

Supplementary Data

The MOF-containing NSL complex associates globally with housekeeping genes, but activates only a defined subset

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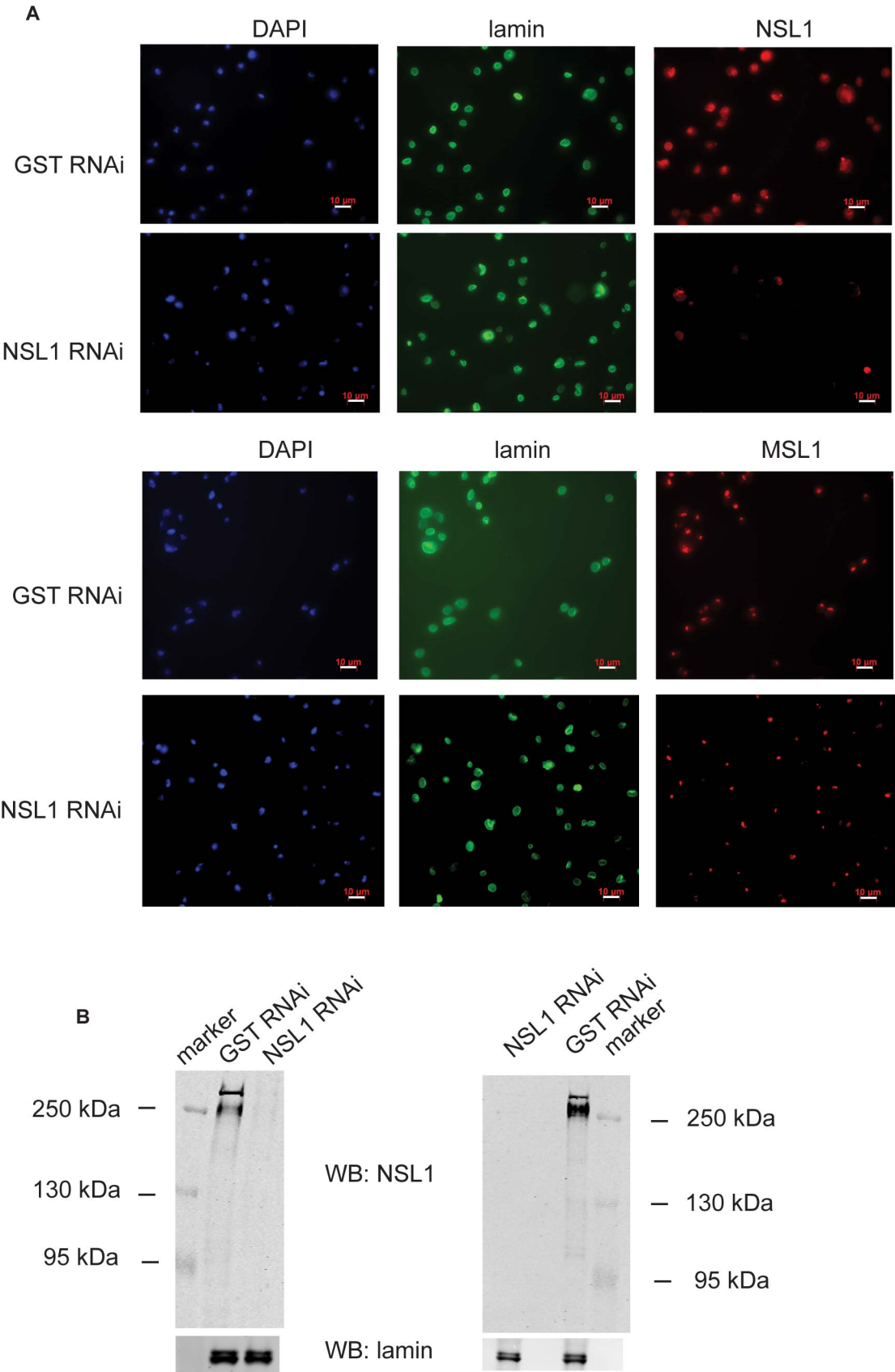
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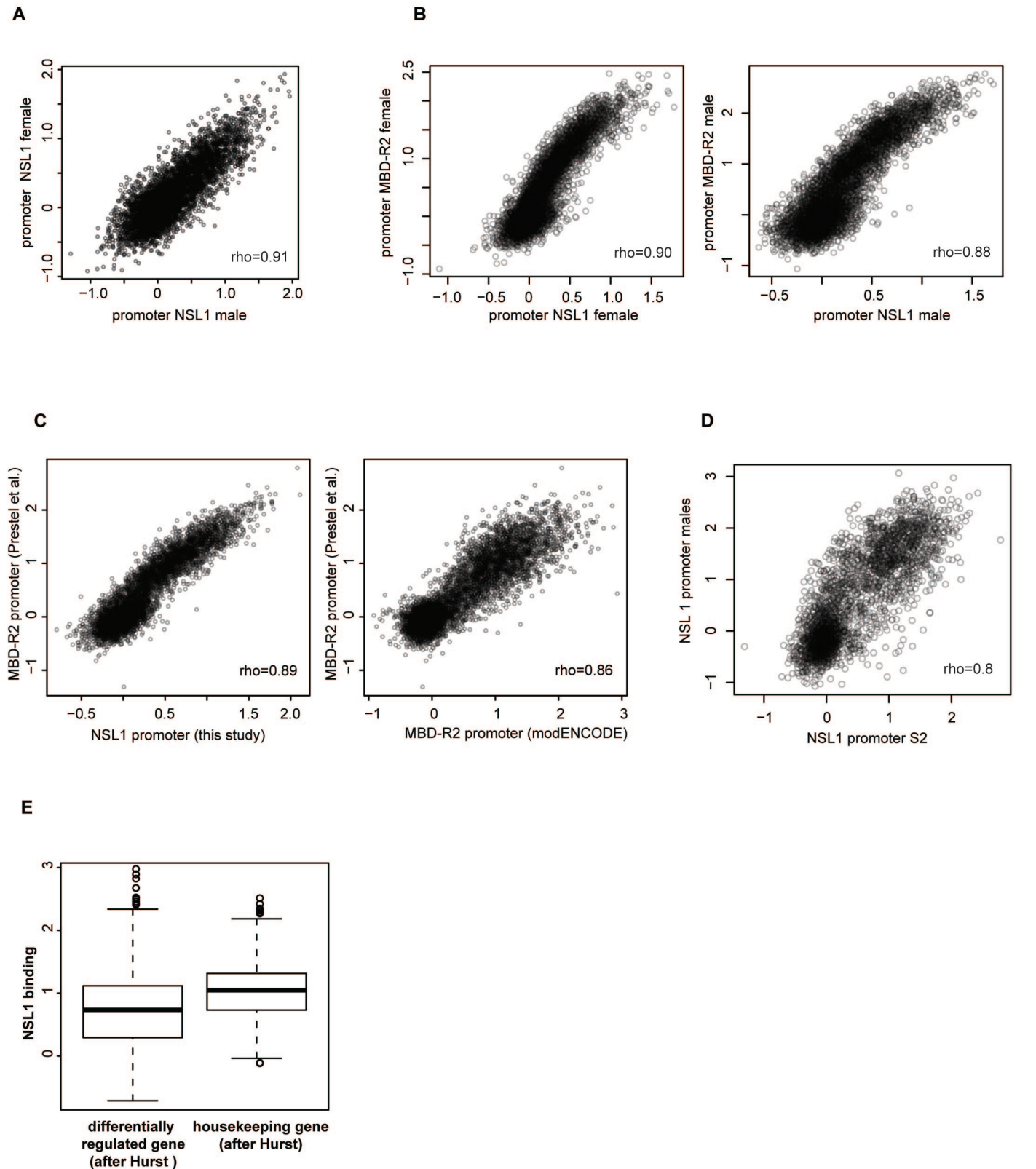
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Supplementary Figure 1: NSL1 antibody specificity control.

