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Lotka–Volterra dynamics kills the Red Queen: population size fluctuations and associated stochasticity dramatically change host-parasite coevolution

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Abstract

Background: Host-parasite coevolution is generally believed to follow Red Queen dynamics consisting of ongoing oscillations in the frequencies of interacting host and parasite alleles. This belief is founded on previous theoretical work, which assumes infinite or constant population size. To what extent are such sustained oscillations realistic?

Results: Here, we use a related mathematical modeling approach to demonstrate that ongoing Red Queen dynamics is unlikely. In fact, they collapse rapidly when two critical pieces of realism are acknowledged: (i) population size fluctuations, caused by the antagonism of the interaction in concordance with the Lotka-Volterra relationship; and (ii) stochasticity, acting in any finite population. Together, these two factors cause fast allele fixation. Fixation is not restricted to common alleles, as expected from drift, but also seen for originally rare alleles under a wide parameter space, potentially facilitating spread of novel variants.

Conclusion: Our results call for a paradigm shift in our understanding of host-parasite coevolution, strongly suggesting that these are driven by recurrent selective sweeps rather than continuous allele oscillations.

Keywords: Host-parasite coevolution, Red Queen hypothesis, Lotka-Volterra dynamics, Genetic drift, Population bottleneck

Background

The Red Queen from Lewis Carroll's tale 'Through the looking glass' is commonly used as a metaphor for selection-induced rapid evolution [1-3]. It is based on the observation that persistence in an environment with changing selective constraints requires ongoing adaptations to the encountered challenges [4]. Host-parasite coevolution with antagonistic and inter-dependent interactions represents one of the role models for such rapid evolutionary change [5,6]. For instance, an increase in host resistance reduces parasite fitness, thus immediately favoring parasite varieties with altered virulence and/or

immune-evasion mechanisms. In turn, a novel parasite attack mechanism decreases host fitness, thus favoring host varieties with new counter-defenses. If the interaction persists, then it will lead to continuous parasite adaptations and host counter-adaptations. The rapid evolutionary dynamics associated with these interactions is very well documented in the literature, ranging from field studies on rabbits and their myxoma viruses [7], snails and their trematode parasites [8], *Daphnia magna* waterfleas and their bacterial parasites [9] to laboratory-based coevolution experiments between bacteria and their phages [10-12], the nematode *Caenorhabditis elegans* and bacterial parasites [13,14], or the red flour beetle *Tribolium castaneum* and its microsporidian parasite [15,16].

It is thus widely accepted that these interactions evolve fast and continuously. Yet, to date, the exact underlying selection dynamics are not always well understood.

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These dynamics can generally be influenced by metapopulation structure and environmental variation [17,18]. Within a particular population and specific environmental context, two main alternatives are thought to be of prime importance: recurrent selective sweeps and negative frequency-dependent selection [5,6,19-21]. Both alternatives are consistent with the above original definition of the Red Queen hypothesis by Van Valen [4], whereas, curiously, only the second alternative is referred to as Red Queen dynamics [5,6,20]. The two alternatives are closely related because both assume a selective advantage of a rare genotype, for example a novel host resistance variant. However, they differ fundamentally in the way in which the new variant originates and spreads within the population. The concept of recurrent selective sweeps (often termed *arms race dynamics*) consists of two steps: the *de novo* appearance of a beneficial allele (e.g., by mutation or immigration) and its subsequent spread through the population to fixation (i.e., the selective sweep). These sweeps occur repeatedly in host and parasite populations, usually each time with a new beneficial allele. They may only lead to fast changes in absolute time if at least one of the following factors applies: new alleles arise frequently, new alleles become immediately visible and thus selectable at the phenotypic level, the new alleles provide a high selective advantage, and/or the organisms have short generation times. This situation is best met in bacteriophage interactions, which are usually characterized by large population sizes (i.e., high likelihood of the occurrence of favorable mutations), short generation times, and haploid genomes (i.e., new mutations are immediately expressed phenotypically) [11,22-24] (but see also [25]).

In contrast, the dynamics for multicellular host systems are traditionally viewed to be determined by negative frequency-dependent selection leading to sustained oscillations of the same alleles (i.e., Red Queen dynamics [6,20]), but not to the fixation of single alleles. In this case, standing genetic variation is required, because the population sizes for these hosts are usually comparatively small, their generation times comparatively long, and their genomes diploid. As a consequence, recurrent selective sweeps are commonly thought to be rather slow in these systems. Instead, if standing genetic variation is available, then negative frequency-dependent selection can produce fast and continuous allele frequency changes even in these host systems. Such negative frequency-dependent dynamics seem to be present in some multicellular host systems, including the freshwater snail *Potamopyrgus antipodarum* [8,26] and the waterflea *Daphnia magna* [9].

Numerous theoretical models have been developed to study the underlying selection dynamics. Interestingly, the current models typically focus on evolutionary change (i.e., the rate of change in host and parasite allele frequencies in response to the type of interaction). These

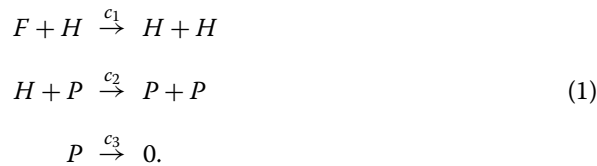
approaches have thus largely neglected ecological dynamics, which can have a huge impact on the evolutionary process. Population size fluctuations deserve particular attention in this context, because they are induced by reciprocal selection among the antagonists and, therefore, represent an inherent property of host-parasite coevolution - irrespective of additional environmental variation [7,10,27-30]. Since selection is reciprocal, population size fluctuations should be coupled between the antagonists, and generally follow Lotka-Volterra dynamics [31,32]. Such demographic variations have the potential to affect the dynamics of host-parasite allele frequency changes by introducing two important effects. Firstly, the rising and falling population sizes produce bottlenecks where selection favours a particular allele. The favored allele may thus reach comparatively high frequencies during the bottleneck, possibly enhancing its spread in the subsequently expanding population. Secondly, the elevated stochasticity during the bottleneck may lead to a further increase and thus spread of the favored allele.

In this manuscript, we aim at understanding in how far Lotka-Volterra population size fluctuations and the associated stochastic effects influence the dynamics of allele frequency changes during host-parasite coevolution. While several previous theoretical models have applied the Lotka-Volterra dynamics to host-parasite coevolution (e.g., [33-37]), their influence on the evolutionary dynamics has not yet been systematically explored by comparison with a model with constant population size. Similarly, stochastic effects during host-parasite coevolution have only been considered in a few theoretical studies (e.g., [38,39]), yet, to our knowledge, with a single exception [40] under constant population size and not in combination with Lotka-Volterra dynamics. Hence, while the previous studies have independently utilised stochastic effects or Lotka-Volterra dynamics, a systematic analysis of the consequences of each of these factors, either alone or in combination, is as yet missing - in spite of their potential importance. The novelty of our study lies in bringing together these two aspects and comparing their influence to the traditional model, in which Lotka-Volterra dynamics and stochastic effects are excluded. More specifically, we here use the standard matching-alleles host-parasite interaction model to assess allele frequency dynamics in the presence versus absence of Lotka-Volterra oscillations for a stochastic versus an analogous deterministic model.

