9 GC chromatogram of the isolated cyclohexane carboxylic acid.

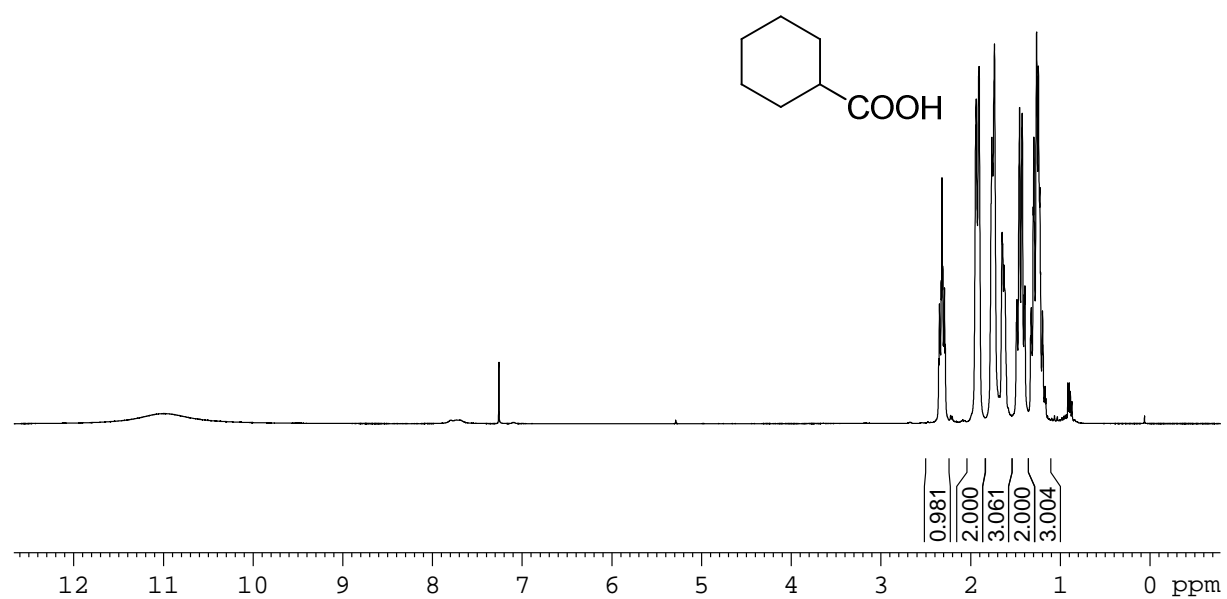
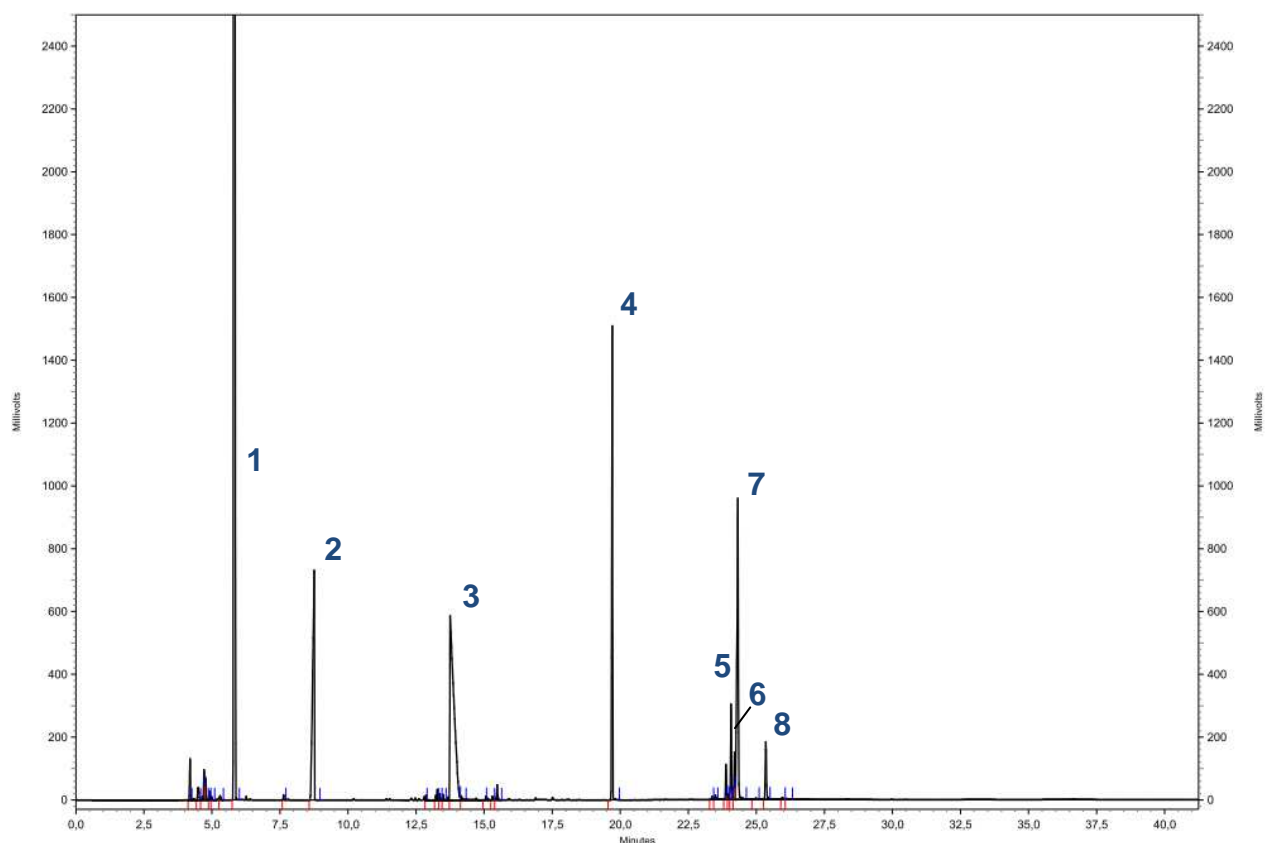


Figure 3.10. ¹H NMR spectrum of the isolated product measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

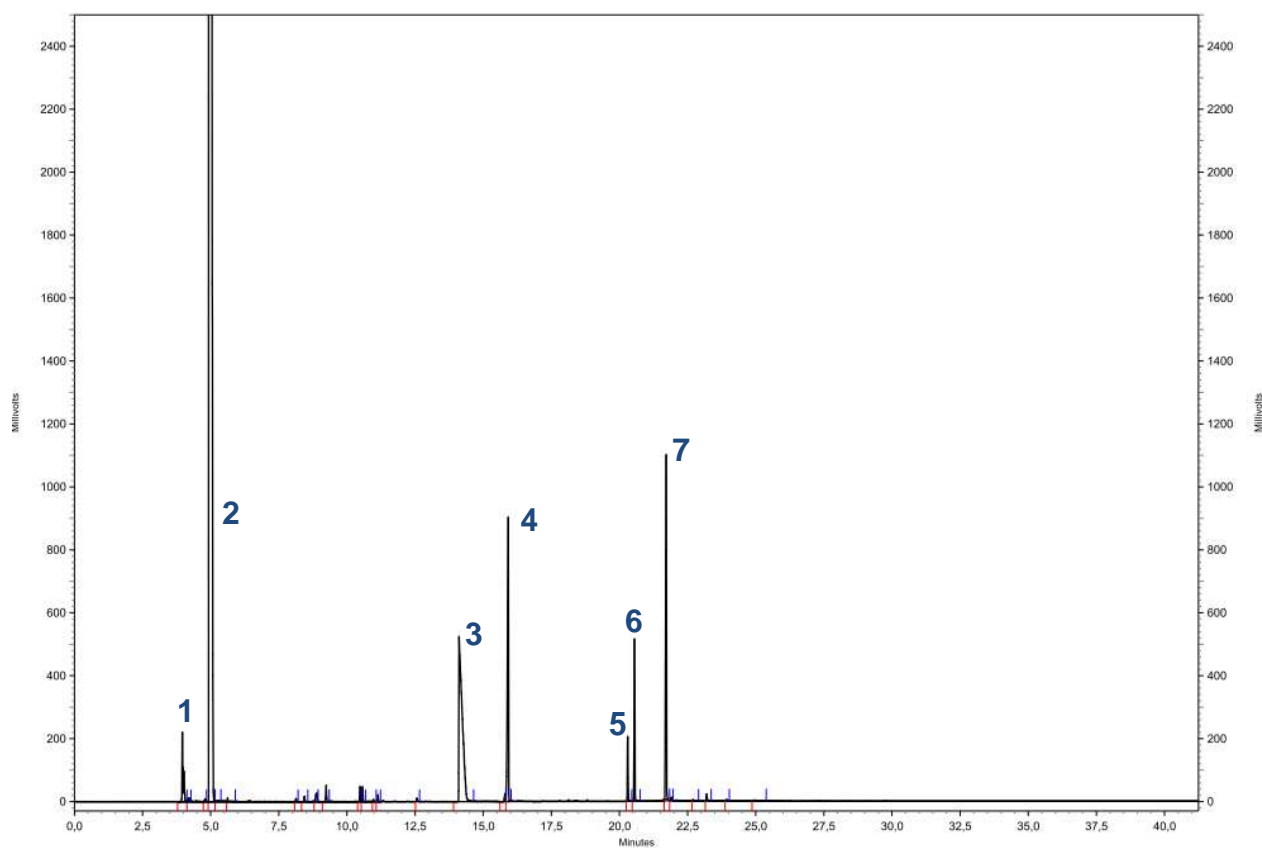
Entry 13



Entry	Retention Time [min]	Substance	Area
1 ^[a]	5.790	CDCl ₃ (Solv.)	367119043
2	8.760	<i>n</i> -Dodecane (Stand.)	33495695
3	13.747	CH ₃ COOH (Solv.)	59139257
4	19.707	1-Phenylethanol (Stand.)	29286436
5	24.073	2-Methyl cyclohexane carboxylic acid	7063099
6	24.198	3-Methyl cyclohexane carboxylic acid	3499874
7	24.325	4-Methyl cyclohexane carboxylic acid	34118403
8	25.350	Carboxymethyl cyclohexane carboxylic acid	5247547

[a]: CDCl₃ was used in this experiment to dilute the reaction mixture in order to avoid signal overlap with the substrate peak and allow for simultaneous NMR analysis.

Entry 14



Entry	Retention Time [min]	Substance	Area
1	3.958	Hexane	5716156
2	4.927	CH ₂ Cl ₂ (Solv.)	556022214
3	14.112	CH ₃ COOH (Solv.)	44565895
4	15.922	<i>n</i> -Octanol (Stand.)	23743985
5	20.303	2-Ethyl pentanoic acid	3152384
6	20.552	2-Methyl hexanoic acid	8133389
7	21.713	<i>n</i>-Heptanoic acid	21904251

[a]: *n*-Octanol was used in this experiment to avoid signal overlap with the substrate peak.