(A) Indirect immunofluorescence with the indicated antibodies in S2 cells treated with control and NSL1 RNAi. MSL1 (subunit of DCC complex) and lamin stainings are not compromised under NSL1 RNAi conditions. DNA was counterstained with DAPI. Scale bar (10 μ m) is indicated in white.

(B) Immunoblot analysis on S2 chromatin extracts (left) and Kc chromatin extracts (right) from NSL1 RNAi, and control RNAi (Glutathion-S-Transferase RNAi) using the NSL1 antibody. Lamin immunoblot was used as a loading control. The NSL1 protein shows aberrant mobility (11).



Supplementary Figure 2: NSL complex members bind similar across conditions.

(A-D) Scatter plots comparing NSL1 promoter binding in adult male and female flies (A), NSL1 vs. MBD-R2 in adult female flies (B, right), NSL1 binding with MBD-R2 binding in adult male flies (B, left), NSL1 binding and MBD-R2 binding (12) in S2 cells (C, left), MBD-R2 binding in S2 cells between the datasets of (12) and the modENCODE consortium (C, right), NSL1 binding in S2 cells with NSL1 binding in adult male cells (D). Rank-based Spearman correlation coefficients (ρ) are indicated.

(E) Genes were categorized in “housekeeping” and “differentially regulated” genes, respectively, based on the classification of Hurst and colleagues (22). Binding of NSL1 was displayed in these two groups and was found to be significantly higher in housekeeping genes than differentially regulated genes (Welch two sample t test, $p < 2.2 \times 10^{-16}$). Similar results were obtained using the S2 MBD-R2 ChIP-chip dataset from (12) and the MBD-R2 dataset generated by the modENCODE consortium (17). Note that the “housekeeping” definition after Hurst and colleagues is more stringent, which results in a number of genes classified as “housekeeping” by our definition but “differentially regulated” by Hurst and colleagues.

