

Automated Glycan Assembly of Complex Oligosaccharides Related to Blood Group Determinants

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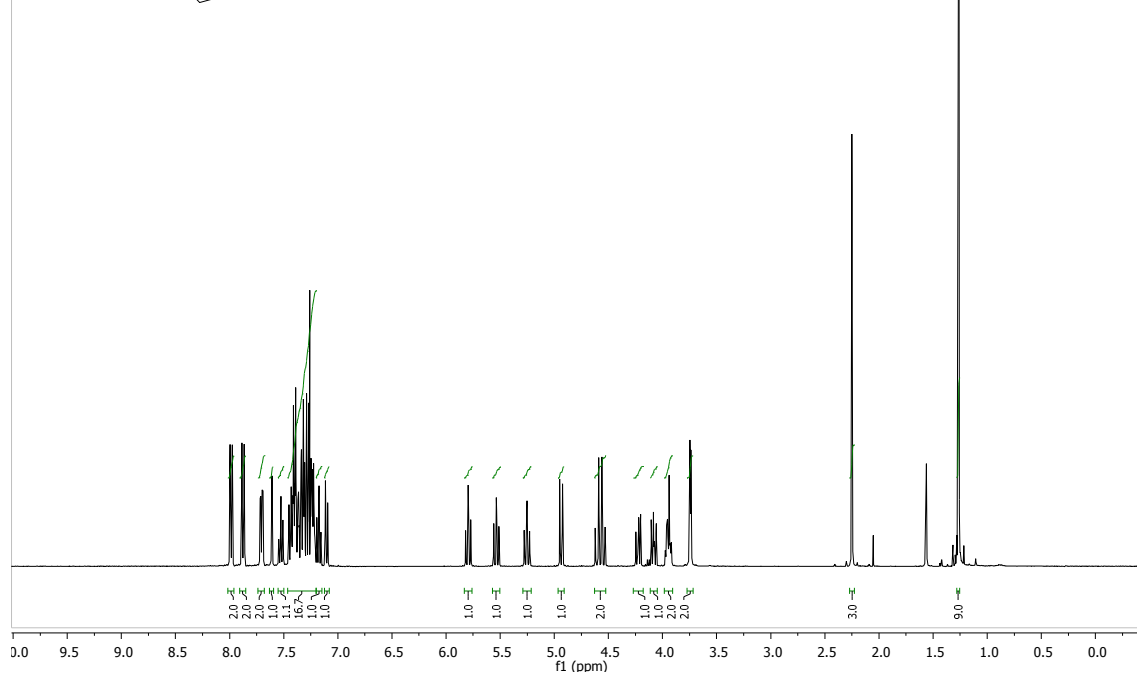
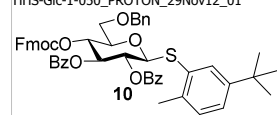
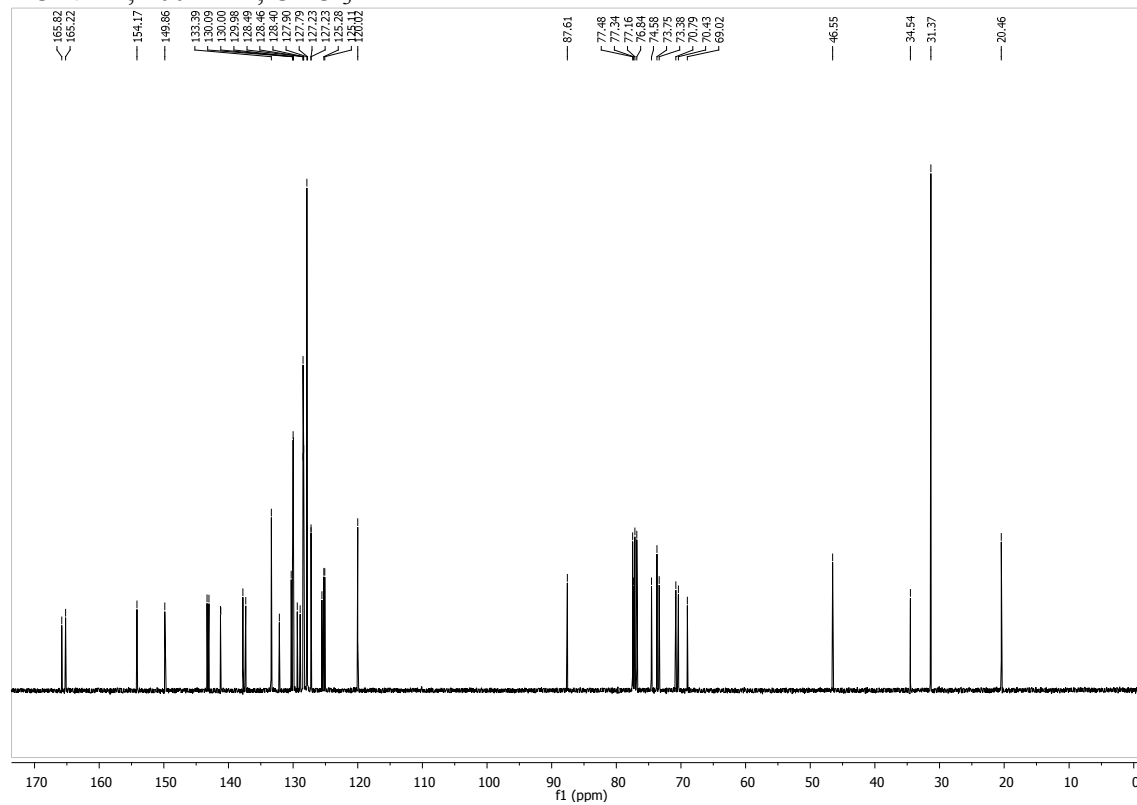
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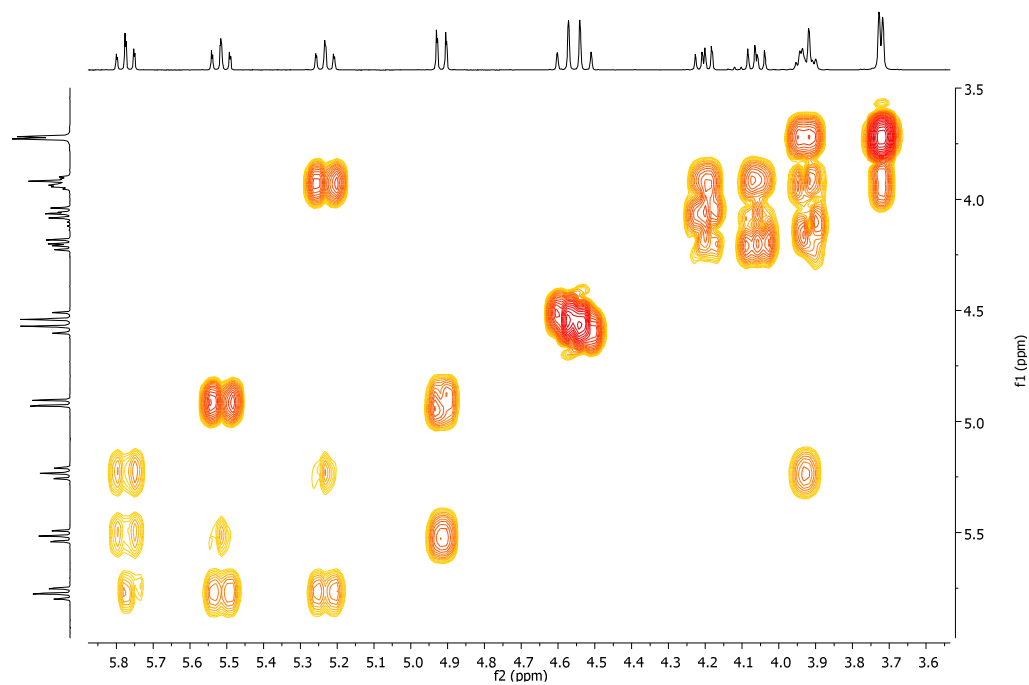
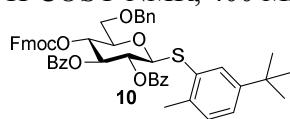
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¹H NMR, 400 MHz, CDCl₃

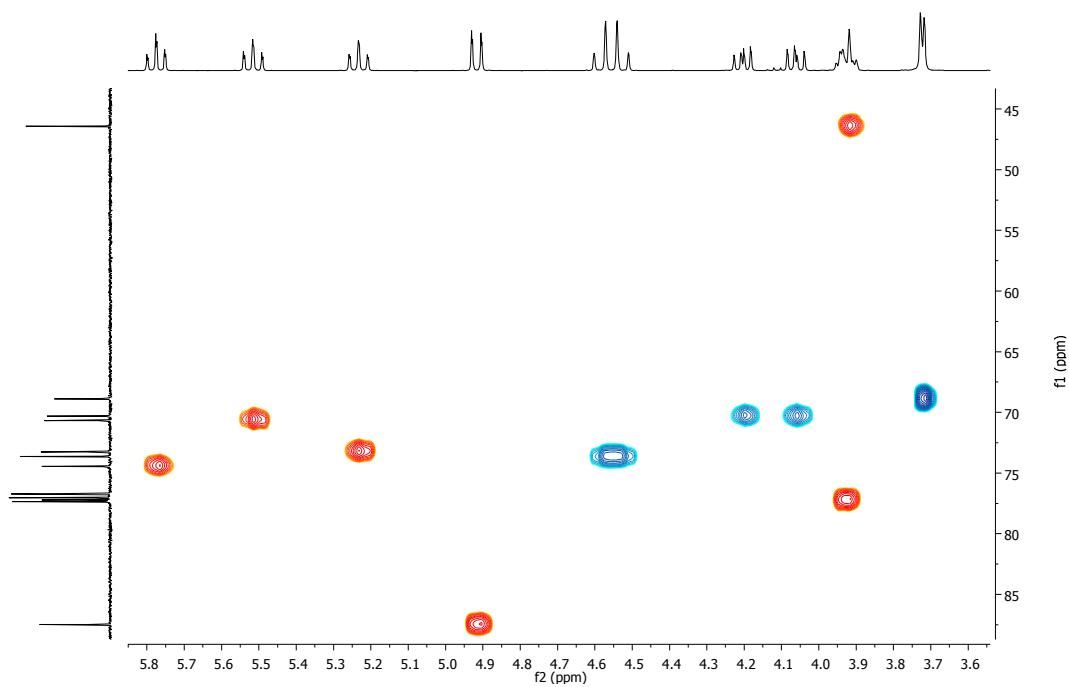
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¹³C NMR, 100 MHz, CDCl₃

^1H -COSY NMR, 400 MHz, CDCl_3

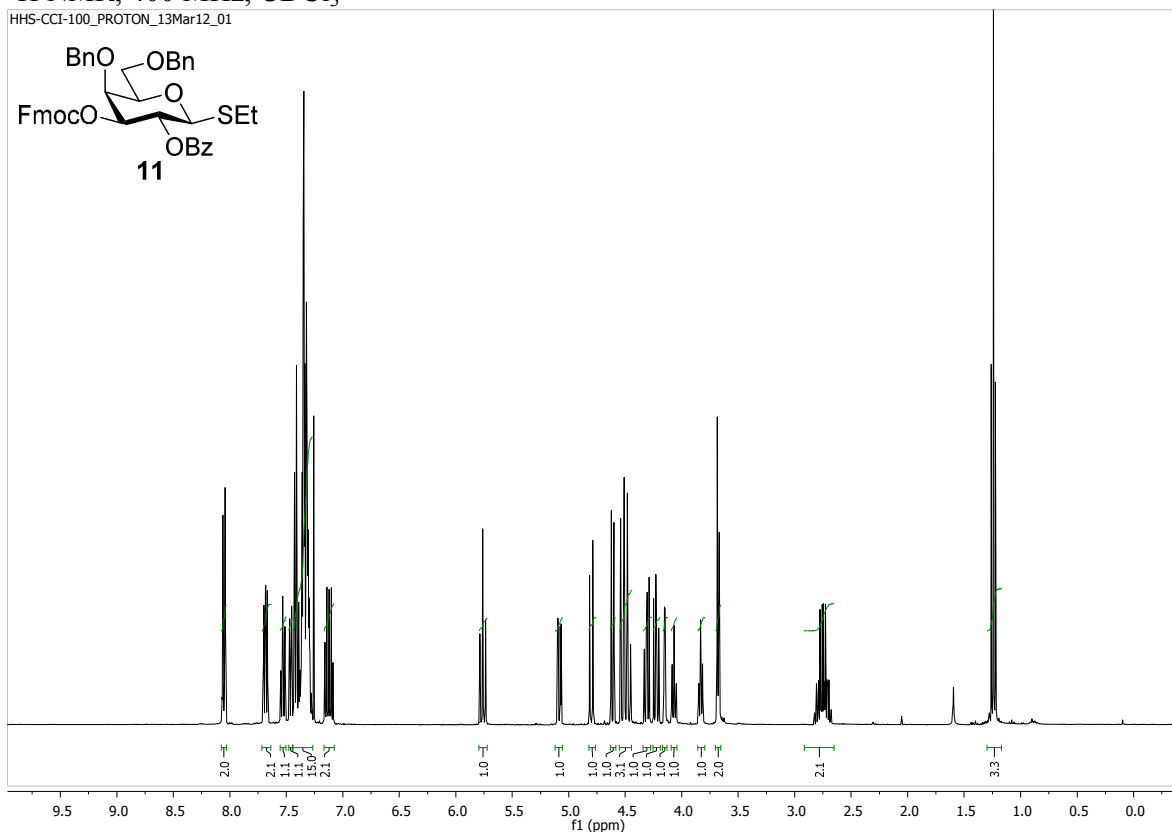
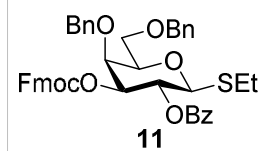
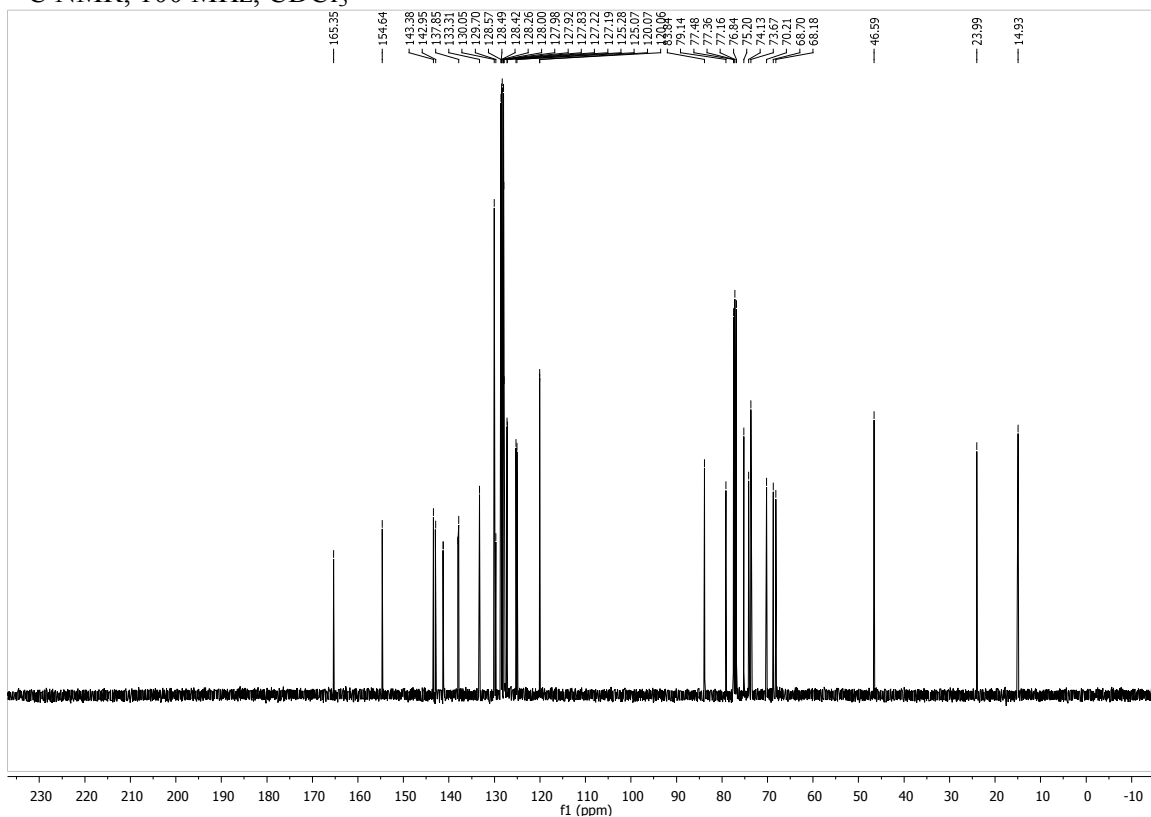


^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3

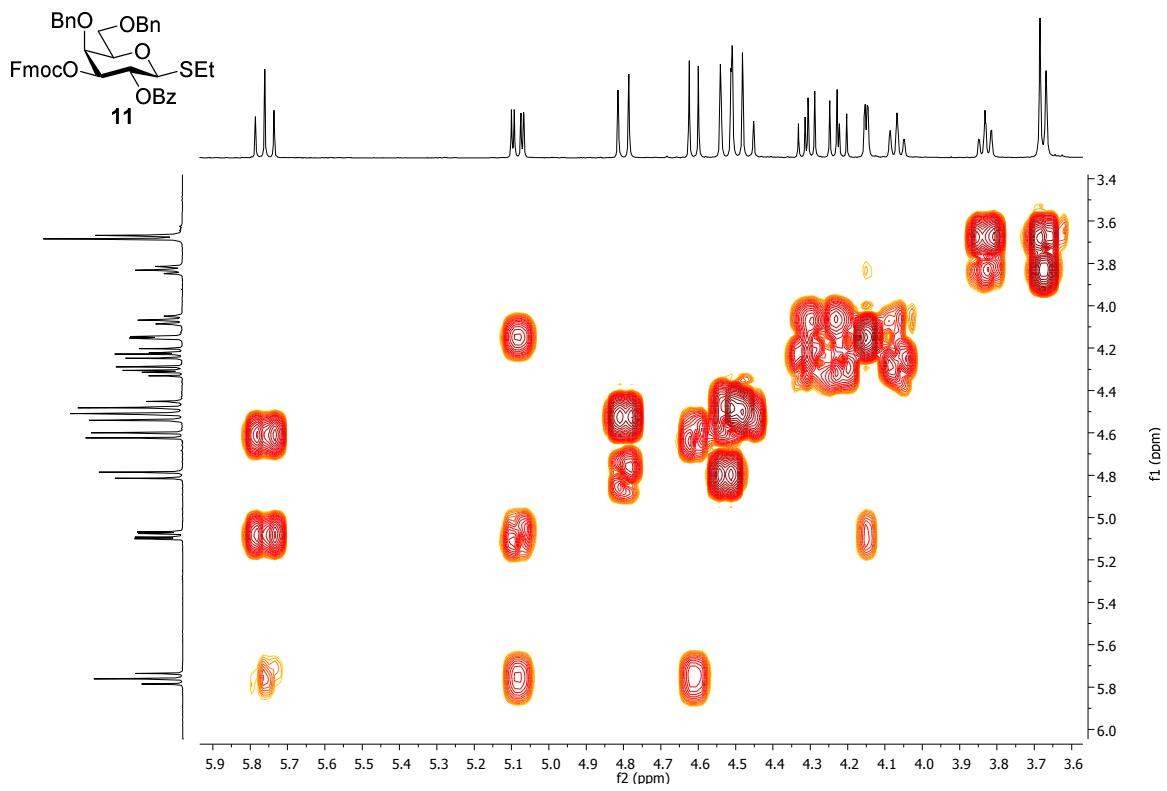


¹H NMR, 400 MHz, CDCl₃

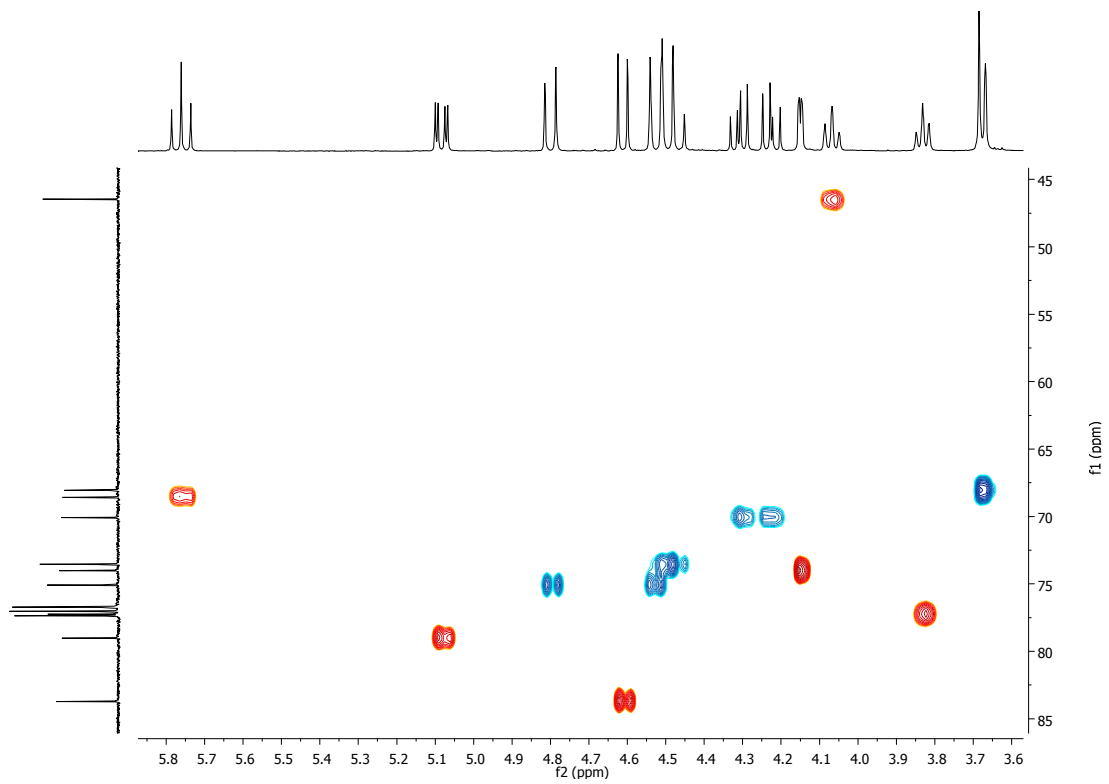
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¹³C NMR, 100 MHz, CDCl₃

^1H -COSY NMR, 400 MHz, CDCl_3

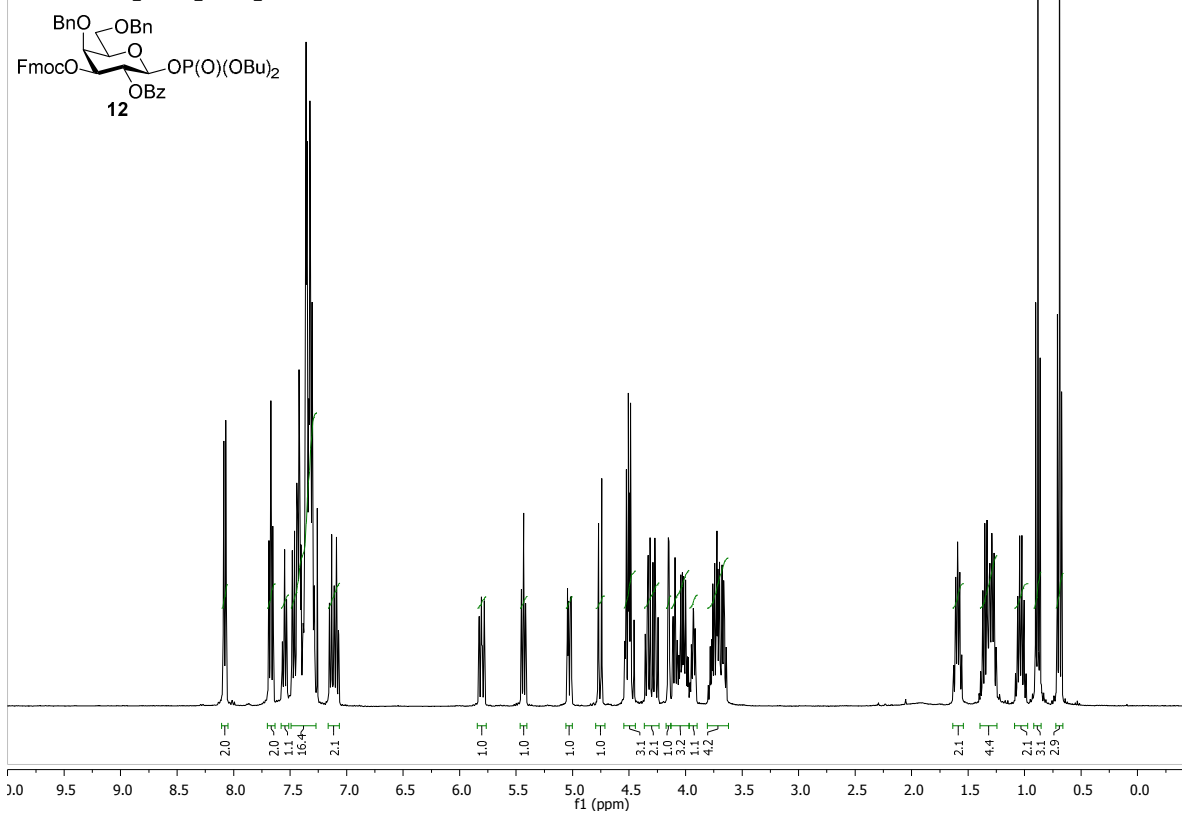
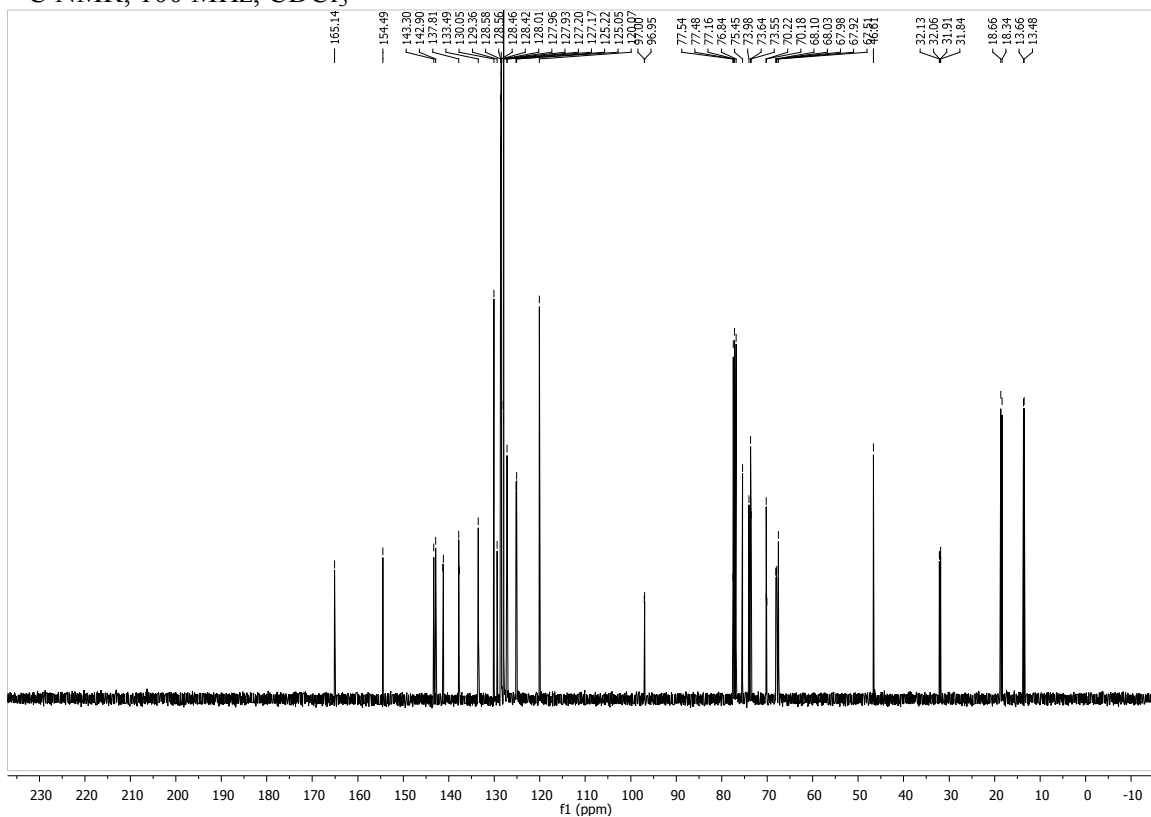