Data of the isolated product to Entry 14

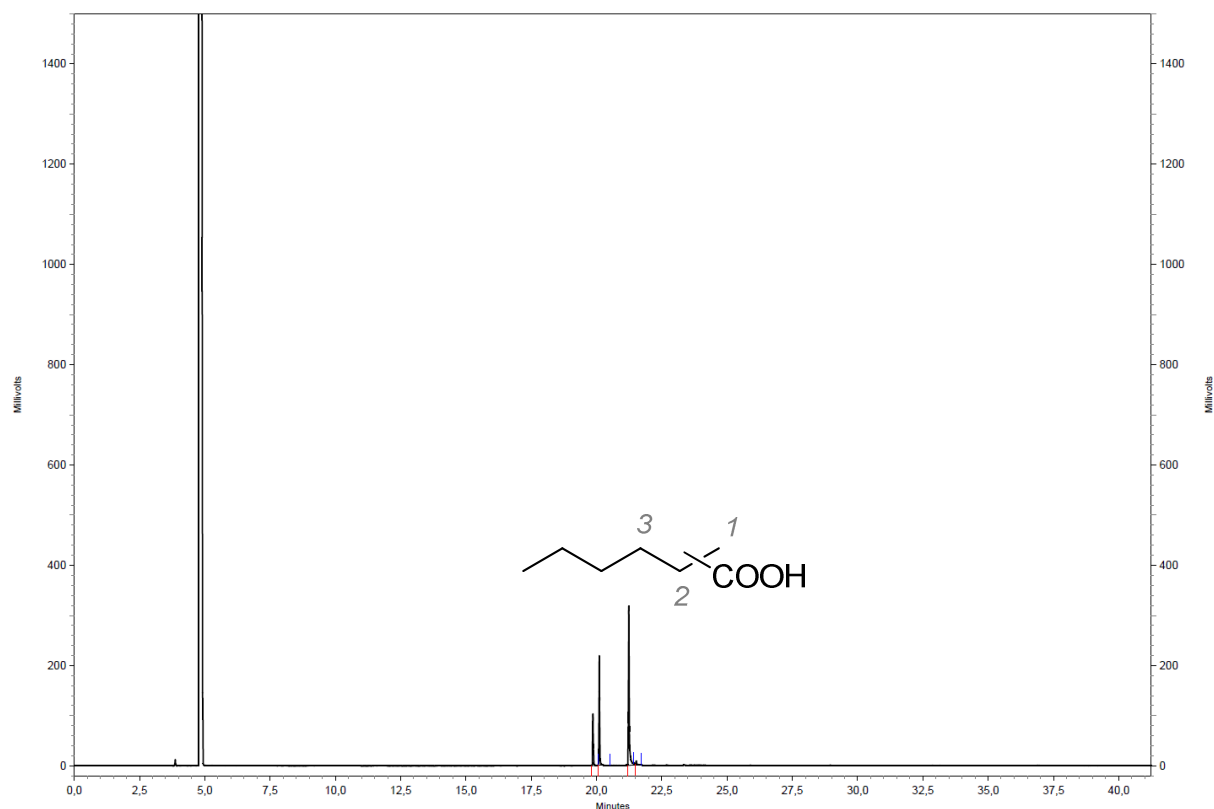


Figure 3.11 GC chromatogram of the isolated product mixture.

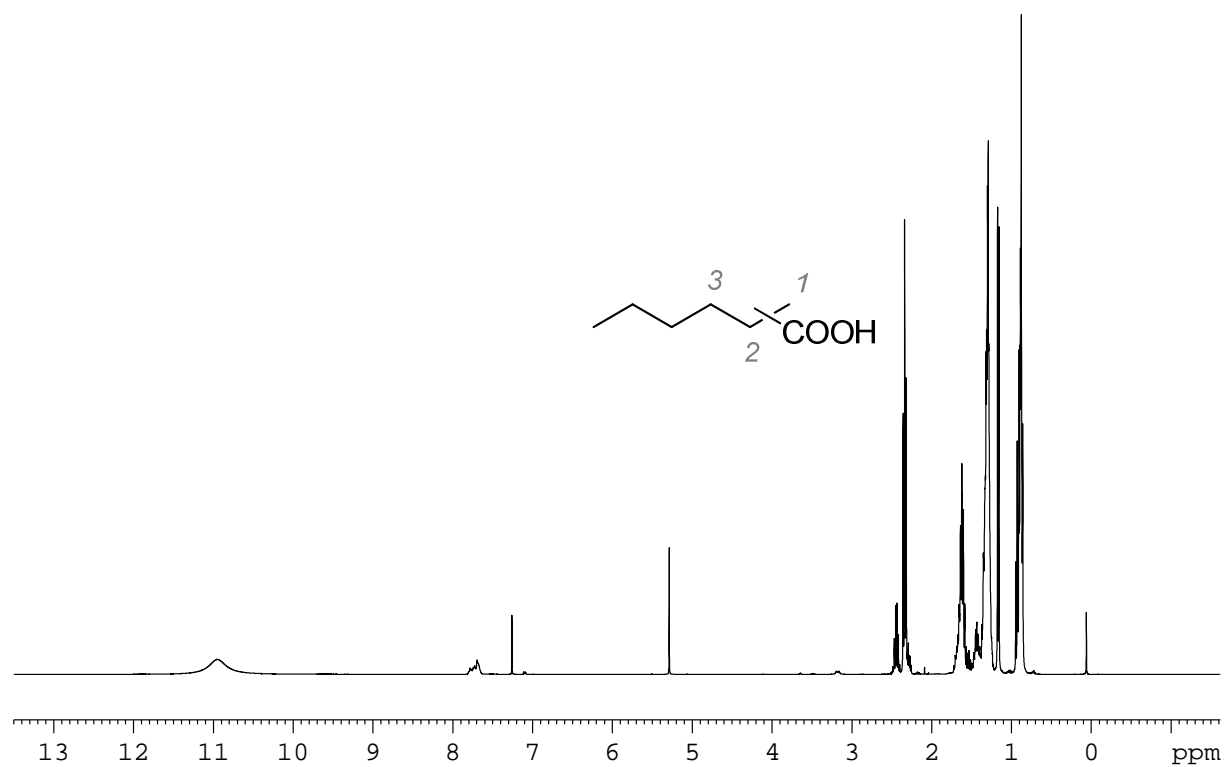
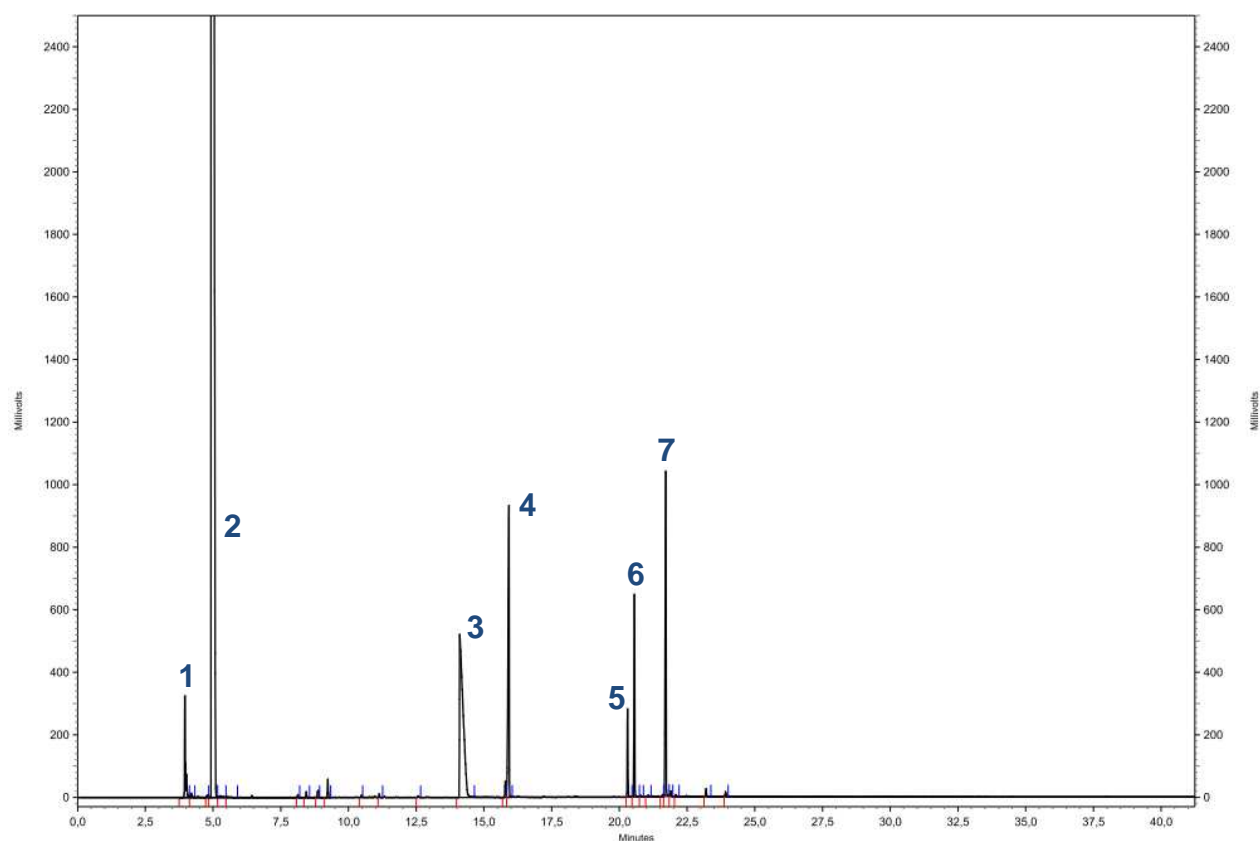


Figure 3.12. ¹H NMR spectrum of the isolated product mixture measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