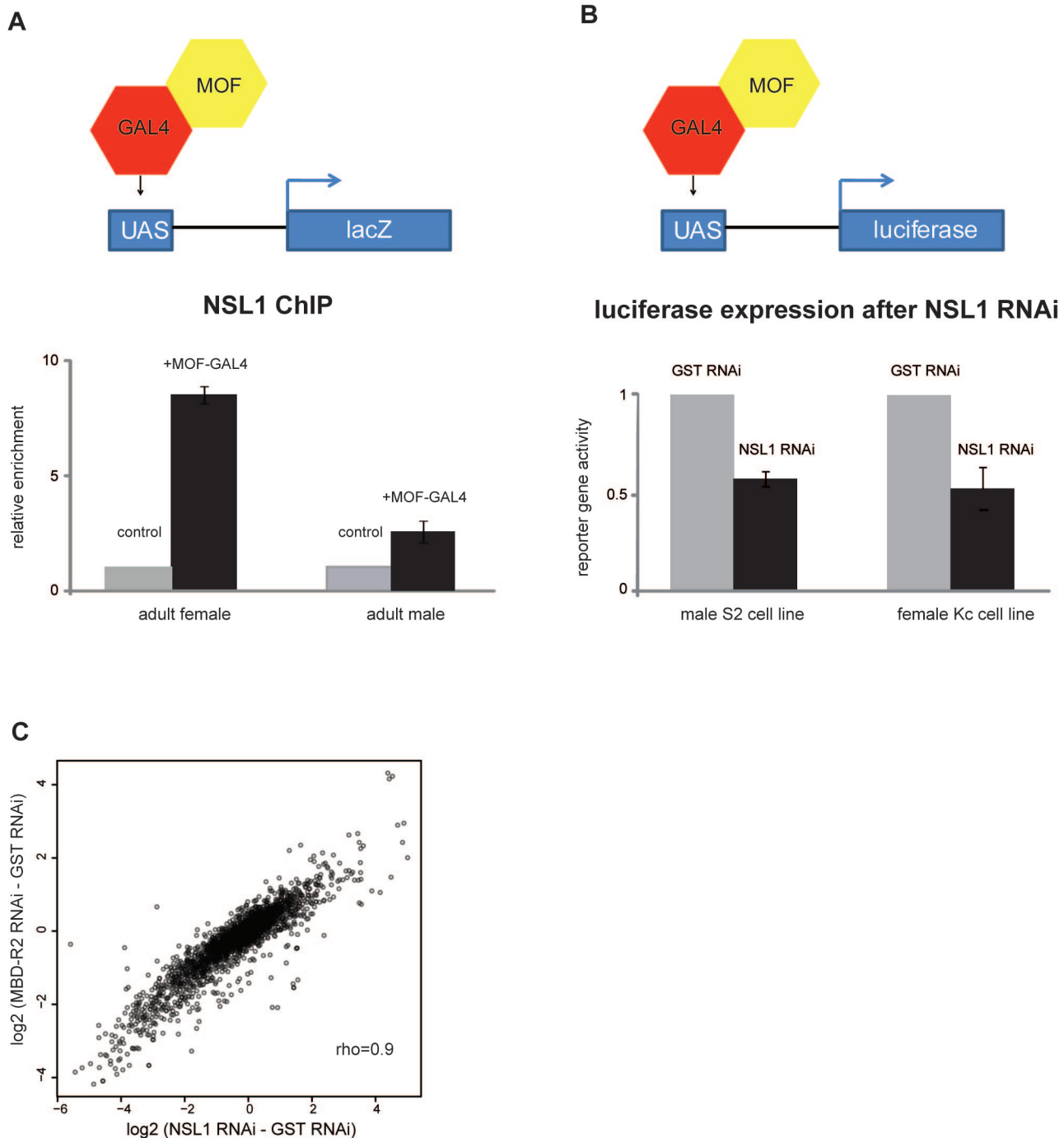
Supplementary Figure 3: Gene Ontology (GO) enriched biological processes of NSL complex bound genes

Term	PValue
GO:0051188~cofactor biosynthetic process	2,33E-12
GO:0031023~microtubule organizing center organization	7,03E-11
GO:0051186~cofactor metabolic process	1,40E-03
GO:0007017~microtubule-based process	1,70E-03
GO:0051297~centrosome organization	1,70E-03
GO:0009108~coenzyme biosynthetic process	3,00E-03
GO:0007098~centrosome cycle	4,50E-03
GO:0000226~microtubule cytoskeleton organization	4,90E-03
GO:0070271~protein complex biogenesis	5,80E-03
GO:0006461~protein complex assembly	5,80E-03
GO:0006732~coenzyme metabolic process	9,70E-03
GO:0016071~mRNA metabolic process	9,80E-03
GO:0043069~negative regulation of programmed cell death	9,90E-03
GO:0060548~negative regulation of cell death	9,90E-03
GO:0006397~mRNA processing	1,03E-02
GO:0022402~cell cycle process	1,17E-02
GO:0000278~mitotic cell cycle	1,17E-02
GO:0051656~establishment of organelle localization	1,50E-02
GO:0033043~regulation of organelle organization	1,55E-02
GO:0007010~cytoskeleton organization	1,82E-02
GO:0006352~transcription initiation	2,25E-02
GO:0043066~negative regulation of apoptosis	2,26E-02
GO:0022403~cell cycle phase	2,40E-02
GO:0007049~cell cycle	2,46E-02
GO:0051298~centrosome duplication	2,63E-02
GO:0007173~epidermal growth factor receptor signaling pathway	2,69E-02
GO:0000279~M phase	2,76E-02
GO:0051130~positive regulation of cellular component organization	2,99E-02
GO:0009314~response to radiation	3,30E-02
GO:0017038~protein import	3,41E-02
GO:0033365~protein localization in organelle	3,44E-02
GO:0006367~transcription initiation from RNA polymerase II promoter	4,07E-02
GO:0006605~protein targeting	4,08E-02
GO:0051640~organelle localization	4,21E-02
GO:0006733~oxidoreduction coenzyme metabolic process	4,92E-02
GO:0048477~oogenesis	5,56E-02
GO:0016044~membrane organization	5,59E-02
GO:0006366~transcription from RNA polymerase II promoter	5,62E-02
GO:0007447~imaginal disc pattern formation	5,73E-02
GO:0007052~mitotic spindle organization	5,77E-02
GO:0006468~protein amino acid phosphorylation	5,88E-02
GO:0031124~mRNA 3'-end processing	6,00E-02
GO:0007276~gamete generation	6,06E-02
GO:0019953~sexual reproduction	6,74E-02
GO:0007051~spindle organization	6,99E-02
GO:0051276~chromosome organization	7,19E-02
GO:0008156~negative regulation of DNA replication	7,27E-02
GO:0007292~female gamete generation	8,03E-02
GO:0016310~phosphorylation	8,32E-02
GO:0007349~cellularization	8,38E-02
GO:0000289~nuclear-transcribed mRNA poly(A) tail shortening	8,59E-02
GO:0000288~nuclear-transcribed mRNA catabolic process, deadenylation-d	8,59E-02
GO:0035222~wing disc pattern formation	8,77E-02
GO:0051493~regulation of cytoskeleton organization	8,77E-02
GO:0009628~response to abiotic stimulus	9,11E-02
GO:0032504~multicellular organism reproduction	9,29E-02
GO:0048609~reproductive process in a multicellular organism	9,29E-02
GO:0051301~cell division	9,52E-02
GO:0006281~DNA repair	9,65E-02
GO:0065003~macromolecular complex assembly	9,66E-02
GO:0006259~DNA metabolic process	9,70E-02
GO:0016072~rRNA metabolic process	9,70E-02
GO:0006612~protein targeting to membrane	9,70E-02
GO:0006364~rRNA processing	9,70E-02

Gene Ontology (GO) enrichment analysis was performed for biological processes using DAVID GO SLIM (57). P values were derived from a Fisher Exact test using the DAVID web-implementation. All NSL1 bound genes were compared to all active genes in S2 cells.