^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3

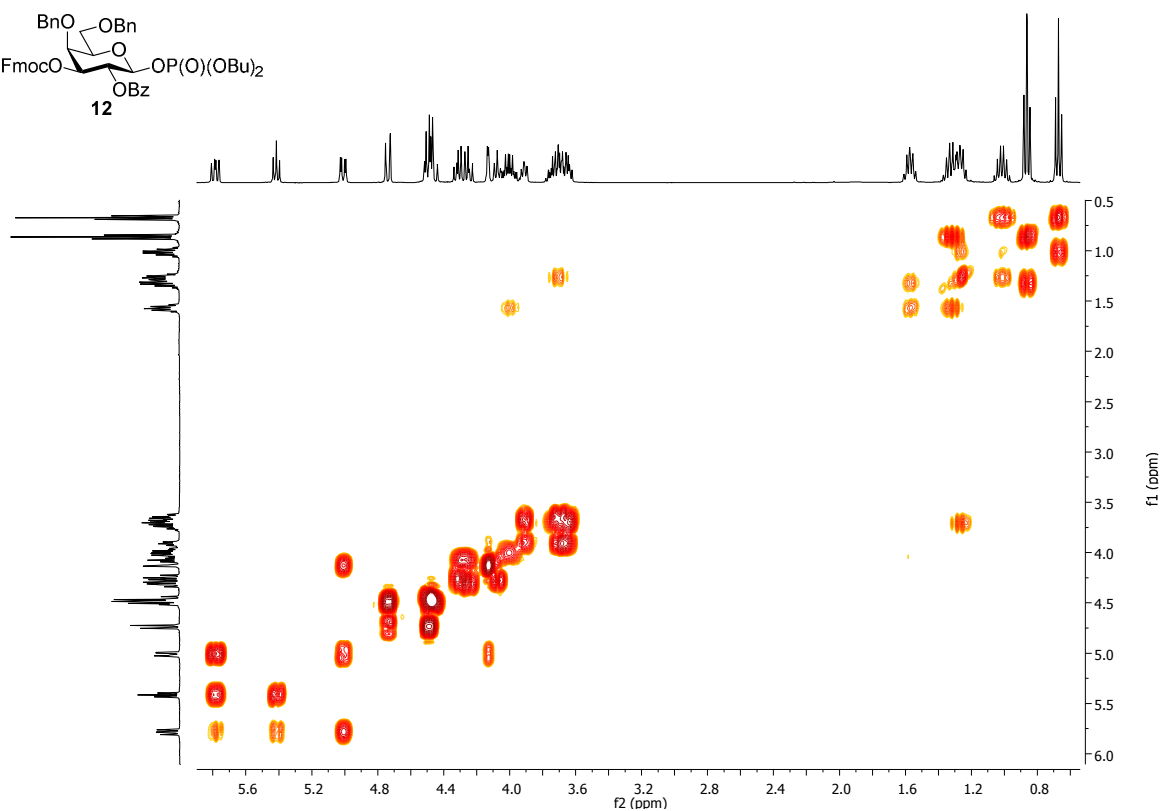
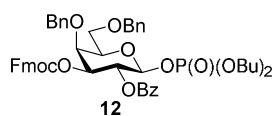


¹H NMR, 400 MHz, CDCl₃

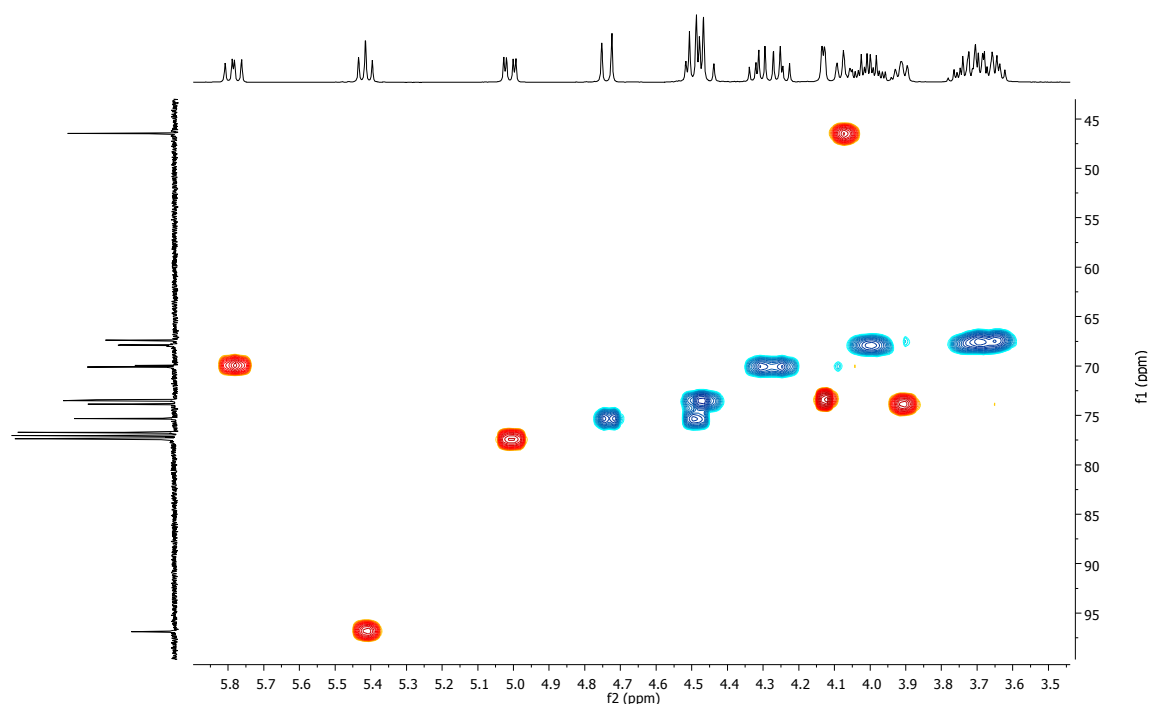
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¹³C NMR, 100 MHz, CDCl₃

^1H -COSY NMR, 400 MHz, CDCl_3

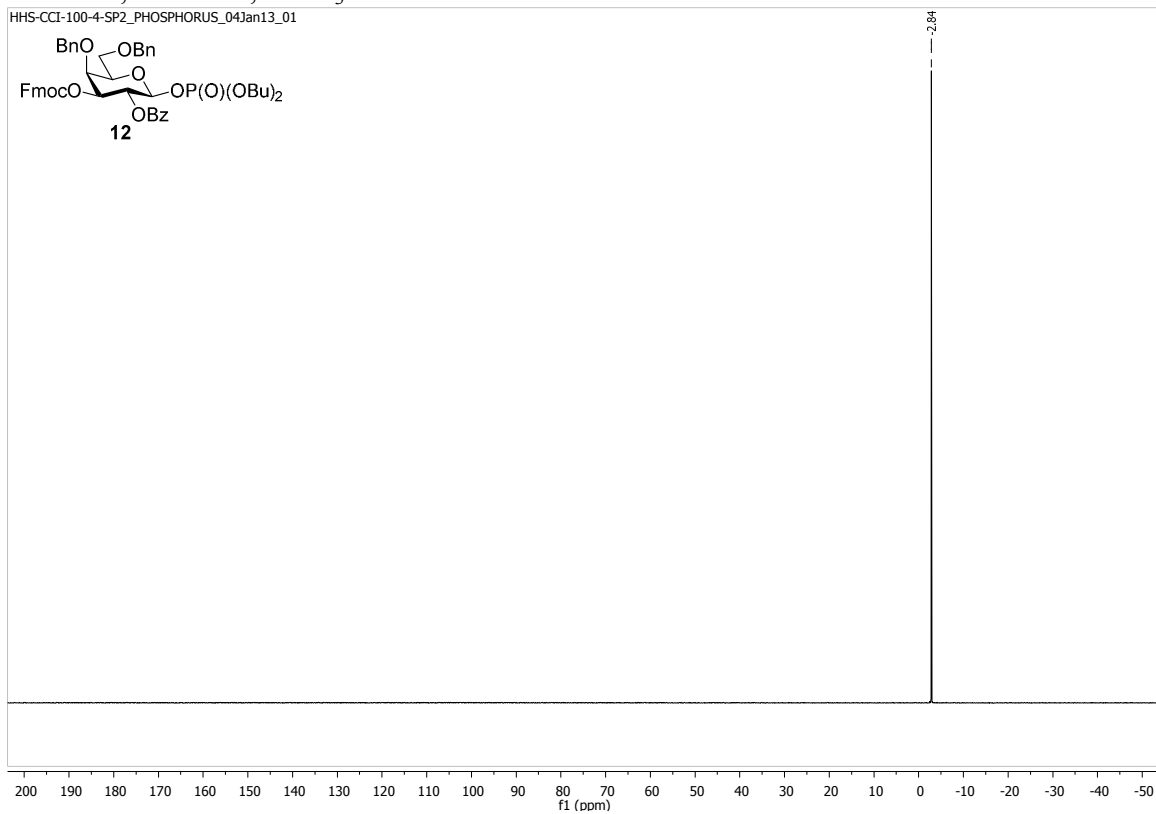
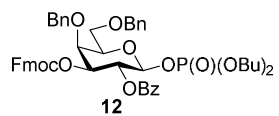


^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3



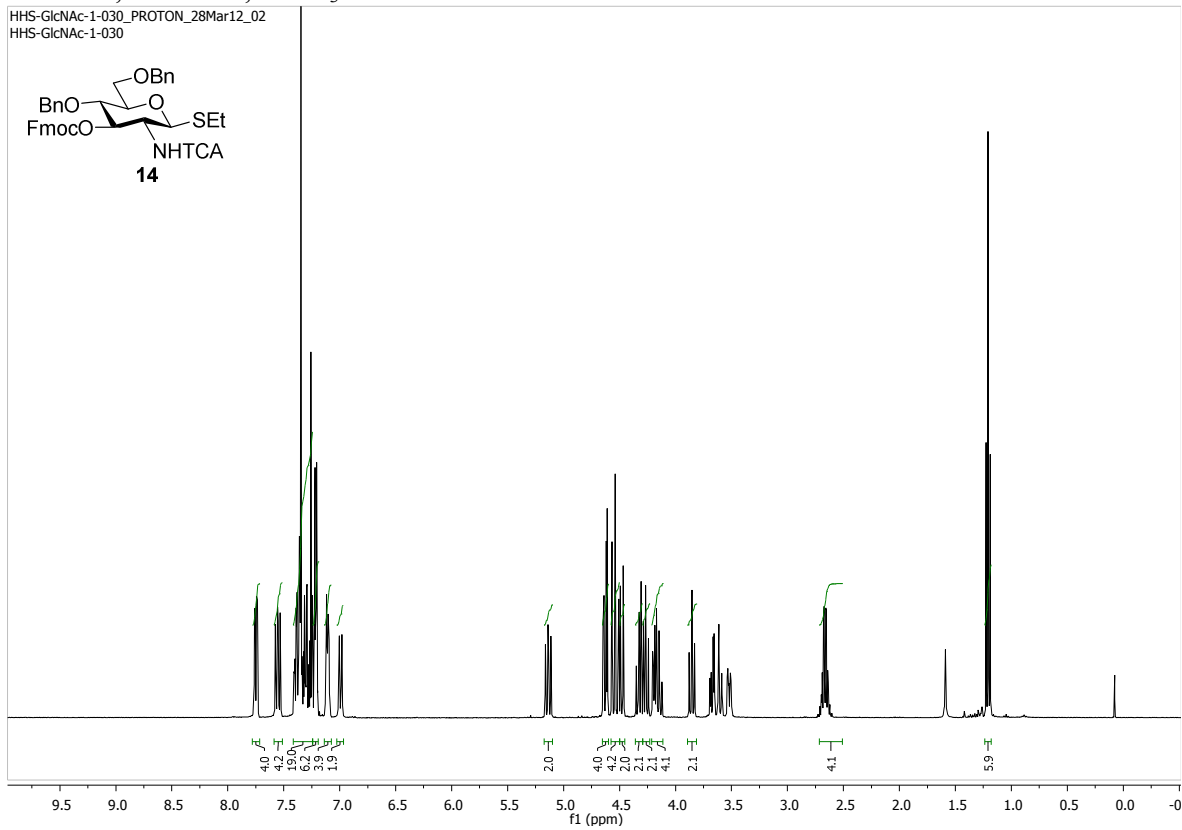
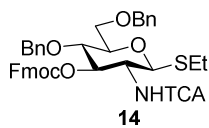
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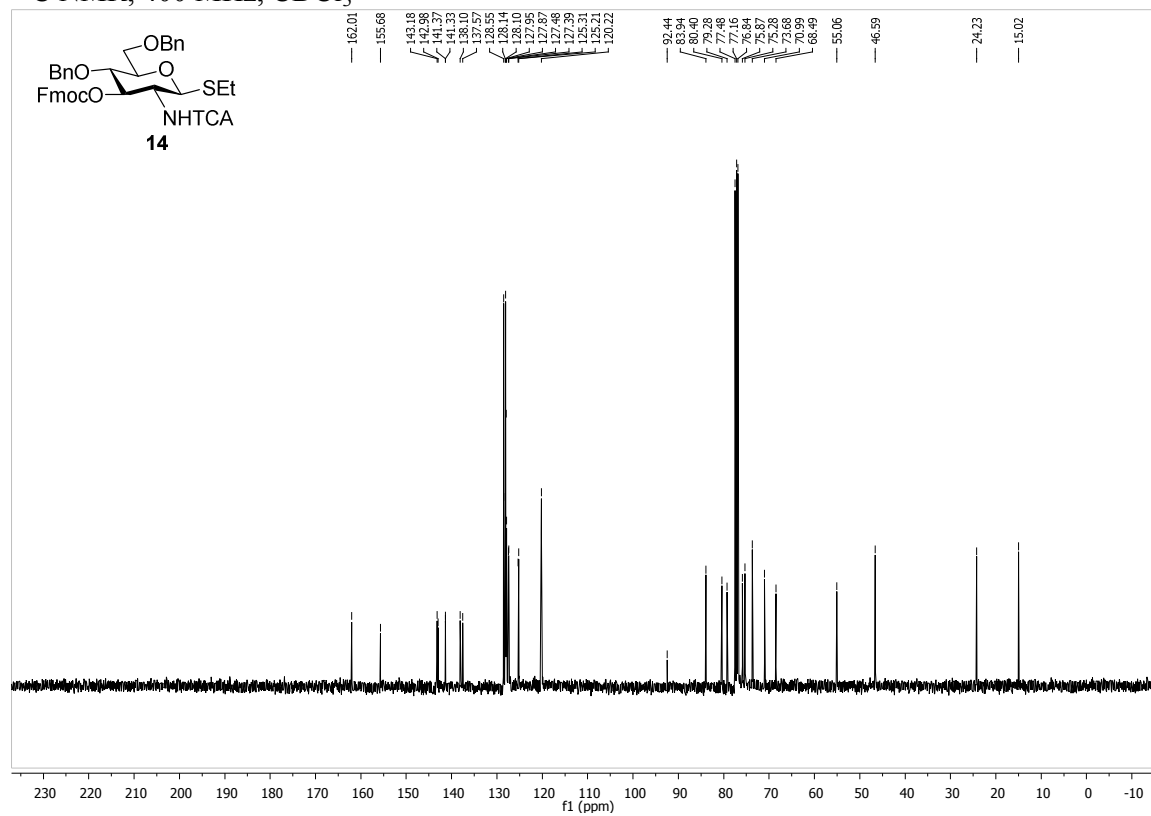
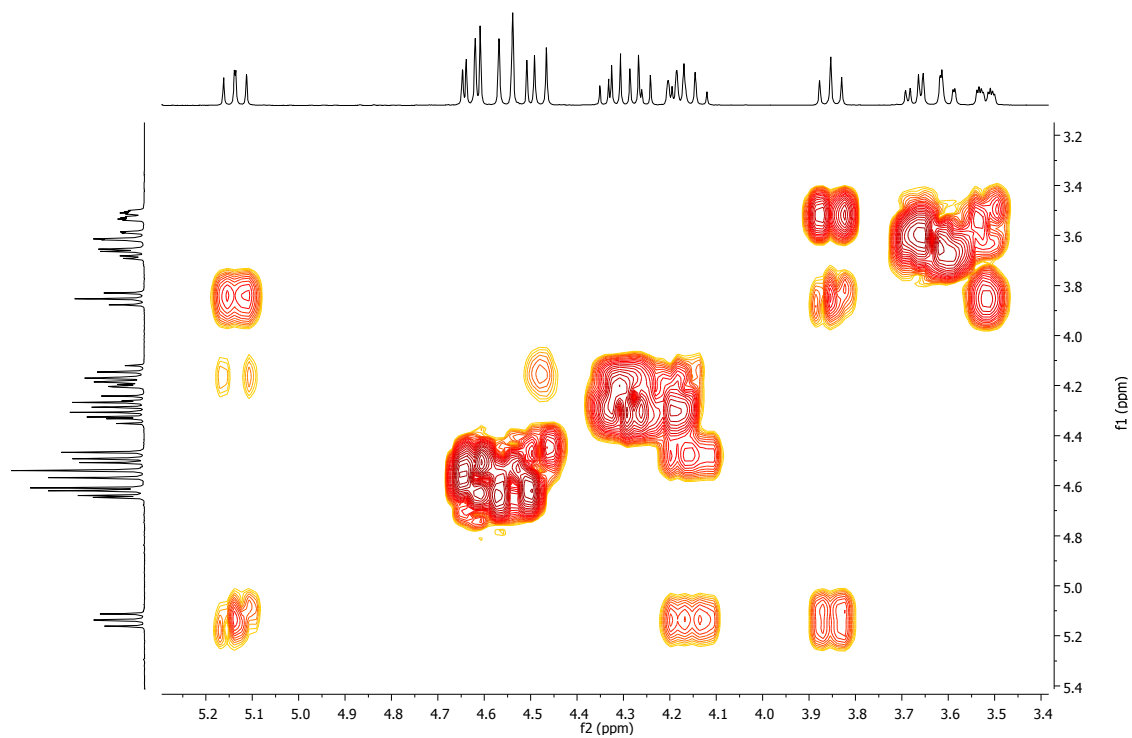
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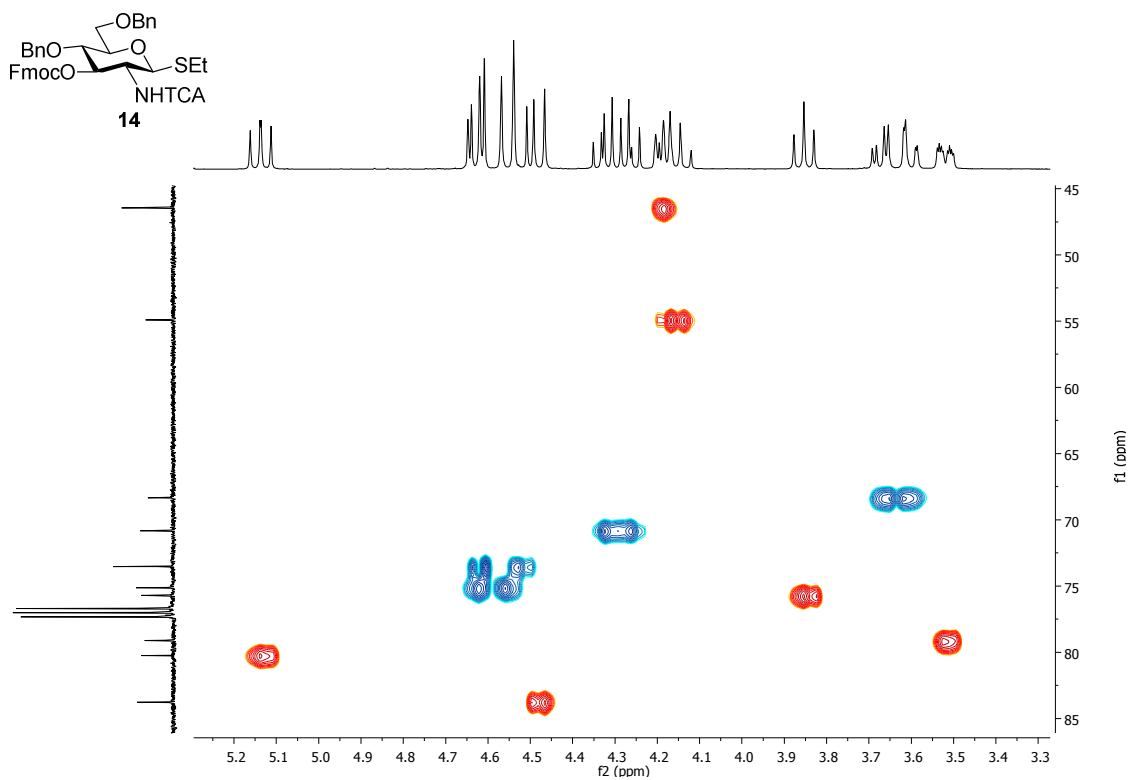
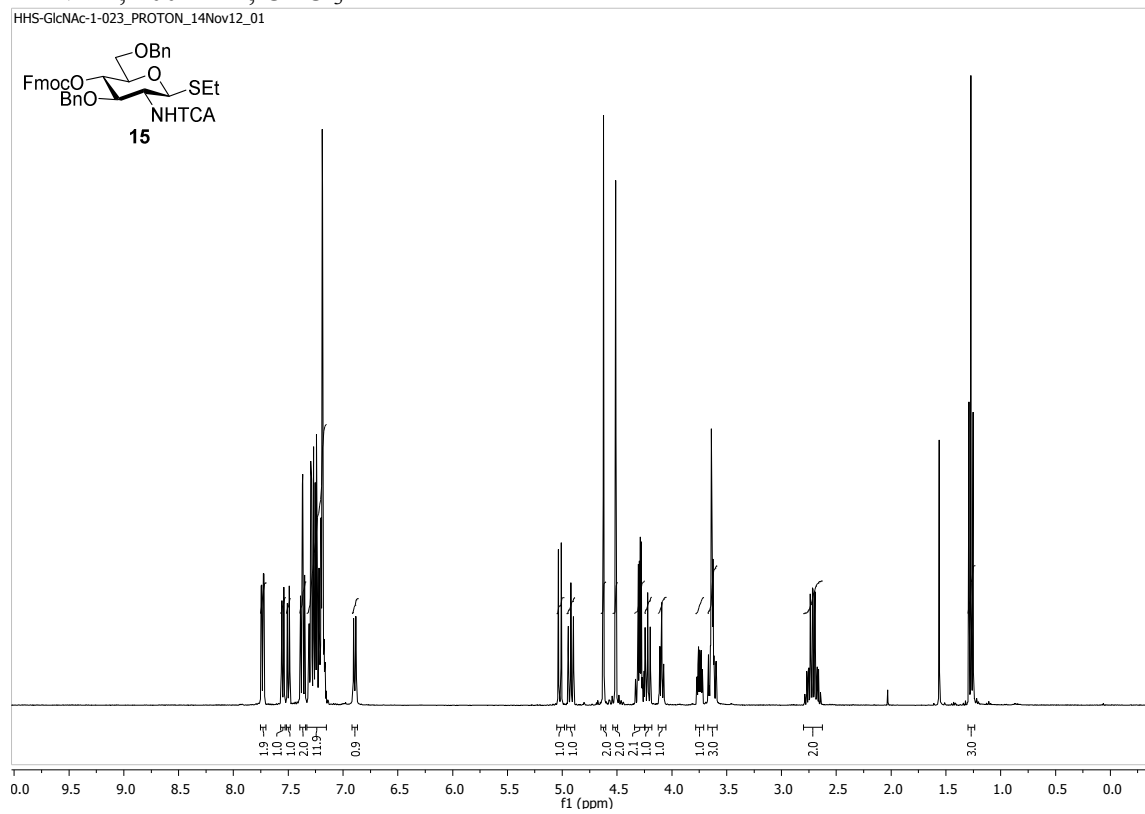
 ^1H NMR, 400 MHz, CDCl_3

