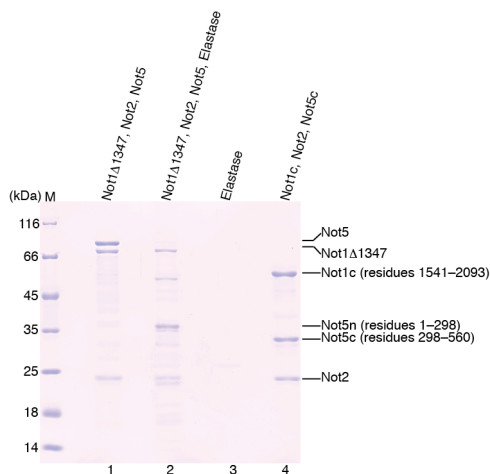


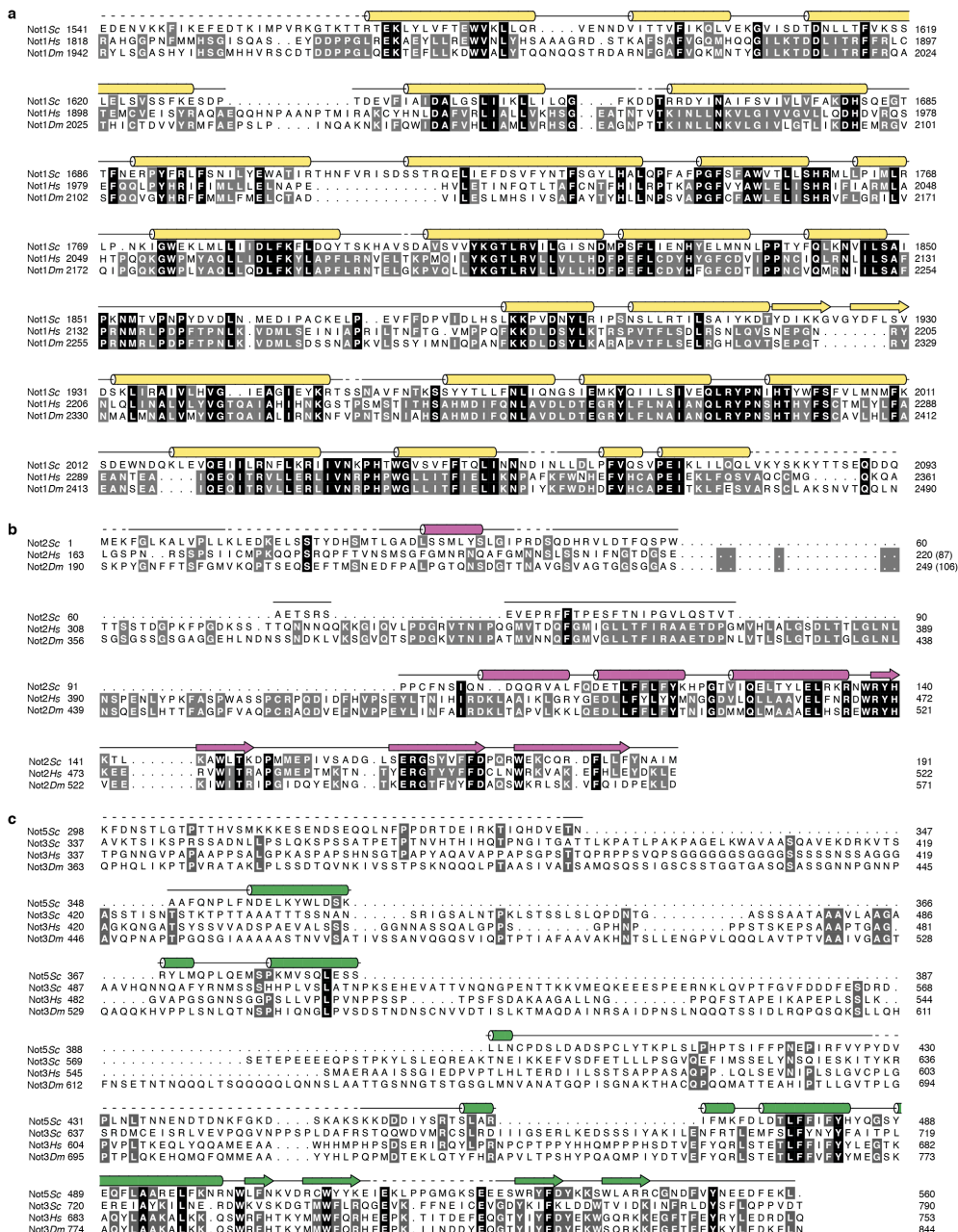
**Structure and RNA-binding properties  
of the Not1–Not2–Not5 module of the yeast Ccr4–Not complex**