Term	PValue	Term	PValue
GO:0007606~sensory perception of chemical stimulus	1,57E-07	GO:0007451~dorsal/ventral lineage restriction, imaginal disc	0,01073
GO:0055114~oxidation reduction	4,48E-07	GO:0048598~embryonic morphogenesis	0,01169
GO:0007600~sensory perception	3,79E-06	GO:0044275~cellular carbohydrate catabolic process	0,01187
GO:0050890~cognition	6,59E-06	GO:0046164~alcohol catabolic process	0,01187
GO:0007155~cell adhesion	1,49E-05	GO:0006508~proteolysis	0,01269
GO:0022610~biological adhesion	1,49E-05	GO:0007476~imaginal disc-derived wing morphogenesis	0,01297
GO:0035218~leg disc development	1,79E-05	GO:0007494~midgut development	0,01349
GO:0006952~defense response	2,93E-05	GO:0009996~negative regulation of cell fate specification	0,01349
GO:0048569~post-embryonic organ development	5,89E-05	GO:0042659~regulation of cell fate specification	0,01349
GO:0035286~leg segmentation	1,03E-04	GO:0010453~regulation of cell fate commitment	0,01349
GO:0035285~appendage segmentation	1,03E-04	GO:0010454~negative regulation of cell fate commitment	0,01349
GO:0046942~carboxylic acid transport	1,03E-04	GO:0007626~locomotory behavior	0,01432
GO:0015849~organic acid transport	1,03E-04	GO:0007472~wing disc morphogenesis	0,01436
GO:0006865~amino acid transport	1,35E-04	GO:0043473~pigmentation	0,01489
GO:0015837~amine transport	1,35E-04	GO:0007164~establishment of tissue polarity	0,0158
GO:0007166~cell surface receptor linked signal transduction	1,41E-04	GO:0001736~establishment of planar polarity	0,0158
GO:0007552~metamorphosis	1,44E-04	GO:0007435~salivary gland morphogenesis	0,01581
GO:0035111~leg joint morphogenesis	1,92E-04	GO:0022612~gland morphogenesis	0,01581
GO:0007444~imaginal disc development	2,05E-04	GO:0001654~eye development	0,01737
GO:0045087~innate immune response	3,64E-04	GO:0019318~hexose metabolic process	0,01837
GO:0007186~G-protein coupled receptor protein signaling pathway	3,97E-04	GO:0009072~aromatic amino acid family metabolic process	0,01867
GO:0048707~instar larval or pupal morphogenesis	3,99E-04	GO:0035167~larval lymph gland hemopoiesis	0,01867
GO:0048563~post-embryonic organ morphogenesis	4,28E-04	GO:0006935~chemotaxis	0,01867
GO:0007560~imaginal disc morphogenesis	4,28E-04	GO:0007469~antennal development	0,01867
GO:0042742~defense response to bacterium	5,24E-04	GO:0007440~foregut morphogenesis	0,01867
GO:0048737~imaginal disc-derived appendage development	5,54E-04	GO:0035166~post-embryonic hemopoiesis	0,01867
GO:0048732~gland development	5,60E-04	GO:0007608~sensory perception of smell	0,01867
GO:0009886~post-embryonic morphogenesis	6,30E-04	GO:0001738~morphogenesis of a polarized epithelium	0,02213
GO:0048736~appendage development	6,39E-04	GO:0006959~humoral immune response	0,02258
GO:0006955~immune response	6,55E-04	GO:0008219~cell death	0,02319
GO:0035214~eye-antennal disc development	7,53E-04	GO:0016265~death	0,02319
GO:0035114~imaginal disc-derived appendage morphogenesis	8,79E-04	GO:0007411~axon guidance	0,02345
GO:0007447~imaginal disc pattern formation	9,21E-04	GO:0035289~posterior head segmentation	0,02396
GO:0048565~gut development	9,88E-04	GO:0042688~crystal cell differentiation	0,02396
GO:0035107~appendage morphogenesis	0,00101	GO:0008587~imaginal disc-derived wing margin morphogenesis	0,02585
GO:0016052~carbohydrate catabolic process	0,00139	GO:0043067~regulation of programmed cell death	0,02652
GO:0009617~response to bacterium	0,00175	GO:0010941~regulation of cell death	0,02652
GO:0009791~post-embryonic development	0,00192	GO:0007478~leg disc morphogenesis	0,02674
GO:0050830~defense response to Gram-positive bacterium	0,00215	GO:0019730~antimicrobial humoral response	0,02721