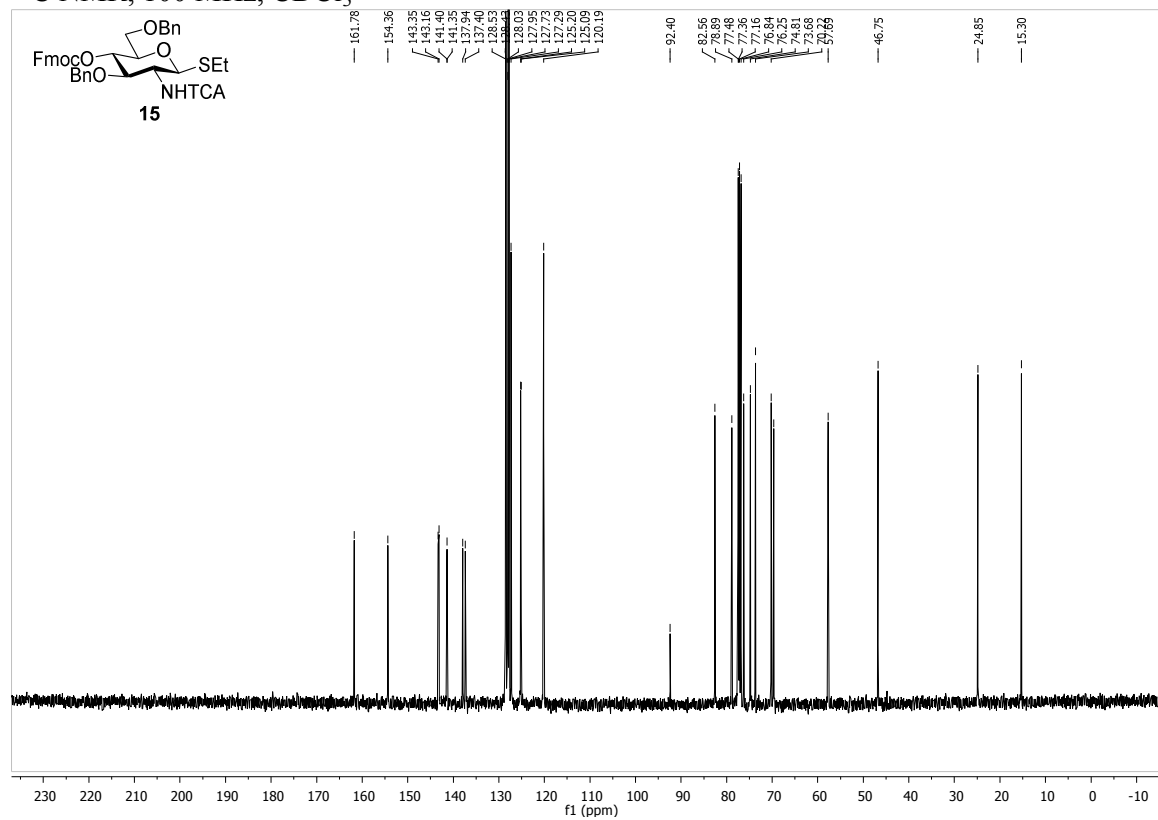
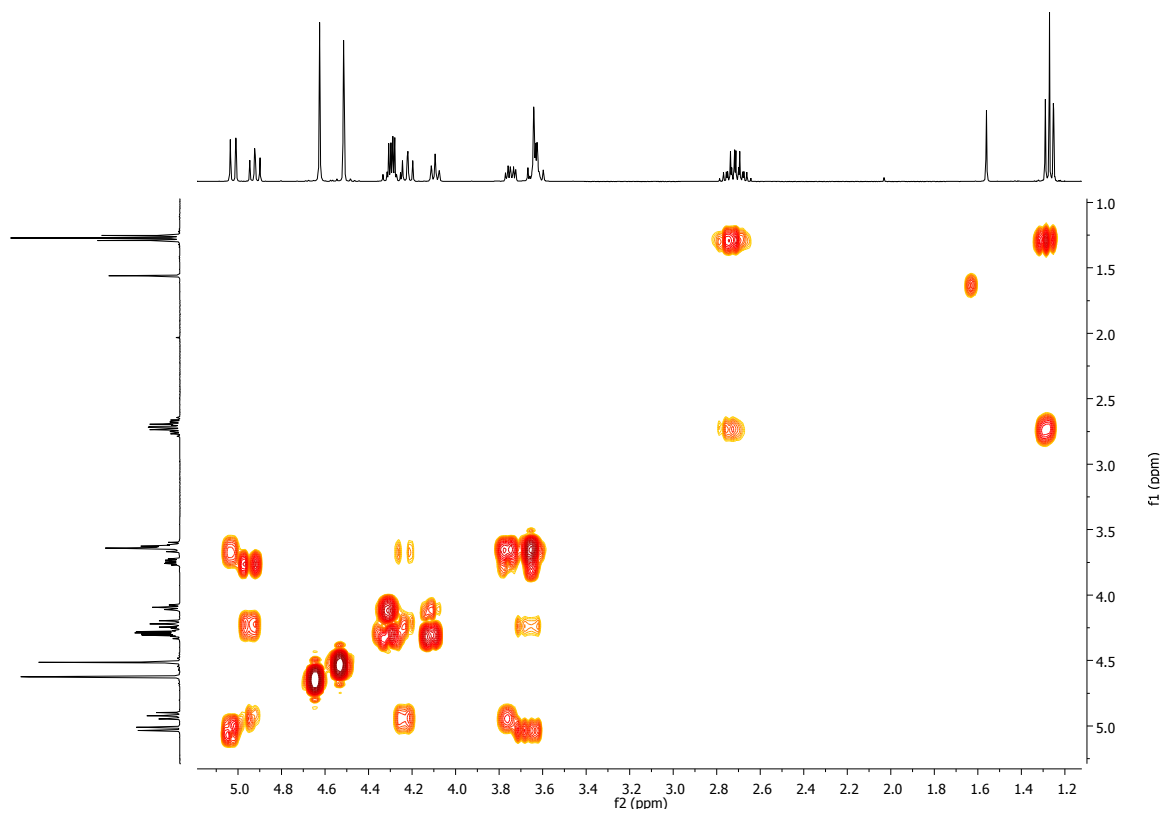
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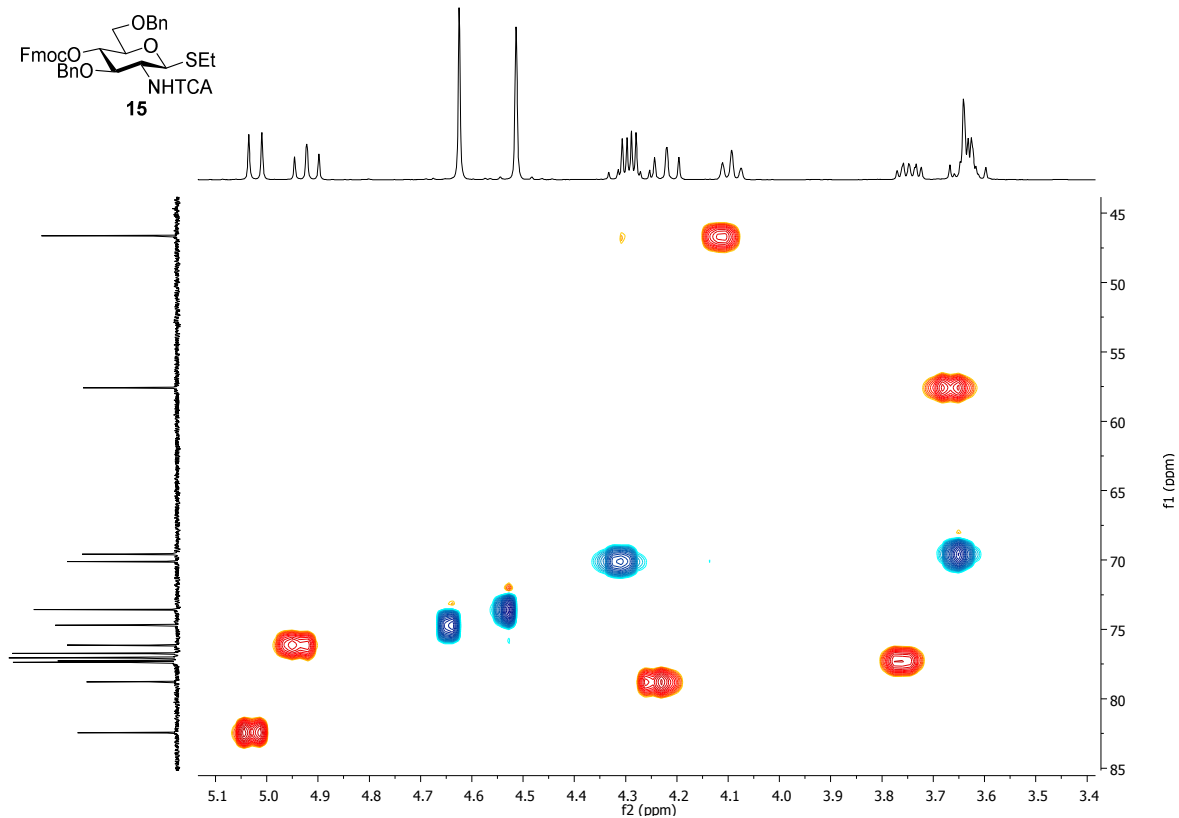
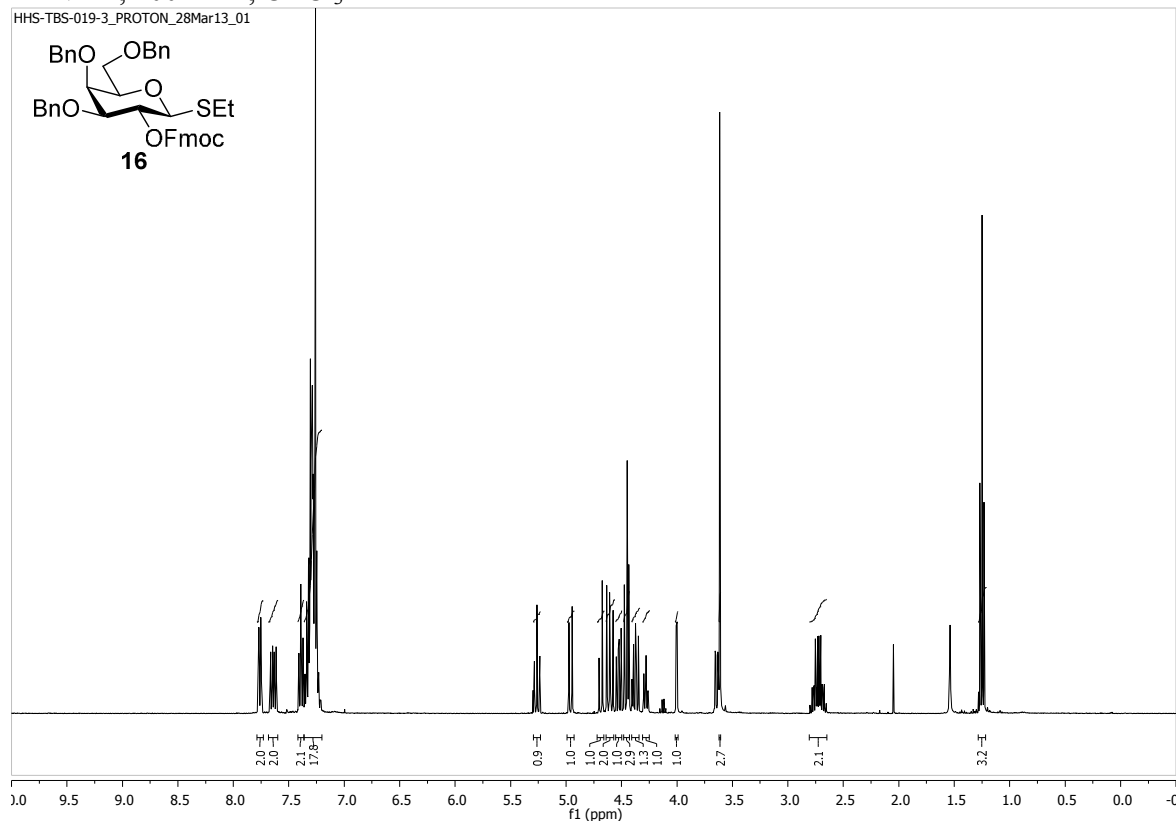
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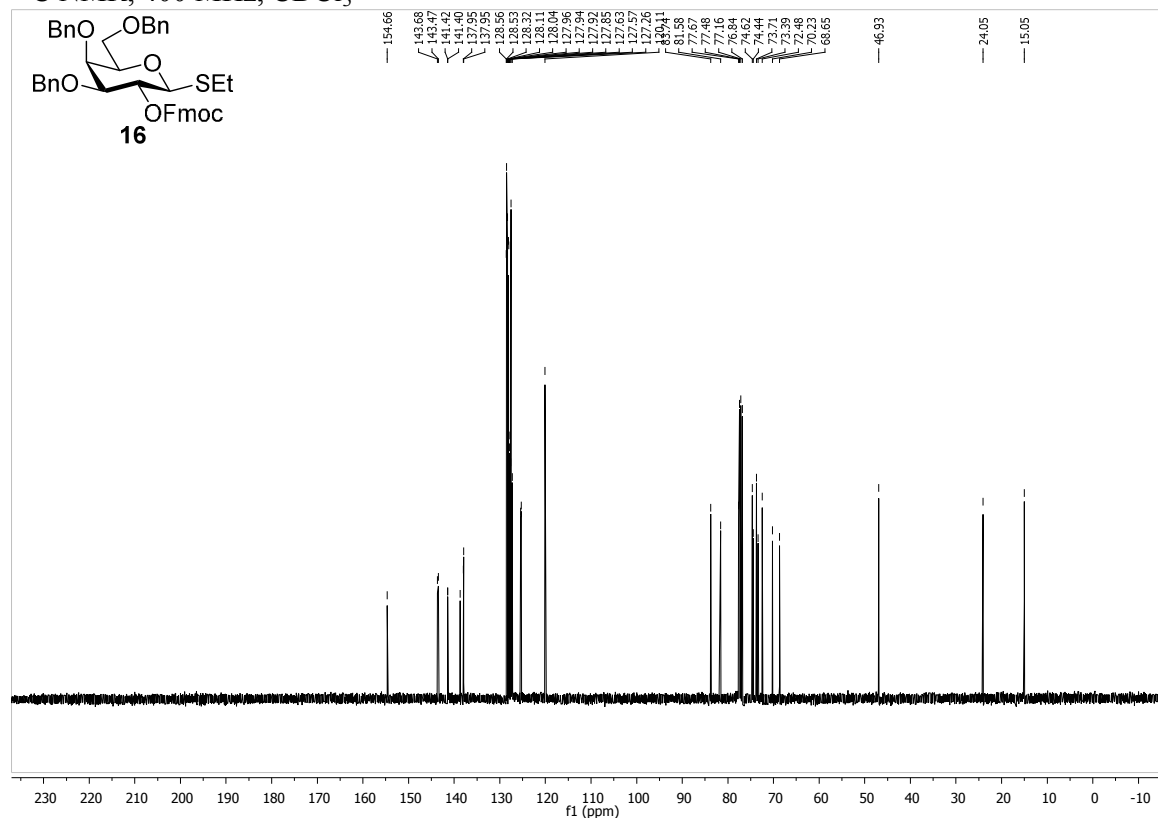
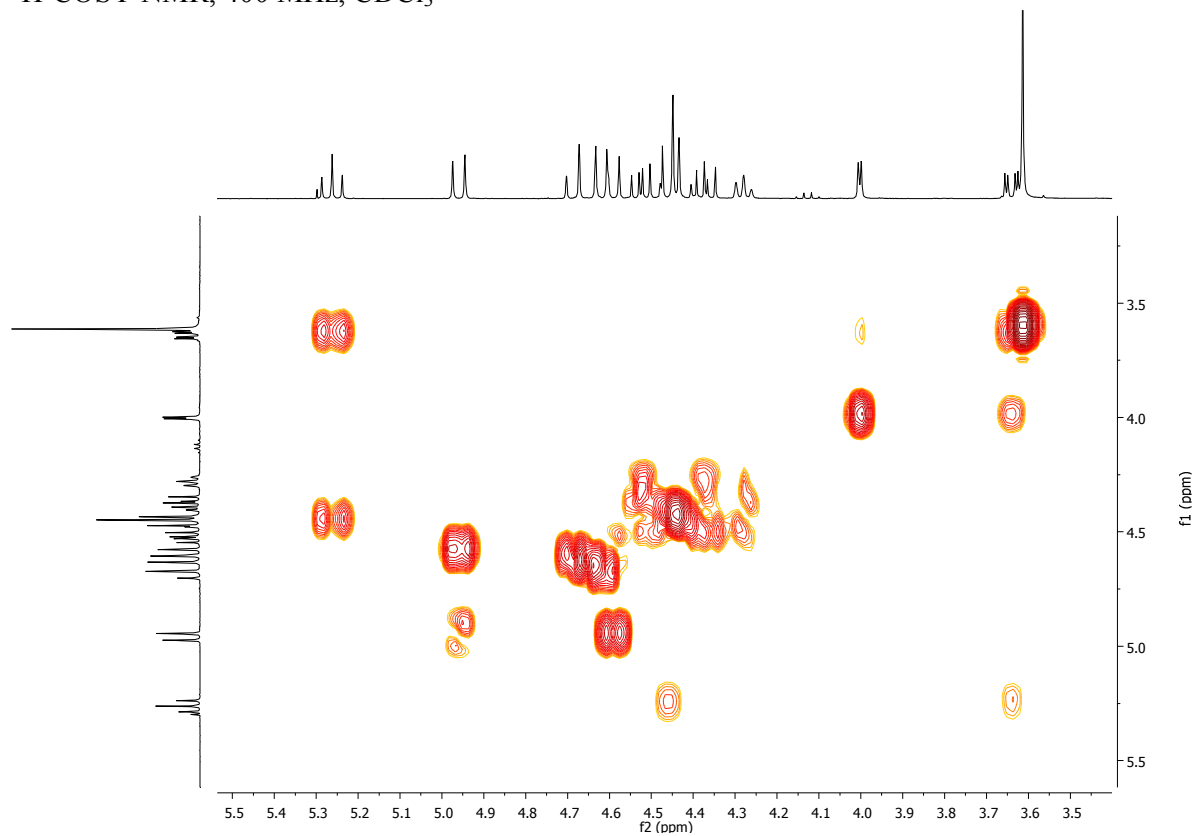


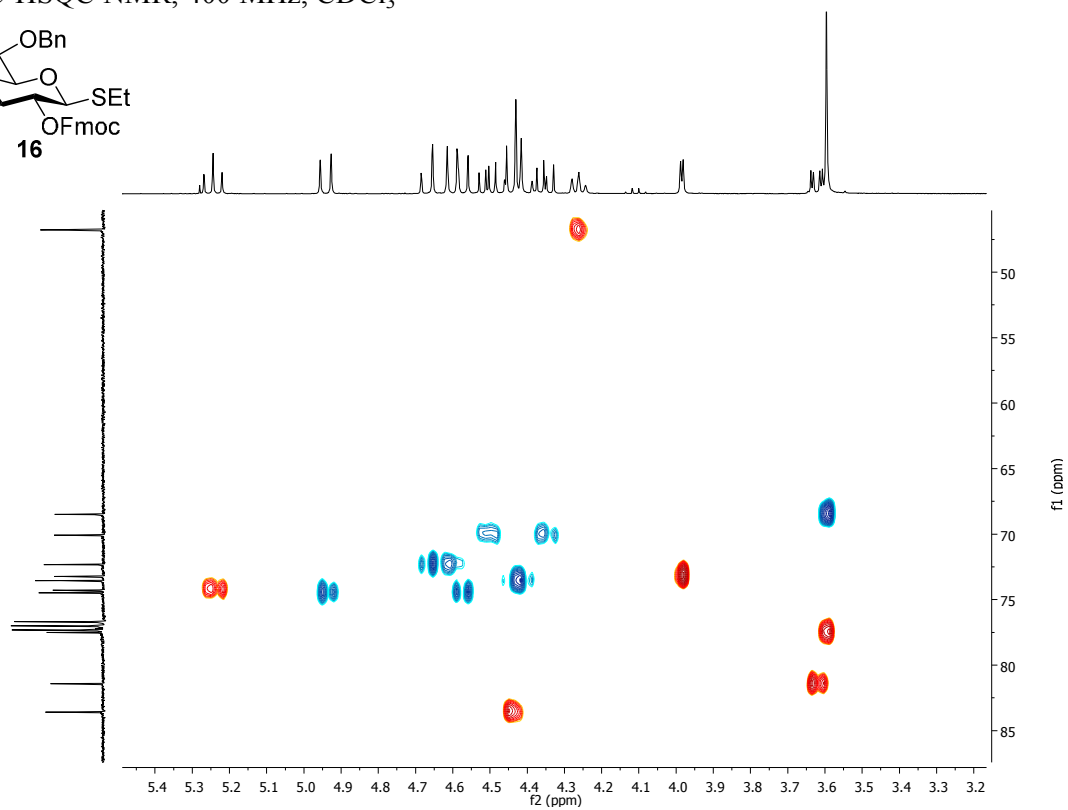
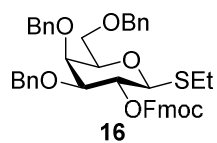
^{13}C NMR, 400 MHz, CDCl_3  ^1H -COSY NMR, 400 MHz, CDCl_3 

^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3  ^1H NMR, 400 MHz, CDCl_3 

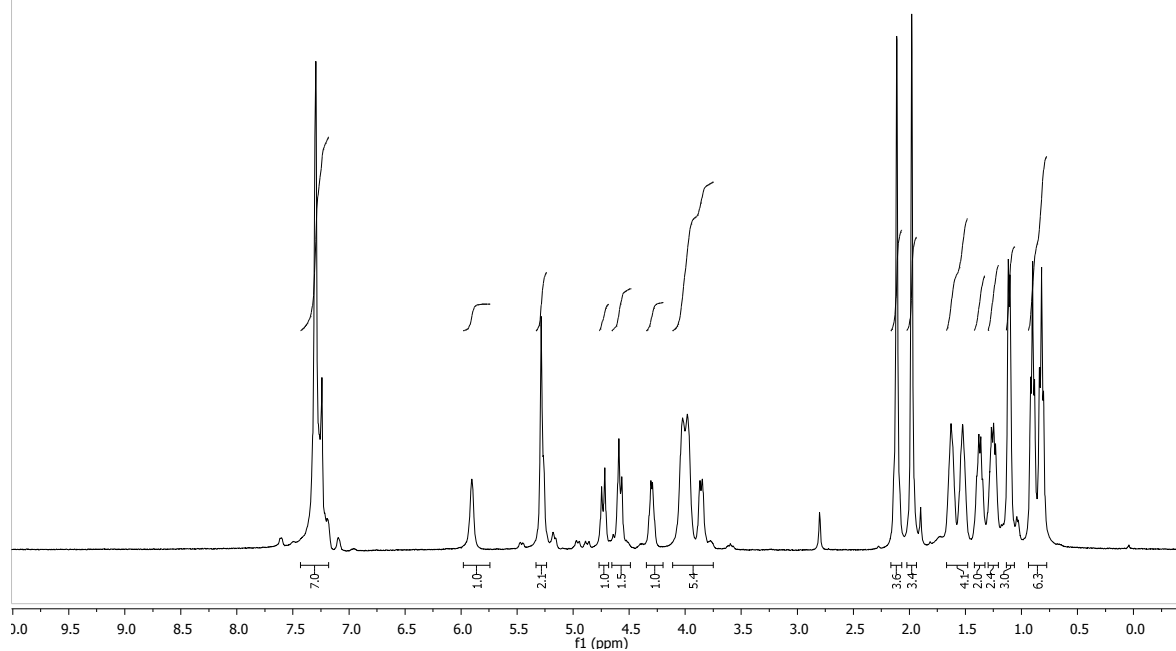
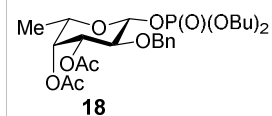
^{13}C NMR, 100 MHz, CDCl_3  ^1H -COSY NMR, 400 MHz, CDCl_3 

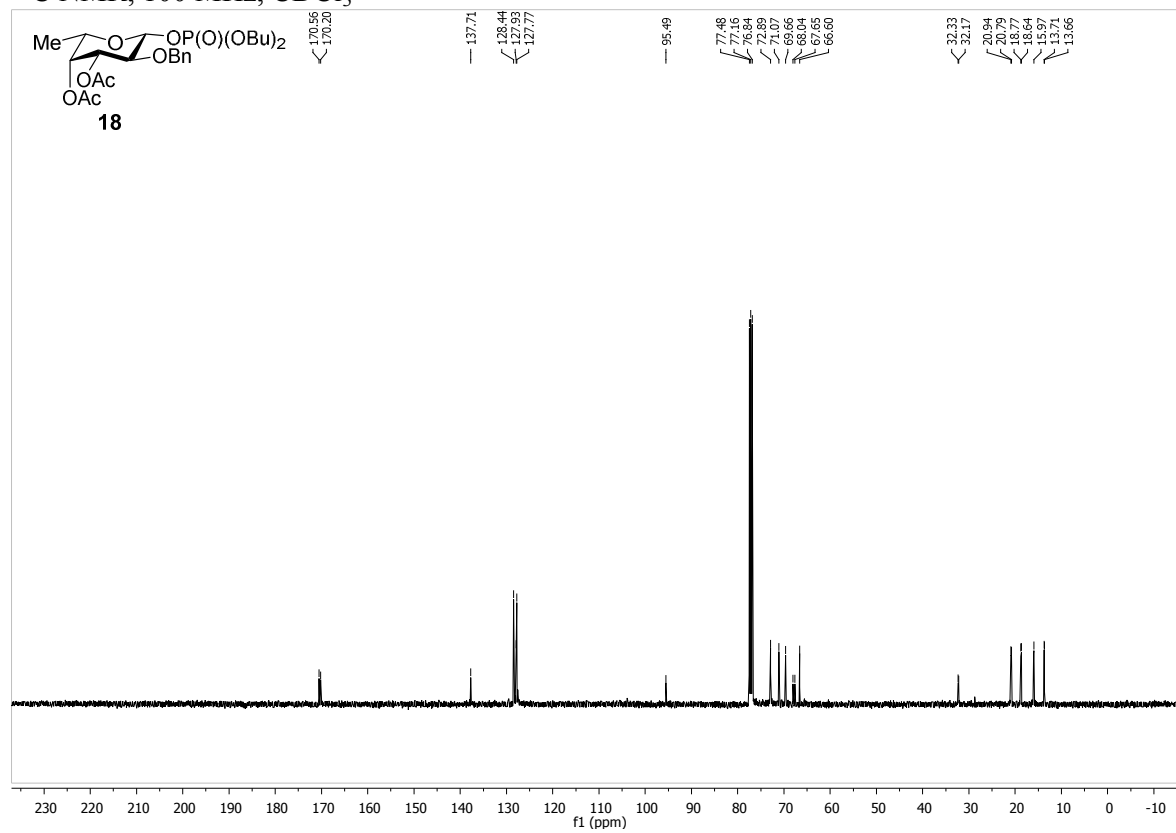
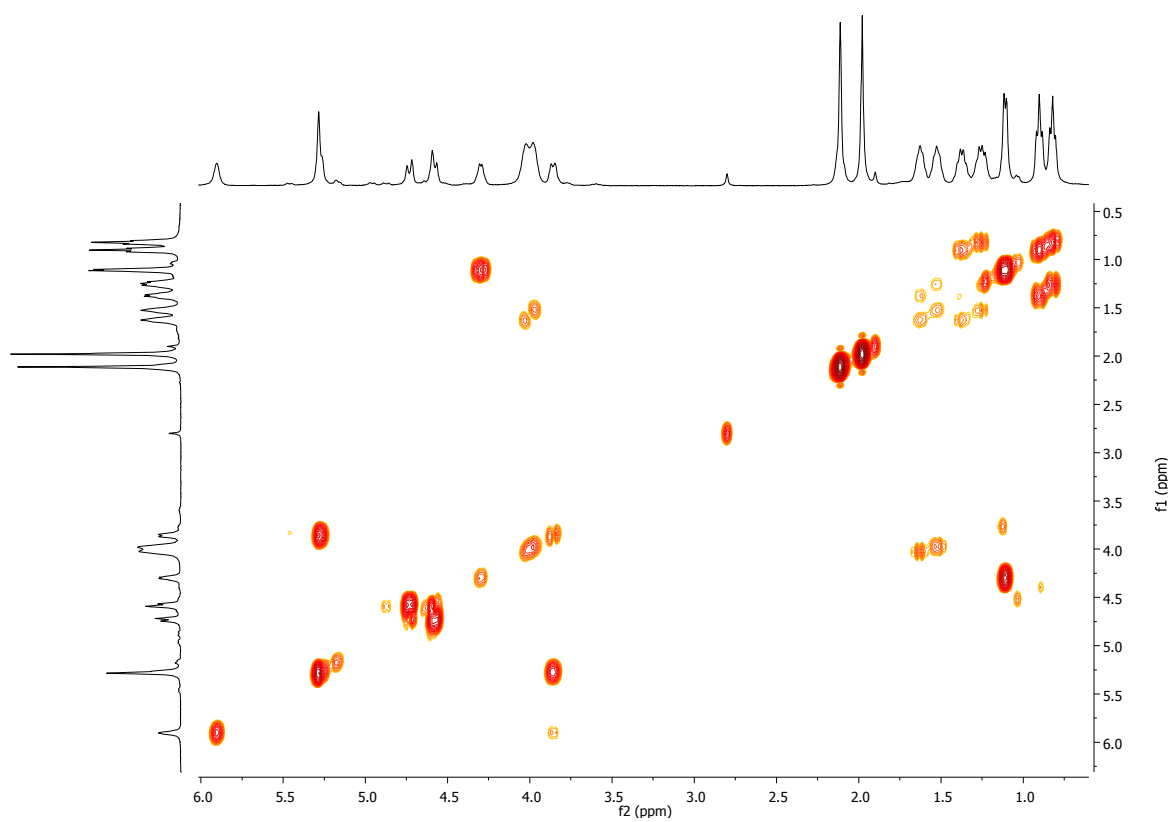
^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3  ^1H NMR, 400 MHz, CDCl_3 