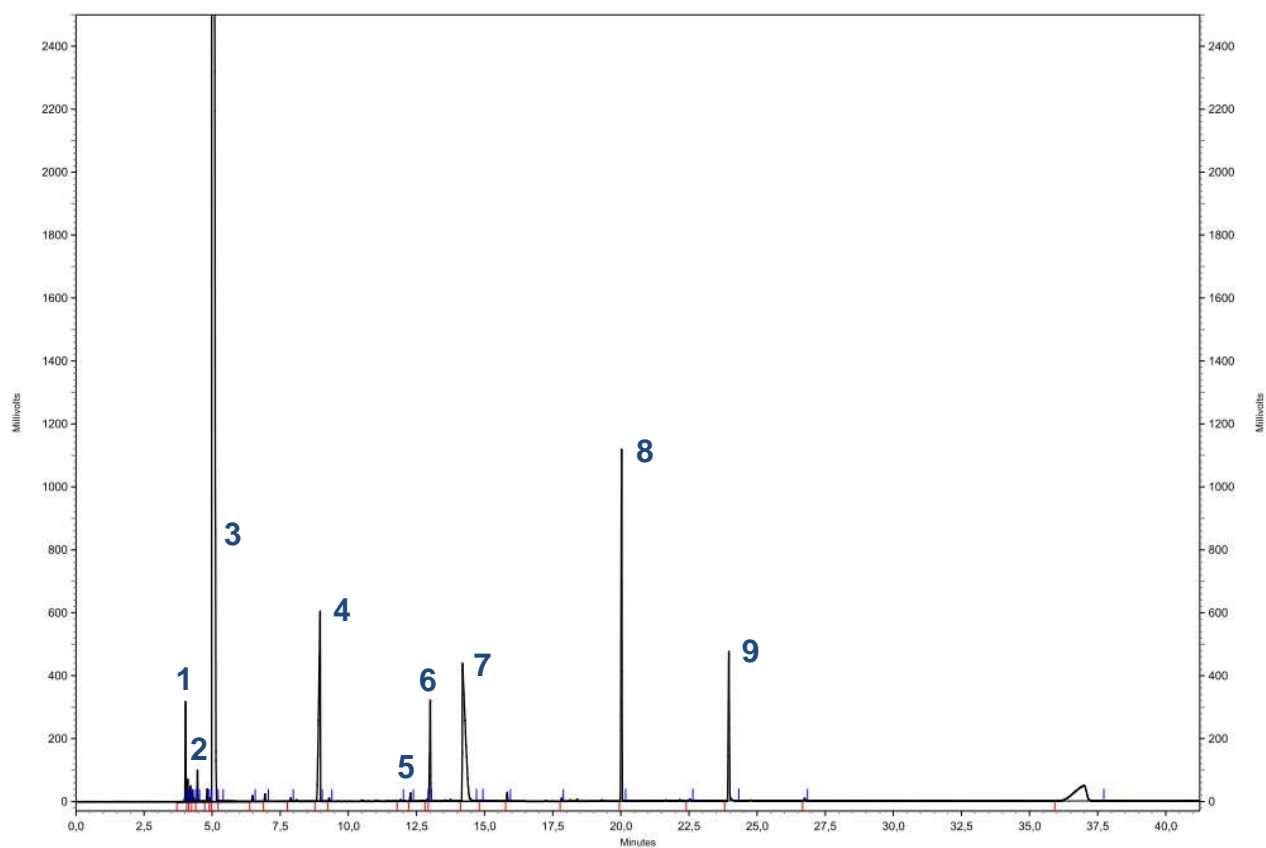
Entry 15



Entry	Retention Time [min]	Substance	Area
1	3.960	Hexane	6557964
2	4.927	CH ₂ Cl ₂ (Solv.)	553192172
3	14.107	CH ₃ COOH (Solv.)	45606092
4 ^[a]	15.923	<i>n</i> -Octanol (Stand.)	25208028
5	20.305	2-Ethyl pentanoic acid	4348062
6	20.555	2-Methyl hexanoic acid	10543606
7	21.712	<i>n</i>-Heptanoic acid	20621202

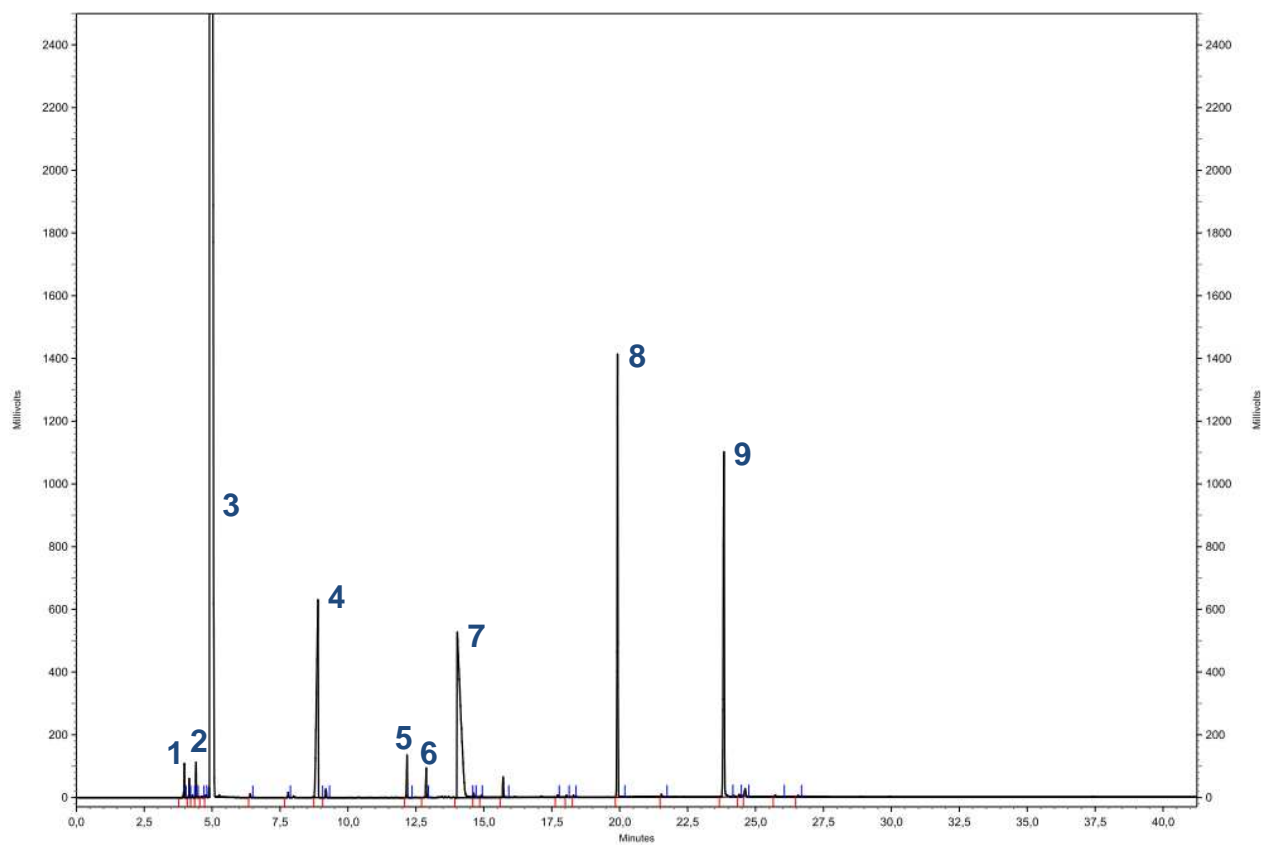
[a]: *n*-Octanol was used in this experiment to avoid signal overlap with the substrate peak.

Entry 16



Entry	Retention Time [min]	Substance	Area
1	4.105	CH	886422
2	4.462	CE	1521631
3	4.993	CH ₂ Cl ₂ (Solv.)	442202356
4	8.965	<i>n</i> -Dodecane (Stand.)	23486393
5	12.275	CAc	469594
6	13.003	CI	6920339
7	14.188	CH ₃ COOH (Solv.)	33074464
8	20.025	1-Phenylethanol (Stand.)	20020794
9	23.965	CA	11015083

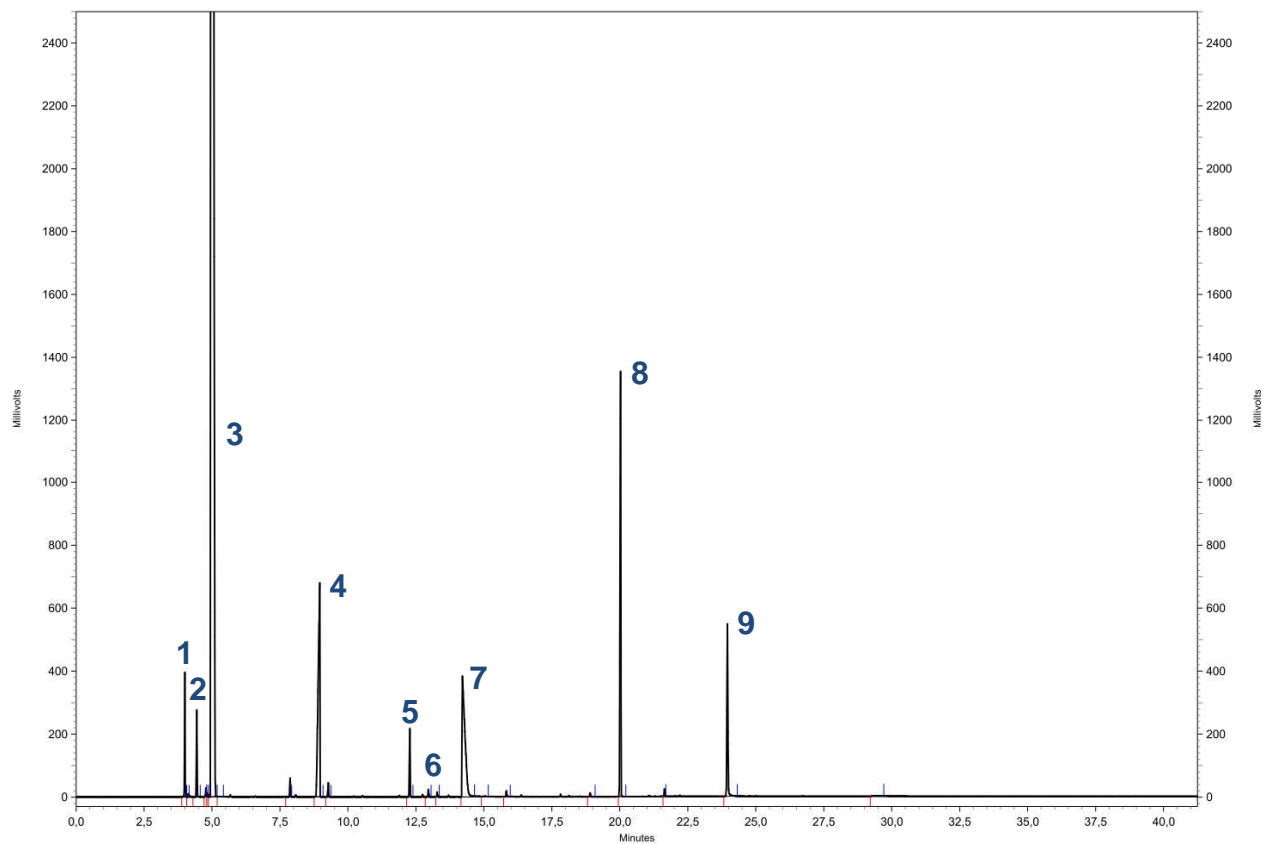
Entry 17



Entry	Retention Time [min]	Substance	Area
1	3.978	CH	1532625
2	4.408	CE	1738139
3	<i>cutted</i>	CH ₂ Cl ₂ (Solv.)	--
4	8.897	<i>n</i> -Dodecane (Stand.)	25850032
5	12.172	CAc	2219896
6	12.877	CI	1660622
7	14.023	CH ₃ COOH (Solv.)	46455180
8	19.925	1-Phenylethanol (Stand.)	27194328
9	23.837	CA	28915927

