

# Distribution of Bubble Lengths in DNA

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The distribution of bubble lengths in double-stranded DNA is presented for segments of varying guanine-cytosine (GC) content, obtained with Monte Carlo simulations using the Peyrard-Bishop-Dauxois model at 310 K. An analytical description of the obtained distribution in the whole regime investigated, i.e., up to bubble widths of the order of tens of nanometers, is available. We find that the decay lengths and characteristic exponents of this distribution show two distinct regimes as a function of GC content. The observed distribution is attributed to the anharmonic interactions within base-pairs. The results are discussed in the framework of the Poland-Scheraga and the Peyrard-Bishop (with linear instead of nonlinear stacking interaction) models.

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Fluctuating thermal openings of the double helix of DNA (bubbles) are known to exist in a regime extending well below the denaturation transition, which includes physiological temperatures. These openings could be exploited by regulatory proteins or functional enzymes to perform their work [1]. It has been recently suggested that there is an increased probability for large bubbles to appear at functionally relevant sites of gene promoter DNA segments [2, 3]. More accurate numerical calculations [4, 5] have unadvertedly reinforced this idea [6], by finding an increased opening probability at functional binding sites of the promoter region upstream of a gene initiation site. Such a picture is in analogy to the functional role of fluctuations in proteins [7]. This has led to an increased interest for equilibrium and dynamical studies of thermally induced bubbles in DNA.

In a different context, recent experiments on cyclization of short DNA fragments [8, 9] have drawn attention toward the possibility that regions of very low rigidity may appear in double stranded DNA [10]. It has been proposed [11, 12] that these *bending kinks* may be caused by the formation of denaturation bubbles, letting the higher flexibility of single stranded DNA explain the sharp bending observed in experiments [8]. In this context, the question of how frequently denaturation bubbles arise is one of the keys for the understanding of this phenomenon.

Here we present the distribution of bubble lengths, for lengths ranging from 0.34 nm (single base-pair openings) up to a few tens of nanometers (corresponding to openings of size several tens of base-pairs). The bubble length distributions have been obtained with Monte Carlo simulations, using the Peyrard-Bishop-Dauxois (PBD) model [13] for describing the energy of base-pair openings. We discuss equilibrium distributions at  $T = 310$  K, averaged over DNA segments with total size of one thousand base-pairs. The bubble length distributions depend on the guanine-cytosine (GC) content, i.e., the fraction of GC base-pairs in the sequence. The variation of the distribu-

tion as a function of the GC content is investigated. Note that the size of a bubble is characterized by its length (width) and its amplitude. Here we fix a threshold for the amplitude and when a bubble of length  $l$  base-pairs (or  $L = 0.34 \times l$  nm) is mentioned, it means that all the  $l$  successive base-pairs have openings larger than the fixed amplitude threshold  $y_{thres}$ , while the limiting base-pairs (the first neighbors to the left and to the right of the stretch of the  $l$  successive base-pairs) have smaller openings than that. Periodic boundary conditions are used, so our study considers only bubbles in the middle of a sequence, disregarding end effects. Fraying, i.e. the opening of the DNA molecule starting at one of its ends, is a very interesting problem that will be addressed elsewhere.

In this Letter we show that the bubble length distributions, obtained within the PBD model, follow a non-exponential law and, further, an analytical description (see Eq. (2) below) is available describing the observed, slower than exponential, decay with the length. This behavior has been quantified for various GC contents and remains qualitatively the same independently on the value of the fixed amplitude. The fact that the same distribution law is also predicted by the completely unrelated Poland-Scheraga [14] model of DNA, strongly suggests that our findings are beyond theoretical speculation and might be a proper description of actual DNA physics.

In recent investigations, numerically exact equilibrium properties [4, 5] and Langevin dynamics up to nanosecond time-scales averaged over several hundred realizations [2, 3, 15] have been achieved taking advantage of the efficiency of the simple PBD model for describing base-pair openings in double-stranded DNA. The PBD model coarse-grains the relatively rigid internal structure of the nucleotides and considers their anharmonic stretching interactions at the single base-pair level [13, 16, 17]. Its accuracy has been demonstrated by several comparisons with different experiments [2, 18, 19].