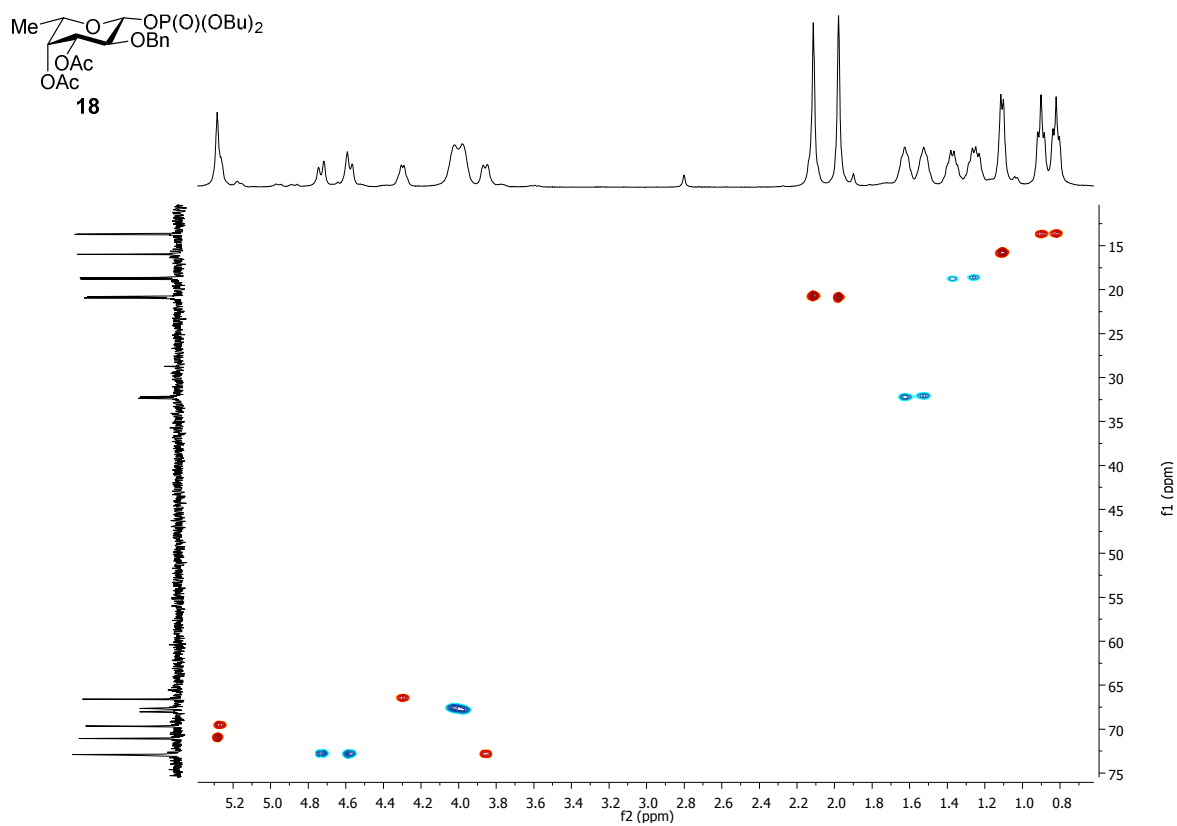
^{13}C NMR, 400 MHz, CDCl_3  ^1H -COSY NMR, 400 MHz, CDCl_3 

^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3  ^1H NMR, 400 MHz, CDCl_3

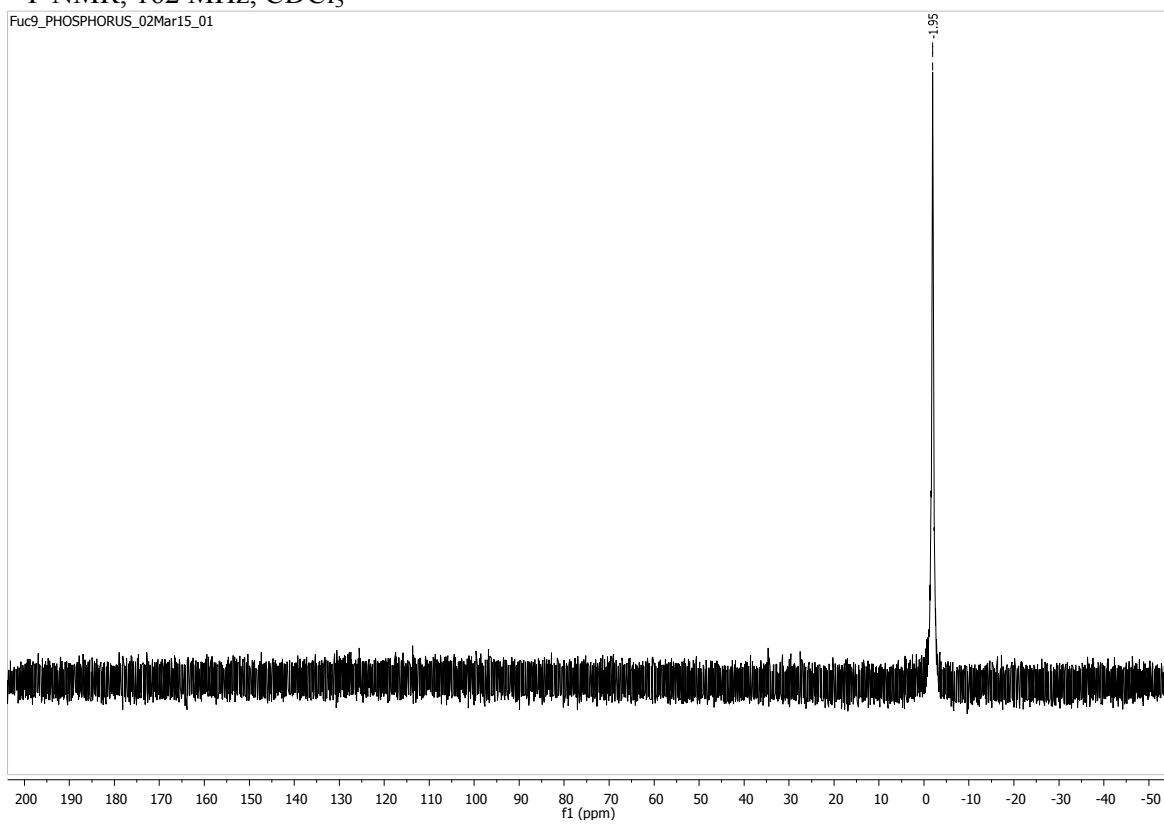
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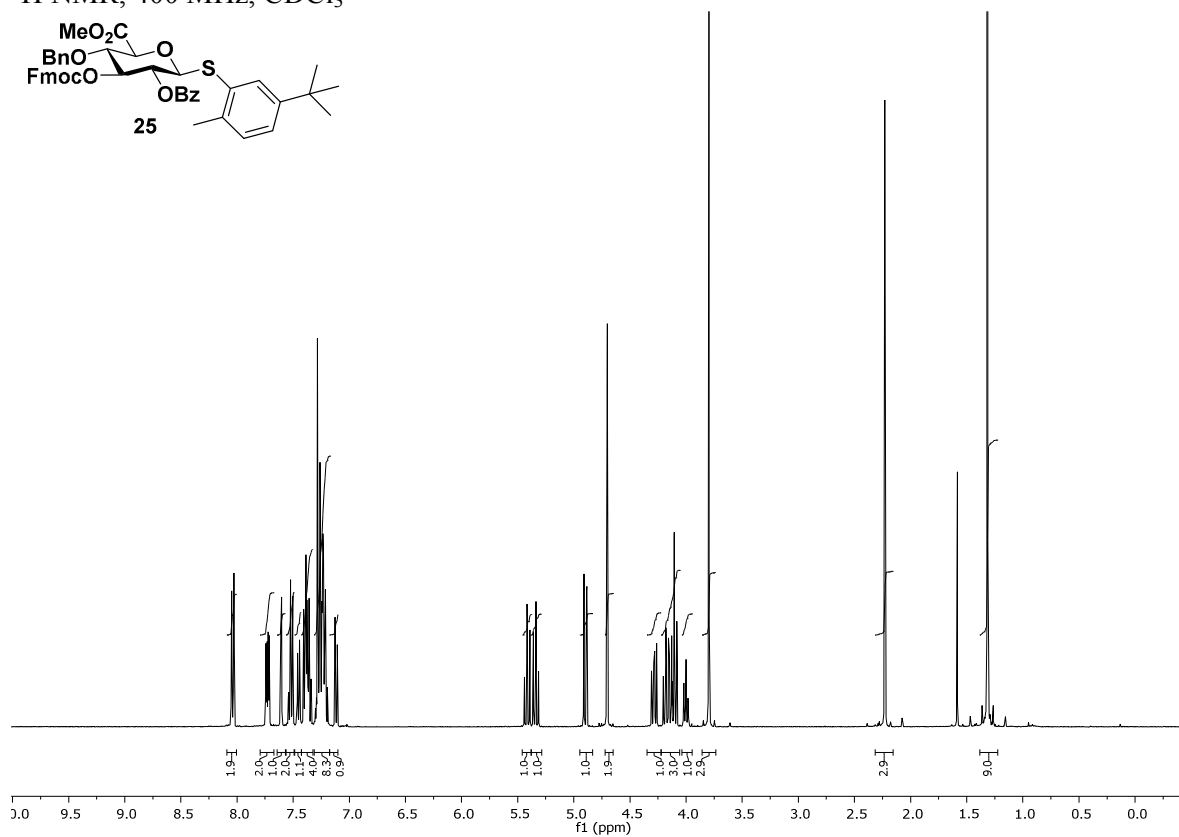
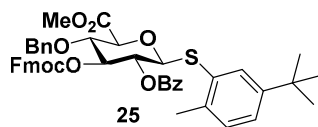
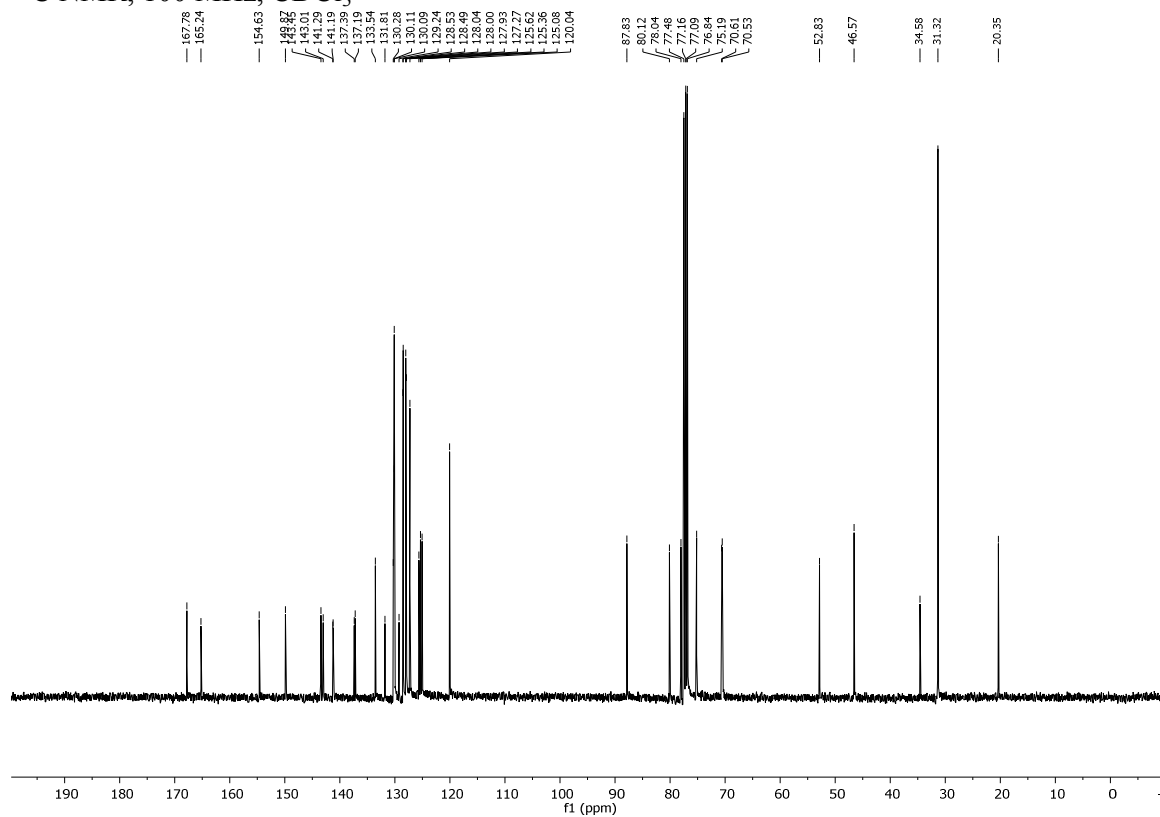


^{13}C NMR, 100 MHz, CDCl_3  ^1H -COSY NMR, 400 MHz, CDCl_3 

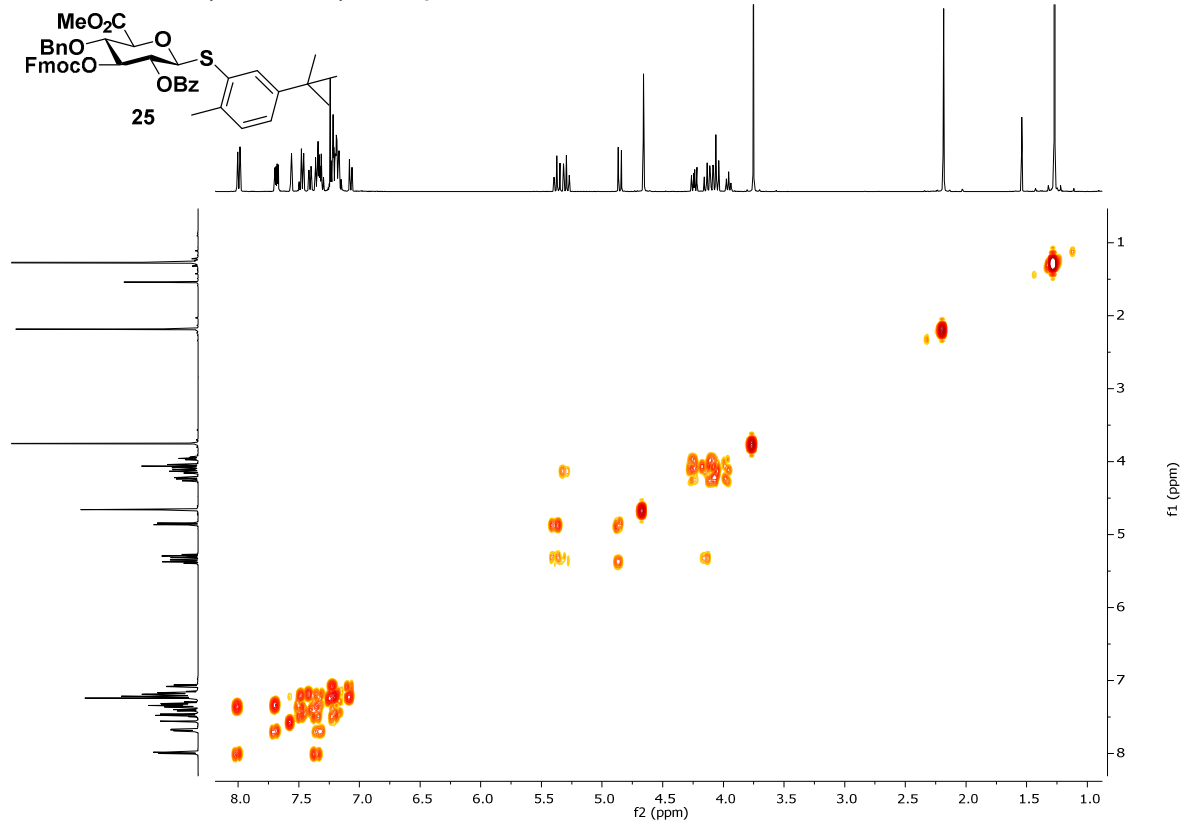
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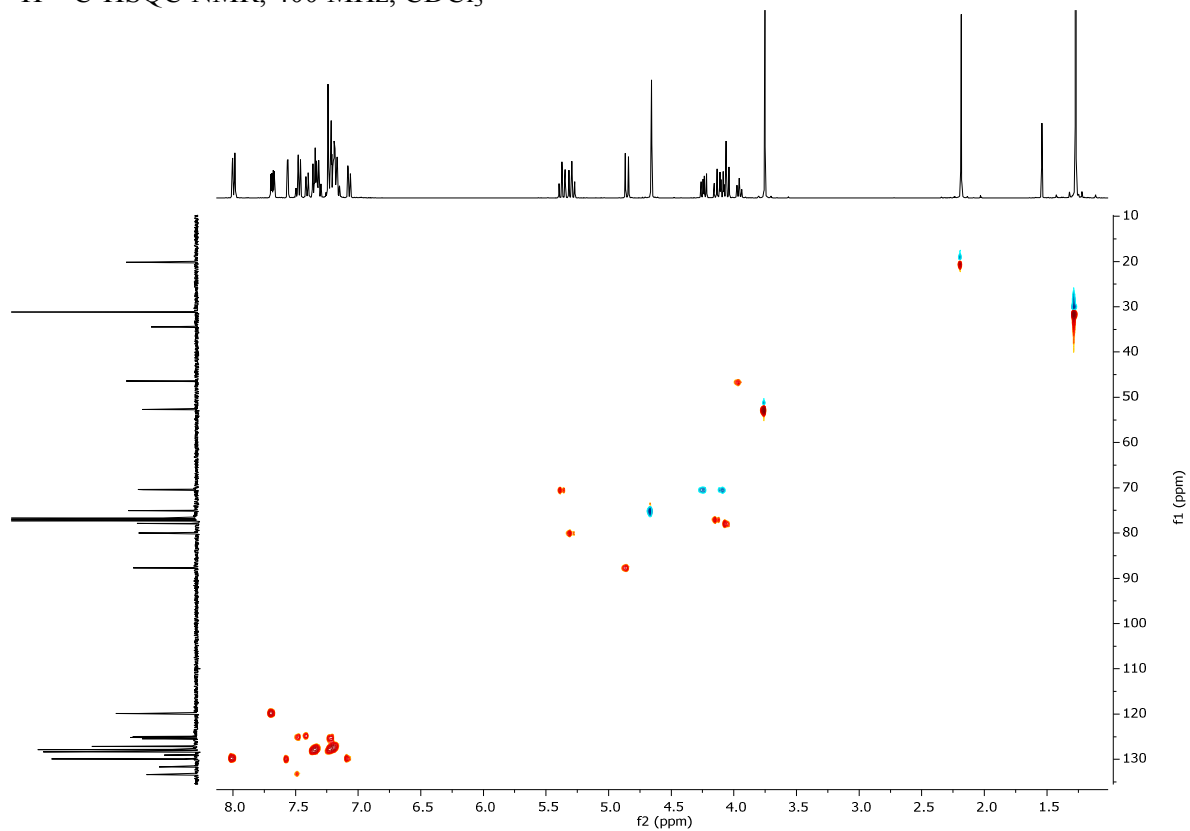


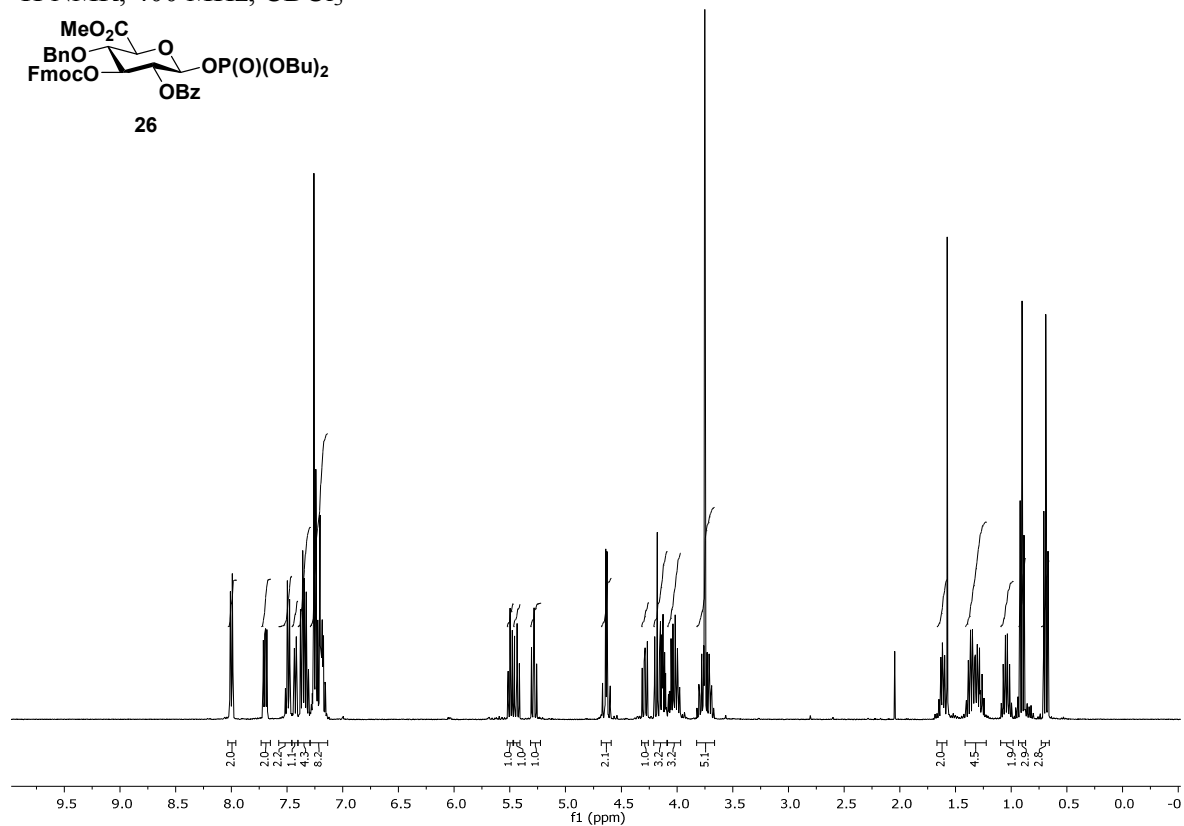
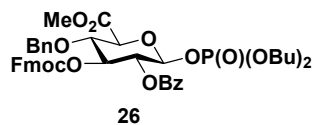
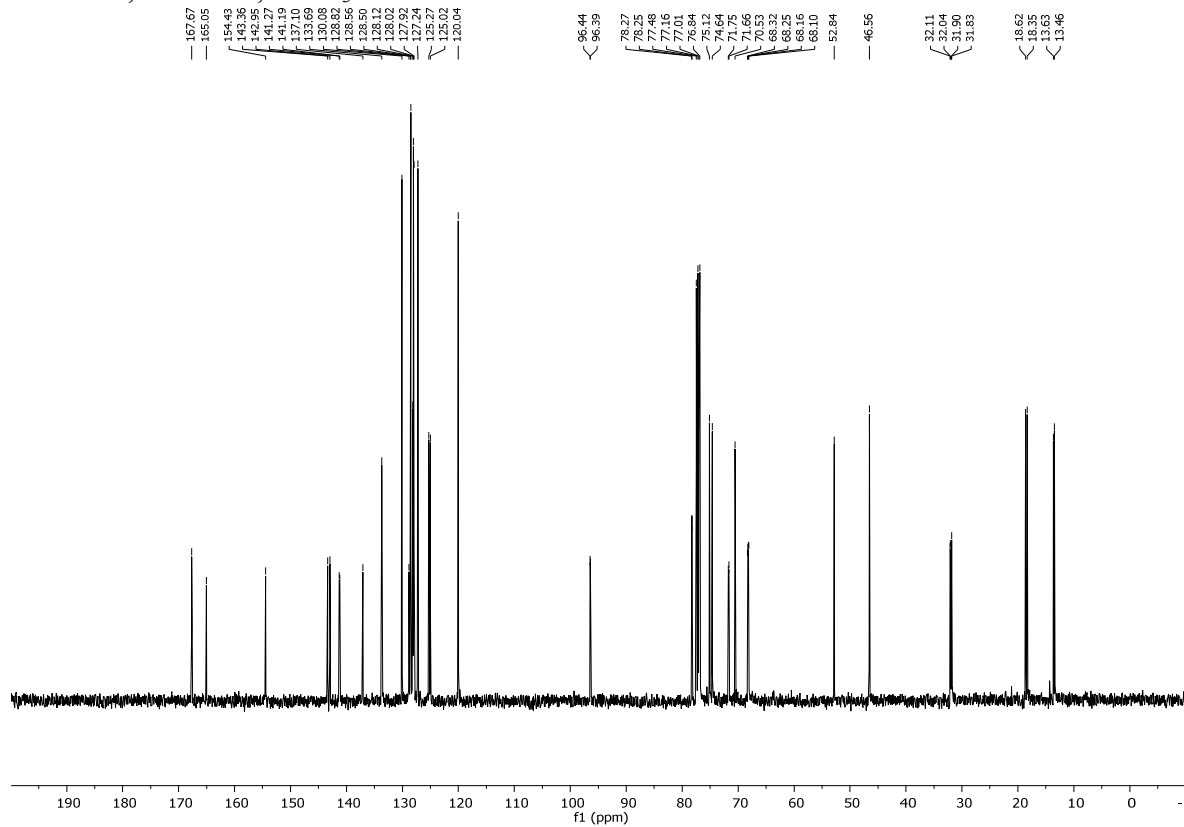
^1H NMR, 400 MHz, CDCl_3  ^{13}C NMR, 100 MHz, CDCl_3 