Methods

Based on the Lotka-Volterra equations [31,32], we address the population dynamics of interacting hosts and parasites. The host corresponds to the prey in the original model, and the parasite to the predator. The host

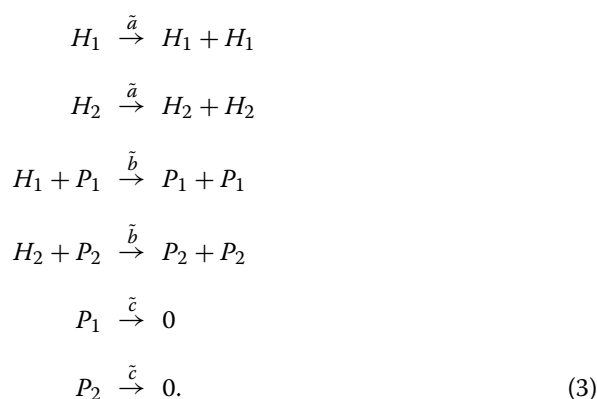
consumes a (constant) food supply F and reproduces at rate c_1 . Parasites infect hosts at rate c_2 , leading to elimination of a host and generation of an additional parasite. Parasites die at rate c_3 . The number of host and parasite individuals are given by H and P . In a stochastic system these interactions can be defined by the following reactions [41,42],



Usage of these specific reactions facilitates tracking of each unit of the interacting antagonists and, thus, it allows a more precise characterization of the resulting dynamics. These reactions can also be used directly for exact stochastic simulations based on the Gillespie algorithm. They further provide a microscopic dynamics from which the deterministic Lotka-Volterra equations emerge in the limit of infinite population size [42],

$$\begin{aligned} \dot{H} &= c_1FH - c_2HP \\ \dot{P} &= c_2HP - c_3P. \end{aligned} \quad (2)$$

Host-parasite coevolution is modeled using the standard matching alleles model [6]. For this, we define two host and two parasite types, H_1 and H_2 for the host and P_1 and P_2 for the parasite. This is equivalent to a haploid system with two antagonists, each of which possesses two alleles at a single locus. The interaction according to the matching alleles model is described with the following six reactions,



In the matching alleles model, the interactions between alternate hosts and parasites (H_1, P_2 and H_2, P_1) are without consequence and thus do not appear here. While the absence of these interactions is the standard assumption in the matching alleles model, allowing a small amount of these interactions does not change our results qualitatively (see Appendix). In the limit of infinite population

size [42], we obtain a set of four coupled nonlinear differential equations,

$$\begin{aligned} \dot{h}_1 &= h_1(a - bp_1) \\ \dot{h}_2 &= h_2(a - bp_2) \\ \dot{p}_1 &= p_1(bh_1 - c) \\ \dot{p}_2 &= p_2(bh_2 - c), \end{aligned} \quad (4)$$

where the frequencies of H_i and P_i are given by h_i and p_i . The above equations consider interdependence of host and parasite demographics, allowing population sizes to vary in response to the interaction with the antagonist, consistent with the Lotka-Volterra model. The precise nature of the resulting oscillations in population size is determined by the parameters, most importantly by b .

As we are interested in the effects of population size variation induced by the Lotka-Volterra equations, we have to compare this to a scenario in which the population size is constant. Such constant population size models are common, e.g. the Wright-Fisher model or the Moran process. However, microscopically these models are distinct from the Lotka-Volterra equations considered above. Therefore, we used the above approach and enforced constant population size by resetting host and parasite population sizes to their initial values after every generation (N_{avg} transition events, see Appendix), while relative allele frequencies were maintained. The dynamics were subsequently assessed for different average population sizes. To ensure comparability of allele frequency fluctuations across population sizes and evolutionary models, we rescaled the interaction parameters with N_{avg} for the deterministic analogues of the considered stochastic scenarios (Appendix).

Results

Host-parasite coevolutionary dynamics are analyzed in the presence and absence of Lotka-Volterra dynamics. Figure 1 illustrates an exemplary result. All models initially produce oscillatory allele frequency changes, but only with Lotka-Volterra dynamics are these accompanied by changes of population size. As a consequence, changes in allele numbers are also more pronounced (top versus bottom in Figure 1). As the deterministic model allows for arbitrary small frequencies of each type, it formally never leads to allele fixation and thus produces continuous oscillations. In contrast, the corresponding stochastic models have absorbing states, making fixation possible. Interestingly, allele fixation appears to be substantially faster in the stochastic model that includes Lotka-Volterra fluctuations (top versus bottom panels, Figure 1). As such, it seems that these conditions favor rapid termination of the Red Queen oscillations.

We next analyze the impact of the average population size on this pattern. In the following, we focus on the time