The potential energy of the PBD model is given by:

$$V = \sum_n \left[ D_n (e^{-a_n y_n} - 1)^2 + \frac{K}{2} (1 + \rho e^{-b(y_n + y_{n-1})}) (y_n - y_{n-1})^2 \right]. \quad (1)$$

The sum is over all the base-pairs of the molecule and  $y_n$  denotes the relative displacement from equilibrium at the  $n^{\text{th}}$  base pair. The first term is an on-site Morse potential, representing the hydrogen bonds between bases in the same pair as well as other effective interactions between complementary nucleotides. The second term is an anharmonic coupling between adjacent base pairs that models the stacking interaction. The heterogeneity of the genetic sequence is taken into account giving different values to the parameters of the Morse potential for adenine-thymine (AT) or guanine-cytosine (GC) base pairs. The values of the parameters we have used [20] were fitted in Ref. [18] to reproduce thermodynamic properties of DNA. The same parameters have been subsequently used, without additional fitting procedures, to successfully describe experimental observations [2, 19].

To study the statistics of bubbles we have performed Monte Carlo simulations of the PBD model using the same procedure introduced in Ref. [19]. The Metropolis algorithm was used to produce equilibrium configurations of the molecule at  $T = 310$  K. Results were averaged, after proper thermalization, over several realizations (typically 25, each one consisting of  $8 \cdot 10^6$  Monte Carlo steps, which makes a total of  $2 \cdot 10^8$  steps) with different initial conditions. As we used quite large DNA sequences (containing 1000 base-pairs) and the temperature studied is well below the melting temperature, the probability that a complete melting occurred during the time of a simulation was so low that no melting events (that, for long enough simulations, would eventually take place for this model at any temperature [16, 21]) were observed. We show results for bubbles of amplitude equal or greater than  $y_{\text{thres}} = 1.5$  Å. We have also studied bubbles of amplitude  $y_{\text{thres}} = 0.5$  Å, 3 Å and 5 Å, finding the same qualitative results presented here. We have checked that our results are independent of the length of the sequences studied, provided they are longer than the longest observed bubble and there is no complete melting.

Figure 1 shows the bubble length distribution per base-pair,  $P(l)$ , obtained from our Monte Carlo simulations. This histogram is defined as  $P(l) = \lim_{L \rightarrow \infty} \langle N(l) \rangle / L$ , where  $N(l)$  is the averaged (during the Monte Carlo simulation) number of bubbles of length  $l$  base pairs on a sequence of total size  $L$ , and the average is over different realizations of an uncorrelated random sequence with a given GC content. The limit  $L \rightarrow \infty$  in the definition of  $P(l)$  has the meaning that the total size of the DNA segment should be sufficiently larger than the considered bubble lengths. Under this condition, the distribution  $P(l)$  should be interpreted as follows: for a given DNA

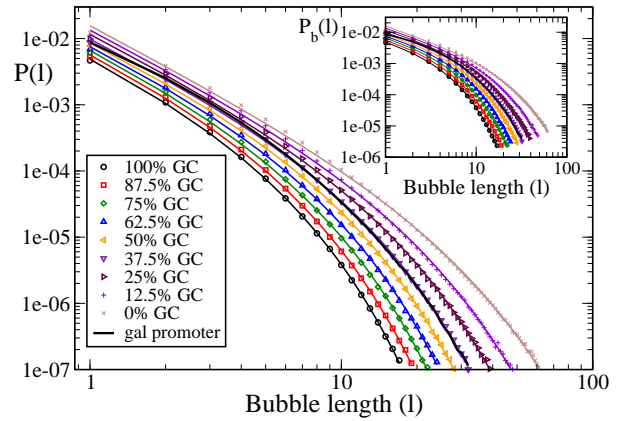


FIG. 1: Distribution per base-pair of bubble sizes  $l$  (in number of base-pairs,  $l \geq 1$ ),  $P(l)$ , for random sequences with different GC contents (points, as indicated in the label). Also shown is the result for a segment from *E. coli*'s *gal* promoter with a 37.45% GC percentage (thick line). Thin lines are fits with the analytical distribution, Eq.(2).  $T = 310$  K,  $y_{\text{thres}} = 1.5$  Å. Inset: Probability  $P_b(l) = lP(l)$  (for  $l \geq 1$ ) that an individual base-pair forms part of a bubble of size  $l$  (see text).

sequence of total length  $L$  base-pairs, the quantity  $P(l) \cdot L$  gives the average number of occurrences of a bubble of length  $l$  base-pairs, in thermal equilibrium.