^1H -COSY NMR, 400 MHz, CDCl_3

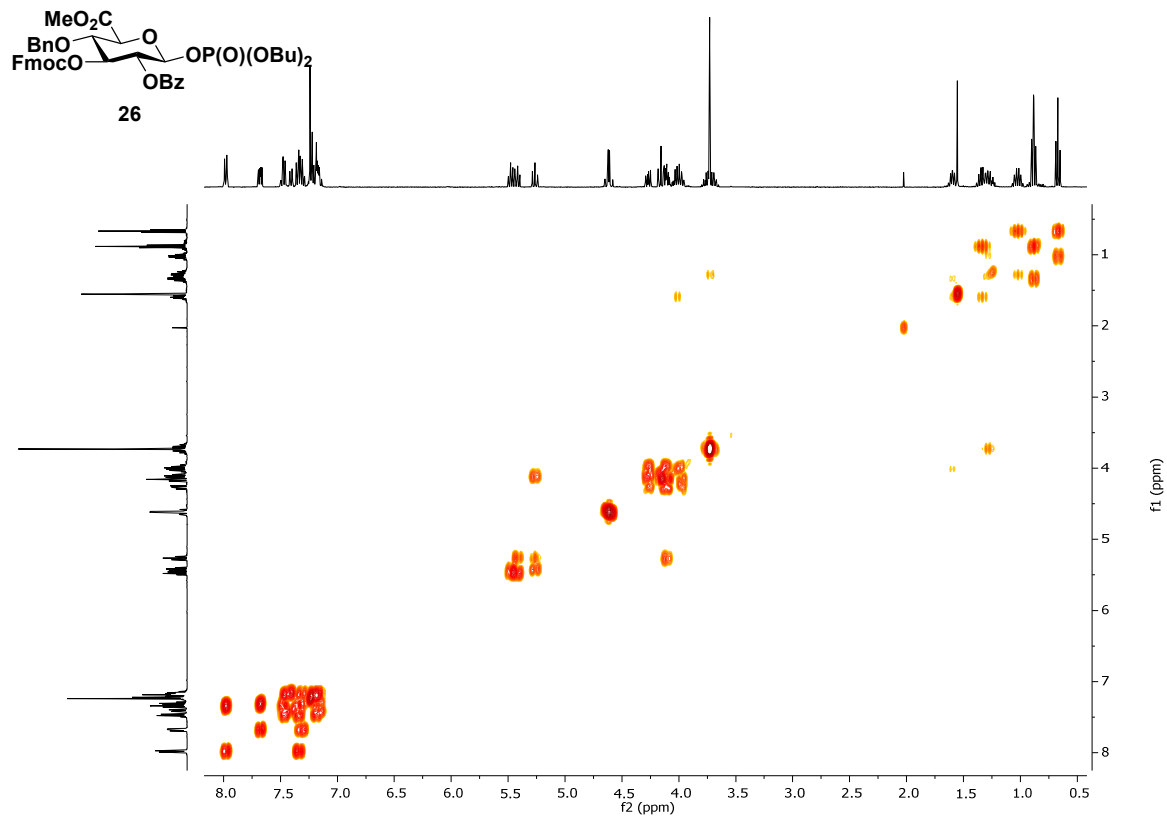


^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3

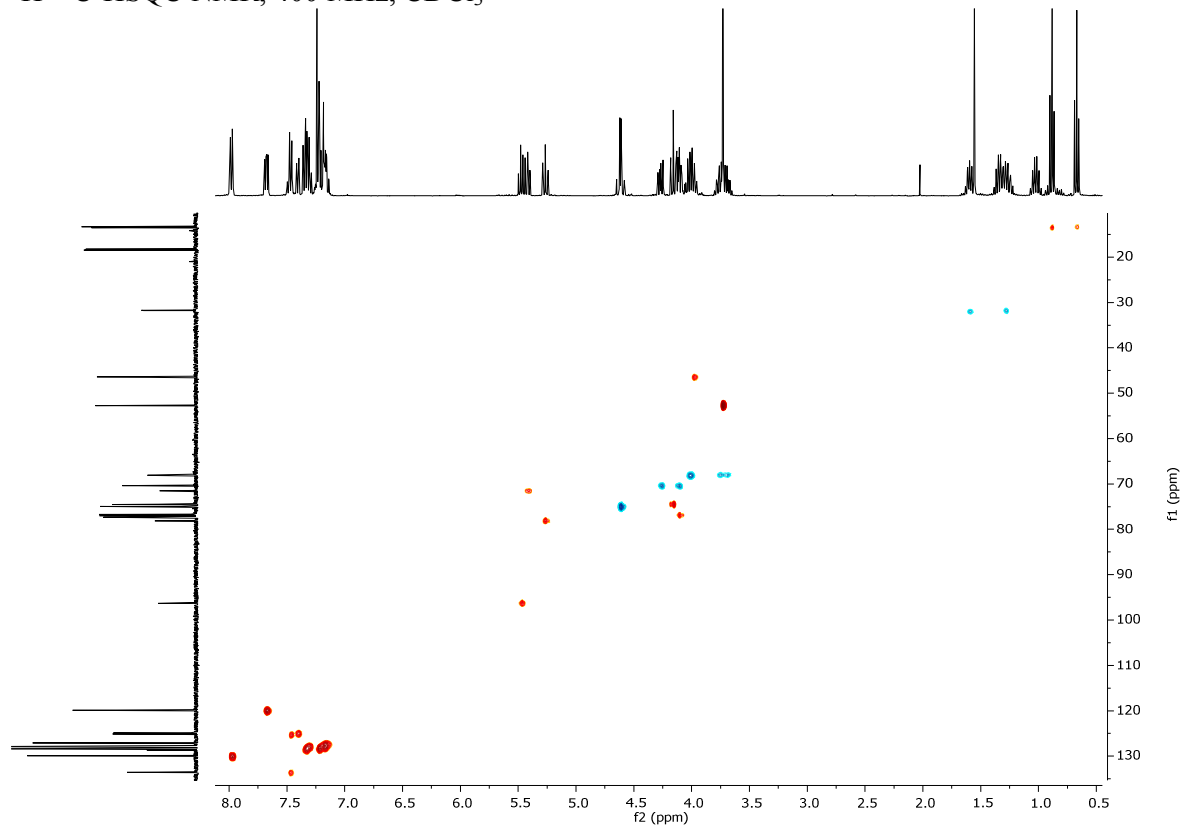


^1H NMR, 400 MHz, CDCl_3  ^{13}C NMR, 100 MHz, CDCl_3 

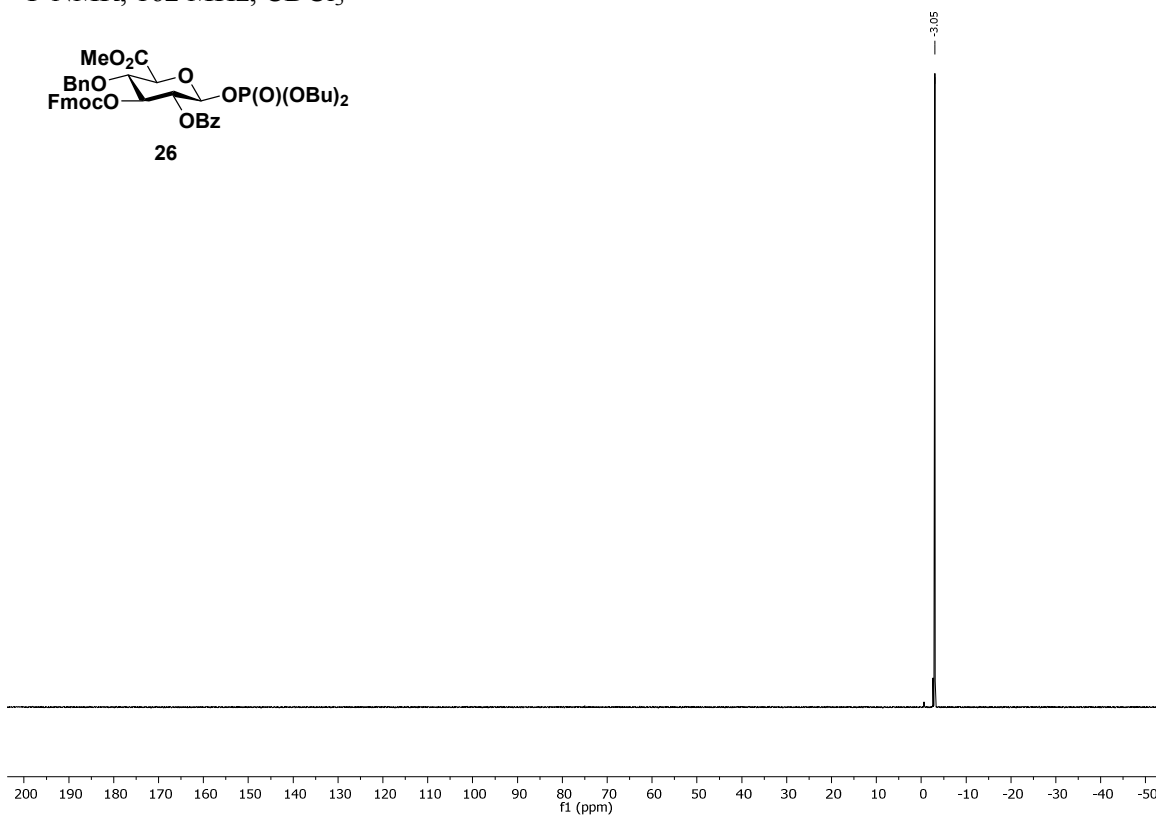
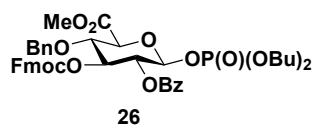
^1H -COSY NMR, 400 MHz, CDCl_3



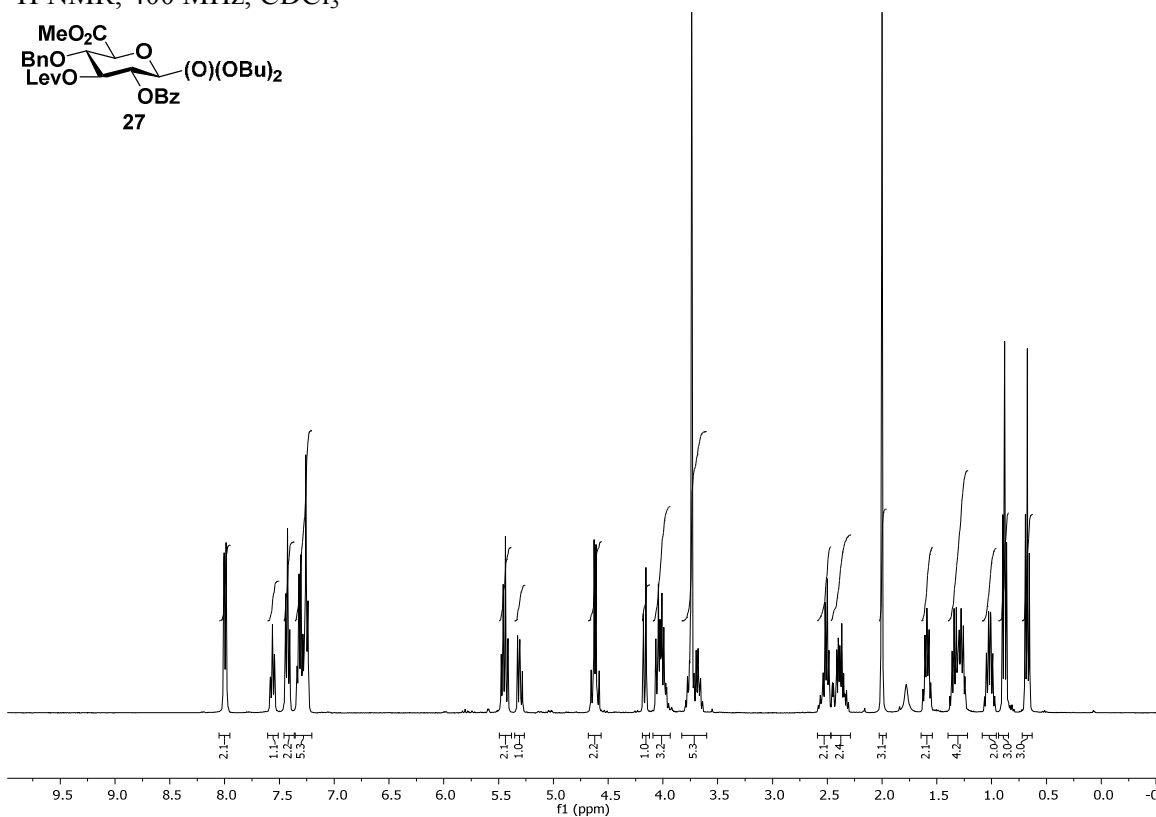
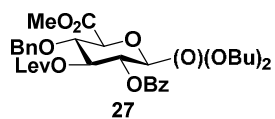
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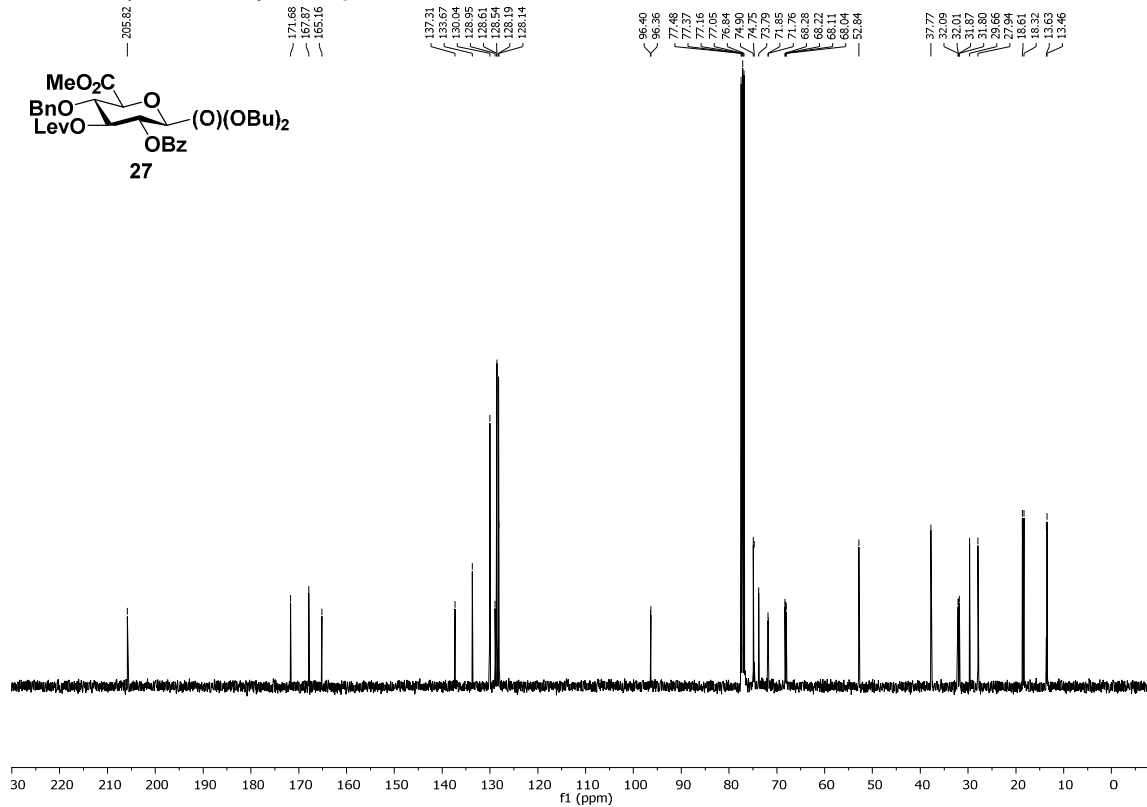
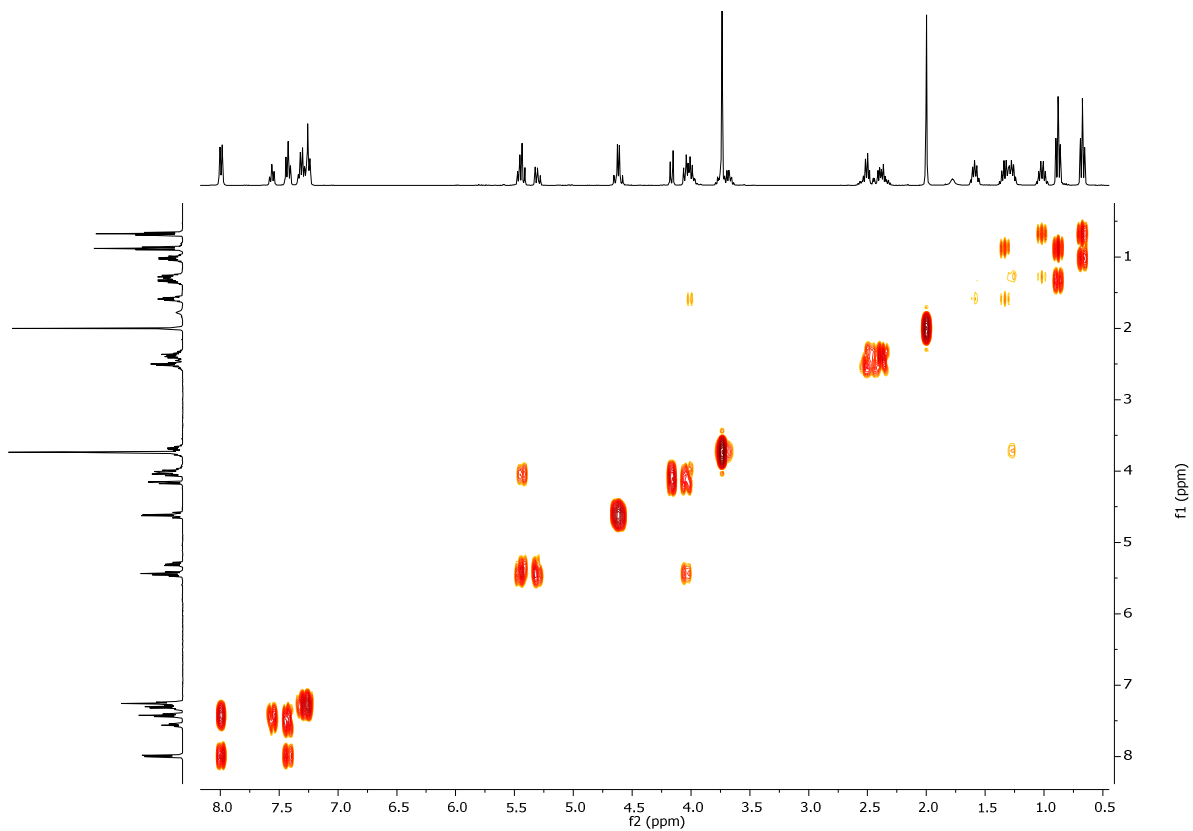


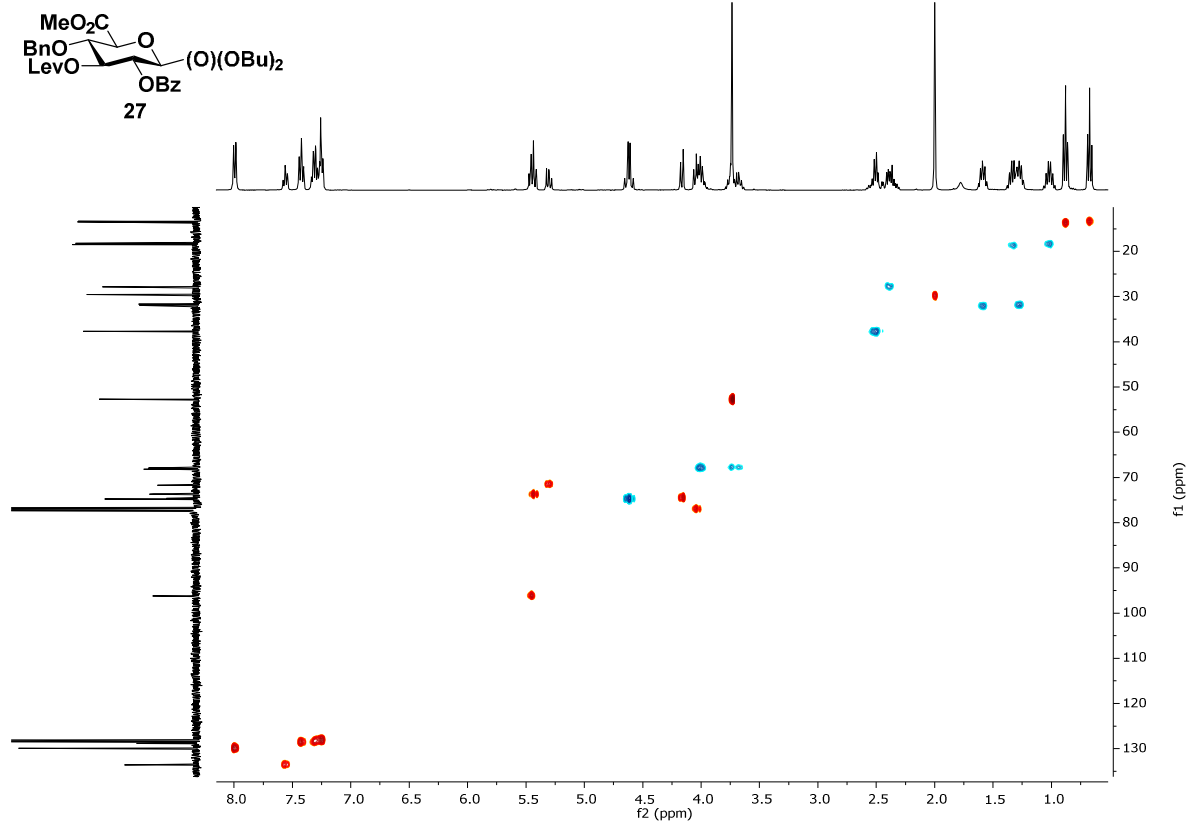
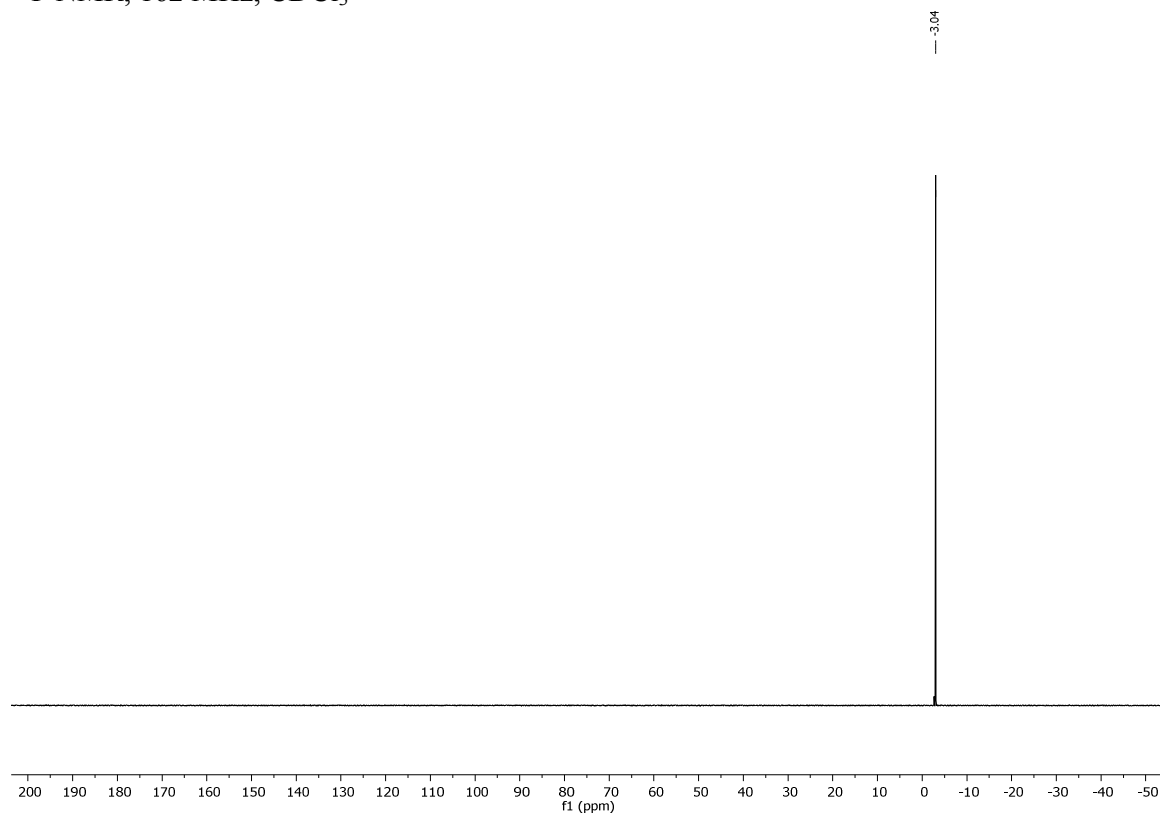
^{31}P NMR, 162 MHz, CDCl_3



^1H NMR, 400 MHz, CDCl_3



^{13}C NMR, 100 MHz, CDCl_3  ^1H -COSY NMR, 400 MHz, CDCl_3 

^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3  ^{31}P NMR, 162 MHz, CDCl_3 

Building Block	Promotor	T_a (°C)	t_1 (min)	T_i (°C)	t_2 (min)
2, 6, and 7	NIS/TfOH	- 30	5	- 10	25
3, 8, 9, 10, and 11		- 40	5	- 20	25
23		- 20	5	- 10	50
4 and 22	TMSOTf	- 40	5	- 20	25
12		- 10	5	0	50
24		- 20	5	- 10	50

Sequence	Module	Details	Condition
I	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	2	5 eq. building block (2, 3, 6, 7, 8, and 23), 5 eq. of NIS Solution	
	3	Fmoc Removal	r.t for 5 min
II	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	4-1	5 eq. building block (4 and 15) , 5 eq. of TMSOTf Solution	
	3	Fmoc Removal	r.t for 5 min
III	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	2	5 eq. building block (9, 10, and 11) 5 eq. of NIS Solution	
III	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	2	5 eq. building block (22), 5 eq. of TMSOTf solution	
IV	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	4-1	5 eq. building block 12 and 24 , 5 eq. of TMSOTf Solution	
	5	Lev Removal	r.t for 5 min
V	1	2.5 eq. of TMSOTf solution	-20 °C, for 1 min
	4-2	5 eq. building block 12 , 5 eq. of TMSOTf Solution	
	5	Lev Removal	r.t for 5 min

Table S1. Sequences of the glycosylation cycle with the corresponding monomers and optimized conditions for “approved building block”. Glycosylation condition: activation temperature (T_a) and time (t_1), incubation temperature (T_i) and time (t_2).