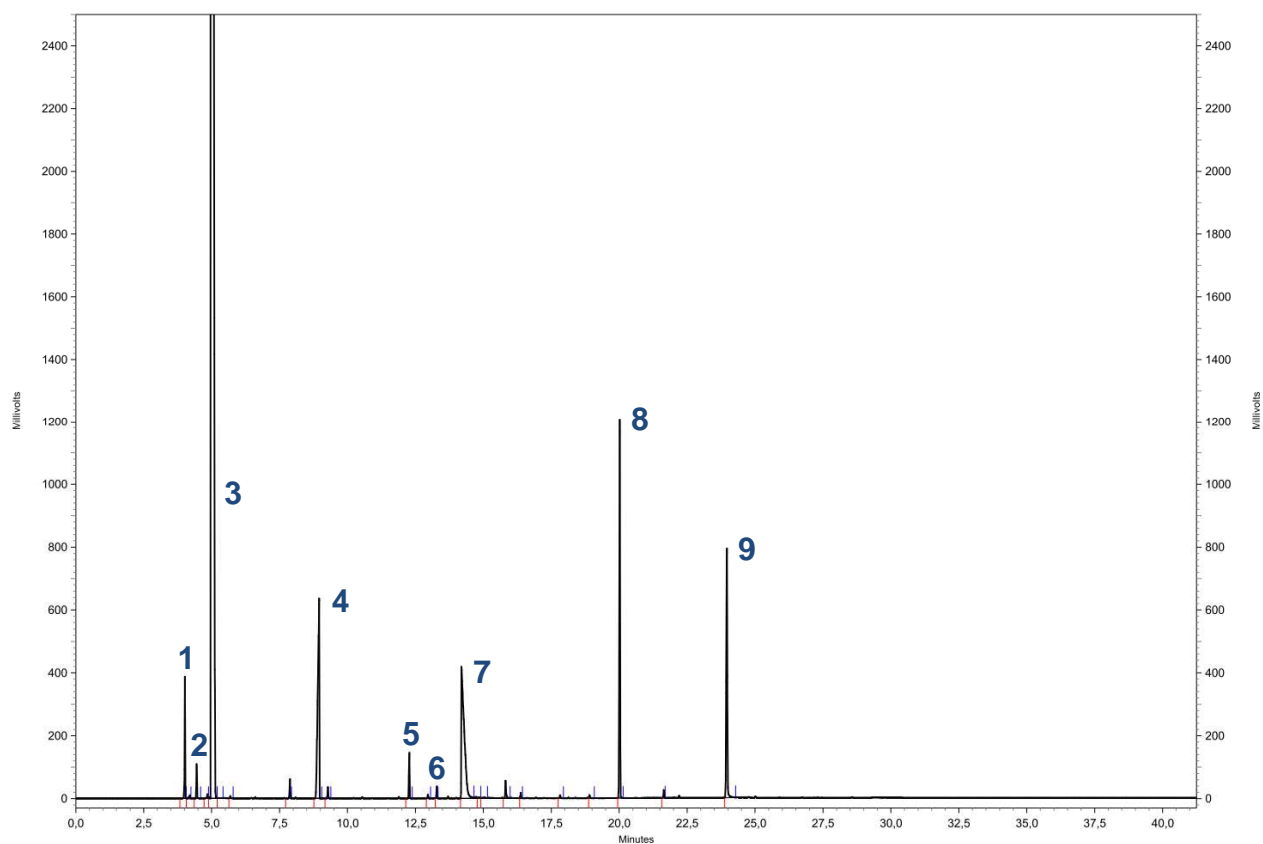
S3.8 Gaschromatograms to Table S2.8

Entry 1



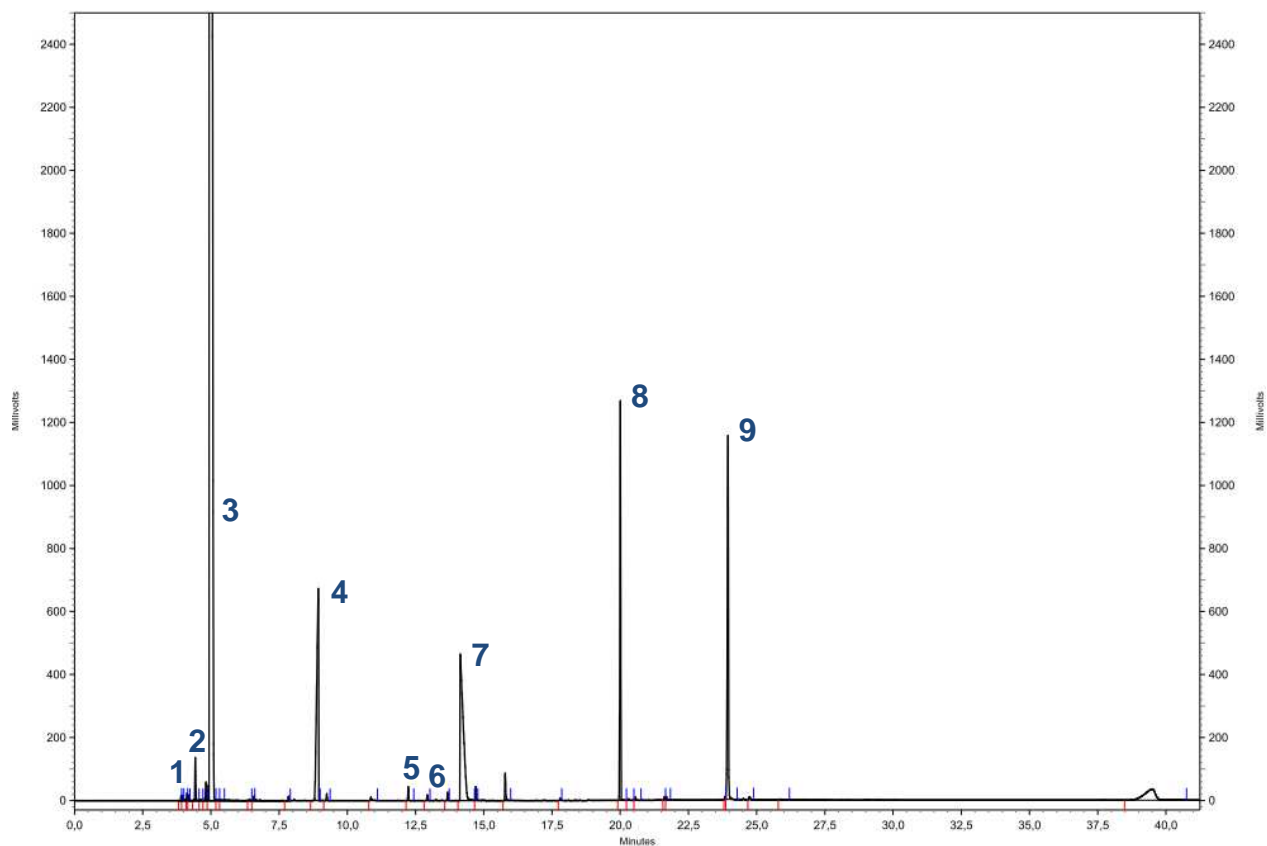
Entry	Retention Time [min]	Substance	Area
1	4.002	CH	4917794
2	4.437	CE	4097124
3	4.955	CH ₂ Cl ₂ (Solv.)	527292253
4	8.963	<i>n</i> -Dodecane (Stand.)	29266189
5	12.277	CAC	3705318
6	12.962	CI	416532
7	14.208	CH ₃ COOH (Solv.)	26990150
8	20.025	1-Phenylethanol (Stand.)	25556688
9	23.960	CA	13083714

Entry 2



Entry	Retention Time [min]	Substance	Area
1	4.013	CH	4803994
2	4.450	CE	1654564
3	4.972	CH ₂ Cl ₂ (Solv.)	515167964
4	8.960	<i>n</i> -Dodecane (Stand.)	25402549
5	12.275	CAc	2397405
6	12.963	CI	218698
7	14.195	CH ₃ COOH (Solv.)	31434402
8	20.022	1-Phenylethanol (Stand.)	22217797
9	23.963	CA	19465837

Entry 3



Entry	Retention Time [min]	Substance	Area
1	4.123	CH	345223
2	4.428	CE	2222789
3	4.935	CH ₂ Cl ₂ (Solv.)	572180609
4	8.937	<i>n</i> -Dodecane (Stand.)	29129720
5	12.233	CAc	716350
6	12.933	CI	322269
7	14.135	CH ₃ COOH (Solv.)	37123096
8	20.007	1-Phenylethanol (Stand.)	23395148
9	23.952	CA	30421755

S4 NMR and Mass Spectra

S4.1 Additional NMR Spectra to Table Table S2.7

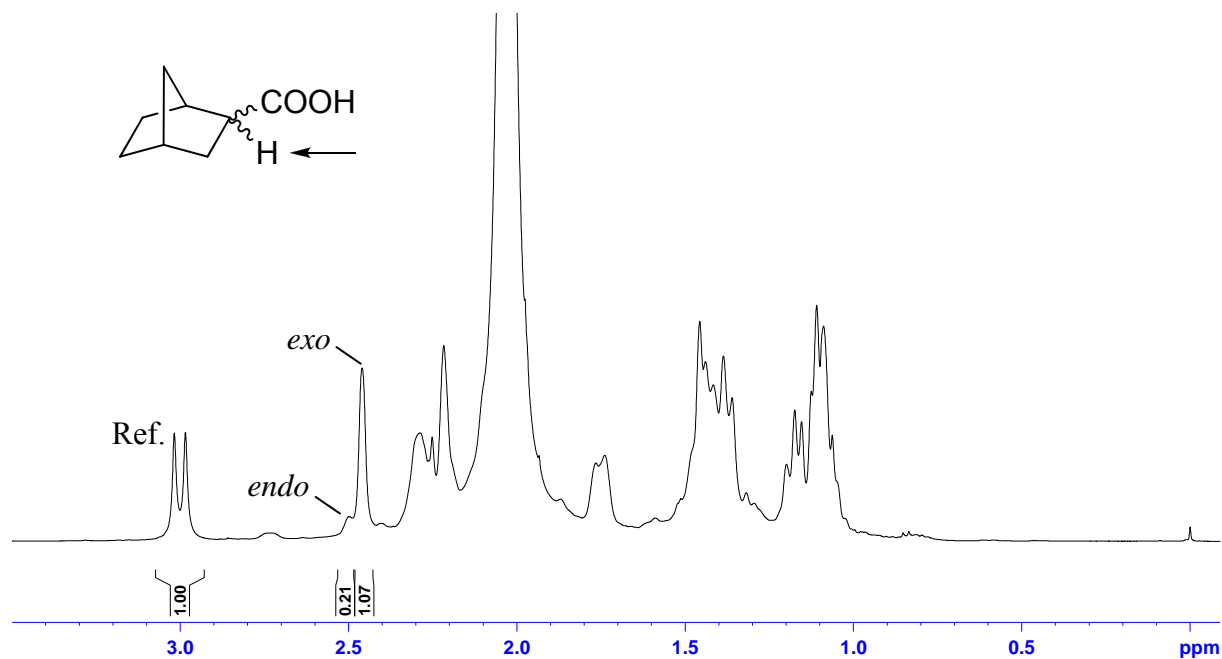


Figure S4.1. ¹H NMR spectrum of the reaction mixture after the catalysis with the substrate norbornene indicating the ratio of the integrals for *exo/endo* substitution. Measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

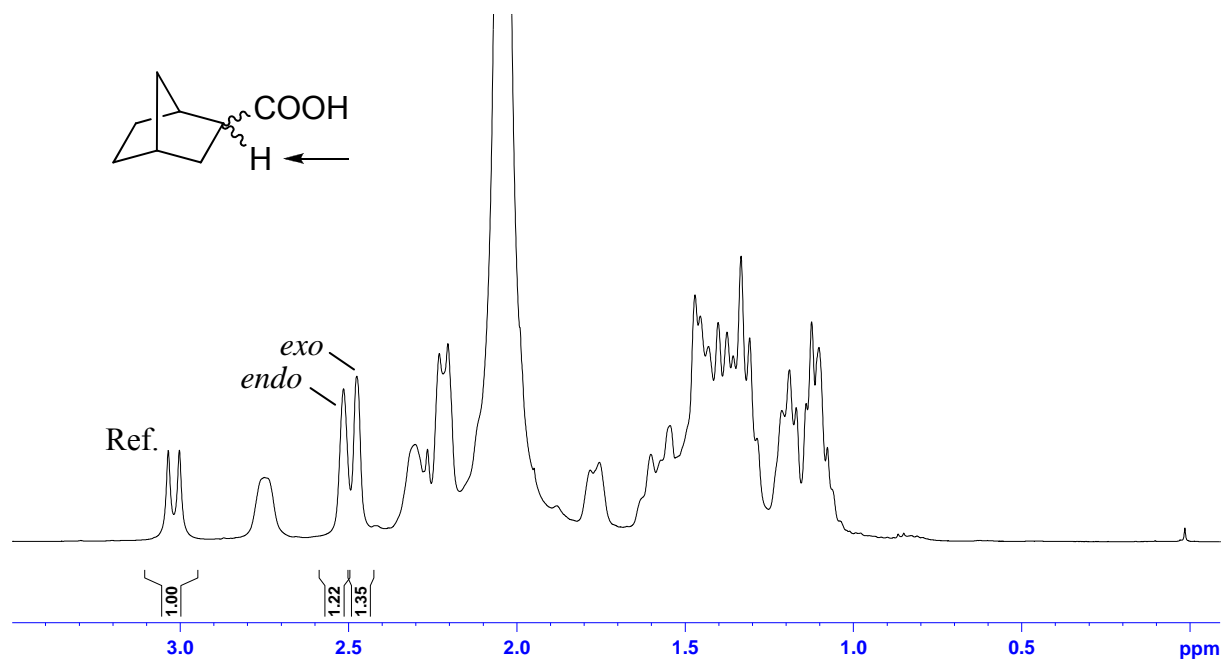


Figure S4.2. ¹H NMR spectrum of the reaction mixture after the catalysis with the substrate norbornene indicating the ratio of the integrals for *exo/endo* substitution enriched with pure *endo* product. Measured in CDCl₃ at ambient temperature with a resonance frequency of 400 Mhz.