From  $P(l)$  we obtain the probability that an individual base-pair forms part of a bubble of length  $l$  as  $P_b(l) = lP(l)$  for  $l \geq 1$ , while the probability of not belonging to any bubble (i.e., that the base-pair has an opening smaller than  $y_{\text{thres}}$ ) is  $P_b(0) = P(0)$ . From the definition of  $P(l)$  follows that  $P(0) = 1 - \sum_{l \geq 1} lP(l)$ , thus ensuring correct normalization  $\sum_l P_b(l) = 1$  of the probability  $P_b$ .

As a check of our simulations, we have also computed for pure GC and AT sequences the probability of i) a single base-pair to have an opening larger than the considered fixed amplitude and ii) two neighboring base-pairs to simultaneously have openings larger than the fixed amplitude, using the transfer integral operator method [22–24]. The probabilities calculated in these two cases, using a different but numerically exact method, are in a very good agreement (up to numerical accuracy) with those obtained through the Monte Carlo simulation from the distributions  $P(l)$ , as  $\sum_{l \geq 1} lP(l) = 1 - P(0)$  and  $\sum_{l \geq 2} (l-1)P(l)$ , respectively.

We find that the distributions per base-pair can be fitted in the whole range of bubble sizes studied [25], with a single function of the form:

$$P(l) = W \frac{e^{-l/\xi}}{l^c}, \quad \text{for } l \geq 1. \quad (2)$$

A fit with a stretched exponential function having a stretching exponent smaller than 1 is also accurate [26], but we have preferred Eq. (2) because this functional form can be derived from existing theories both for a

simplified version of the used model [27] (the Peyrard-Bishop model, with linear stacking interactions [28]) and for the independent Poland-Scheraga model [14].

In order to investigate which interactions of those present in the model are responsible for the observed behavior of the distribution, we have performed similar calculations varying different terms of the potential of Eq.(1). When  $\rho = 0$ , i.e. the stacking interaction is linearized, the distribution remains nonexponential, although it is described by smaller  $c$ , in complete agreement with previous studies [27]. However, when the on-site Morse potential is linearized (substituting the Morse potential by its harmonic approximation), then the obtained distribution is exponential. This happens independently of setting  $\rho = 0$  or  $\rho = 2$ . Therefore, we infer that the nonlinear interactions between the bases within base-pairs result in the observed bubble length distribution. These anharmonic on-site interactions have been found to qualitatively affect other properties of the model as well [29]. This kind of interaction appears also in modified models where the stacking interaction has a qualitatively different shape [30].

In Figure 1 is also shown, for sake of comparison, a result for a natural sequence; a segment from *Escherichia coli's gal* promoter [31], extending from the position  $-160$  (upstream) up to the site  $+91$  (downstream) around the transcription start site of *galE* gene. This sequence has a GC fraction of 37.45%, and, as can be seen from Figure 1, its results are indistinguishable from those of a random sequence with almost the same GC content (37.5%). This is expected, as it is in agreement with the known fact [32] that, despite statistical correlations in the sequence and local properties, equilibrium averaged physical properties of large DNA molecules are basically similar to those of random sequences.

DNA molecules with higher GC content exhibit a faster decay (see Figure 1), as the stronger bound GC pairs make large bubbles less likely. The change of the distribution with the GC content is quantified in Figure 2, where the dependence of the parameters of Eq. (2) on the GC fraction is shown. All the parameters decrease monotonically with the GC percentage. The decay of  $W$  signifies that the higher the GC content the more difficult it is to excite large openings in the double strand, therefore in AT rich sequences bubbles have a higher statistical weight. The decay length  $\xi$  is smaller for GC-rich sequences, in which the distribution decays faster. The variation of all these parameters can be fitted by a bilinear or a biexponential function (see Figure 2), revealing two distinct regimes on the GC-fraction dependency: above and below a GC content of about 40%. Previous studies [5] have shown that the nucleation of bubbles depends strongly on the sequence: the weaker AT base-pairs have to go over the potential barrier imposed by their GC neighbors in order to break the bonds. Our findings suggest that for GC concentrations over 40%,