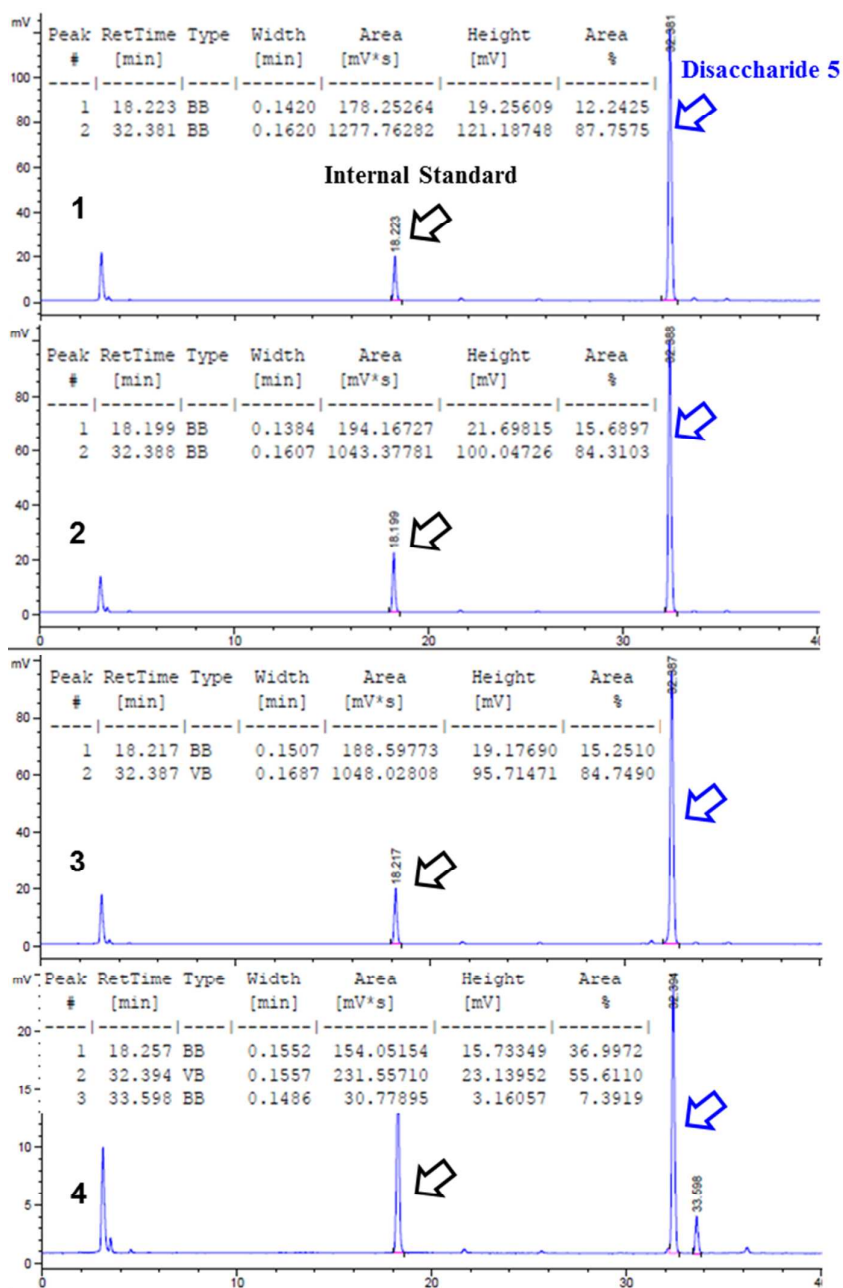
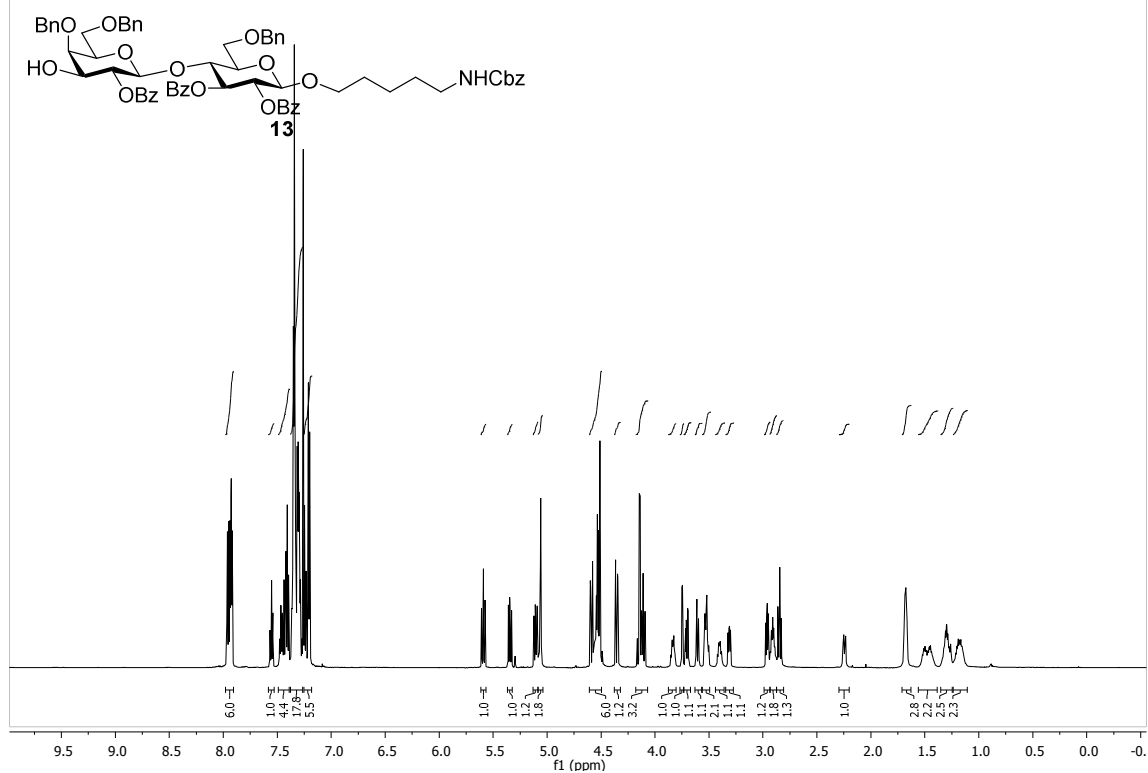
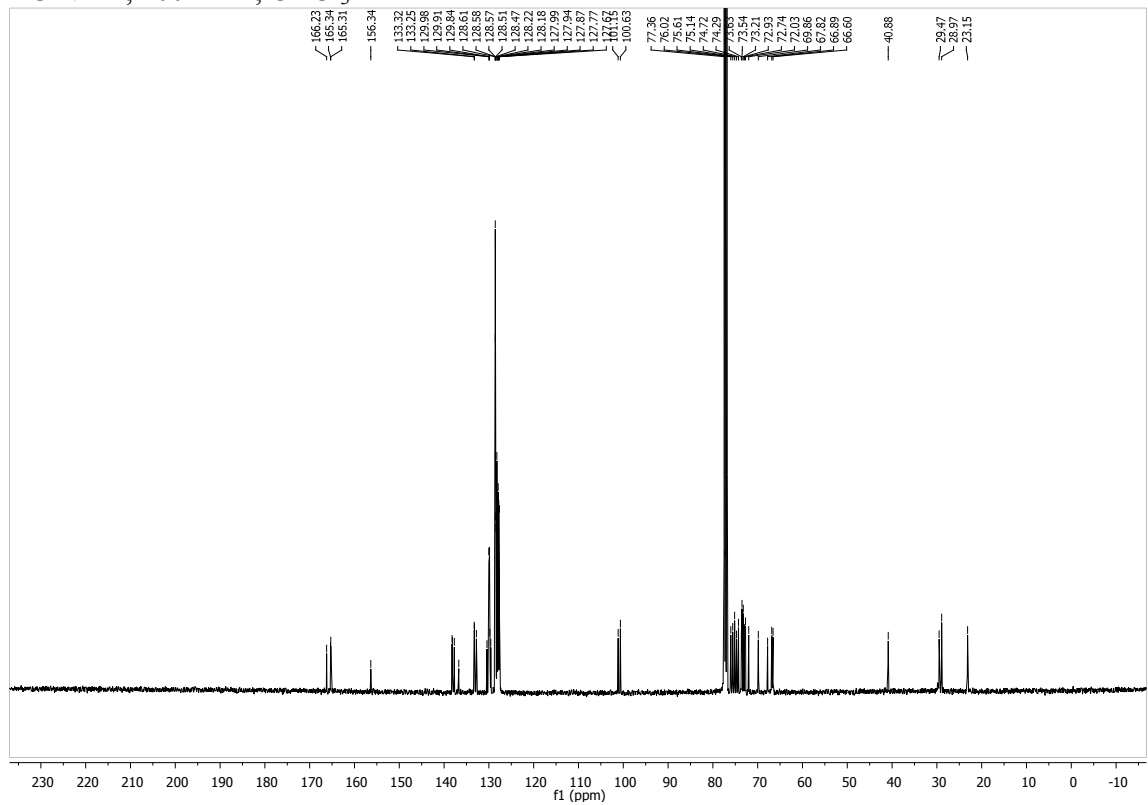


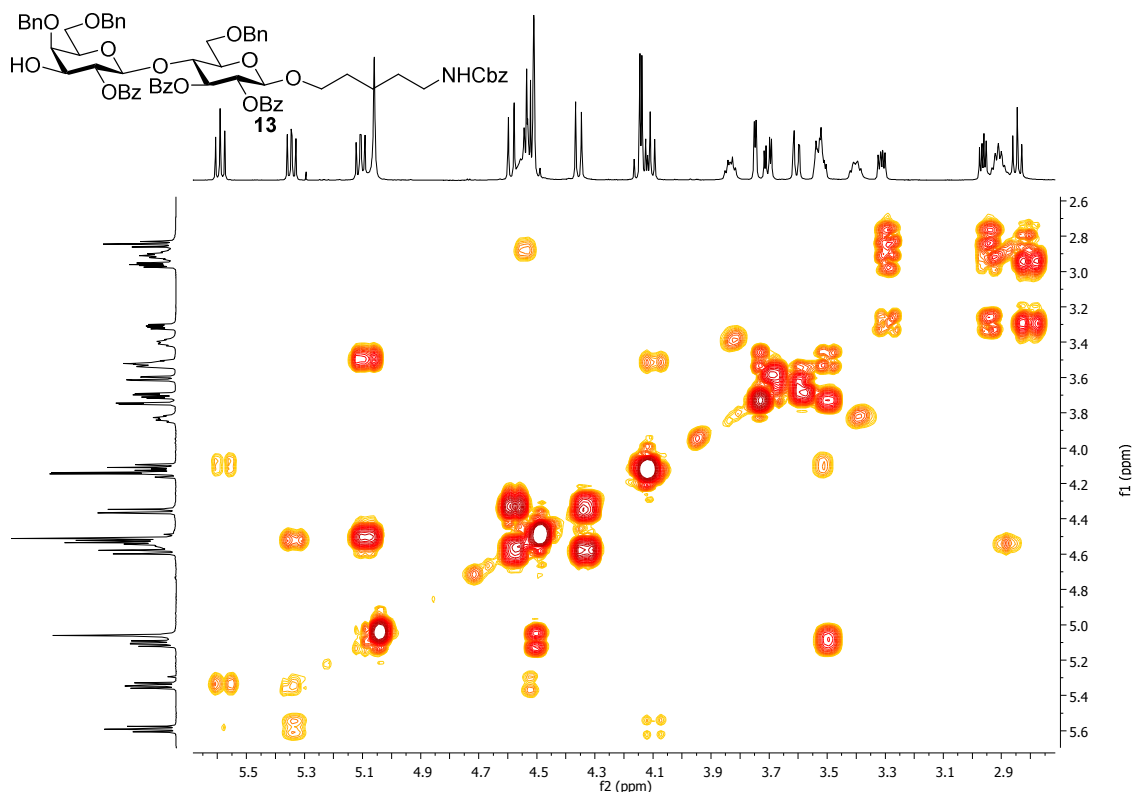
Figure S1. LC-MS of disaccharide **13** (blue arrow) including building block **11** (black arrow) as an Internal standard.

^1H NMR, 400 MHz, CDCl_3

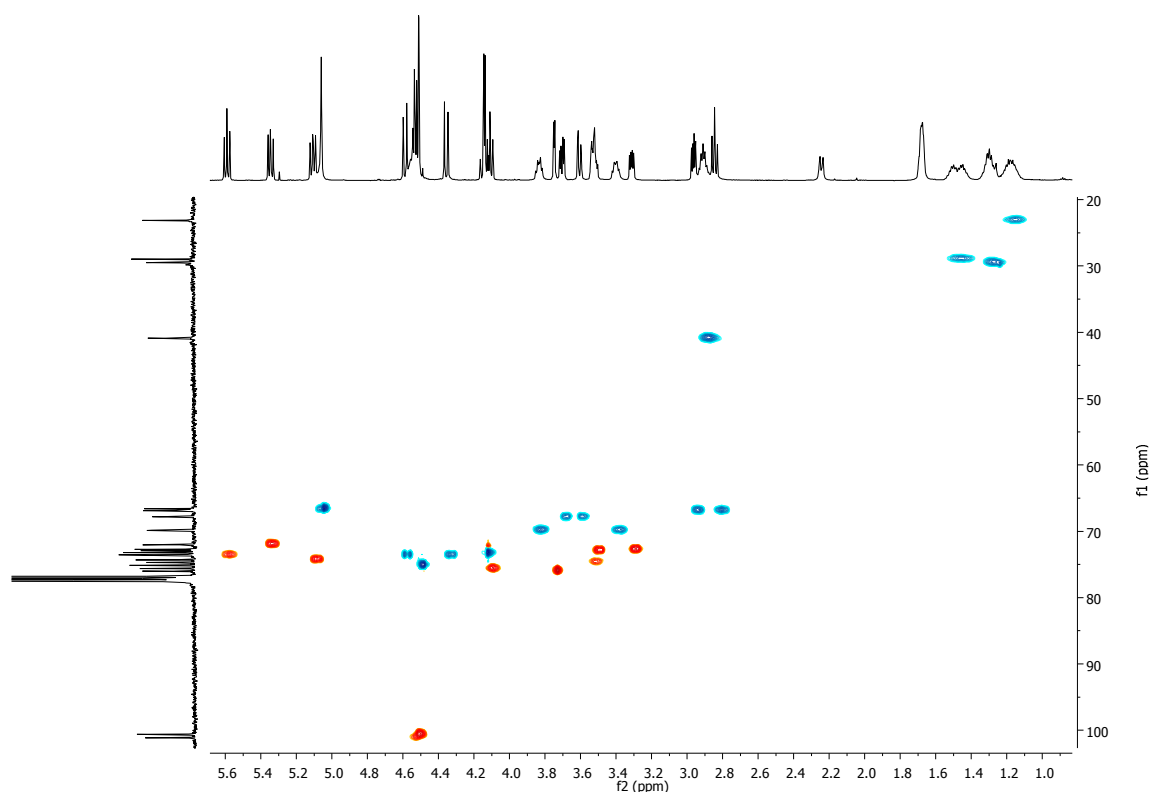
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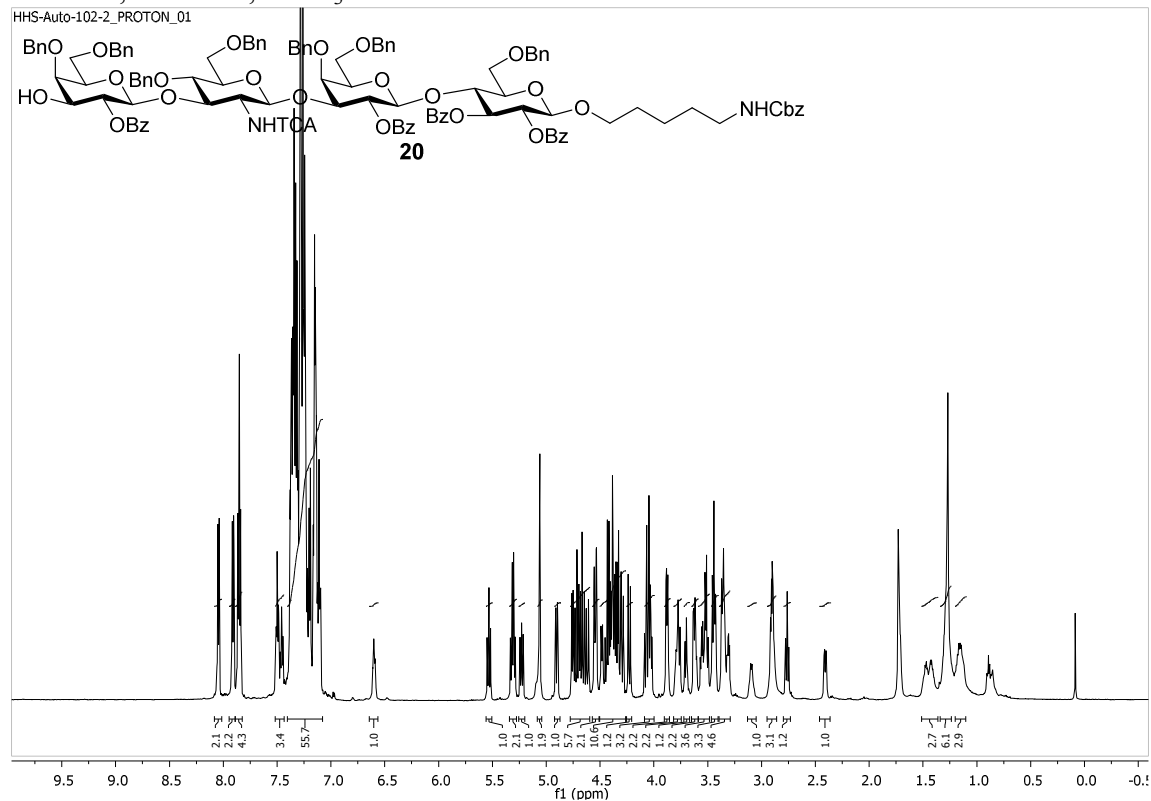
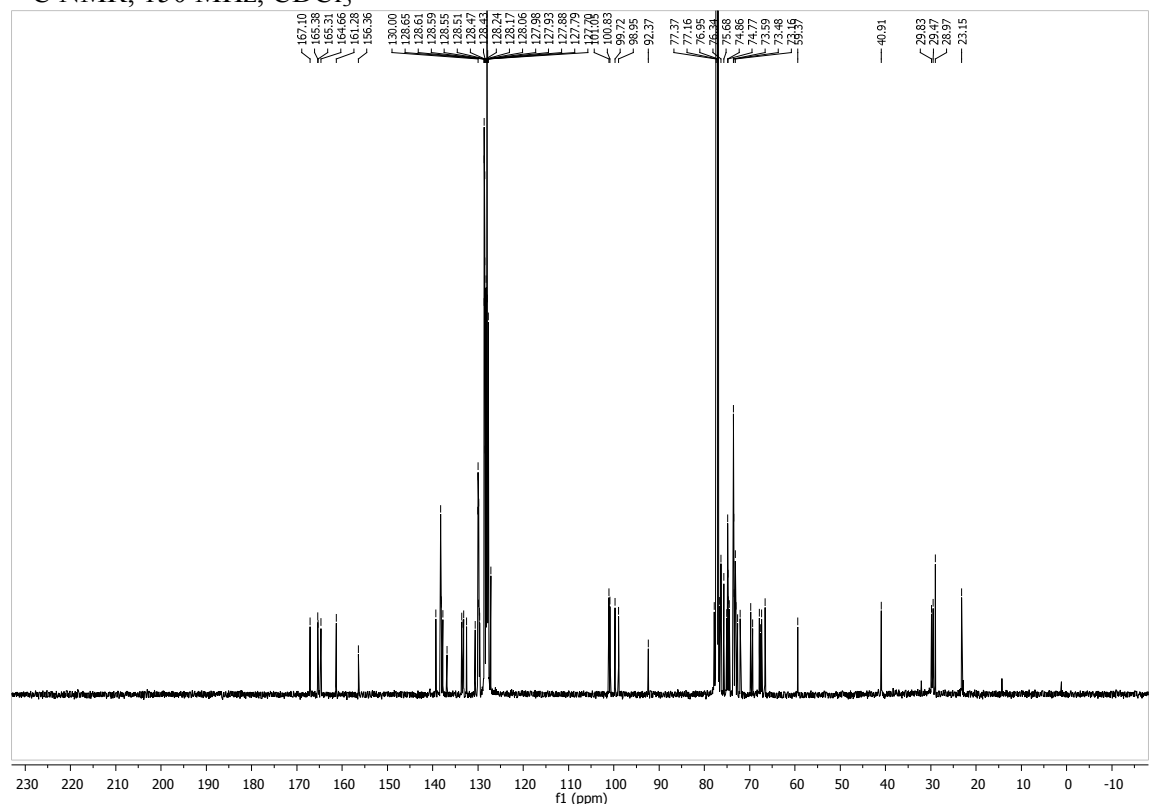
 ^{13}C NMR, 100 MHz, CDCl_3 

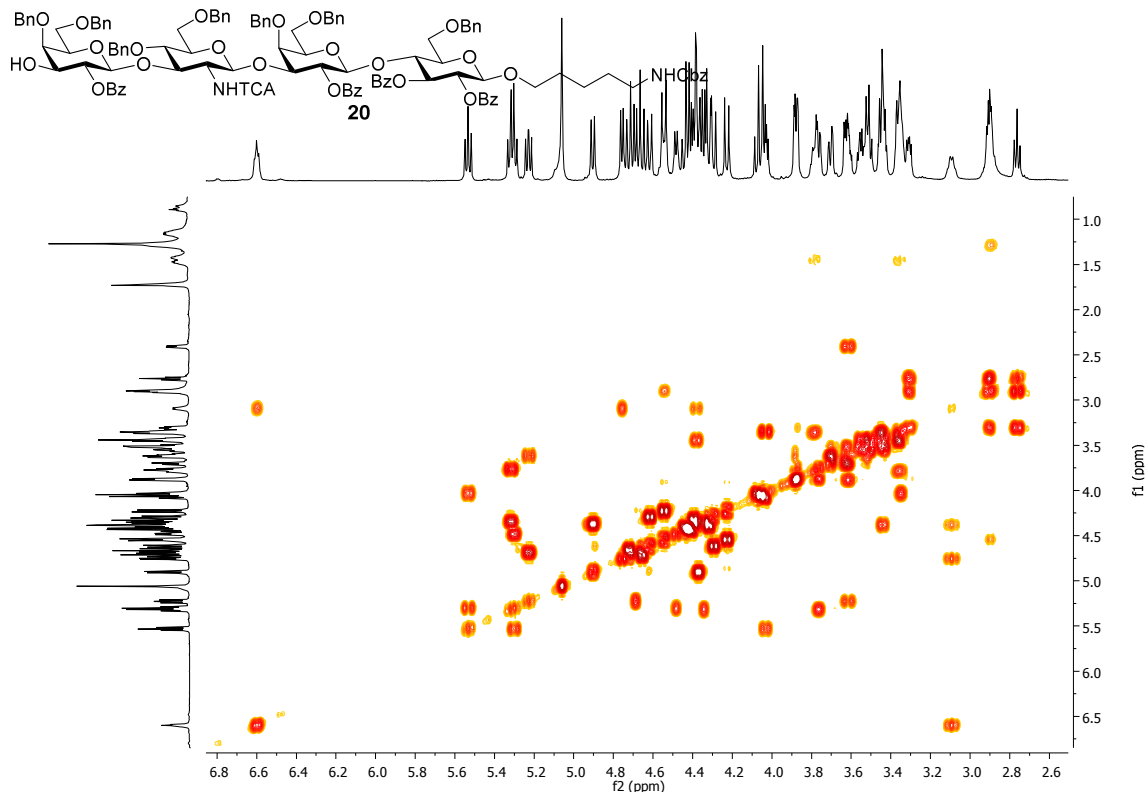
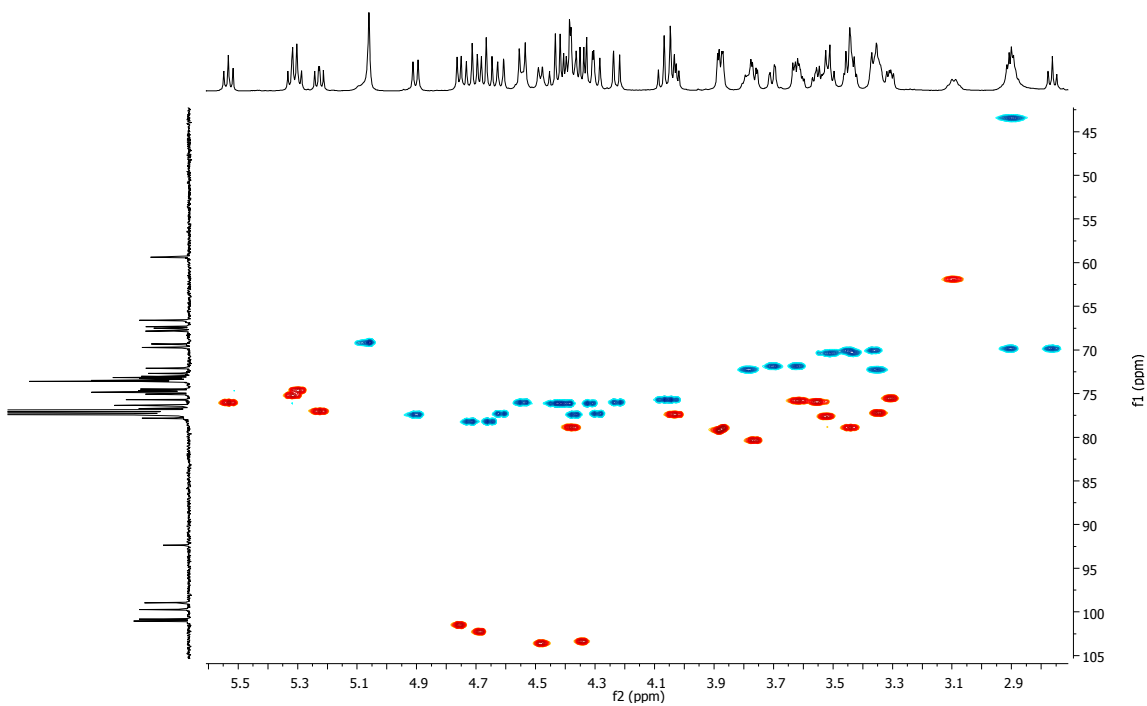
^1H -COSY NMR, 400 MHz, CDCl_3