GO:0007354~zygotic determination of anterior/posterior axis, embryo	0,00215	GO:0042060~wound healing	0,02983
GO:0035108~limb morphogenesis	0,00221	GO:0012501~programmed cell death	0,03034
GO:0060173~limb development	0,00221	GO:0009310~amine catabolic process	0,03038
GO:0035110~leg morphogenesis	0,00221	GO:0006007~glucose catabolic process	0,03224
GO:0007157~heterophilic cell adhesion	0,00238	GO:0019320~hexose catabolic process	0,03224
GO:0007449~proximal/distal pattern formation, imaginal disc	0,00238	GO:0048066~pigmentation during development	0,03293
GO:0035220~wing disc development	0,00258	GO:0016054~organic acid catabolic process	0,03293
GO:0007431~salivary gland development	0,00261	GO:0046395~carboxylic acid catabolic process	0,03293
GO:0035272~exocrine system development	0,00261	GO:0006576~biogenic amine metabolic process	0,03535
GO:0035120~post-embryonic appendage morphogenesis	0,00276	GO:0050829~defense response to Gram-negative bacterium	0,03535
GO:0042067~establishment of ommatidial polarity	0,00283	GO:0006334~nucleosome assembly	0,03535
GO:0010623~developmental programmed cell death	0,00378	GO:0035222~wing disc pattern formation	0,03789
GO:0007450~dorsal/ventral pattern formation, imaginal disc	0,00433	GO:0030030~cell projection organization	0,03796
GO:0007610~behavior	0,00438	GO:0000902~cell morphogenesis	0,03819
GO:0035215~genital disc development	0,00444	GO:0005976~polysaccharide metabolic process	0,03902
GO:0007219~Notch signaling pathway	0,00492	GO:0046365~monosaccharide catabolic process	0,03963
GO:0007366~periodic partitioning by pair rule gene	0,00509	GO:0042386~hemocyte differentiation	0,04036
GO:0001708~cell fate specification	0,00608	GO:0035162~embryonic hemopoiesis	0,04036
GO:0019731~antibacterial humoral response	0,00617	GO:0000272~polysaccharide catabolic process	0,04036
GO:0050877~neurological system process	0,00683	GO:0048568~embryonic organ development	0,04036
GO:0048190~wing disc dorsal/ventral pattern formation	0,00808	GO:0048749~compound eye development	0,04235
GO:0007362~terminal region determination	0,00834	GO:0046667~compound eye retinal cell programmed cell death	0,04283
GO:0050909~sensory perception of taste	0,00834	GO:0035161~imaginal disc lineage restriction	0,04283
GO:0009954~proximal/distal pattern formation	0,00834	GO:0009253~peptidoglycan catabolic process	0,04283
GO:0006006~glucose metabolic process	0,00866	GO:0006027~glycosaminoglycan catabolic process	0,04283
GO:0006928~cell motion	0,00924	GO:0000270~peptidoglycan metabolic process	0,04283
GO:0016337~cell-cell adhesion	0,00933	GO:0006206~pyrimidine base metabolic process	0,04283
GO:0002165~instar larval or pupal development	0,00964	GO:0035287~head segmentation	0,04283
GO:0009065~glutamine family amino acid catabolic process	0,01053	GO:0035223~leg disc pattern formation	0,04283
GO:0006584~catecholamine metabolic process	0,01053	GO:0007479~leg disc proximal/distal pattern formation	0,04283
GO:0009636~response to toxin	0,01053	GO:0014070~response to organic cyclic substance	0,04416
GO:0040034~regulation of development, heterochronic	0,01053	GO:0007419~ventral cord development	0,04416
GO:0018958~phenol metabolic process	0,01053	GO:0043279~response to alkaloid	0,04416
GO:0034311~diol metabolic process	0,01053	GO:0006096~glycolysis	0,04557
GO:0009712~catechol metabolic process	0,01053	GO:0045596~negative regulation of cell differentiation	0,0459
		GO:0042981~regulation of apoptosis	0,04595
		GO:0043068~positive regulation of programmed cell death	0,04734
		GO:0030031~cell projection assembly	0,04734
		GO:0010942~positive regulation of cell death	0,04734