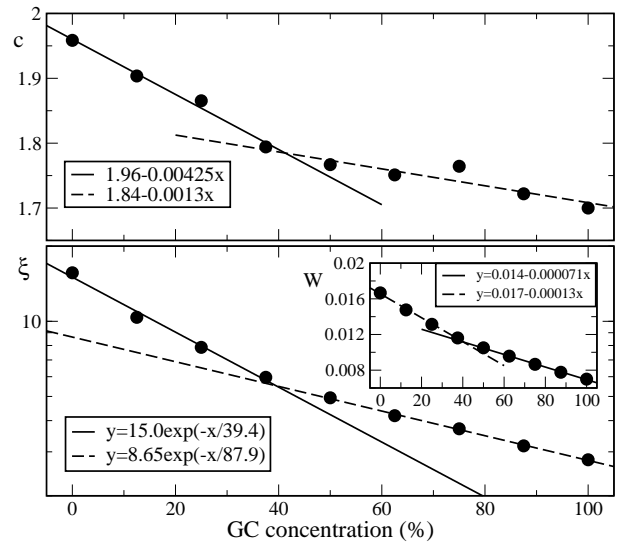


FIG. 2: Dependence of the exponent  $c$  (top), decay length  $\xi$  (bottom), and preexponential coefficient  $W$  (bottom, inset) of the distribution, Eq. (2), on the GC content of the sequence (circles). Lines show fits of two distinct regimes with linear (exponential for  $\xi$ ) functions.  $T = 310$  K,  $y_{thres} = 1.5$  Å.

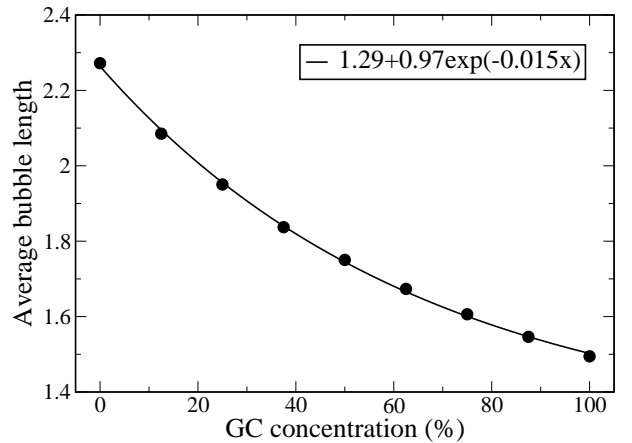


FIG. 3: Dependence of the average bubble length  $L_B$ , Eq. (3), on the GC content of the sequence (points). Continuous line shows a fitting with an exponential decay.  $T = 310$  K,  $y_{thres} = 1.5$  Å.

the formation of bubbles is a GC-dominated process, as AT base pairs are not in average free to melt without GC opposition. But below 40% GC content, large AT regions are possible that form bubbles freely, hence dominating the bubble formation process.

Figure 3 presents the average bubble length  $L_B$ , which is given by the total number of base-pairs in bubble states divided by the total number of bubbles:

$$L_B = \lim_{L \rightarrow \infty} \frac{\sum_l l \langle N(l) \rangle}{\sum_{l \geq 1} \langle N(l) \rangle} = \frac{\sum_l l P(l)}{\sum_{l \geq 1} P(l)}. \quad (3)$$

$L_B$  depends strongly on the GC content, showing an ex-

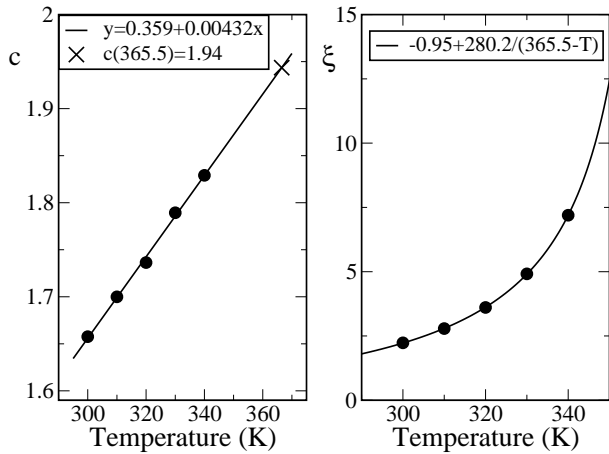


FIG. 4: Temperature dependence of the parameters of distribution, Eq. (2), for pure GC sequences. Left: exponent  $c$ , the line is a linear fit. Right: decay length  $\xi$ , the line is a fit with a divergent behavior as  $(T_c - T)^{-1}$ , where the melting temperature is predicted to be  $T_c = 365.5$  K, in agreement with transfer integral calculations.

ponential decay. This stresses the importance the sequence has on the typical size of denaturation bubbles.