^1H - ^{13}C -HSQC NMR, 400 MHz, CDCl_3

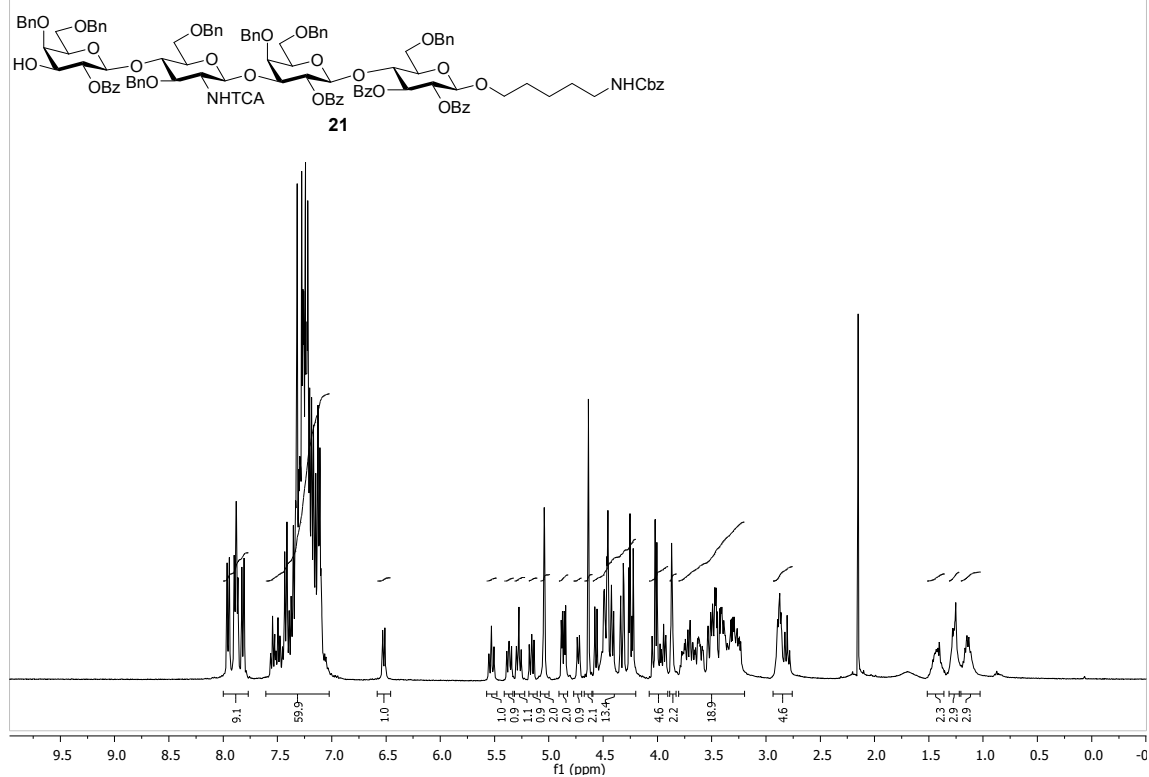
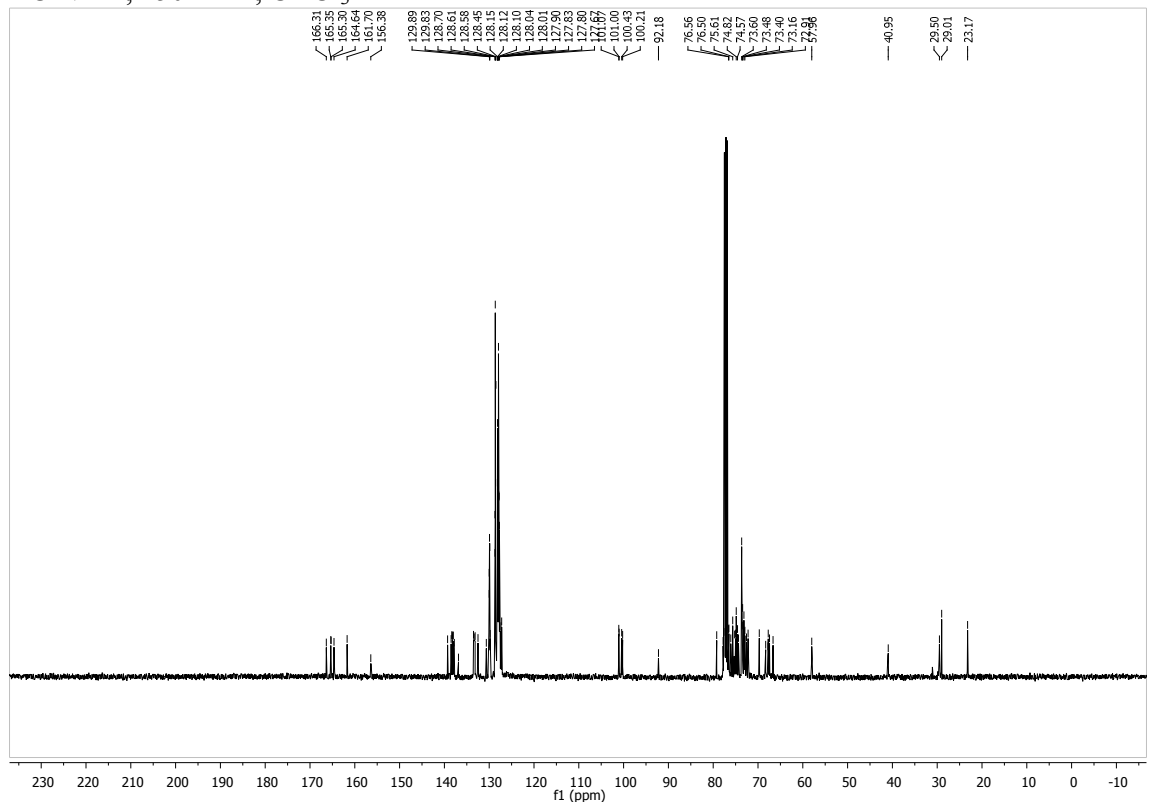


¹H NMR, 600 MHz, CDCl₃¹³C NMR, 150 MHz, CDCl₃

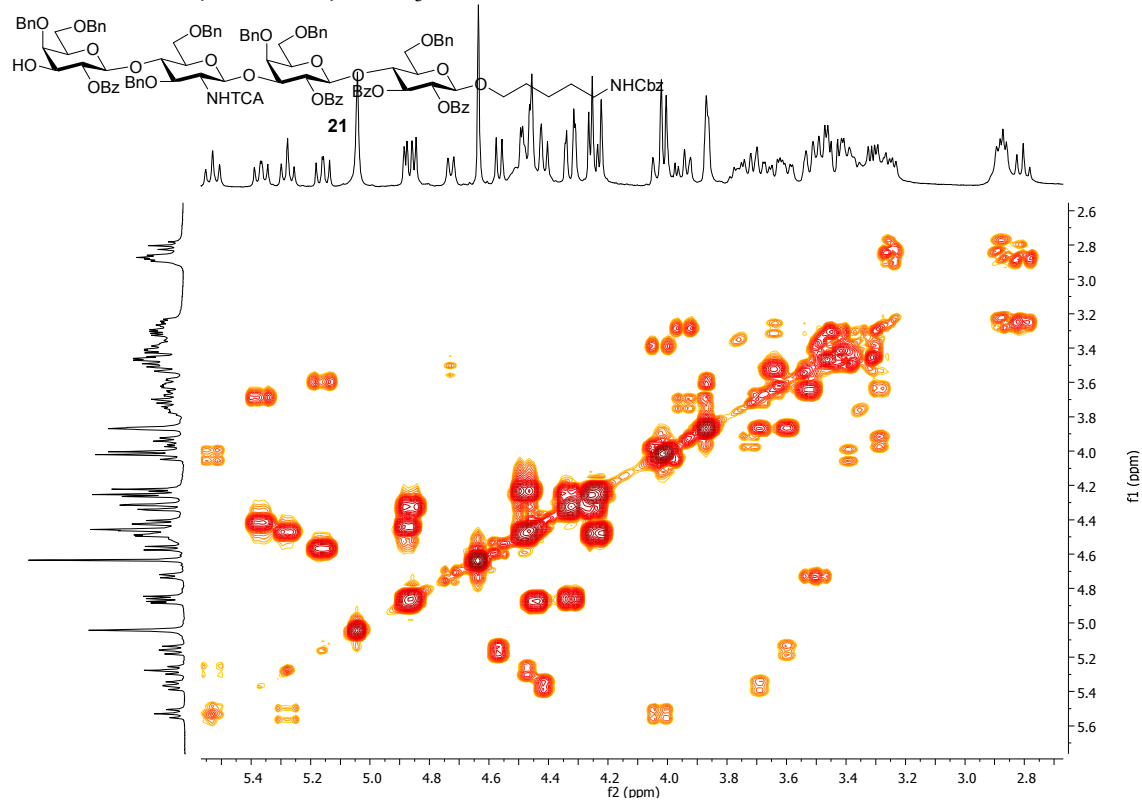
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^1H NMR, 600 MHz, CDCl_3

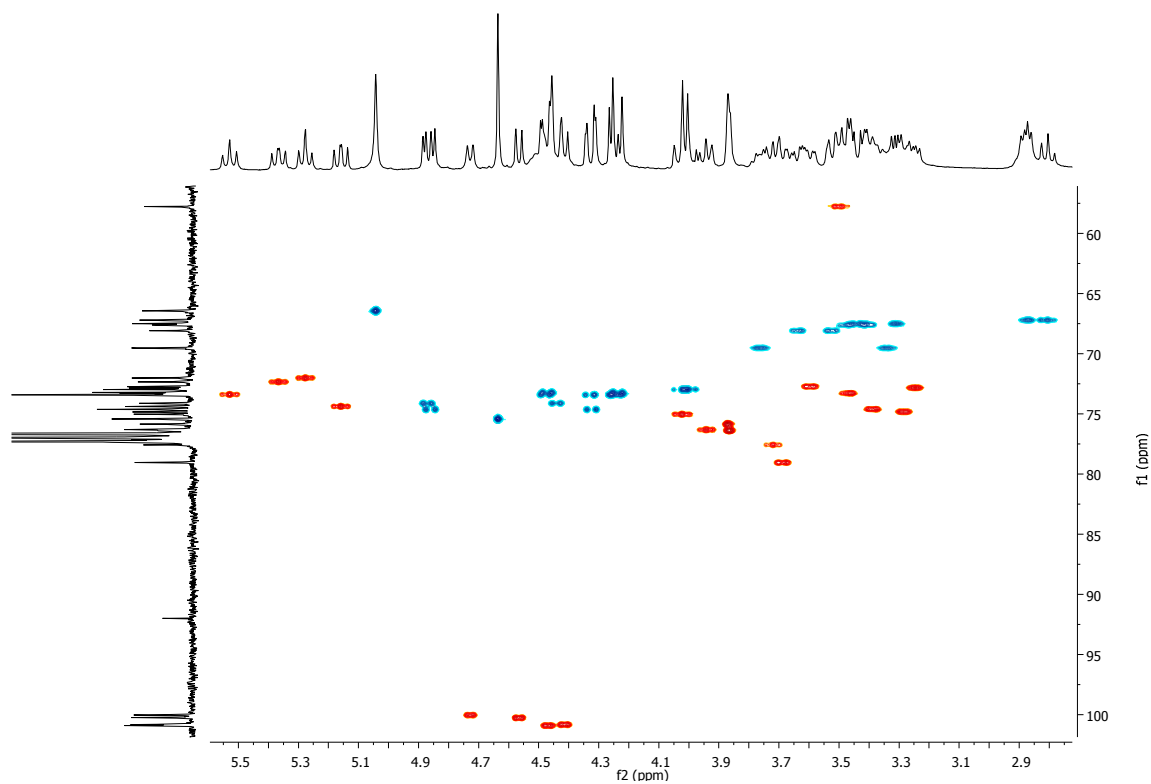
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 ^{13}C NMR, 150 MHz, CDCl_3 

^1H -COSY NMR, 600 MHz, CDCl_3

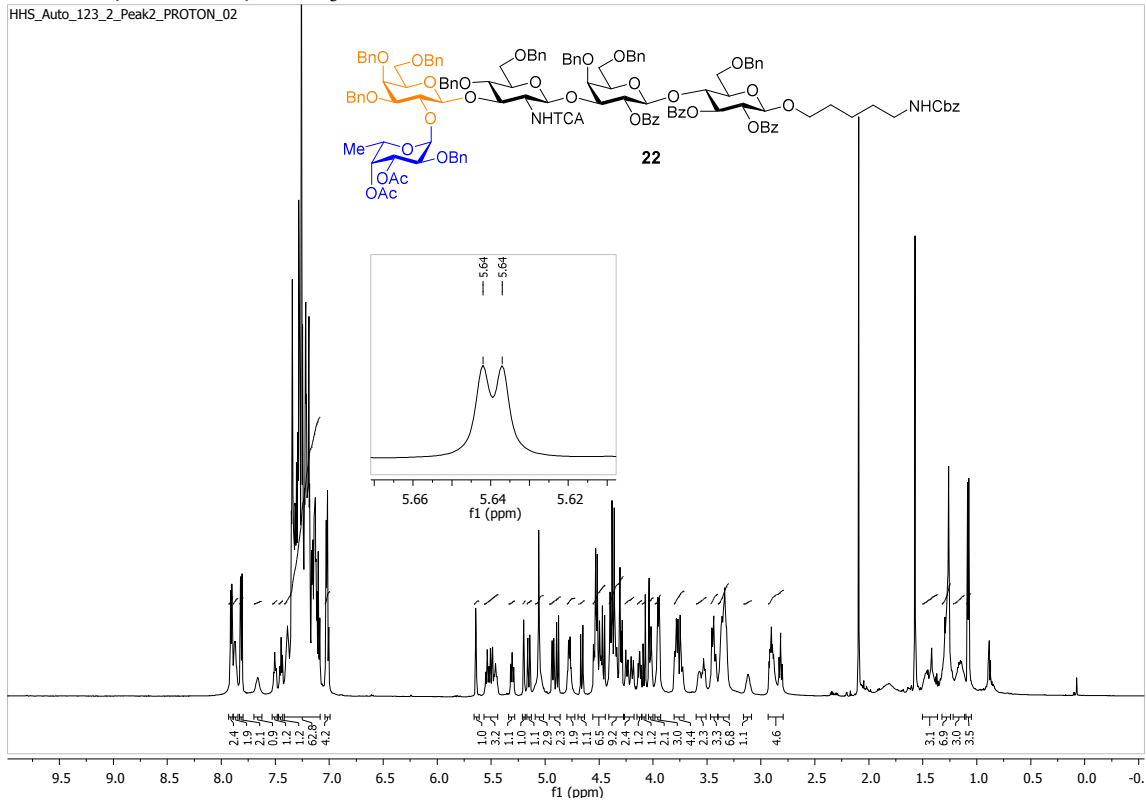
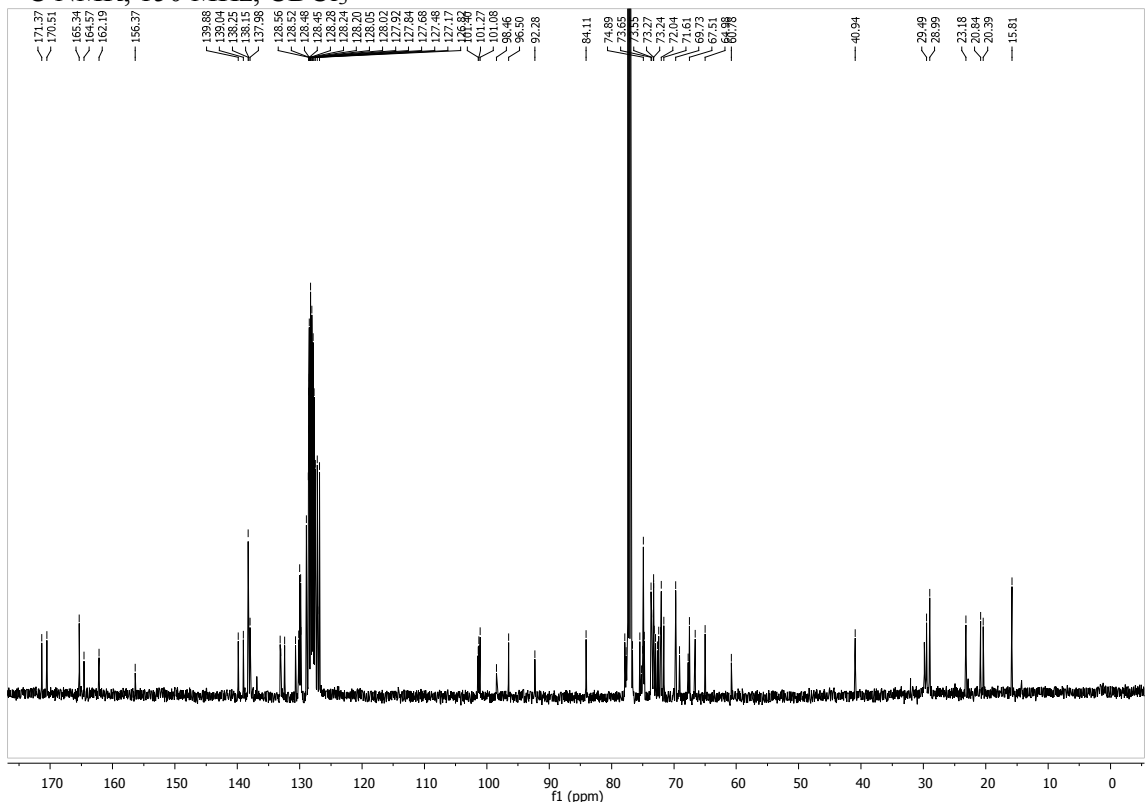


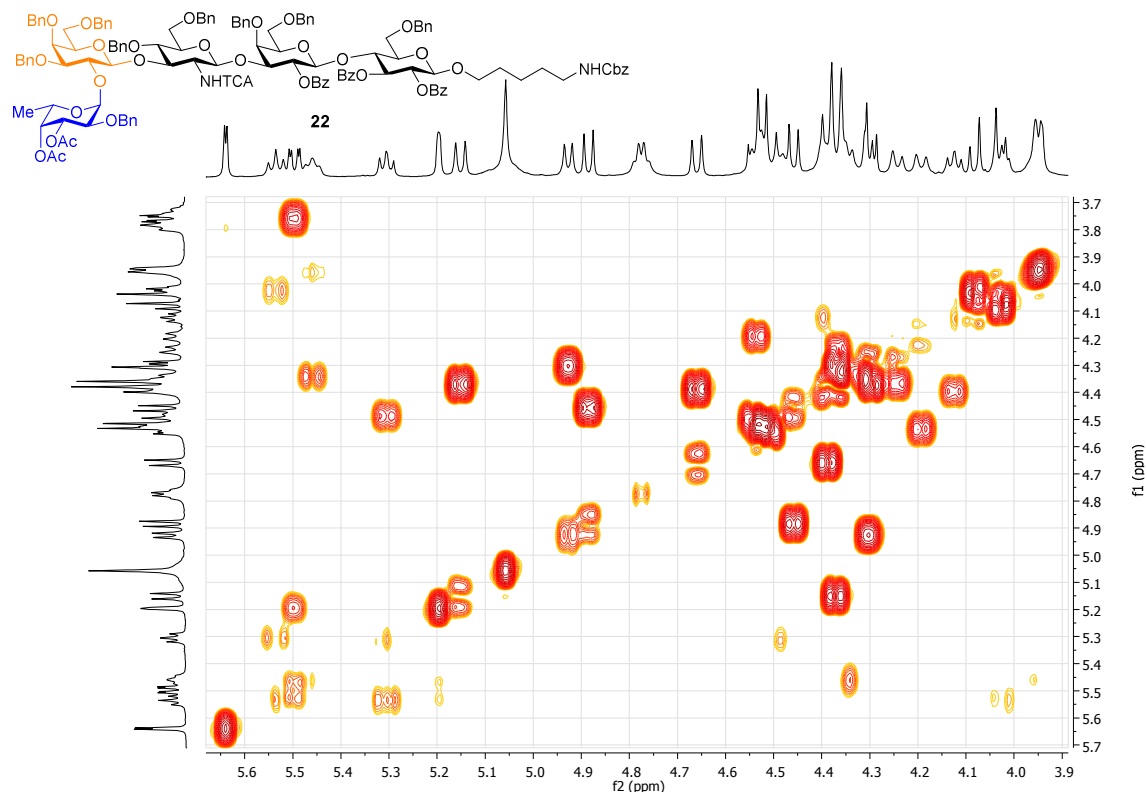
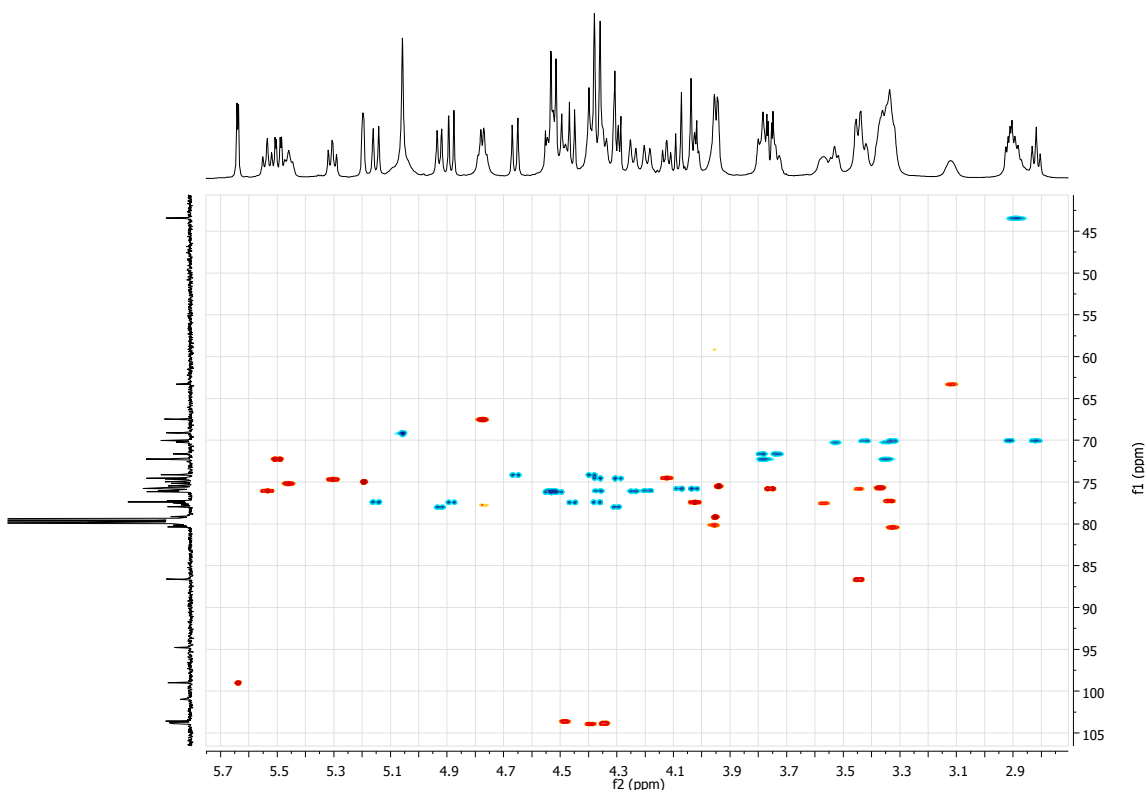
^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3



^1H NMR, 600 MHz, CDCl_3

HHS_Auto_123_2_Peak2_PROTON_02

 ^{13}C NMR, 150 MHz, CDCl_3 

^1H -COSY NMR, 600 MHz, CDCl_3  ^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3 

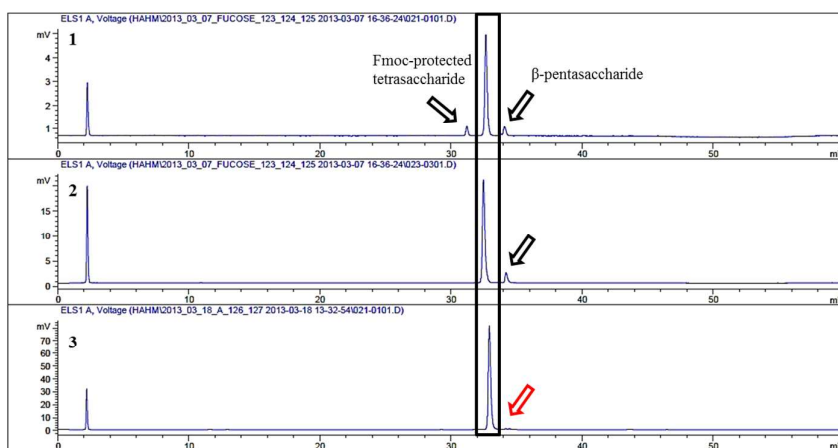
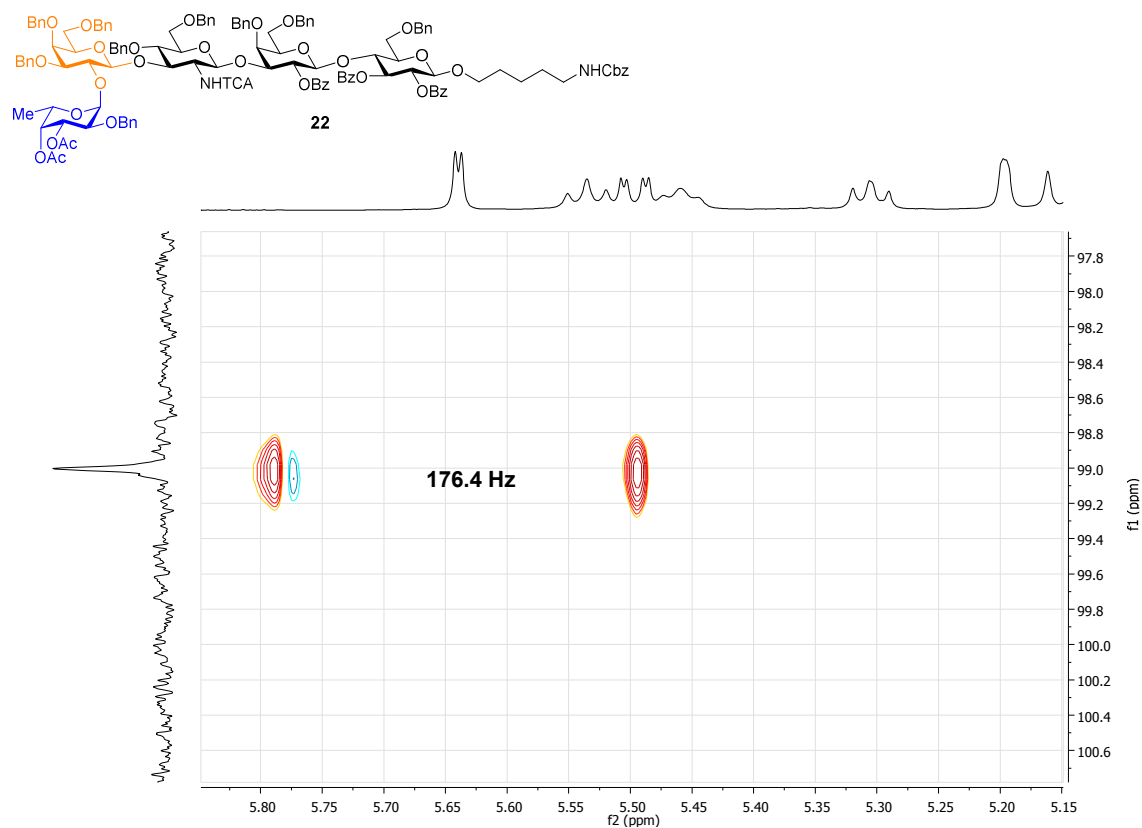
^1H - ^{13}C -coupled-HSQC NMR, 600 MHz, CDCl_3 

Figure S2. LC-MS of pentasaccharide **22**. Condition: **17** dissolved in DCM and Et_2O (v/v, 1/3) for entries 1 and 2. **17** dissolved in DCM for entry 3.