Gene Ontology (GO) enrichment analysis was performed using active but NSL1 unbound genes against the background of all active genes. DAVID GO SLIM (57) terms were used for biological processes. P values were derived from a Fisher Exact test using the DAVID web-implementation.



Supplementary Figure 5: NSL1 stimulates transcription on a reporter gene and genome-wide.

(A) NSL1 binding correlates with stronger activation in female flies as compared to male flies after MOF recruitment to a reporter gene. We previously showed that recruitment of the MOF-GAL4 fusion protein to an UAS-element upstream of the lacZ reporter gene leads to stronger activation in adult female flies as compared to adult male flies (12). MBD-R2 recruitment as measured by ChIP at the lacZ gene follows this trend (12). Here, we tested NSL1 binding at the reporter gene and find a similar recruitment of NSL1 which is significantly ($p < 0.05$, 2-sided unpaired t-test) increased in adult female flies when compared to adult male flies. We conclude that the higher activation potential of MOF in female flies correlates with an increased NSL complex recruitment. Error bars represent SEM of at least three independent biological replicates.

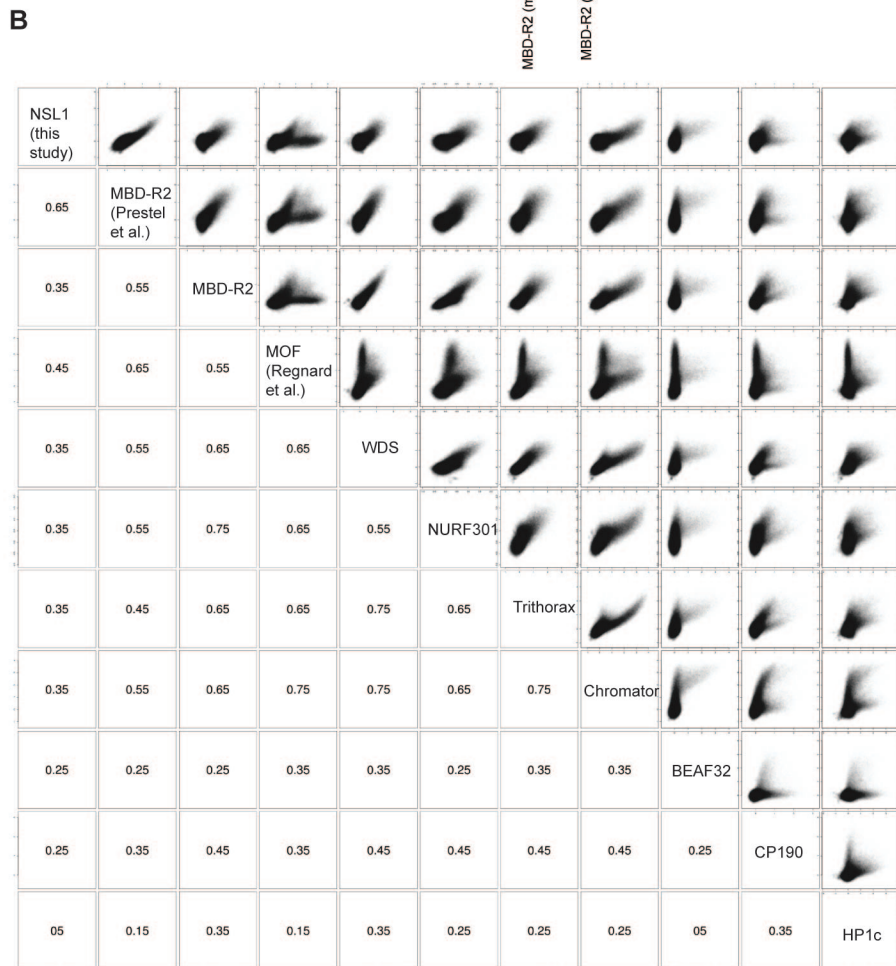
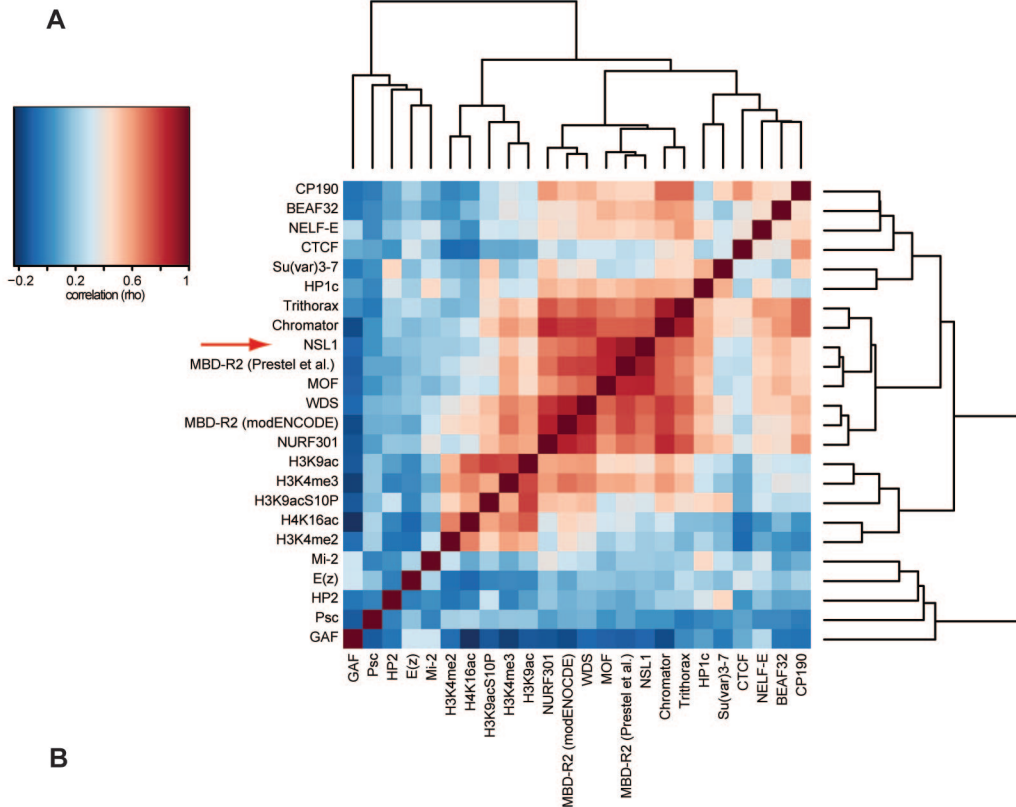
(B) RNAi mediated ablation of NSL1 leads to a diminished MOF-dependent activation of a luciferase reporter gene in male S2 cells and female Kc cells. Transiently co-transfection of MOF-GAL4 fusion gene, UAS-luciferase reporter and renilla reporter (used for normalization) lead to an activation of the luciferase reporter. NSL1 RNAi in male S2 and female Kc cells significantly diminished ($p < 0.05$, 2-sided unpaired t-test) the luciferase reporter expression when compared to a control RNAi treatment. Error bars represent SEM of at least three independent biological replicates.

(C) Transcriptome changes are similar in response to NSL1 RNAi and MBD-R2 RNAi. Scatter plot of transcriptome changes after NSL1 (this study) and MBD-R2 RNAi (12), respectively, relative to control RNAi. Rank-based Spearman correlation coefficient (ρ) is indicated in the corner.