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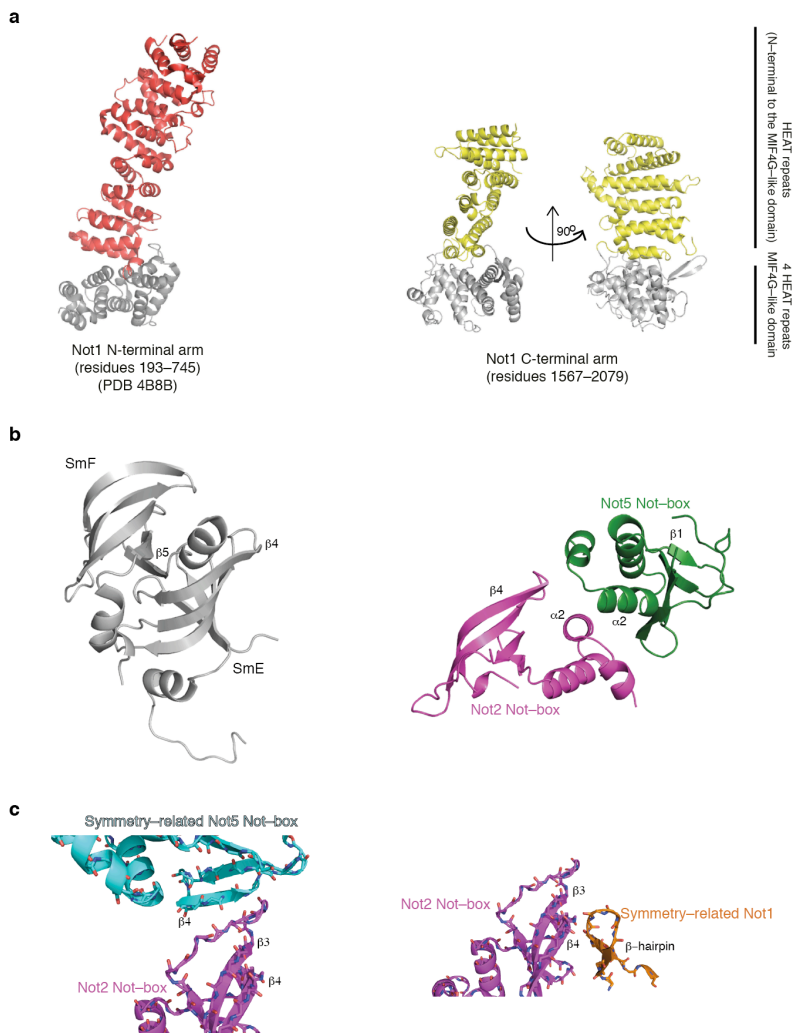
**Supplementary Information**