A recent work reports exponential decay of the bubble length probability [15]. Although this seems to contradict our present results, this is not the case because the study [15] considers only a restricted regime of lengths, extending from  $l = 3$  up to  $l = 12 - 14$ . Therefore, a restricted portion of the distribution may appear as a seemingly straight line in a semilogarithmic plot, indicating exponential decay. However, if one looks for either smaller ( $l = 1, 2$ ) or larger lengths, below or above this regime, the exponential law fails. On the contrary the formula (2), or a stretched exponential law [26], describes accurately the whole regime from  $l = 1$  up to several tens of base-pairs. This means that large bubbles are more probable than what it would have been implied by exponential decay. Earlier studies of a simplified version of the PBD model (using a linearized stacking interaction) have also shown a slower than exponential decay of the distribution function [27, 33].

Importantly, the Poland-Scheraga (PS) model of DNA melting predicts a bubble length distribution described by Eq.(2) [34–36]. Hence our results establish a bridge between these two different theoretical approaches, which could help to future better understanding of the principles underlying the models, and henceforth a more effective modeling approach to DNA. In the PS model, the exponent  $c$  has a very important physical meaning, as its value at the critical temperature (where  $\xi$  diverges and the distribution is given by a pure power-law) indicates the order of the melting transition: for  $c < 1$  there is no phase transition, the transition is smooth for  $1 < c < 3/2$ , second order for  $3/2 \leq c < 2$  and first order for  $c > 2$ . It is interesting to characterize this ex-

ponent also for the PBD model; work in this direction is in progress and a detailed temperature dependence of bubble length distributions will be presented elsewhere [37]. Independently of the model, for  $c > 2$  the average bubble length  $L_B \propto \sum l P(l)$  remains finite as  $T \rightarrow T_c^-$ , and it diverges for  $c < 2$ , thus presenting a discontinuous or a continuous transition, respectively.

As an advance of our work in progress, in Figure 4 we depict a preliminary result for the exponent  $c$  and the decay length  $\xi$  obtained for pure GC sequences at different temperatures. As happens in the PS model [34], also here the exponent increases linearly with temperature below the critical temperature. The PS model also predicts that  $\xi$  diverges as  $(T_c - T)^{-1}$  [34]. Using this to fit the  $\xi$  data, we obtain a value  $T_c(\text{GC}) = 365.5$  K, in agreement with transfer integral calculations we have performed. Hence the predicted value of the exponent  $c$  at  $T_c$  is  $c = 1.94$  (compatible with the value  $c = 1.95$  used in biological studies [38]): we would have a very sharp transition, but still not first order, if the PS scheme applies. But we have to be very careful: for AT at  $T = 310$  K, Figure 2 shows that  $c = 1.96$ . A transfer integral calculation yields  $T_c(\text{AT}) = 325.5$  K, so the exponent at  $T_c$  is expected to be higher than 2. Work in this direction is necessary, but the possibility to speculate that not only the transition temperature but also the nature of the phase transition itself depends on the sequence, is too tempting to let it go. The role that the parameters of the model play in this has also to be carefully investigated. Interestingly, previous work using the Peyrard-Bishop model without the anharmonic stacking interaction ( $\rho = 0$ ) yields a value of  $c$  much lower than 2 [27]. Thus, regardless whether the transition in the full PBD model is first order or not, the analysis of bubble distributions shows once again [13, 16] that the anharmonic stacking interaction is responsible for the sharp ( $c \approx 2$ ) transition in the model.

We are not aware of any experimental technique able to track individual bubbles in long sequences in order to study their size distributions. However, with study of the probabilities of complete melting of molecules of different sizes, the power law behavior we have described may appear. The probability of complete melting of a given sequence has already been experimentally measured [39] using a novel technique based on hairpin quenching. This same technique, applied at constant temperature to sequences of varying length but similar composition, should clarify how the formation of bubbles depends on their size below the melting temperature. However, it should be noted that this experiment measuring complete melting is not exactly equivalent to our study of bubble formation: sequence, finite-size and boundary effects may play an important role.

In summary, using the Peyrard-Bishop-Dauxois model we have shown that at physiological temperatures the formation of thermally induced bubbles of different sizes follows a nonexponential distribution with long tails, due to

nonlinear interactions within base-pairs. The occurrence of these openings may have an important effect, which should be taken into account in biological processes involving the opening of double stranded DNA. Moreover, structural properties of the DNA molecule may also depend on the frequency of occurrence of these bubbles.

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