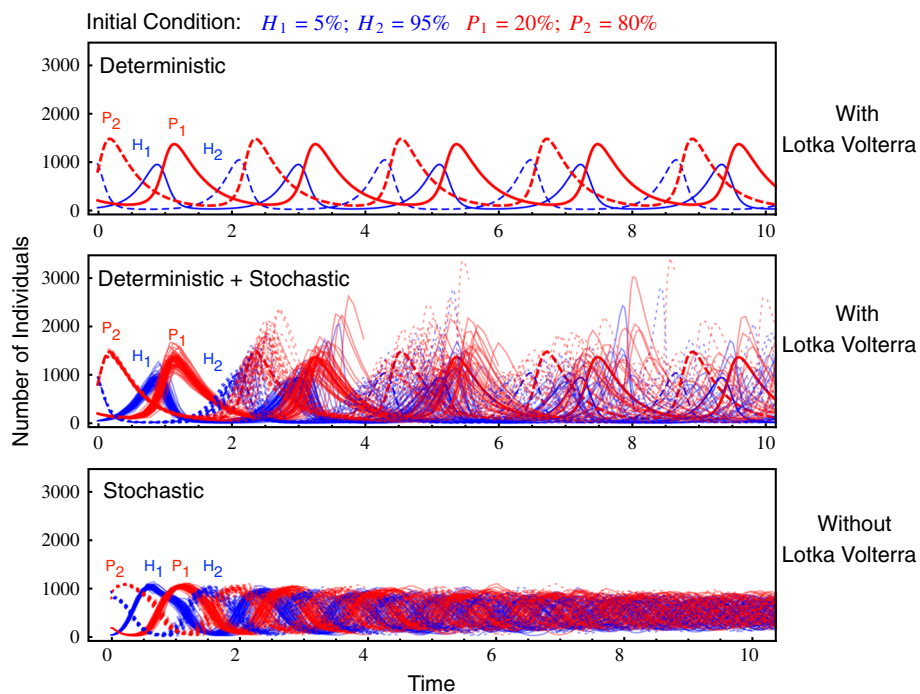


Figure 1 Example of allele frequency dynamics with and without Lotka-Volterra population size fluctuations. Top: Lines show the deterministic Lotka-Volterra dynamics, as often considered in theoretical studies, cf. Eqs. (4). Middle: When stochasticity is included (thin lines show the results of 50 individual stochastic Gillespie simulations), then simulations may initially produce allele oscillations as above and below. However, alleles usually spread to fixation (or go extinct) at a much faster rate. Bottom: Dynamics without Lotka-Volterra cycles, fixing the average population size of both species to $N_{avg} = 1000$ by resetting it after every N_{avg} reactions, while maintaining the ratio between the alleles. The 50 individual stochastic simulations now only rarely reach fixation. The figure illustrates the scenario where the rare host allele (H_1) is more likely to reach fixation than the frequent host allele (see Figure 3). This fixation probability decreases with increasing initial frequency (cf. Figure 3). The simulation parameters are $a = 5$, $c = 2.5$, $b = 10/N_{avg} = 0.01$ with $H_1 = 5\%$, $H_2 = 95\%$, $P_1 = 20\%$, $P_2 = 80\%$ as initial condition.

until one of the alleles from either of the antagonists has reached fixation in order to compare evolutionary rates across population sizes and models. In general, Lotka-Volterra dynamics cause a substantial increase in allele fixation rate (Figure 2). Interestingly, in this case, allele fixation rates depend only weakly on average population size. Figure 1 suggests that this is because allele frequencies can become very small during the Lotka-Volterra demographic fluctuations. In contrast, average population size has a much stronger effect when it is artificially kept constant. Here, the time until allele fixation increases exponentially with increasing population size (Figure 2).

Figure 2 explores the time to fixation of any of the alleles in either the host or the parasite using a specific combination of initial allele frequencies (i.e., the rare host allele is present at 5%, the common at 95%, whereas the parasite alleles are at 20% and 80% respectively). How does this depend on the initial allele frequencies in both antagonists? For instance, the selective advantage of a rare allele is not only the result of its own frequency, but also determined by the abundance of the corresponding allele in the antagonist. Allele fixation rates were thus explored as a function of initial allele frequencies in the

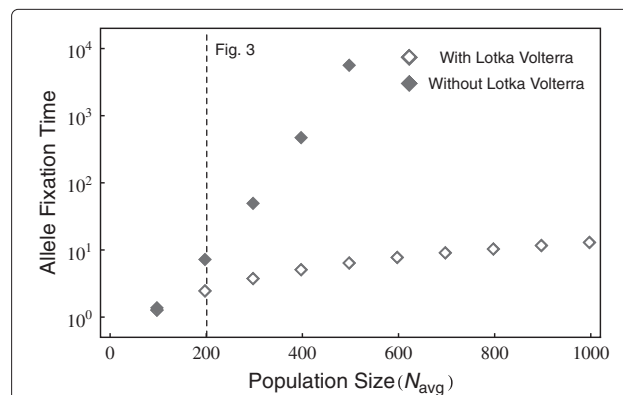


Figure 2 The time until fixation of one allele, either host or parasite, is shown in dependence of the initial population size N_{avg} . Including Lotka-Volterra fluctuations, the fixation time is only weakly affected by increases in N_{avg} . Excluding Lotka-Volterra cycles maintains allele frequency oscillations, leading to an exponential increase in fixation time as N_{avg} increases. For all simulations the initial condition were $H_1 = 5\%$, $H_2 = 95\%$, $P_1 = 20\%$, $P_2 = 80\%$ of the N_{avg} , and the parameters $\mu = 5$, $c = 2.5$, $b = 10/N_{avg}$ with averages over 10^6 realizations). The vertical dotted line shows the population size employed in Figure 3.

two antagonists. Most impressively, Lotka-Volterra fluctuations cause much faster allele fixation under almost all initial conditions (Figure 3, left column, top versus bottom panel). The detailed analysis then suggests that, in case of Lotka-Volterra fluctuations, host alleles can have a high fixation probability even if initially rare (Figure 3, middle panel in top row). This is true across a relatively wide distribution of initial frequencies for the corresponding parasite allele. Interestingly, it even applies when the corresponding parasite allele has high initial frequencies (Figure 3, top left corner in top middle panel). This counterintuitive result can be explained by consideration of the dynamics that ensue from these initial conditions. In this particular case, the low initial frequency of host allele 1 means that host allele 2 is initially common, whereas the high initial frequency of parasite allele 1 means that parasite allele 2 is rare. High host allele 2 abundance then specifically favors parasite allele 2, which rapidly increases in frequency. The unexpected consequence of these starting conditions is that these two interacting alleles subsequently engage in highly pronounced frequency oscillations that show larger amplitudes than those observed for host and parasite alleles 1 (Figure 4). If during these oscillations low allele 2 frequencies coincide with a Lotka-Volterra bottleneck and associated stochastic effects, then host allele 2 has a very high likelihood of going extinct, resulting in fixation of host allele 1 (see Figure 4).

The results also highlight that the dynamics are usually determined by fixation of one of the host alleles (red colour is mainly found in middle rather than right panel of the top row of Figure 3). Note that the simulations are stopped as soon as either one of the host or one of the parasite alleles reaches fixation and, thus, the fixation probabilities of both host as well as both parasite alleles sum up to one. In our case, fixation of the host allele is more likely than fixation of the parasite allele because for our parameter combination and initial condition, it is usually the host that first experiences a Lotka-Volterra bottleneck and consequentially a drop in the frequency of one of the alleles (see also Figure 4). Nevertheless, if both the parasite and corresponding host allele are common, then it is the parasite allele that has a high probability of fixation (Figure 3, top right).

The overall pattern looks different in the absence of Lotka-Volterra fluctuations (Figure 3, bottom row). Host allele fixation probability increases with its own high initial frequency and, at the same time, low initial abundance of the corresponding parasite allele (Figure 3 middle panel in bottom row). Parasite allele fixation is enhanced when both parasite and corresponding host alleles are initially common (Figure 3 bottom right). Under these conditions, fixation probabilities of both host and parasite alleles are almost identical at initially intermediate frequencies, most likely due to negative frequency dependent selection, as illustrated in Figure 1.