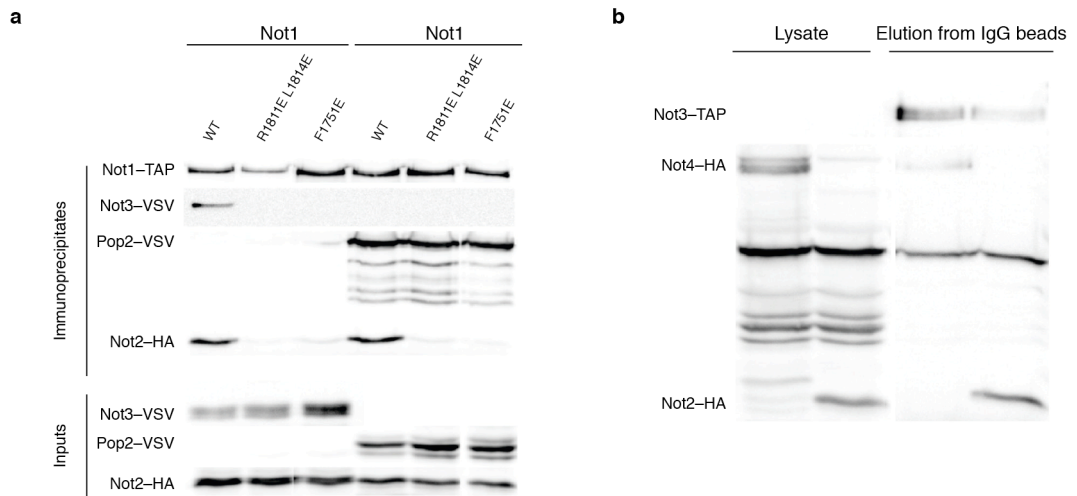
**Supplementary Figure 1** Identification of the core of the *S. cerevisiae* Not1–Not2–Not5 interaction. The 12% SDS PAGE gel shows in lane 1 the larger complex that we initially purified (Not1 starting at residue 1348, Not2 f.l. and Not5 f.l.). Lane 2 shows the result of the limited proteolysis of the complex in lane 1 with elastase. Lane 3 shows the protease alone as a control. Lane 4 shows the complex reconstituted with the minimal interacting regions of Not1, Not2 and Not5 that yielded diffracting crystals (Not1c, Not2 and Not5c).



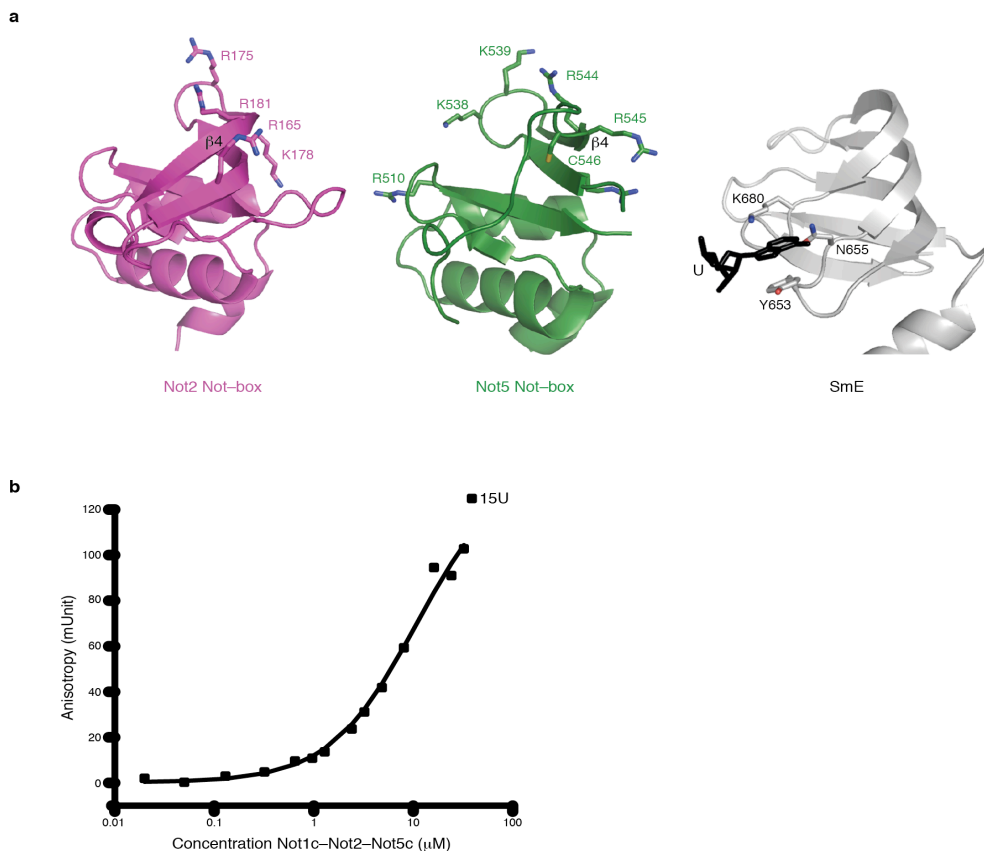
**Supplementary Figure 2** Structure-based sequence alignments of Not1c, Not2 and Not5c. The sequence alignments include the polypeptides of *S. cerevisiae* (*Sc*) Not1, Not2 and Not5 we crystallized in a complex and their orthologues from *H. sapiens* (*Hs*) and *D. melanogaster* (*Dm*). Not5*Sc* is a homologue of Not3. Secondary structure elements are shown above the sequences and colored in yellow for Not1 (**a**), magenta for Not2 (**b**) and green for Not5 (**c**). Straight lines refer to extended regions and dotted lines refer to disordered regions in the structure of the *S. cerevisiae* complex. Sequence conservation is highlighted in shades of gray.