S4.2 NMR Spectra to Table S2.9

Entry 1

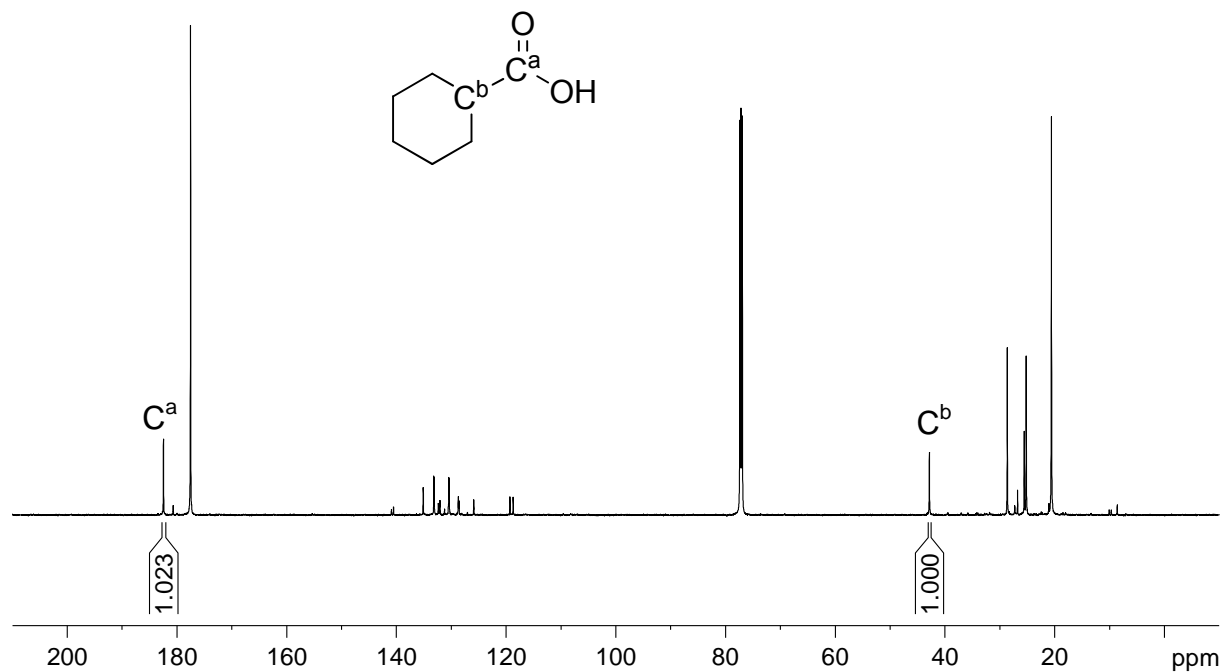


Figure S4.3. Quantitative ¹³C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for C^a:C^b. Measured in CDCl₃ at ambient temperature with a resonance frequency of 151 Mhz.

Entry 2

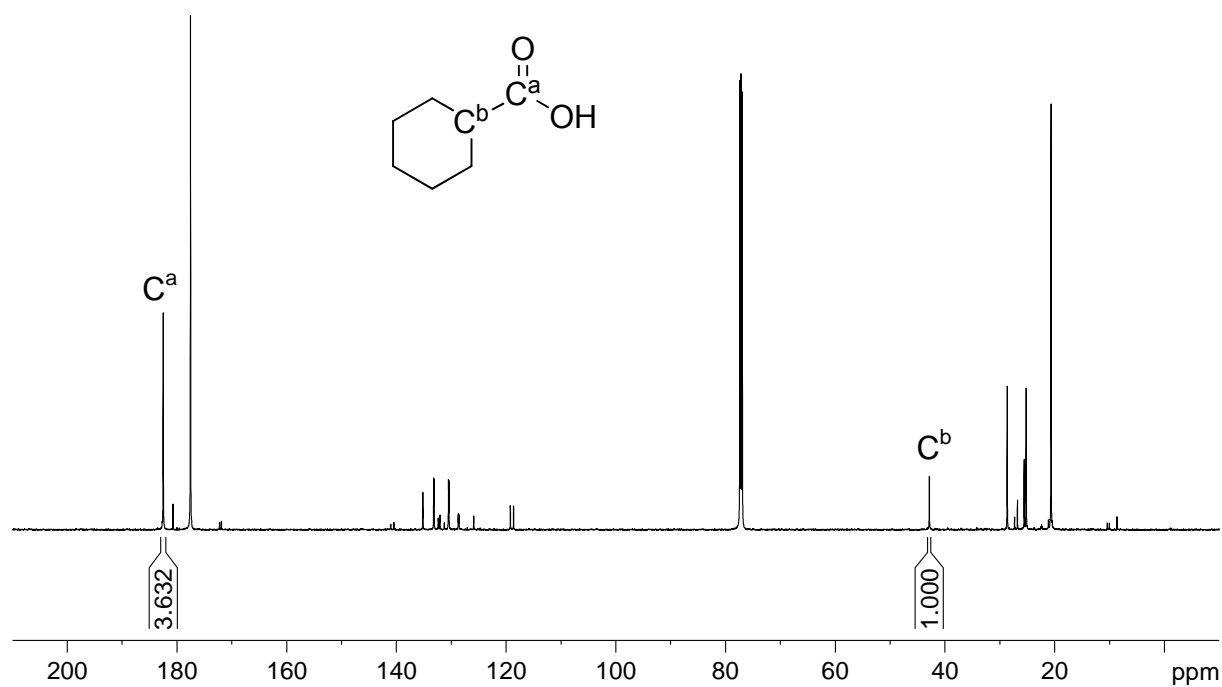


Figure S4.4. Quantitative ¹³C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for C^a:C^b. Measured in CDCl₃ at ambient temperature with a resonance frequency of 151 Mhz.

Entry 3

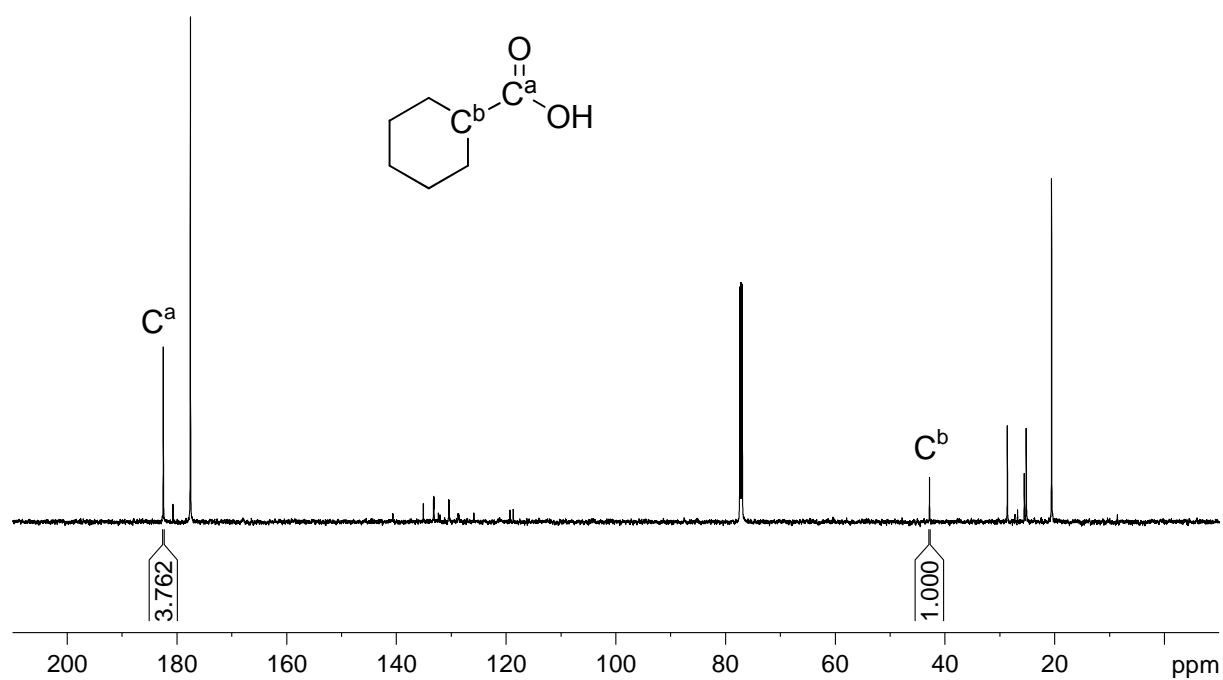


Figure S4.5 . Quantitative ^{13}C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for $C^a:C^b$. Measured in CDCl_3 at ambient temperature with a resonance frequency of 151 Mhz.