Supplementary Figure 6: Gene Ontology enrichment analysis of NSL1 RNAi downregulated genes

Term	Pvalue
GO:0006259~DNA metabolic process	1,30E-05
GO:0006281~DNA repair	2,70E-04
GO:0034660~ncRNA metabolic process	9,00E-04
GO:0006412~translation	7,23E-02
GO:0033554~cellular response to stress	0,001
GO:0006399~tRNA metabolic process	0,001
GO:0006974~response to DNA damage stimulus	0,001
GO:0008033~tRNA processing	0,002
GO:0034470~ncRNA processing	0,005
GO:0006284~base-excision repair	0,006
GO:0006366~transcription from RNA polymerase II promoter	0,011
GO:0006367~transcription initiation from RNA polymerase II promoter	0,012
GO:0006351~transcription, DNA-dependent	0,014
GO:0006352~transcription initiation	0,015
GO:0032774~RNA biosynthetic process	0,016
GO:0006396~RNA processing	0,017
GO:0009451~RNA modification	0,023
GO:0006260~DNA replication	0,032

Downregulated genes (fold change < -2) were compared to all active genes (Affymetrix log₂-scale > 4). Gene Ontology enrichment analysis was performed using DAVID (57), testing for biological processes using the GO SLIM terms. P values were derived from a Fisher Exact test using the DAVID web-implementation. A similar result was obtained using the R/Bioconductor package GStats (data not shown).



Supplementary Figure 7: Co-localization of NSL complex members with Trithorax and NURF301.

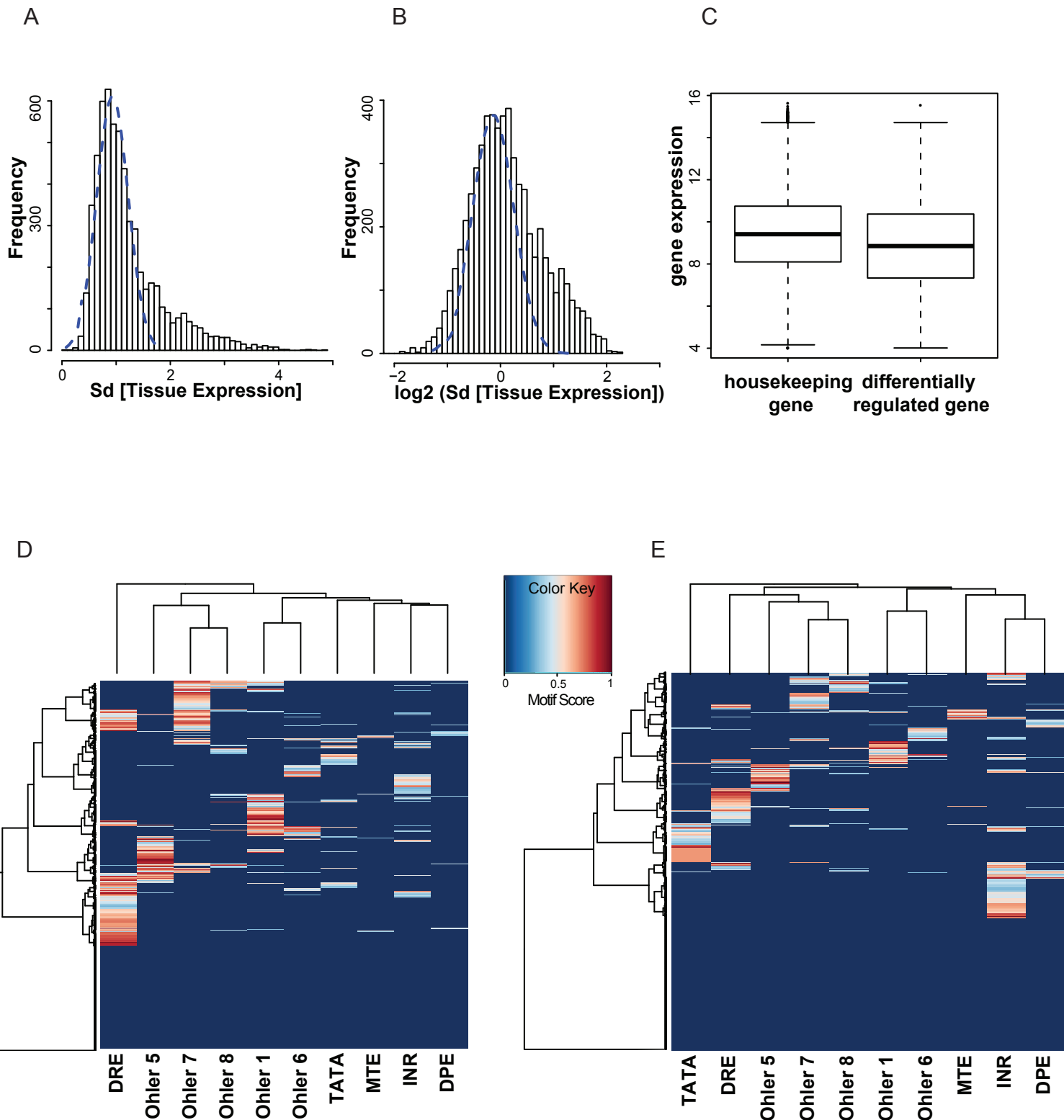
(A) Heatmap was generated as in Figure 3A but included in addition to the profiles shown in Figure 3A the ChIP-chip datasets of MBD-R2 (12), MBD-R2 (modENCODE) and MOF (modENCODE).

(B) Probe-wise scatter plot indicated co-localization of the NSL complex with Trithorax and NURF301. ChIP-chip datasets were used from the modENCODE consortium database (17) unless otherwise indicated. Probe-wise enrichments were calculated, smoothed and genomic positions were compared in 100 base pair intervals across samples. Note that all probes are shown including the inactive unbound probes and the X chromosomal active probes where NSL1 and MBD-R2 binding deviate from MOF binding towards the transcribed regions of genes (compare to (12)). Spearman correlation coefficient is indicated.