**Supplementary Figure 3** The HEAT and Sm folds of Not1–Not2–Not5. **(a)** Comparison of the HEAT-repeat architecture in the C-terminal arm of Not1 (on the right) and the N-terminal arm of Not1 (on the left, PDB code 4B8B<sup>1</sup>). The MIF4G-like folds are shown in gray. The longer HEAT-repeat units perpendicular to the MIF4G-like folds are shown in yellow and red for the C-terminal and N-terminal arms, respectively. **(b)** Dimerization properties of Not-box domains. The subcomplex of SmF and SmE (PDB code 2Y9A<sup>2</sup>) is shown on the left in gray. The  $\beta 4$  strand of one monomer (SmE) interacts with strand  $\beta 5$  of the other monomer (SmF). On the right, dimerization of Not2–Not5 leaves strand  $\beta 4$  exposed to solvent. **(c)** Lattice contacts are reminiscent of Sm–Sm interactions. In the upper panel, the loop between strands  $\beta 2$  and  $\beta 3$  of a Not2 molecule (in magenta) has an extended conformation and interacts both with the  $\beta 4$  strand of a symmetry-related Not5 molecule (in cyan). In the lower panel, the  $\beta 4$  strand of a Not2 molecule (in magenta) interacts with the  $\beta$ -hairpin of a symmetry-related Not1 molecule (in orange).



**Supplementary Figure 4** *In vivo* interactions of Not proteins **(a)** Immunoprecipitation of Not1 from yeast strains harbouring a TAP tagged Not1 wild-type (WT) or indicated mutants and carrying tagged chromosomal variant of Not2 and Not3 (BSY1230), or Not2 and Pop2/Caf1 (BSY1231). Coimmunoprecipitation of Not2-HA, Not3-VSV or Pop2-VSV was assayed by western blotting. As a control, the presence of the tagged protein in the starting extracts was also assayed. **(b)** Immunoprecipitation of Not3 from yeast strains carrying tagged chromosomal variant of Not3, Pop2/Caf1 and Not4 (BSY1240), or Not3, Caf1/Pop2 and Not2 (BSY1242). Coimmunoprecipitation of Not2-HA or Not4-HA was assayed by western blotting. As a control, the level of the tagged protein in the starting extracts was also assayed.



**Supplementary Figure 5** Protein and RNA interactions at the Not-boxes **(a)** Surface features of the Not2 and Not5 Not-boxes. On the left is the structure of Not2, showing Arg165 (putative ADA2-binding residue) as well as positively charged residues at a similar position as in Not5. In the central panel is the Not-box of Not5, in the same orientation, showing the uridine-crosslinked residue Cys546 as well as the surrounding positively charged residues (as in **Fig. 5d**). On the right is the structure of RNA-bound SmE (U4 snRNP, PDB code 2Y9A<sup>2</sup>) oriented in a similar view as the structures in the left and central panels (after optimal superimposition), showing a bound uridine nucleotide (in black). **(b)** Quantification of the RNA-binding properties of Not1c-Not2-Not5c by fluorescence anisotropy. The  $K_d$  of the Not1c-Not2-Not5c complex to 6-FAM-labeled U<sub>15</sub> RNA under these conditions was found to be  $9.47 \pm 0.95 \mu\text{M}$ .

## Reference

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- 2 Leung, A. K. W., Nagai, K. & Li, J. Structure of the spliceosomal U4 snRNP core domain and its implication for snRNP biogenesis. *Nature* **473**, 536–539 (2011).