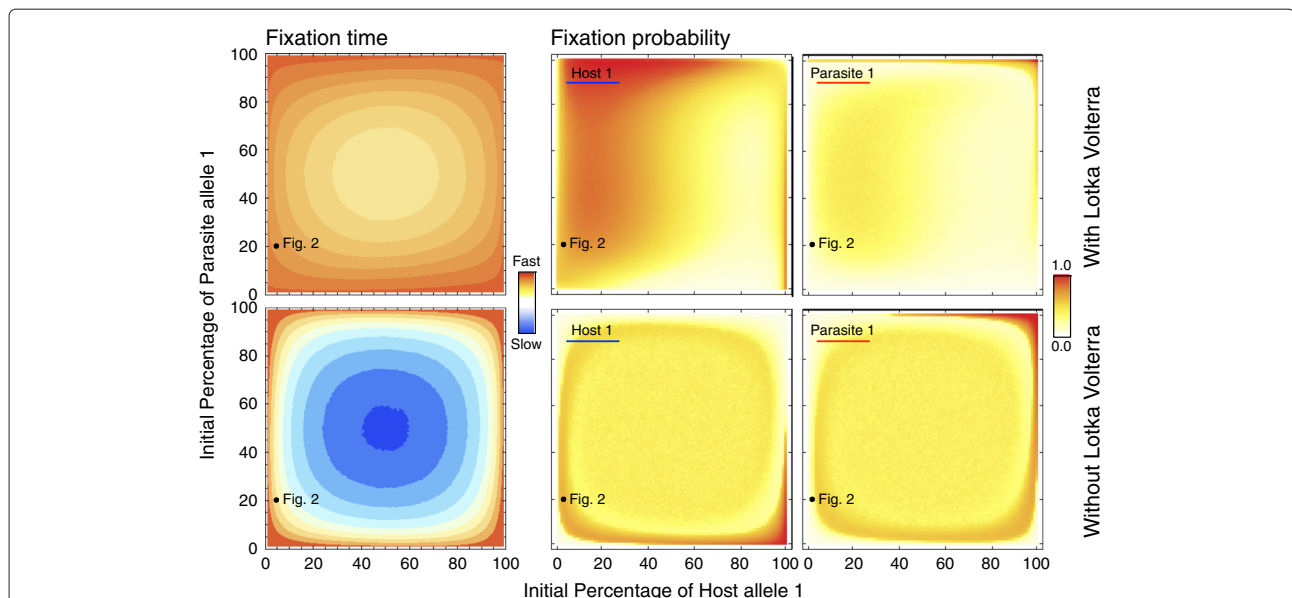


Figure 3 The influence of initial allele frequency on fixation time and probability for the stochastic models. For an $N_{avg} = 200$, we plot the time until any of the four alleles goes to fixation (left column) and the probability of fixation of one of the host and parasite alleles for all possible initial conditions (middle and right columns) (averages over 10^6 realizations). Lotka-Volterra fluctuations lead to substantially faster allele fixations (top left panel) and high fixation probability for the host allele across a wide range of initial conditions (top middle panel). The simulations were always stopped when either one of the host or one of the parasite alleles reached fixation. Thus, the sum of the fixation probabilities of all alleles sums up to 1. The specific initial conditions used in Figure 2 are indicated.

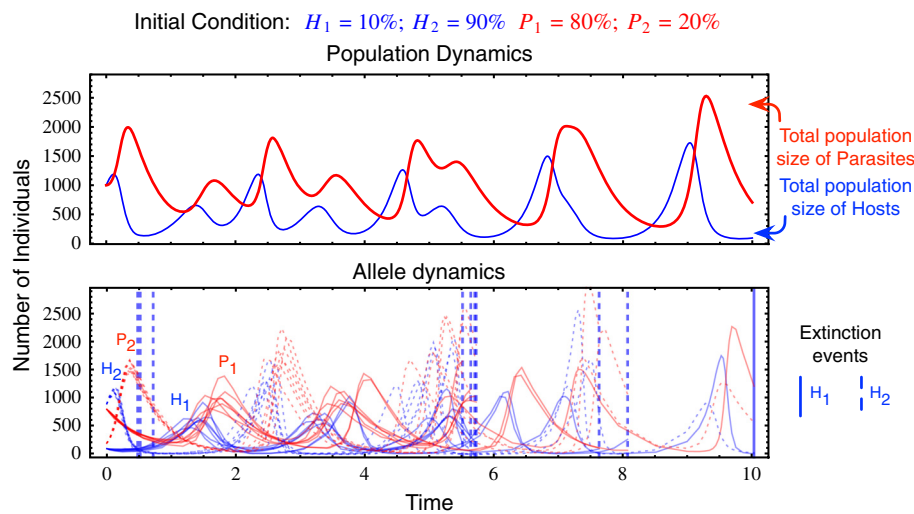


Figure 4 Detailed dynamics explaining the seemingly counterintuitive result of the high fixation probability of Host allele 1 in spite of the high prevalence of Parasite allele 1 (in Figure 3 top middle panel) due to the inclusion of Lotka-Volterra dynamics. In this figure we depict the dynamics that occur at the initial conditions with a low H_1 frequency and a high P_1 frequency. In this particular case, the low initial frequency of H_1 means that H_2 is initially common, which in turn favours P_2 . This parasite allele thus rapidly increases in frequency, subsequently causing highly pronounced H_2 and P_2 frequency oscillations that show larger amplitudes than the interacting H_1 and P_1 alleles. If low H_2 frequencies coincide with a Lotka-Volterra bottleneck in the hosts, then the associated stochastic effects lead to a higher likelihood of H_2 going extinct, resulting in an overall higher fixation probability of H_1 . The top panel shows the average population dynamics, whereas the bottom panel shows the frequency changes for the indicated host and parasite alleles across the ten independent simulations. The vertical lines in the bottom panel denote the time points where the simulation is terminated due to a loss of an allele. Out of the 10 simulation runs 9 are stopped due to the allele H_2 going extinct and only one due to H_1 going extinct. The interaction parameters are $a = 5$, $c = 2.5$, $b = 10/N_{avg} = 0.01$.

Discussion

Population size fluctuations represent an inherent property of host-parasite interactions. Unequivocal evidence for such interaction-dependent demographic variations was obtained from controlled host-parasite mesocosm experiments, for example with *E. coli* and its phage [10,30], *Hydra* hosts and its *Hydramoeba* parasite [43], house fly and its parasitic wasps [44,45], or azuki bean weevil *Callosobruchus chinensis* and its parasitoid *Heteropilus prosopidis* [46]. Similar observations were made under field conditions, for example for rabbits and their myxoma viruses [7], or red grouse and its nematode parasite [47]. Additional examples are summarized by [28] and [29]. As population size fluctuations produce regular bottlenecks, random genetic drift is likely to influence allele frequencies. Previous theoretical models, developed in a different context, strongly suggest that even large populations are influenced by such stochastic processes [48,49]. More generally, under natural conditions in a finite population, it is difficult to imagine that changes in population size do not affect evolutionary dynamics. Consequently, an in-depth understanding of the evolution of host-parasite interactions should take account of the associated ecological processes based on Lotka-Volterra fluctuations.