Entry 4

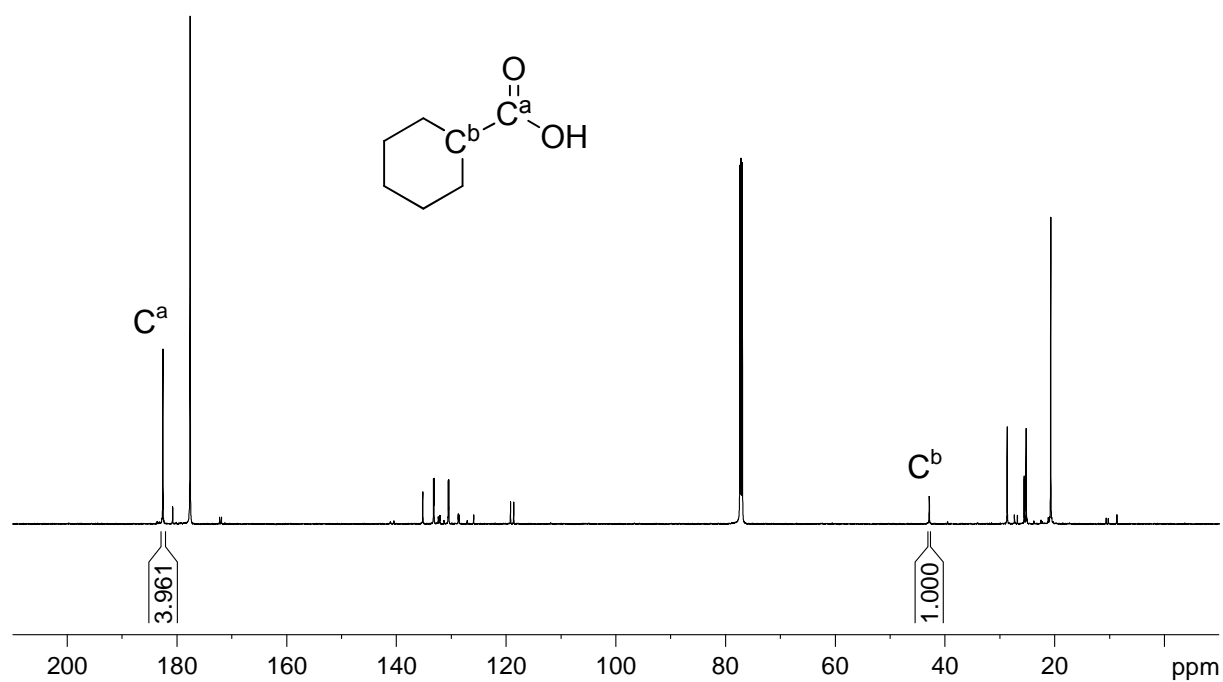


Figure S4.6. Quantitative ^{13}C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for $C^a:C^b$. Measured in CDCl_3 at ambient temperature with a resonance frequency of 151 Mhz.

Entry 5

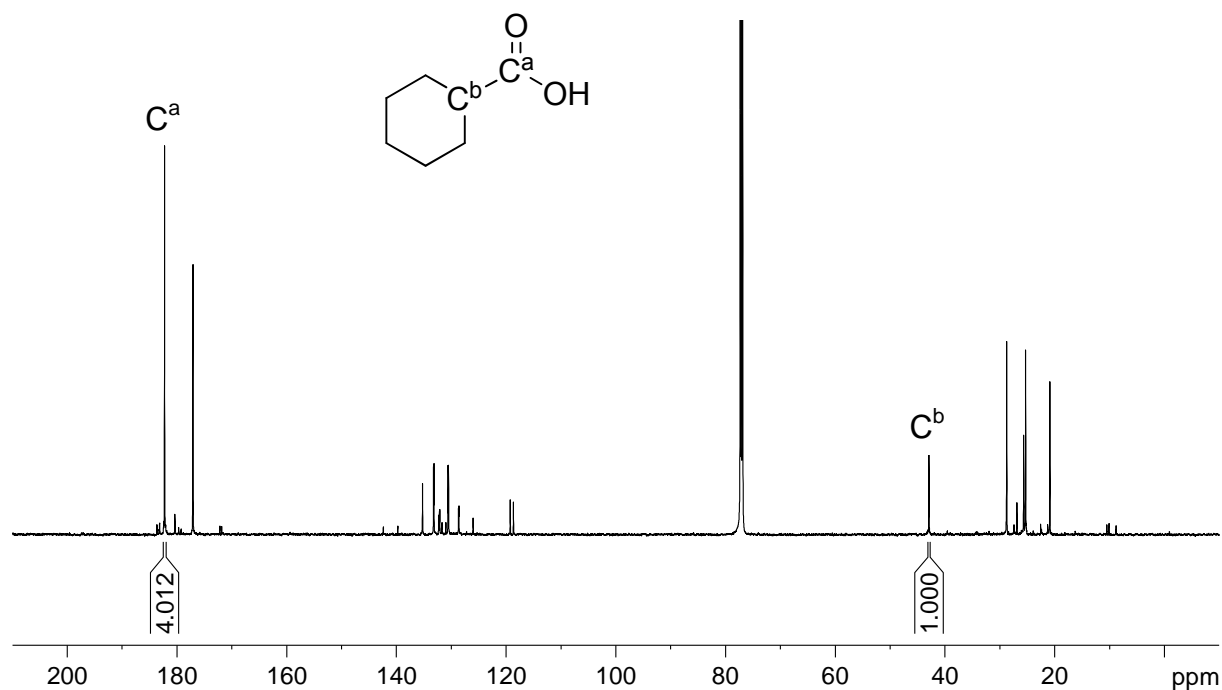


Figure S4.7. Quantitative ¹³C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for C^a:C^b. Measured in CDCl₃ at ambient temperature with a resonance frequency of 151 Mhz.

Entry 6

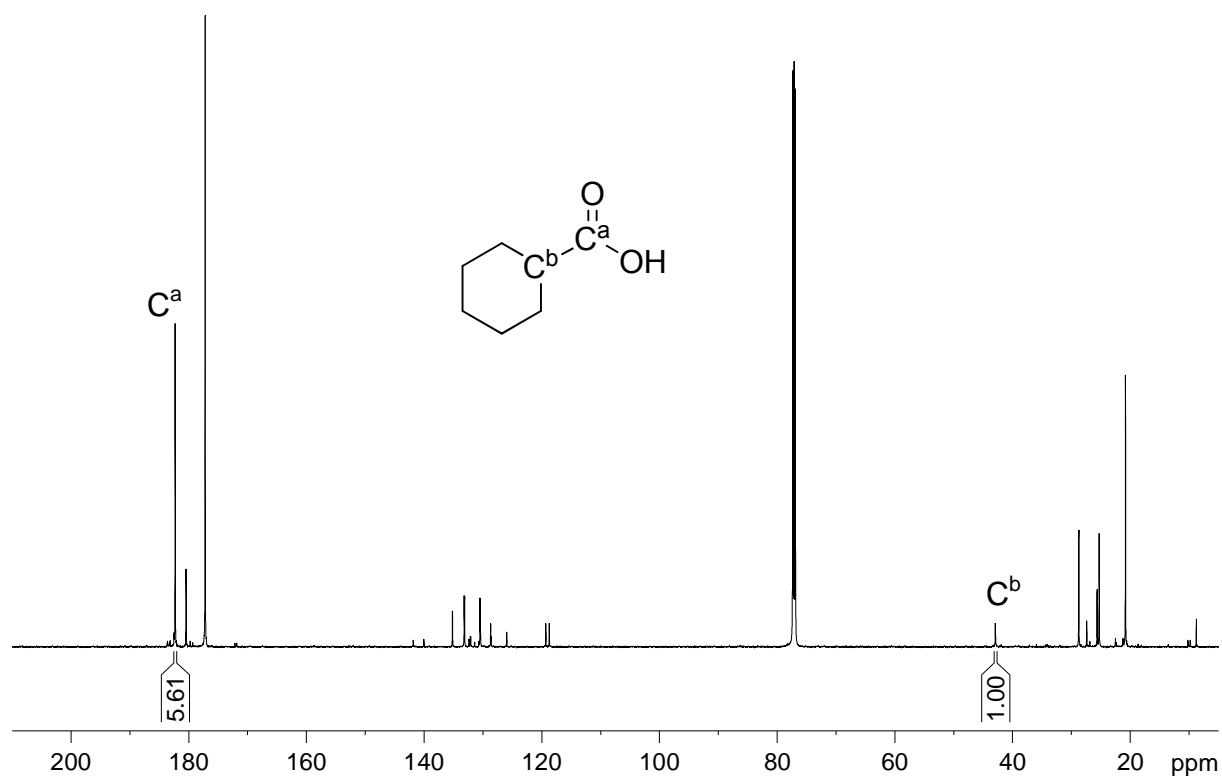


Figure S4.8. Quantitative ¹³C NMR spectrum of the reaction mixture after the catalysis indicating the ratio of the integrals for C^a:C^b. Measured in CDCl₃ at ambient temperature with a resonance frequency of 151 Mhz.

S4.3 NMR Spectra to the D₂ labelling experiment

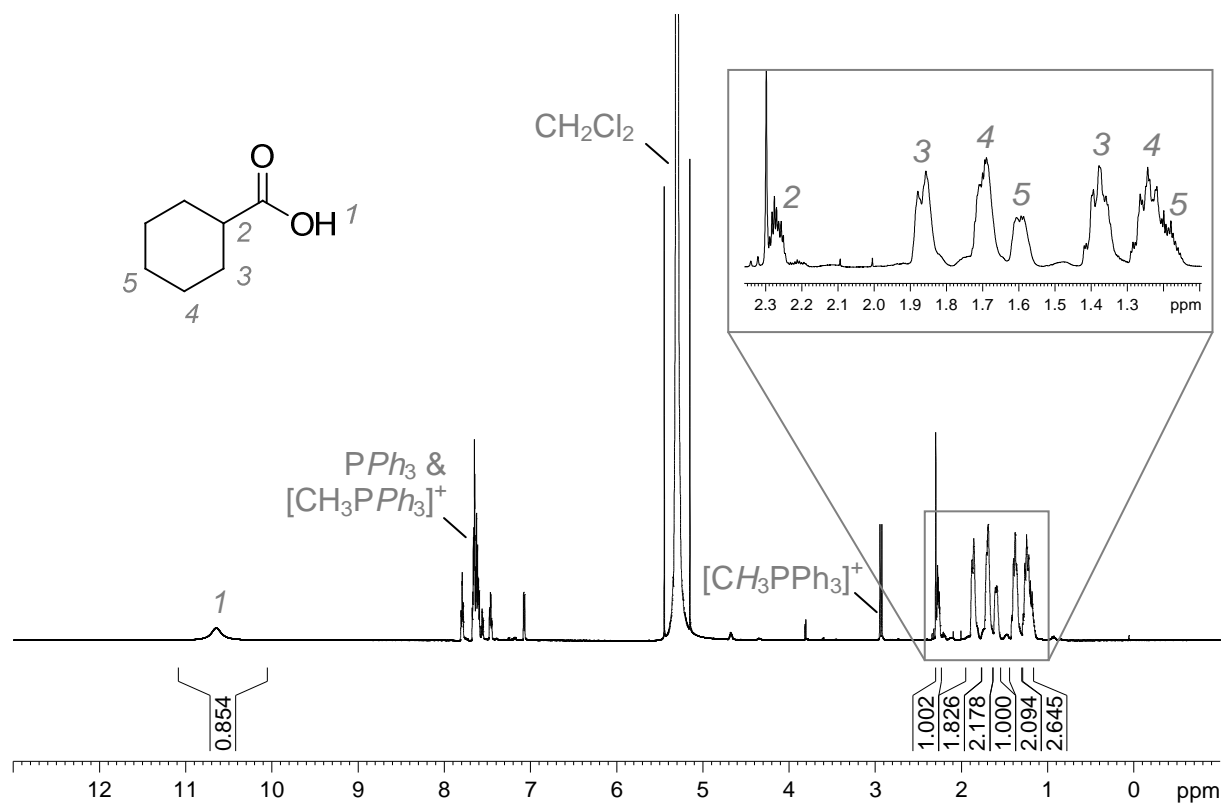


Figure S4.9. ¹H NMR spectrum of the reaction mixture after the catalysis with integrals for the cyclohexanoic acid product CA. Measured in CH₂Cl₂ at ambient temperature with a resonance frequency of 600 Mhz. Signal assignment based on ¹H-¹³C HSQC and HMBC NMR experiment.