Supplementary Figure 8: Summary of factor and sequence occupancies across different conditions.

	% Binding to all			% Binding to all		Enrichment Active/ Inactive	Enrichment Housekeeping/ /DRG
	Genes	Active Genes	Inactive Genes	active Housekeeping	active DRG		
BEAF32	17,9	33,5	1,1	39,9	17	31,7	2,3
Ohler 1	7,6	12,7	2,1	15	6,7	5,9	2,2
HP2	2	3,2	0,7	3,8	1,7	4,8	2,2
DRE	14,4	21,2	7,2	24,6	12,3	2,9	2
Ohler 5	8,6	12,1	4,7	14	7,2	2,6	2
Chromator	44,7	80,7	6	92,7	49,4	13,4	1,9
Ohler 7	10,4	15,3	5,1	17,6	9,5	3	1,9
MBD-R2	38,9	71,7	3,7	82,1	44,7	19,4	1,8
WDS	39,6	73,5	3,1	83,9	46,7	23,5	1,8
Trithorax	40,6	74,4	4,4	84,7	47,5	16,7	1,8
CP190	29	47,5	9,2	54	30,5	5,2	1,8
NURF301	42,2	77,5	4,2	87,8	51,1	18,4	1,7
Ohler 6	6	7,6	4,2	8,5	5,1	1,8	1,7
HP1c	15,5	29,1	1	32,6	20	29,1	1,6
H3K4me3	47,5	87,1	5,1	94,9	67	17,2	1,4
Su(var)3-7	1,8	2,6	1,1	2,8	2	2,4	1,4
H3K9ac	46,9	84,5	6,6	92	65,3	12,8	1,4
Nelf-E	28,8	54,9	0,9	59,3	43,3	59,6	1,4
H4K16ac	50,0	80,0	17,5	78,9	58,6	9,7	1,3
Nelf-B	30,7	58,6	0,9	62,7	47,7	62,3	1,3
CTCF	4	4,4	3,5	4,5	4,2	1,2	1,1
Ohler 8	5,8	6,8	4,7	6,9	6,6	1,4	1
TATA	9,7	8,2	11,4	7,9	8,9	0,7	0,9
E(z)	1,3	0,8	1,9	0,7	1,1	0,4	0,6
INR	18,4	15,6	21,5	12,7	23,2	0,7	0,5
H3.3	20,8	32,3	8,4	26,2	48,3	3,8	0,5
MTE	2	1,9	2,2	1,5	3	0,9	0,5
Psc	1,6	1,1	2	0,8	1,9	0,6	0,4
RING	2,9	3,3	2,4	2,4	5,8	1,4	0,4
DPE	3,6	3,4	3,8	2,5	6	0,9	0,4
Mi-2	2,6	4,7	0,3	2,9	9,2	15,8	0,3
GAF	8,3	14,8	1,4	8,6	30,7	10,6	0,3
H3K27me3	35,8	4,1	69,8	1,9	9,8	0,1	0,2
Pc	2	0,5	3,6	0,2	1,2	0,1	0,1

Factor and sequence occupancy is given relative to all genes in the *D. melanogaster* genome (FlyBase dmel5.32 annotation) (column 2), all active genes (column 3, gene activity measured by modENCODE polymerase II dataset), all inactive genes (column 4), all active housekeeping genes (column 5) and all active “differentially regulated” genes (DRG, column 6). Column 7 indicates the ratio of active genes over inactive genes, and column 8 indicates the ratio of active housekeeping genes over active “differentially regulated” genes. The list was sorted according to the enrichment towards active housekeeping genes (column 8) and overrepresented and underrepresented factors are labeled in red and green, respectively. Only modENCODE ChIP-chip profiles were used.



Supplementary Figure 9: Characterization of core promoter motifs at active genes.

(A) Histogram depicting the distribution of standard deviations of active genes across 40 *Drosophila* tissues (21). Normal distribution is overlaid with a blue dashed line. (B) Same as (A) but \log_2 transformed standard deviations. (C) Comparison of gene expression values in housekeeping genes (left box) and differentially regulated genes (right box). Gene expression values were taken from the \log_2 transformed Affymetrix control RNAi transcriptome dataset. (D) Core promoter motif distribution of active housekeeping genes. Heatmap presentation of motif score (columns) along active housekeeping genes (rows). The dendrogram to the right was generated by hierarchical clustering of gene promoters according to their motif score using the 'complete' method implemented in the R package 'hclust'. The dendrogram above the heatmap was generated by hierarchical clustering ('ward method') of the Spearman correlated motif scores. Note that 1269 of the 4523 active housekeeping genes do not show any significant signature of at least one of the investigated core promoter motifs. See Material and Methods for detailed information on the definition of the 'motif score'. (E) Same as (D) but for active differentially regulated genes. Note that 480 of the 1367 active differentially regulated genes do not show any significant signature of at least one of the investigated core promoter motifs.

Supplementary Figure 10: NSL1 peaks are centered over core promoter motifs characteristic for housekeeping genes

motif	total number sequences enriched for motif	sequence fraction enriched for motif
DRE	1106	19.9
Ohler 1	703	12.7
Ohler 5	556	10.0
Ohler 7	510	9.2
Ohler 6	251	4.5
Ohler 8	109	2.0
INR	84	1.5
TATA	38	0.7
MTE	33	0.6
DPE	12	0.2

De-novo sequence search (Hartmann and Soeding, in preparation) of the sequence surrounding +/- 50 base pairs centered on the NSL1 ChIP-Seq (11) peak summit. Column 2 indicates the number of identified sequences containing the respective motif. Column 3 displays the fractions of each identified motif relative to all identified motifs.