Very few previous theoretical models on host-parasite interactions have considered Lotka-Volterra fluctuations

[33-37]. These studies usually used a deterministic approach and thus excluded stochastic effects, which are most prominent during bottlenecks. Similarly, only few theoretical studies considered stochastic effects in this context [38,39], yet under constant population size, but not in combination with Lotka-Volterra dynamics. We are aware of only one study that looked at host-parasite coevolution in consideration of Lotka-Volterra interactions and stochasticity [40]. However, this study had a different focus, and thus, it did not include a systematic comparison to models without Lotka-Volterra cycles or without stochasticity. Consequently, the interaction of these two aspects for host-parasite coevolution is so far unexplored. At the same time, their relevance was demonstrated for evolution of only one of the antagonists, namely the parasite. For example, the probability of fixation of a beneficial mutation in a bacterial population was shown to be enhanced by periodical bottlenecks [50-52]. Similar results were obtained in a model that explored the effect of bottlenecks during pathogen transmission [53]. Our study explicitly evaluates the influence of both Lotka-Volterra fluctuations and stochastic effects on the dynamics of host-parasite interactions using a comparison to a model with constant population size and/or absence of stochasticity. As the demographic variations are an inherent property of such antagonistic interactions, their influence should apply across a wide range of environmental

conditions and thus be of general relevance for our understanding of host-parasite coevolution.

Based on our approach, we obtained evidence that both Lotka-Volterra fluctuations and associated stochastic effects significantly affect the course and pace of coevolutionary adaptations. In particular, both factors facilitate selective sweeps (i.e., the spread and fixation of an allele). Most impressively, this effect appears to be independent of average population size (Figure 2) and occur at a substantially faster rate (Figure 3, left column). Moreover, allele frequency changes are not exclusively due to drift, which should favor fixation of initially frequent alleles and loss of initially rare alleles. In contrast, our results indicate that initially rare host alleles can spread to fixation across a relatively wide range of conditions (Figure 3, top middle panel). Rare parasite alleles may not necessarily go extinct, but have a certain likelihood of spreading contingent on the frequency of the corresponding host allele (Figure 3, top right panel). Based on these results we propose that selective sweeps rather than oscillatory negative frequency-dependent selection may represent the main driving force during host-parasite coevolution.

Recurrent selective sweeps have been repeatedly suggested to determine coevolutionary dynamics for parasite or host systems with large population sizes such as bacterial hosts or microbial parasites, where novel mutations are frequent and often directly exposed to selection because of a haploid genetic system. If these selective dynamics also apply to multicellular host and parasite systems, then two contrasting effects may be expected on the coevolutionary process. On the one hand, these systems usually have much smaller population sizes, facilitating spread of alleles in spite of the often diploid genetic system. On the other hand, continuous coevolution may become difficult because it is usually assumed that small population size results in a reduced likelihood of the occurrence of advantageous novel mutations [6]. However, the latter assumption may not always be true. It is possible that new alleles become available for example by frequent immigration or a high rate of gene duplication. These processes may further favor the formation of novel genotypes if they act in combination with recombination and/or mutation. Interestingly, the possible impact of gene duplications is usually not addressed in theoretical work on host-parasite coevolution, even though such duplications are known to be common in almost all organisms [54-56] and often affect genes of relevance for the interaction such as virulence genes in parasites [57,58] or immunity genes in animal hosts [54,59,60].

Several additional factors may favor ongoing coevolution. One of these is founded on a more complex genetic architecture underlying host-parasite co-adaptation that

consists of several interacting loci across the genome (e.g., [61-64]). In this case, allele fixation at one locus may still permit maintenance of variation at other loci, which could then mediate evolutionary responses to the antagonist. Yet another possibility may depend on metapopulation structure, consisting of coevolutionary hot and cold spots and migration among demes, as evidenced for flax and its parasitic rust fungus [65] or the above mentioned snail-trematode interaction [17]. Such an interconnected network could then maintain allelic diversity across the entire metapopulation, even if alleles become temporarily fixed within single demes. Moreover, environmental gradients or perturbations are known to influence host-parasite coevolutionary dynamics [66]. They could similarly prevent loss of alleles, even if the coevolutionary interaction itself would specifically favour only one of the alleles. Obviously, the above processes act in combination with each other in natural systems. Therefore, it is indeed conceivable that recurrent selective sweeps shape long-lasting coevolutionary dynamics even in multicellular host systems.

Conclusion

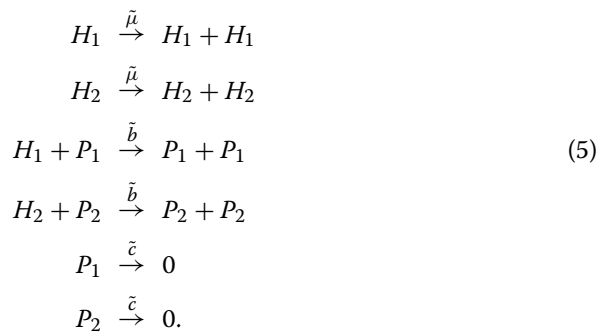
In conclusion, decades of empirical efforts have tried to demonstrate the presence of Red Queen dynamics during host-parasite coevolution. This has led to most ingenious experiments which repeatedly and independently confirmed negative frequency dependence as a driving force [8,9,26,67,68] and such a trend continues to date [21,69]. These studies yielded impressive evidence that parasite abundance typically increases first and, once the host evolves a defense mechanism, it decreases again. However, sustained allele frequency oscillations of a particular allele, as predicted by numerous theoretical models assuming constant population size in the absence of any stochastic effects, have not been reported. We here propose that Lotka-Volterra population size fluctuations and the associated stochastic effects represent an inherent property of host-parasite interactions that can lead to rapid fixation of alleles, even those initially rare, thus preventing sustained oscillations. Consequently, Lotka-Volterra population size fluctuations have the potential to stop the Red Queen - unless novel variants are introduced into the population and/or additional selective constraints maintain allelic diversity. In retrospect, our findings may not be entirely unexpected. However, to date, they have not yet been directly demonstrated using a systematic analysis approach, as implemented here. More importantly, they are generally neglected in the numerous current empirical studies on the topic, in spite of their potential importance. They clearly deserve specific attention in future theoretical and empirical work aimed at an improved understanding of host-parasite coevolutionary dynamics.

Appendix

Relating stochastic and deterministic dynamics

Stochastic models are often developed starting from deterministic formulations [42]. Since the same deterministic formulation can be the limiting case of many individual based models, this procedure may be problematic. Instead, beginning from a stochastic, individual based description and then calculating the deterministic analogue will provide only a single direct link between the two approaches and allows for their direct comparison.