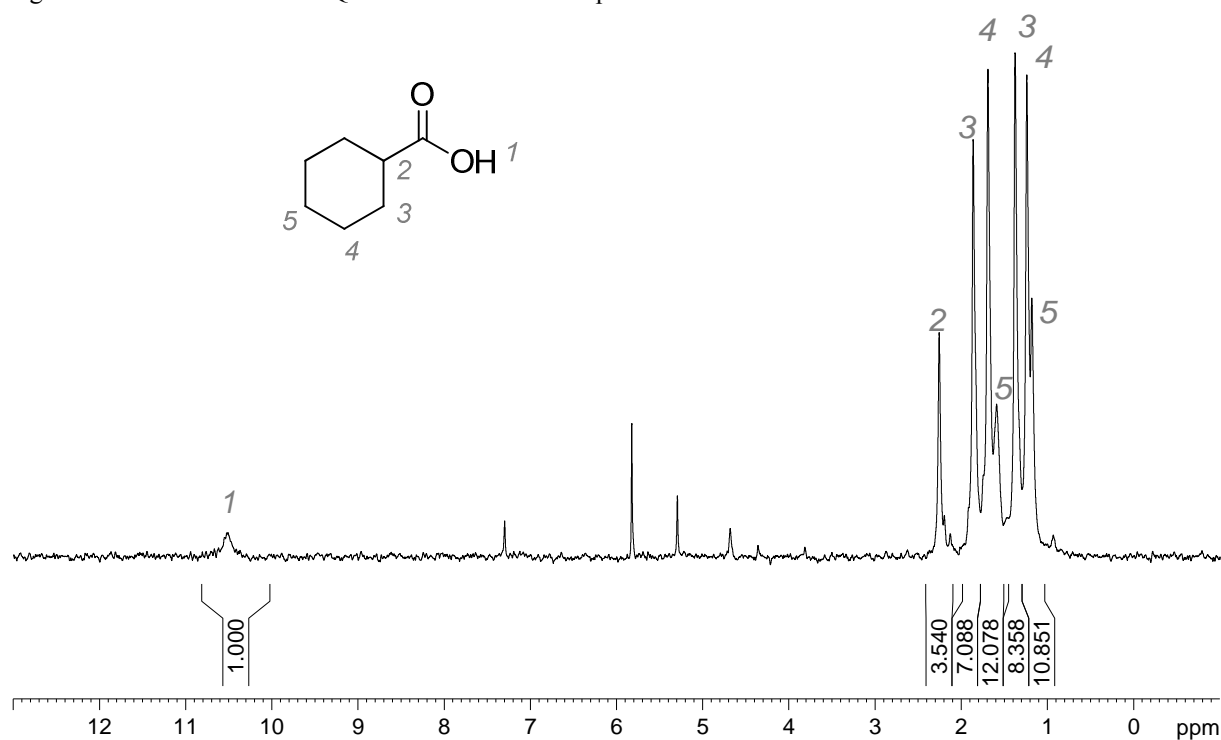


Figure S4.10. ²H NMR spectrum of the reaction mixture after the catalysis with integrals for the cyclohexanoic acid product CA. Measured in CH₂Cl₂ at ambient temperature with a resonance frequency of 92 Mhz.

S4.4 NMR Spectra to the D₂O labelling experiments

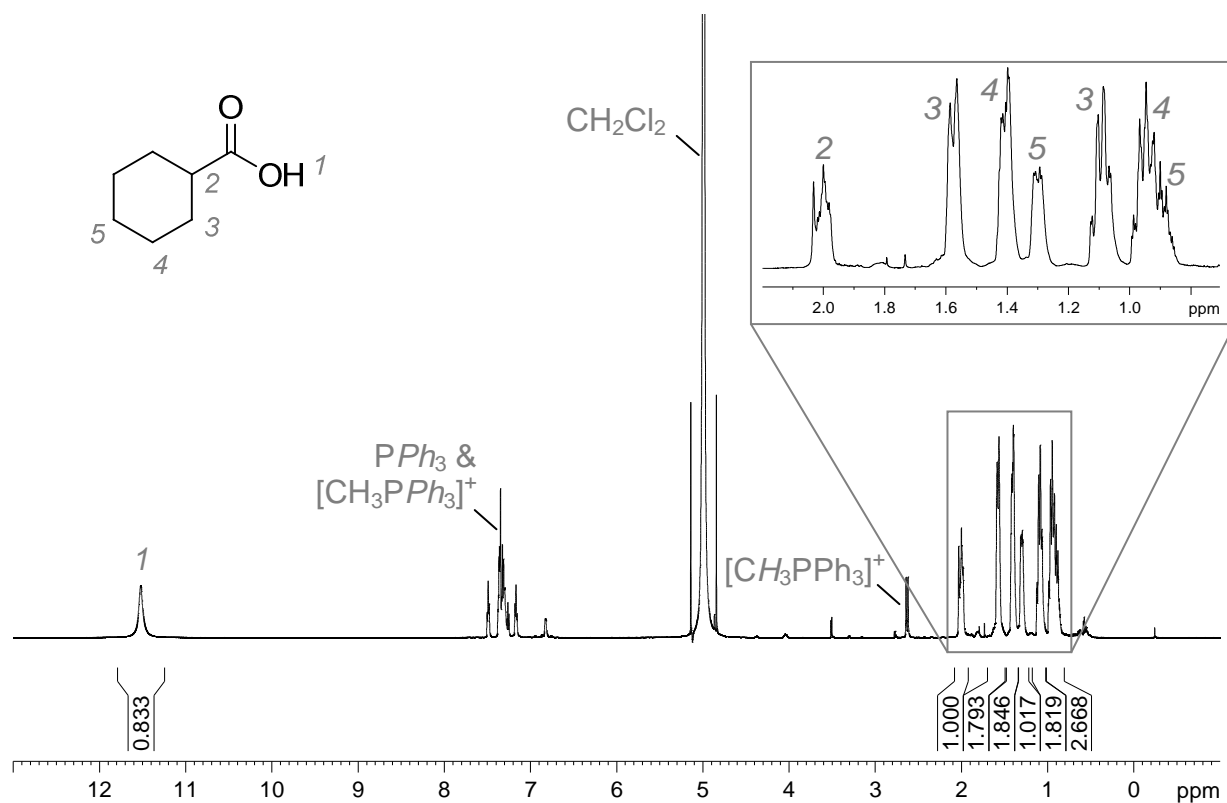


Figure S4.11. ¹H NMR spectrum of the reaction mixture after the catalysis with integrals for the cyclohexanecarboxylic acid product CA. Measured in CH₂Cl₂ at ambient temperature with a resonance frequency of 600 Mhz. Signal assignment based on ¹H-¹³C HSQC and HMBC NMR experiment.

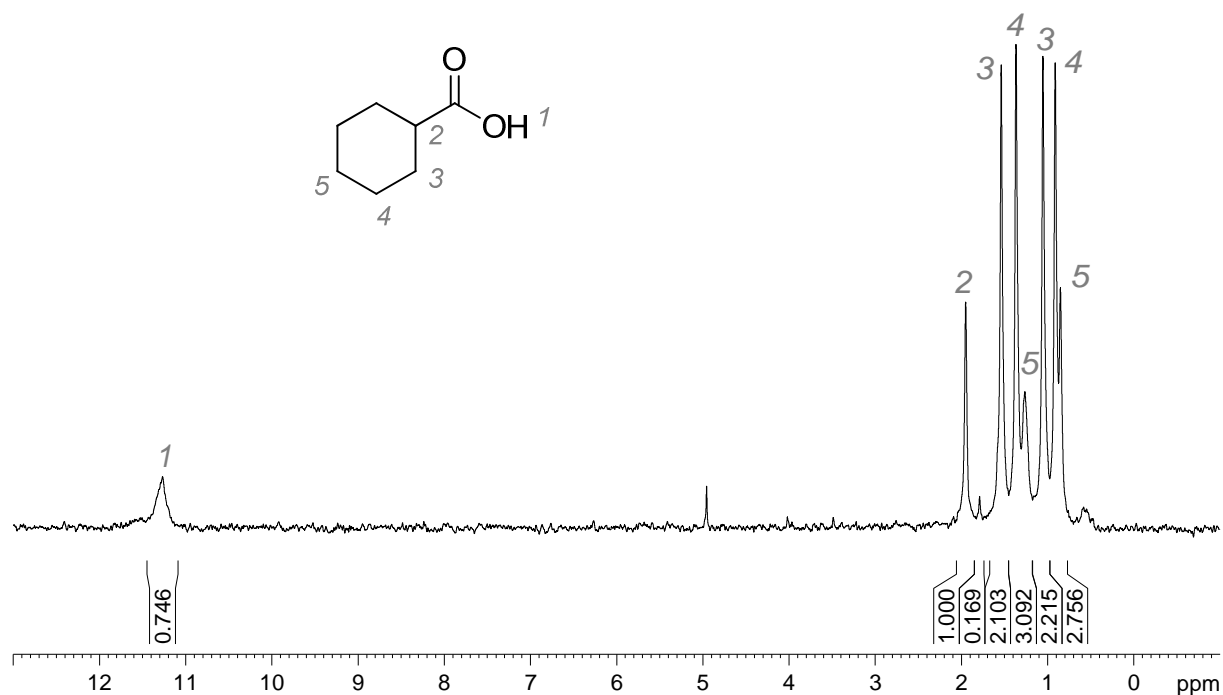


Figure 4.12. ²H NMR spectrum of the reaction mixture after the catalysis with integrals for the cyclohexanecarboxylic acid product CA. Measured in CH₂Cl₂ at ambient temperature with a resonance frequency of 92 Mhz.

S4.5 Mass Spectra to the H₂¹⁸O labelling experiments

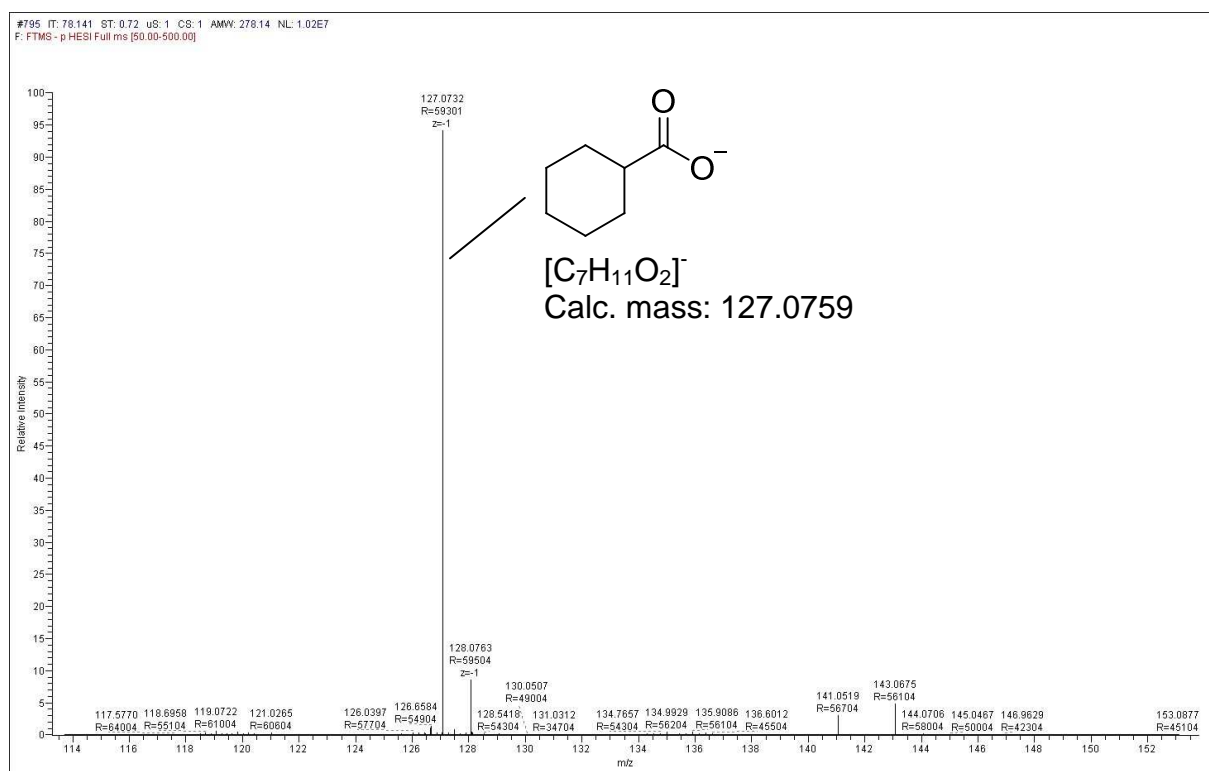


Figure S4.13. High resolution mass spectrum of the reaction mixture after the catalysis without addition of H₂¹⁸O for the cyclohexanoic acid product CA. Measured as ESI(-) in Methanol at ambient temperature.