We consider a haploid system involving two antagonistic pairs, two alleles in hosts and parasites each. Firstly, all possible changes are written in the form of simple chemical reactions. In our particular case we have eight such possible reactions. We denote the two hosts and the two parasites by H_1 and H_2 and P_1 and P_2 respectively. Thus we have,



For instance, a parasite 2 individual dies with the rate \tilde{c} . From these rate reactions, we obtain the transition rates of the system. Depending on the number of individuals of the different types namely $\mathbf{n} = \{n_{H_1}, n_{H_2}, n_{P_1}, n_{P_2}\}$, we write the rates as,

$$\begin{aligned}
 T(n_{H_1} + 1, n_{H_2}, n_{P_1}, n_{P_2} | \mathbf{n}) &= \tilde{\mu} \frac{n_{H_1}}{N_{\text{avg}}} \\
 T(n_{H_1}, n_{H_2} + 1, n_{P_1}, n_{P_2} | \mathbf{n}) &= \tilde{\mu} \frac{n_{H_2}}{N_{\text{avg}}} \\
 T(n_{H_1} - 1, n_{H_2}, n_{P_1} + 1, n_{P_2} | \mathbf{n}) &= 2\tilde{b} \frac{n_{H_1}}{N_{\text{avg}}} \frac{n_{P_1}}{N_{\text{avg}}} \\
 T(n_{H_1}, n_{H_2} - 1, n_{P_1}, n_{P_2} + 1 | \mathbf{n}) &= 2\tilde{b} \frac{n_{H_2}}{N_{\text{avg}}} \frac{n_{P_2}}{N_{\text{avg}}} \\
 T(n_{H_1}, n_{H_2}, n_{P_1} - 1, n_{P_2} | \mathbf{n}) &= \tilde{c} \frac{n_{P_1}}{N_{\text{avg}}} \\
 T(n_{H_1}, n_{H_2}, n_{P_1}, n_{P_2} - 1 | \mathbf{n}) &= \tilde{c} \frac{n_{P_2}}{N_{\text{avg}}}
 \end{aligned} \tag{6}$$

where the reaction rates, have been corrected by each reactions combinatorial possibility [70,71] and N_{avg} is the average population size which we consider to be the same for the hosts as well as the parasites (the difference in the average population size can be interpreted as the ratio between the growth rate of hosts and the death rate of

parasites). Using these rates, we can write down deterministic differential equations for the change in the average number of a certain type, e.g.

$$\frac{d\langle n_{H_1} \rangle}{dt} = \tilde{\mu} \frac{n_{H_1}}{N_{\text{avg}}} - 2\tilde{b} \frac{n_{H_1}}{N_{\text{avg}}} \frac{n_{P_1}}{N_{\text{avg}}}. \tag{7}$$

Introducing rescaled reaction rates, $\mu = \frac{\tilde{\mu}}{N_{\text{avg}}}$, $b = 2\frac{\tilde{b}}{N_{\text{avg}}}$ and $c = \frac{\tilde{c}}{N_{\text{avg}}}$, we obtain

$$\frac{d\langle n_{H_1} \rangle}{dt N_{\text{avg}}} = \mu \left\langle \frac{n_{H_1}}{N_{\text{avg}}} \right\rangle - b \left\langle \frac{n_{H_1}}{N_{\text{avg}}} \frac{n_{P_1}}{N_{\text{avg}}} \right\rangle. \tag{8}$$

In the limit of a large population size we recover the mean field approximation or the population level model [71],

$$\dot{h}_1 = h_1(\mu - bp_1) \tag{9}$$

where $\dot{h}_1 = \frac{d\langle n_{H_1} \rangle}{dt N_{\text{avg}}}$ and the frequencies of H_i and P_i are given by h_i and p_i . In the same way, we can derive deterministic differential equations for the frequencies of the other types,

$$\dot{h}_1 = h_1(\mu - bp_1) \tag{10}$$

$$\dot{h}_2 = h_2(\mu - bp_2) \tag{11}$$

$$\dot{p}_1 = p_1(bh_1 - c) \tag{12}$$

$$\dot{p}_2 = p_2(bh_2 - c) \tag{13}$$

Stochastic simulations

The Gillespie algorithm gives an exact numerical solution of the Master equation of the system [41,70,71]. Our stochastic simulations are implementations of this

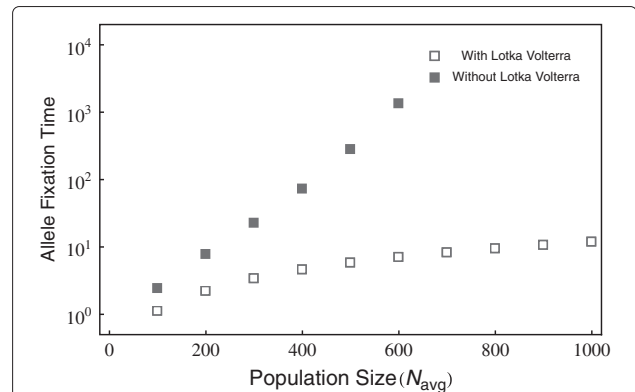


Figure 5 Allele fixation/extinction times for any of the interacting types when we do include a slight interaction between the otherwise independent Lotka-Volterra cycles. As compared to Figure 2 the fixation times in the case without Lotka-Volterra oscillations reduce with slight interaction between independent cycles. However for the case with Lotka-Volterra oscillations the fixation times are practically unchanged. For all simulations the initial condition were $H_1 = H_2 = N_{\text{avg}}/2$, $P_1 = 90N_{\text{avg}}/100$, $P_2 = 10N_{\text{avg}}/100$, and the parameters $\mu = 5$, $c = 2.5$, $b = 10/N_{\text{avg}}$ and $\varepsilon = 0.1b$ with averages over 10^6 realizations).

algorithm with the transition rates as defined in Eqs. 6. Since the population size is not constrained, this simulation method includes a stochastic analogue of the Lotka-Volterra cycles.

We computationally remove the Lotka-Volterra cycles by culling the population of each species after N_{avg} transitions have taken place. During the N_{avg} transitions the types within a species can evolve to different frequencies. But in the end they are reset to sum up to N_{avg} while maintaining the relative abundances. The Gillespie method is discrete in the number of individuals but continuous in time. The unit of time is the same as in the deterministic system.

Alternatively we can consider a small amount ε of interactions between the otherwise independent Lotka-Volterra interactions. This is then represented by the following set of differential equations,

$$\dot{h}_1 = h_1(\mu - bp_1 - \varepsilon p_2) \quad (14)$$

$$\dot{h}_2 = h_2(\mu - bp_2 - \varepsilon p_1) \quad (15)$$

$$\dot{p}_1 = p_1(bh_1 + \varepsilon h_2 - c) \quad (16)$$

$$\dot{p}_2 = p_2(\varepsilon h_1 + bh_2 - c). \quad (17)$$

Even for this case, including Lotka-Volterra interactions causes a faster extinction of the Red Queen cycles involving all four types. As an example we provide simulation results where in addition to similar parameters as in Figure 2 we add a $\varepsilon = 0.1b$ (Figure 5). Although the fixation time is elevated as compared to the case with no interactions (Figure 2), they are still not comparable to the extremely high fixations times observed when Lotka-Volterra dynamics is excluded.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

HS and AP conceived the project. CSG and AT developed the model and performed simulations. All authors analysed the results and wrote the manuscript. All authors read and approved the final manuscript.

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References

1. Carroll L: *Through the Looking-glass, and what Alice Found there*. London: Macmillan; 1871. Reprinted: Bloomsbury, London, 2001.
2. Ridley M: *Evolution*, 3rd edn. Oxford: Wiley-Blackwell; 2003.
3. Barton NH, Briggs DEG, Eisen JA, Goldstein DB, Patel NH: *Evolution*, 1st edn. NY: Cold Spring Harbor Laboratory Press; 2007.
4. van Valen L: **A new evolutionary law**. *Evol Theory* 1973, **1**:1–30.
5. Woolhouse MEJ, Webster JP, Domingo E, Charlesworth B, Levin BR: **Biological and biomedical implications of the co-evolution of pathogens and their hosts**. *Nat Genet* 2002, **32**(4):569–577.
6. Schmid-Hempel P: *Evolutionary Parasitology: The Integrated Study of Infections, Immunology, Ecology, and Genetics*. USA: Oxford Biology. Oxford University Press; 2011.
7. Fenner F, Fantini B: *Biological Control of Vertebrate pests. The History of Myxomatosis—an Experiment in Evolution*. Oxfordshire: CABI Publishing; 1999.
8. Jokela J, Dybdahl MF, Lively CM: **The maintenance of sex, clonal dynamics, and host-parasite coevolution in a mixed population of sexual and asexual snails**. *Am Nat* 2009, **174**(s1):43–53.
9. Decaestecker E, Gaba S, Raeymaekers JAM, R. Stoks LvK, Ebert D, Meester LD: **Host-parasite 'Red queen' dynamics archived in pond sediment**. *Nature* 2007, **450**:870–873.
10. Bohannan BJM, Lenski RE: **Linking genetic change to community evolution: insights from studies of bacteria and bacteriophage**. *Ecol Lett* 2000, **3**(4):362–377.
11. Buckling A, Rainey PB: **Antagonistic coevolution between a bacterium and a bacteriophage**. *Proc R Soc B* 2002, **269**:931–936.
12. Brockhurst MA, Morgan AD, Fenton A, Buckling A: **Experimental coevolution with bacteria and phage: The pseudomonas fluorescens-phi2 model system**. *Infect Genet Evol* 2007, **7**(4):547–552.
13. Schulte RD, Makus C, Hasert B, Michiels NK, Schulerburg H: **Multiple reciprocal adaptations and rapid genetic change upon experimental coevolution of an animal host and its microbial parasite**. *Proc Natl Acad Sci USA* 2010, **107**(16):7359–7364.
14. Morran LT, Schmidt OG, Gelarden IA, Parrish-II RC, Lively CM: **Running with the red queen: Host-parasite coevolution selects for biparental sex**. *Science* 2011, **333**:216–218.
15. Bérénos C, Schmid-Hempel P, Wegner KM: **Experimental coevolution leads to a decrease in parasite-induced host mortality**. *J Evol Biol* 2011, **24**(8):1777–1782.
16. Fischer O, Schmid-Hempel P: **Selection by parasites may increase host recombination frequency**. *Biol Lett* 2005, **1**(2):193–195.
17. King KC, Delph LF, Jokela J, Lively CM: **Coevolutionary hotspots and coldspots for host sex and parasite local adaptation in a snail-trematode interaction**. *Oikos* 2011, **120**(9):1335–1340.
18. Koskella B, Thompson JN, Preston GM, Buckling A: **Local biotic environment shapes the spatial scale of bacteriophage adaptation to bacteria**. *Am Nat* 2011, **177**(4):440–451.
19. Gandon S, Buckling A, Decaestecker E, Day T: **Host-parasite coevolution and patterns of adaptation across time and space**. *J Evol Biol* 2008, **21**:1861–1866.
20. Gaba S, Ebert D: **Time-shift experiments as a tool to study antagonistic coevolution**. *Trends Ecol Evol* 2009, **24**(4):226–232.
21. Brockhurst MA, Koskella B: **Experimental coevolution of species interactions**. *Trends Ecol Evol* 2013, **28**(6):367–375.
22. Lenski RE, Levin BR: **Constraints on the coevolution of bacteria and virulent phage: A model, some experiments and predictions for natural communities**. *Am Nat* 1985, **125**:585–602.
23. Scanlan PD, Hall AR, Lopez-Pascua LDC, Buckling A: **Genetic basis of infectivity evolution in a bacteriophage**. *Mol Ecol* 2011, **20**:981–989.
24. Meyer JR, Dobias DT, Weitz JS, Barrick JE, Quick RT, Lenski RE: **Repeatability and contingency in the evolution of a key innovation in phage lambda**. *Science* 2012, **335**:428–432.
25. Hall AR, Scanlan PD, Morgan AD, Buckling A: **Host-parasite coevolutionary arms races give way to fluctuating selection**. *Ecol Lett* 2011, **14**:635–642.
26. Lively CM, Dybdahl MF: **Parasite adaptation to locally common host genotypes**. *Nature* 2000, **405**:679–681.
27. Anderson RM, May RM: *Infectious Diseases of Humans: Dynamics and Control*. USA: Oxford University Press; 1992.
28. Dobson AP, Hudson PJ: **Microparasites: observed patterns in wild animal populations**. In *Ecology of Infectious Diseases in Natural Populations*. Edited by Grenfell BT, Dobson AP. Cambridge: Cambridge University Press; 1995:52–89.
29. Hudson PJ, Dobson AP: **Macroparasites: observed patterns in naturally fluctuating animal populations**. In *Ecology of Infectious Diseases in Natural Populations*. Edited by Grenfell BT, Dobson AP. Cambridge: Cambridge University Press; 1995:144–176.
30. Bohannan BJM, Lenski RE: **Effect of prey heterogeneity on the response of a model food chain to resource enrichment**. *Am Nat* 1999, **153**:73–82.