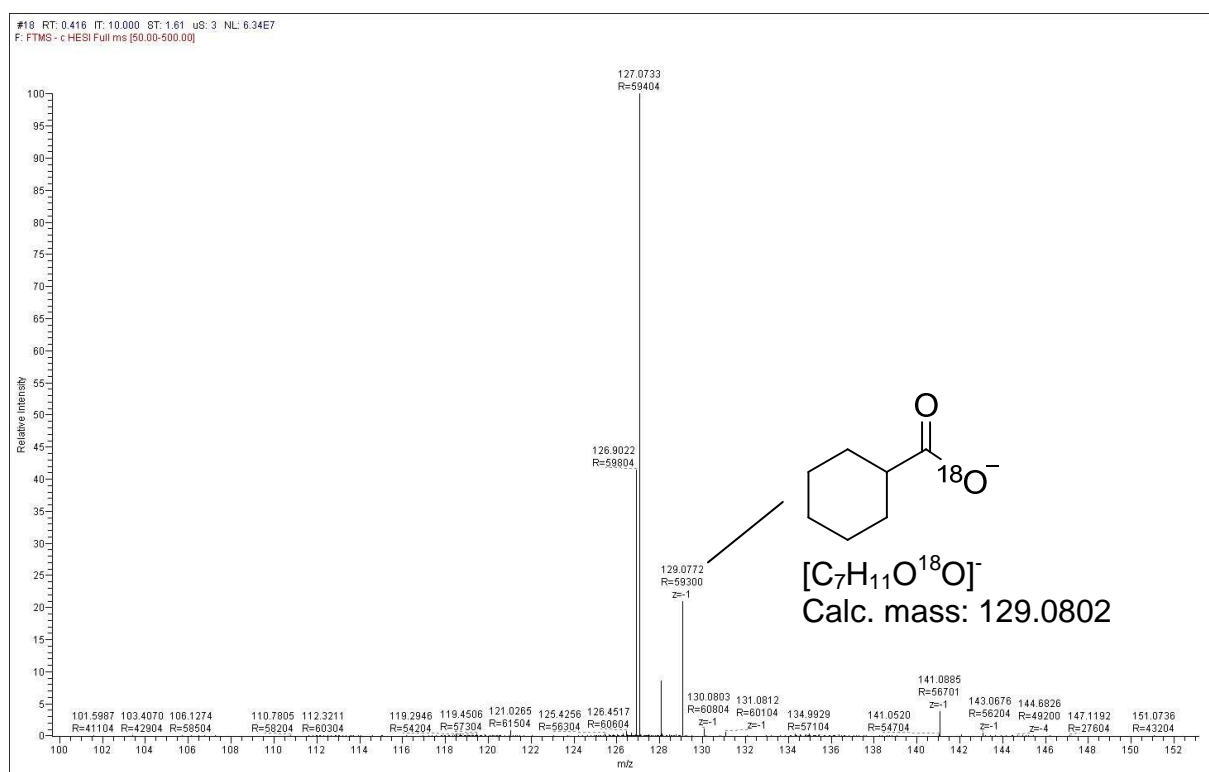


Figure S4.14. High resolution mass spectrum of the reaction mixture after the catalysis with addition of H₂¹⁸O for the cyclohexanoic acid product CA. Measured as ESI(-) in Methanol at ambient temperature.

S4.6 Mass Spectra to the H₂¹⁸O control experiment

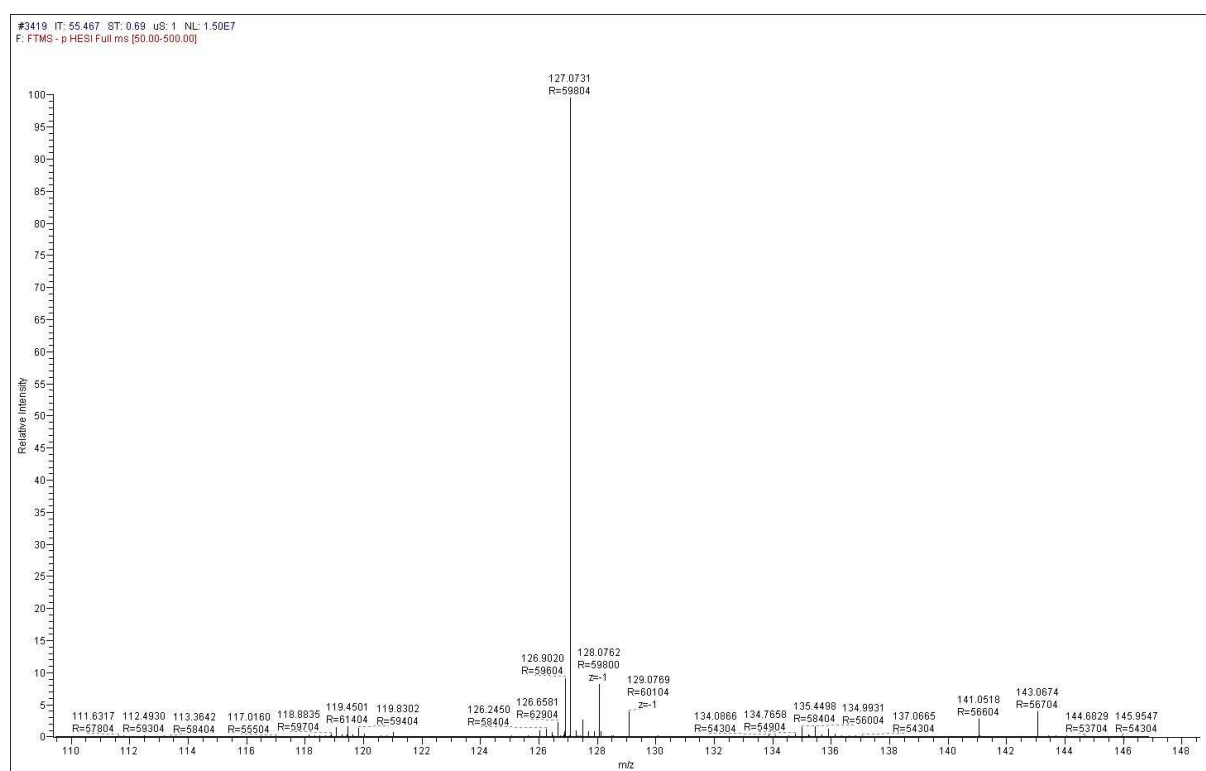


Figure 4.15. High resolution mass spectrum of the reaction mixture after the control experiment with addition of H₂¹⁸O for the cyclohexanoic acid product CA. Measured as ESI(-) in Methanol at ambient temperature.

S5 Crystallographic Details

Crystal data and refinement results have been compiled in Table S5.1. Intensity data were collected at 100 K with a *Bruker APEX* area detector equipped with an *Incoatec microsource* (Mo-K α , $\lambda = 0.71073$ Å, multilayer optics). Temperature was controlled with an *Oxford Cryostream 700* instrument. Intensities were integrated with *SAINTE*^[4] and corrected for absorption by multi-scan methods with *SADABS*^[5]. The structure was solved by direct methods.^[6] The structures were refined by full matrix least squares procedures as implemented in *SHELXL-97*.^[6] All non-hydrogen atoms in the target molecule were assigned anisotropic displacement parameters. The hydrogen atoms were included as riding. Isotropic displacement parameters were assigned to all atoms with fractional site occupancies.

Table S5.1. Crystallographic data to the structure.

Parameter		Parameter	
Empirical formula	C ₁₉ H ₁₅ I ₄ OPRh, C ₁₉ H ₁₈ P	V/Å ³	3814.5(6)
M/g mol ⁻¹	1178.09	Z	4
Crystal dimensions/mm	0.01 x 0.12 x 0.30	μ (Mo K α)/mm ⁻¹	3.798
Crystal shape	Block	Scan range (θ)/°	1.71 / 30.82
Crystal color	Dark brown	Total reflections	56229
Crystal system	Monoclinic	Unique reflections	11273
Space group (no.)	P 2 ₁ /n	Variables refined	416
a/Å	15.4392(15)	R _{int}	0.0400
b/Å	15.0715(14)	wR ₂ (all reflections)	0.0747
c/Å	16.5045(16)	R ₁ (all/obs.)	0.0376 / 0.0291
α /°	90.00	GOF on F ²	1.084
β /°	96.6600(10)	Diff. peak/hole [e/ Å ⁻³]	1.671 / -0.550
γ /°	90.00		

Further details on the crystallographic studies including fractional coordinates, displacement parameters and molecular geometry are given in the CIF format. Crystallographic data (excluding structure factors) for all data collections will be deposited at the Cambridge Crystallographic Data Centre as supplementary publications numbers when the manuscript is accepted for publication. Copies of the data can be obtained free of charge on application to The Director, CCDC, 12 Union Road, Cambridge, CB21EZ, UK (Fax: int. code +44-1223-336-033; E-Mail: deposit@ccdc.cam.ac.uk; web, www:http://www.ccdc.cam.ac.uk).

S6 References

- [1] A. Vij, R. L. Kirchmeier, J. n. M. Shreeve, R. D. Verma, *Coord. Chem. Rev.* **1997**, *158*, 413-432.
- [2] "Dissociation constants of organic acids and bases" in *CRC Handbook of Chemistry and Physics*, 89th ed. (Ed.: D. R. Lide), CRC Press/Taylor and Francis, Boca Raton, FL, **2009**.
- [3] J. P. Guthrie, *Can. J. Chem.* **1978**, *56*, 2342-2354.
- [4] SAINT, Bruker AXS, *Program for Reduction of Data collected on Bruker CCD Area Detector Diffractometer V.6.02*, Bruker AXS Inc., Madison, WI, USA, **1999**.
- [5] SADABS, *Program for Empirical Absorption Correction of Area Detector Data V 2004/1*, Bruker AXS Inc., Madison, WI, USA, **2004**.
- [6] a) G. M. Sheldrick, *SHELXL-97 Program for Crystal Structure Refinement*, Universität Göttingen, **1997**; b) G. Sheldrick, *Acta Crystallogr., Sect. A* **2008**, *64*, 112-122.