31. Lotka AJ: **Analytical note on certain rhythmic relations in organic systems.** *Proc Nat Acad Sci USA* 1920, **6**:410–415.
32. Volterra V: **Variations and fluctuations of the number of individuals in animal species living together.** *J du conseil international pour l'exploration de la mer* 1928, **3**(1):3–51.
33. May RM, Anderson RM: **Epidemiology and genetics in the coevolution of parasites and hosts.** *Proc R Soc Lond B Biol Sci* 1983, **219**(1216):281–313.
34. Frank SA: **Ecological and genetic models of host-pathogen coevolution.** *Heredity* 1991, **67**:73–83.
35. Heesterbeek JAP, Roberts MG: **Mathematical models for microparasites of wildlife.** In *Ecology of Infectious Diseases in Natural Populations*. Edited by Grenfell BT, Dobson AP. Cambridge: Cambridge University Press; 1995:90–122.
36. Roberts MG, Smith G, Grenfell BT: **Mathematical models for macroparasites of wildlife.** In *Ecology of Infectious Diseases in Natural Populations*. Edited by Grenfell BT, Dobson AP. Cambridge: Cambridge University Press; 1995:177–208.
37. Gandon S, Nuismer SL: **Interactions between genetic drift, gene flow, and selection mosaics drive parasite local adaptation.** *Am Nat* 2009, **173**(2):212–224.
38. Kirby GC, Burdon JJ: **Effects of mutation and random drift on Leonard's gene-for-gene coevolution model.** *Phytopathology* 1997, **87**(5):488–493.
39. Salathé M, Scherer A, Bonhoeffer S: **Neutral drift and polymorphism in gene-for-gene systems.** *Ecol Lett* 2005, **8**:925–932.
40. Quigley BJZ, López DG, Buckling A, McKane AJ, Brown SP: **The mode of host-parasite interactions shapes coevolutionary dynamics and the fate of host cooperation.** *Proc R Soc B* 2012, **279**(1743):3742–3748.
41. Gillespie DT: **Exact stochastic simulation of coupled chemical reactions.** *J Phys Chem* 1977, **81**(25):2340–2361.
42. Black AJ, McKane AJ: **Stochastic formulation of ecological models and their applications.** *Trends Ecol Evol* 2012, **27**:337–345.
43. Stiven AE: **Experimental studies on the epidemiology of the host parasite system, hydra and hydramoeba hydroxena (Entz). II. the components of a simple epidemic.** *Ecol Monogr* 1964, **34**(2):119–142.
44. Bach PD, Smith HS: **Are population oscillations inherent in the host-parasite relation?** *Ecology* 1941, **22**(4):363–369.
45. Pimentel D: **Population regulation and genetic feedback evolution provides foundation for control of herbivore, parasite, and predator numbers in nature.** *Science* 1968, **159**(3822):1432–1437.
46. Utida S: **Cyclic fluctuations of population density intrinsic to the host-parasite system.** *Ecology* 1957, **38**(3):442–449.
47. Hudson PJ, Dobson AP, Newborn D: **Prevention of population cycles by parasite removal.** *Science* 1998, **282**(5397):2256–2258.
48. Gillespie JH: **Genetic drift in an infinite population: the pseudohitchhiking model.** *Genetics* 2000, **155**(2):909–919.
49. Lenormand T, Roze D, Rousset F: **Stochasticity in evolution.** *Trends Ecol Evol* 2009, **24**(3):157–165.
50. Campos PRA, Wahl LM: **The effects of population bottlenecks on clonal interference, and the adaptation effective population size.** *Evolution* 2009, **63**(4):950–958.
51. Wahl LM, Krakauer DC: **Models of experimental evolution: the role of genetic chance and selective necessity.** *Genetics* 2000, **156**(3):1437–1448.
52. Wahl LM, Gerrish PJ: **The probability that beneficial mutations are lost in populations with periodic bottlenecks.** *Evolution* 2001, **55**(12):2606–2610.
53. Handel A, Bennett MR: **Surviving the bottleneck: transmission mutants and the evolution of microbial populations.** *Genetics* 2008, **180**(4):2193–2200.
54. Ponting CP: **The functional repertoires of metazoan genomes.** *Nat Rev Genet* 2008, **9**:689–698.
55. Levasseur A, Pontarotti P: **The role of duplications in the evolution of genomes highlights the need for evolutionary-based approaches in comparative genomics.** *Biol Direct* 2011, **6**(11):1–12.
56. Andersson DI, Hughes D: **Gene amplification and adaptive evolution in bacteria.** *Ann Rev Genet* 2009, **43**:167–195.
57. Moran GP, Coleman DC, Sullivan DJ: **Comparative genomics and the evolution of pathogenicity in human pathogenic fungi.** *Eukaryotic Cell* 2011, **10**(1):34–42.
58. Merhej V, Raoult D: **Rickettsial evolution in the light of comparative genomics.** *Biol Rev* 2011, **86**:379–405.
59. Iskow RC, Gokcumen O, Lee C: **Exploring the role of copy number variants in human adaptation.** *Trends Genet* 2012, **28**(6):245–257.
60. Korbel JO, Kim PM, Chen X, Urban AE, Weissman S, Snyder M, Gerstein MB: **The current excitement about copy-number variation: how it relates to gene duplications and protein families.** *Curr Opin Struct Biol* 2008, **18**:366–374.
61. Lively CM, Apanius V: **Genetic diversity in host-parasite interactions.** In *Ecology of Infectious Diseases in Natural Populations*. Edited by Grenfell BT, Dobson AP. Cambridge: Cambridge University Press; 1995:421–449.
62. Frank SA: **Specific and non-specific defense against parasitic attack.** *J Theor Biol* 2000, **202**(4):283–304.
63. Agrawal AF, Lively CM: **Modelling infection as a two-step process combining gene-for-gene and matching-allele genetics.** *Proc R Soc B Biol Sci* 2003, **270**(1512):323–334.
64. Fenton A, Antonovics J, Brockhurst MA: **Two-step infection processes can lead to coevolution between functionally independent infection and resistance pathways.** *Evolution* 2012, **66**(7):2030–2041.
65. Thrall PH, Laine A-L, Ravensdale M, Nemri A, Dodds PN, Barrett LG, Burdon JJ: **Rapid genetic change underpins antagonistic coevolution in a natural host-pathogen metapopulation.** *Ecol Lett* 2012, **15**(5):425–435.
66. Wolinska J, King KC: **Environment can alter selection in host-parasite interactions.** *Trends Parasitol* 2009, **25**(5):236–244.
67. Dybdahl MF, Lively CM: **Host-parasite coevolution: evidence for rare advantage and time-lagged selection in a natural population.** *Evolution* 1998, **52**(4):1057–1066.
68. Eizaguirre C, Lenz TL, Kalbe M, Milinski M: **Rapid and adaptive evolution of MHC genes under parasite selection in experimental vertebrate populations.** *Nat Commun* 2012, **3**(621):1–6.
69. Luijckx P, Fienberg H, Duneau D, Ebert D: **A matching-allele model explains host resistance to parasites.** *Curr Biol* 2013, **23**(12):1085–1088.
70. Gillespie D: **A general method for numerically simulating the stochastic time evolution of coupled chemical reactions.** *J Comput Phys* 1976, **22**:403–434.
71. McKane AJ, Newman TJ: **Stochastic models in population biology and their deterministic analogs.** *Phys Rev E* 2004, **70**:19.

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