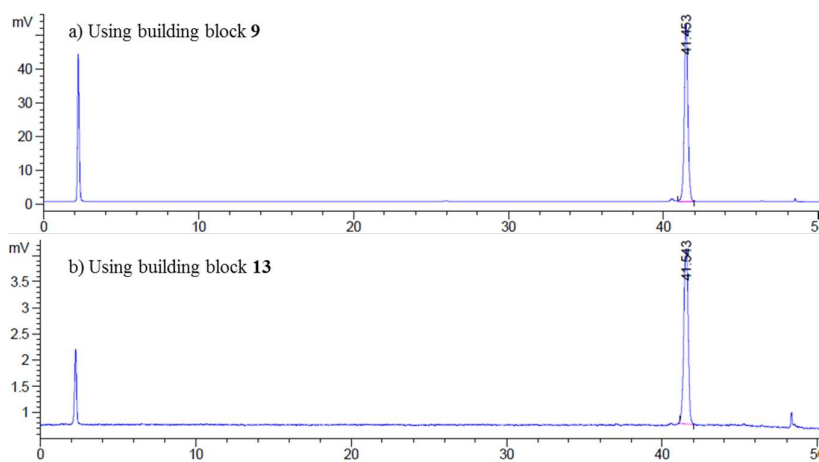


Figure S3. LC-MS of H-type II using fucose building block 17 and 18.

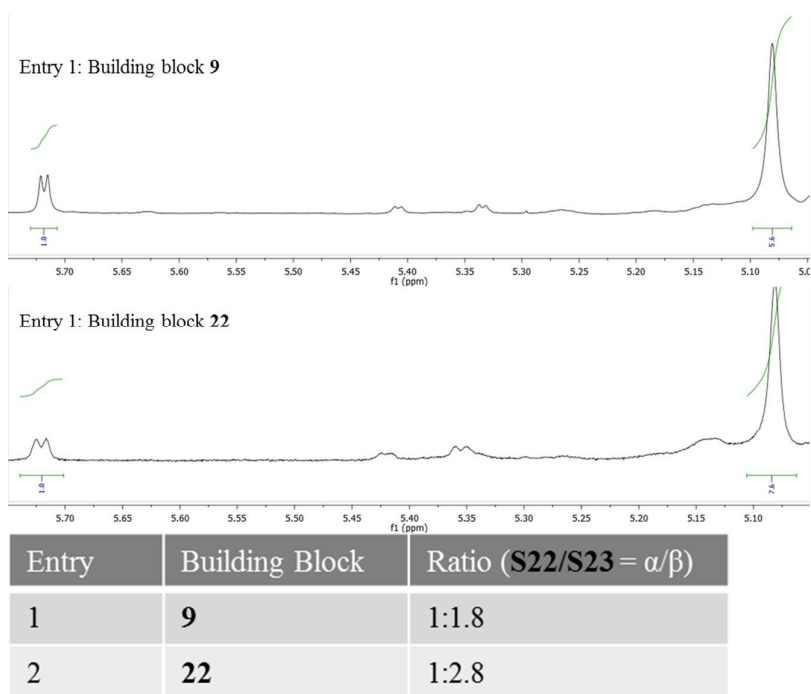


Figure S5. Stereoselectivity of H-type II determined by ^1H NMR.

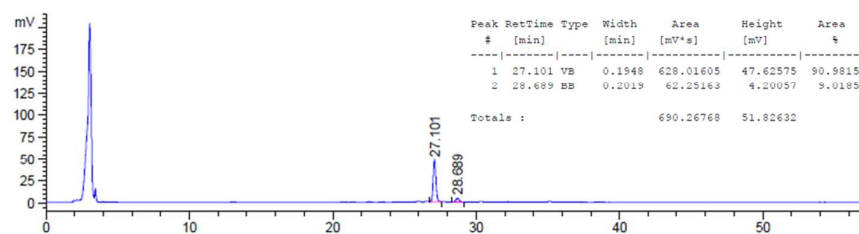
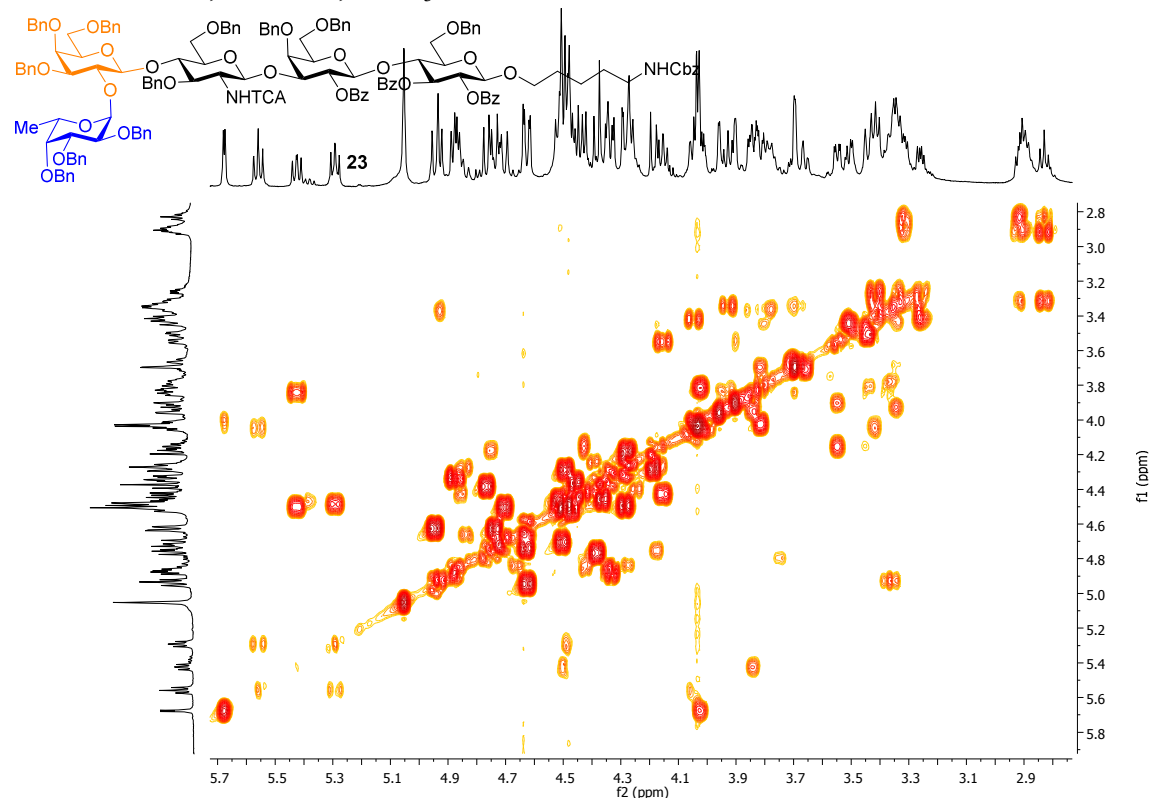
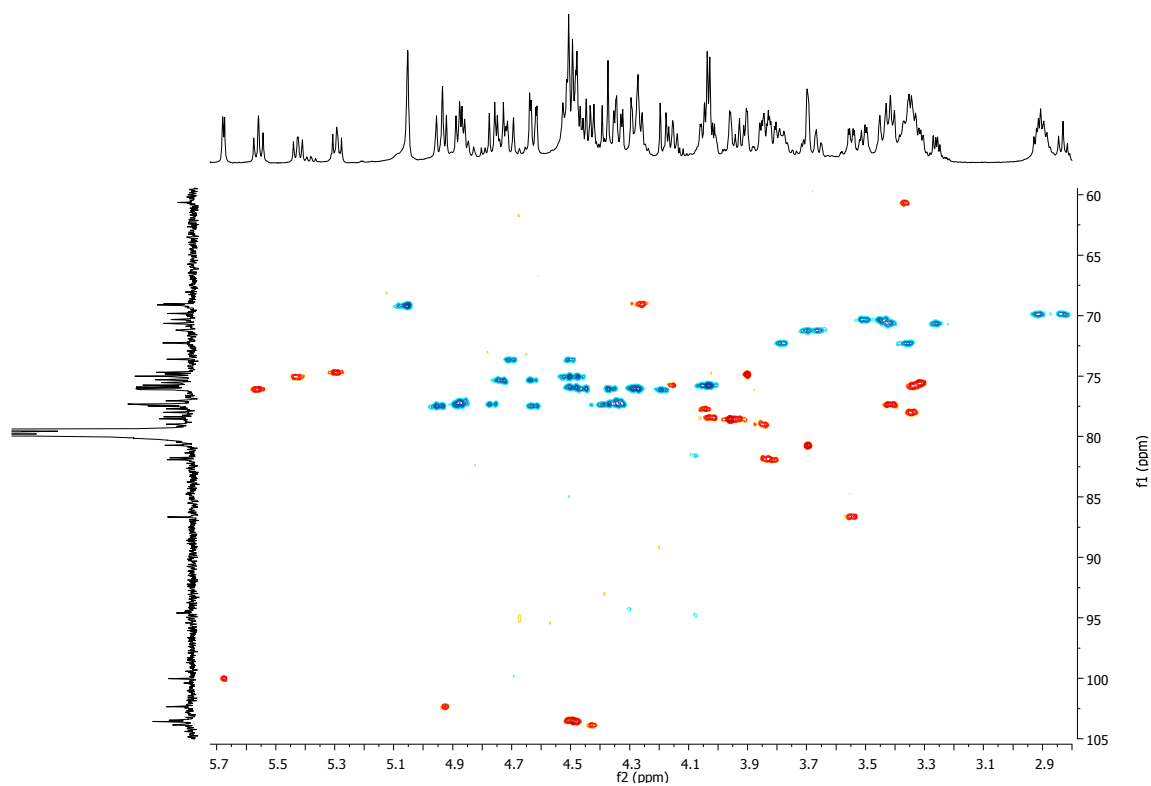
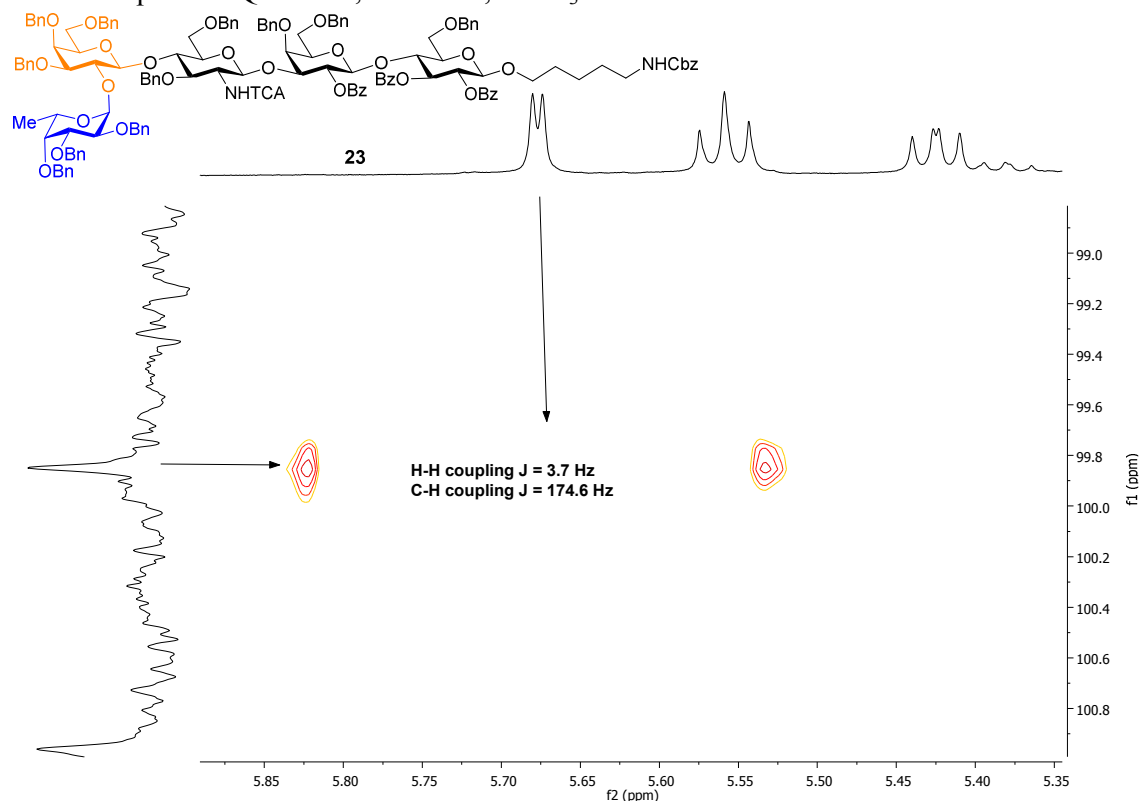
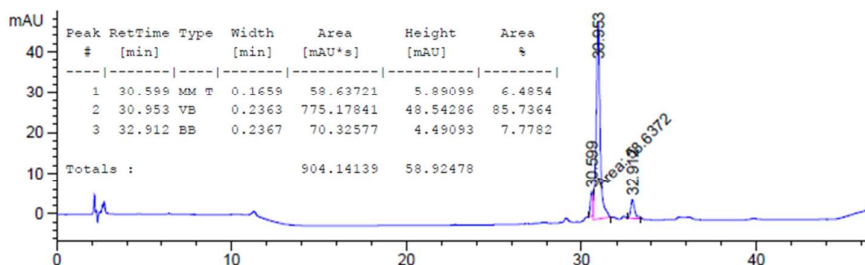


Figure S6. LC-MS of 23.

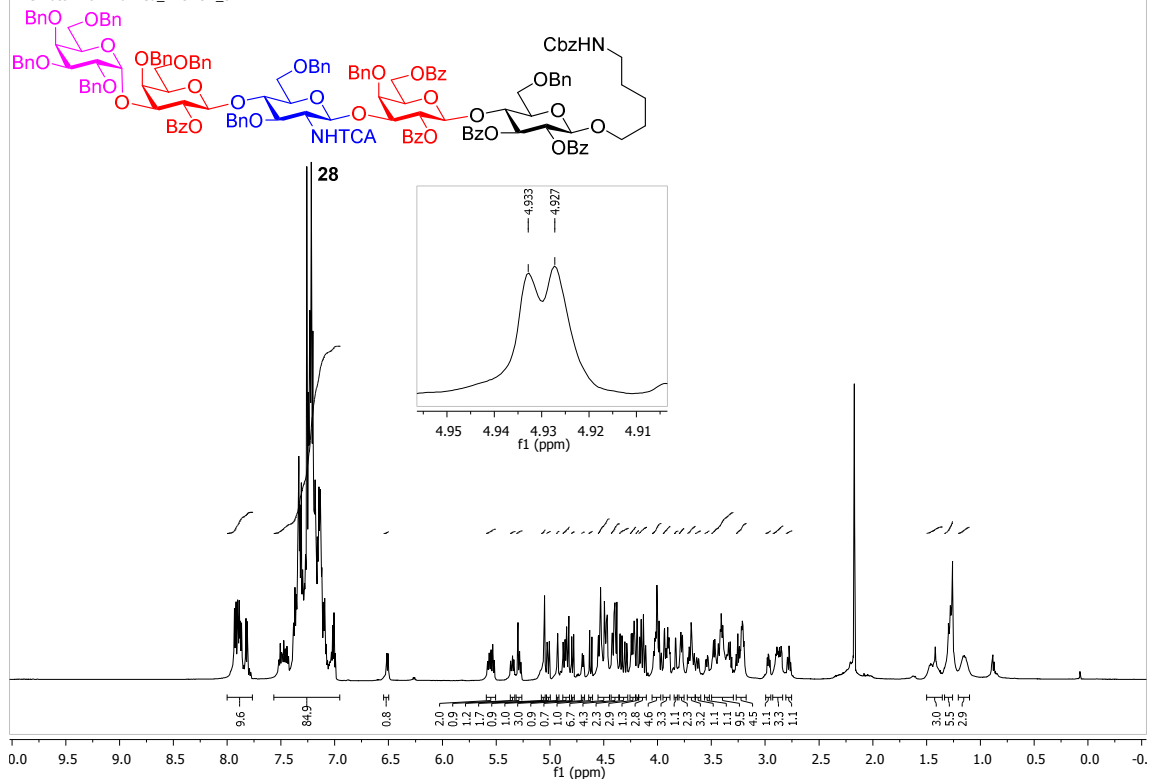
^1H -COSY NMR, 600 MHz, CDCl_3  ^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3 

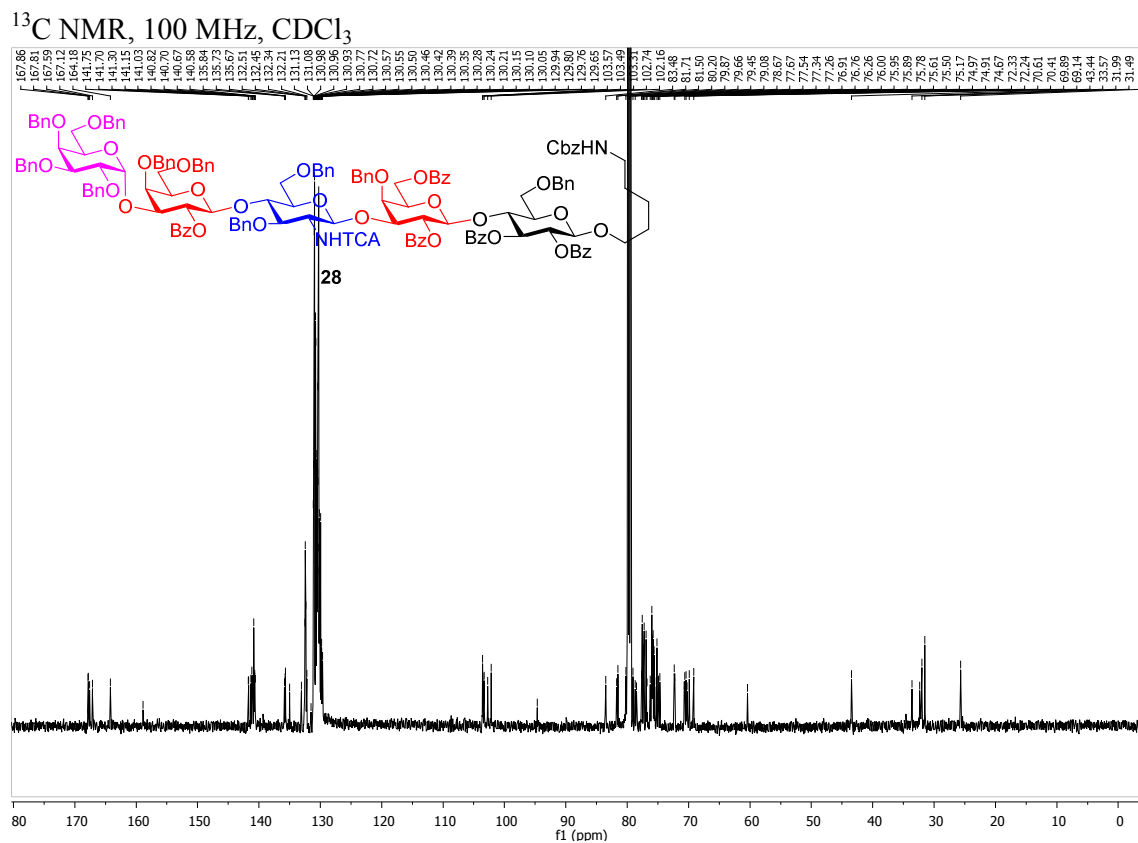
^1H - ^{13}C -coupled-HSQC NMR, 600 MHz, CDCl_3



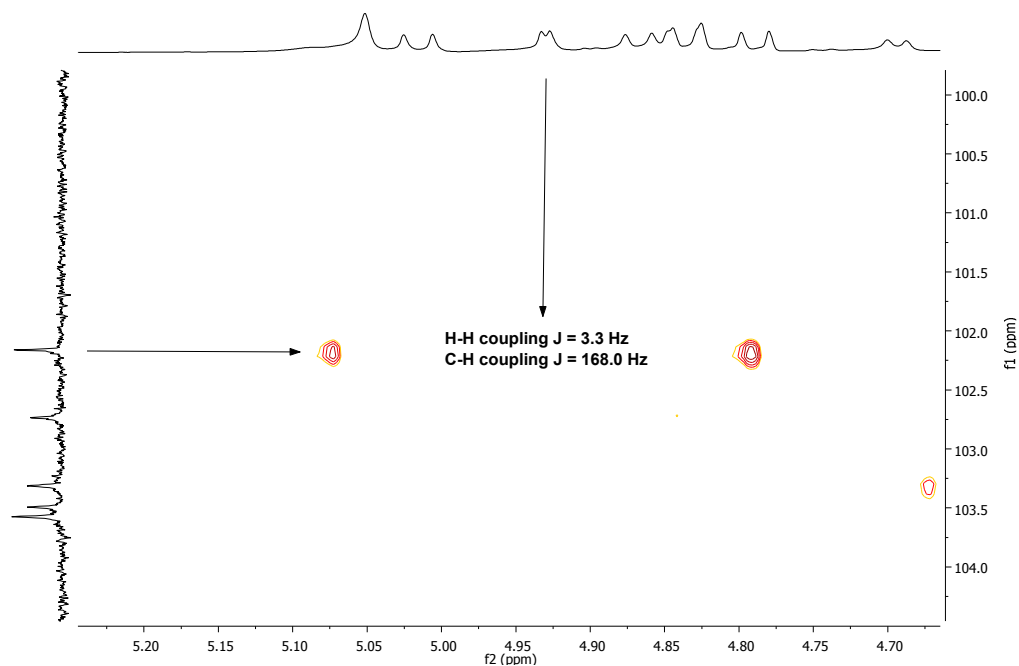
**Figure S7. LC-MS of 28.** ^1H NMR, 600 MHz, CDCl_3

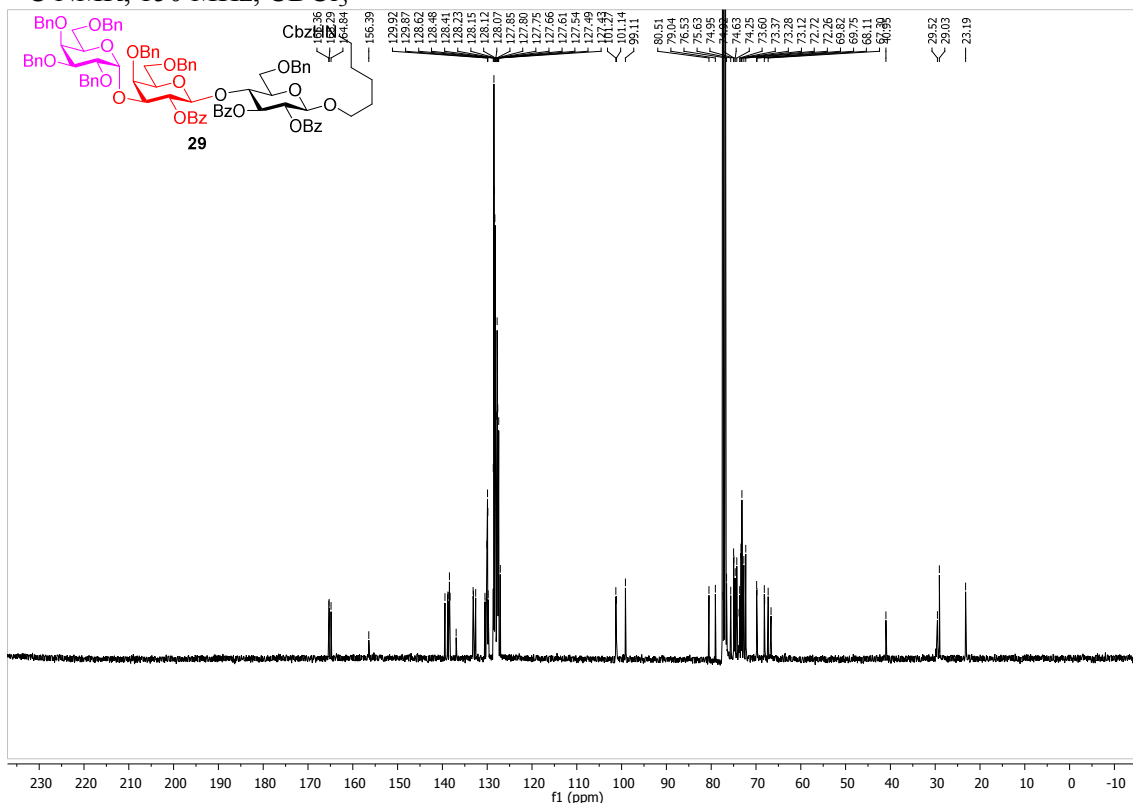
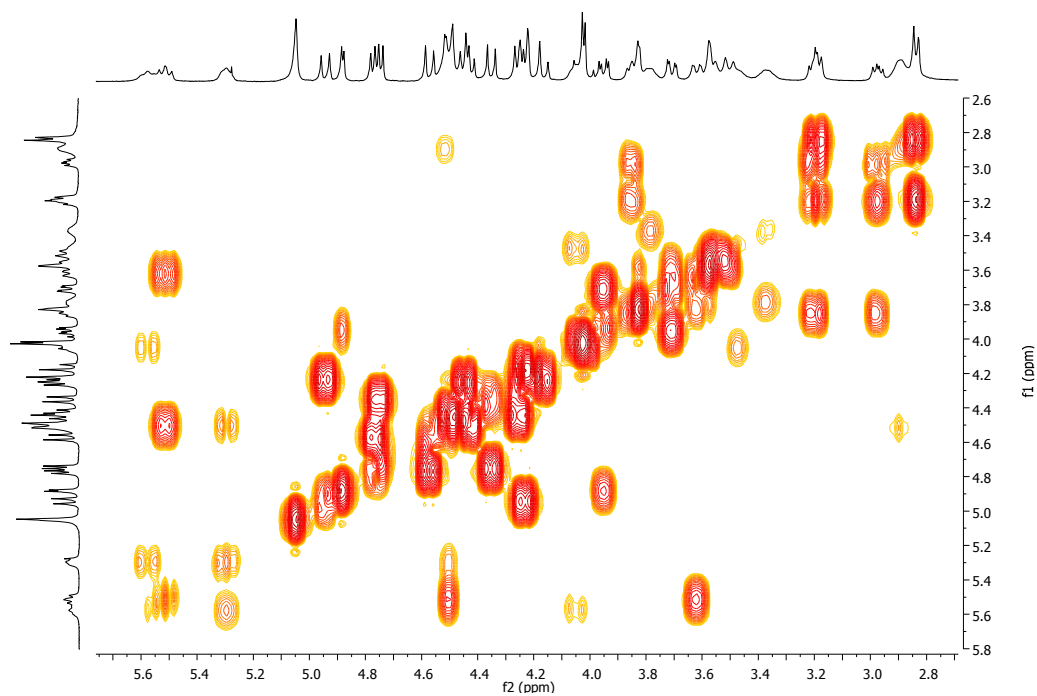
HHS-Auto-120-2-Purified_PROTON_01



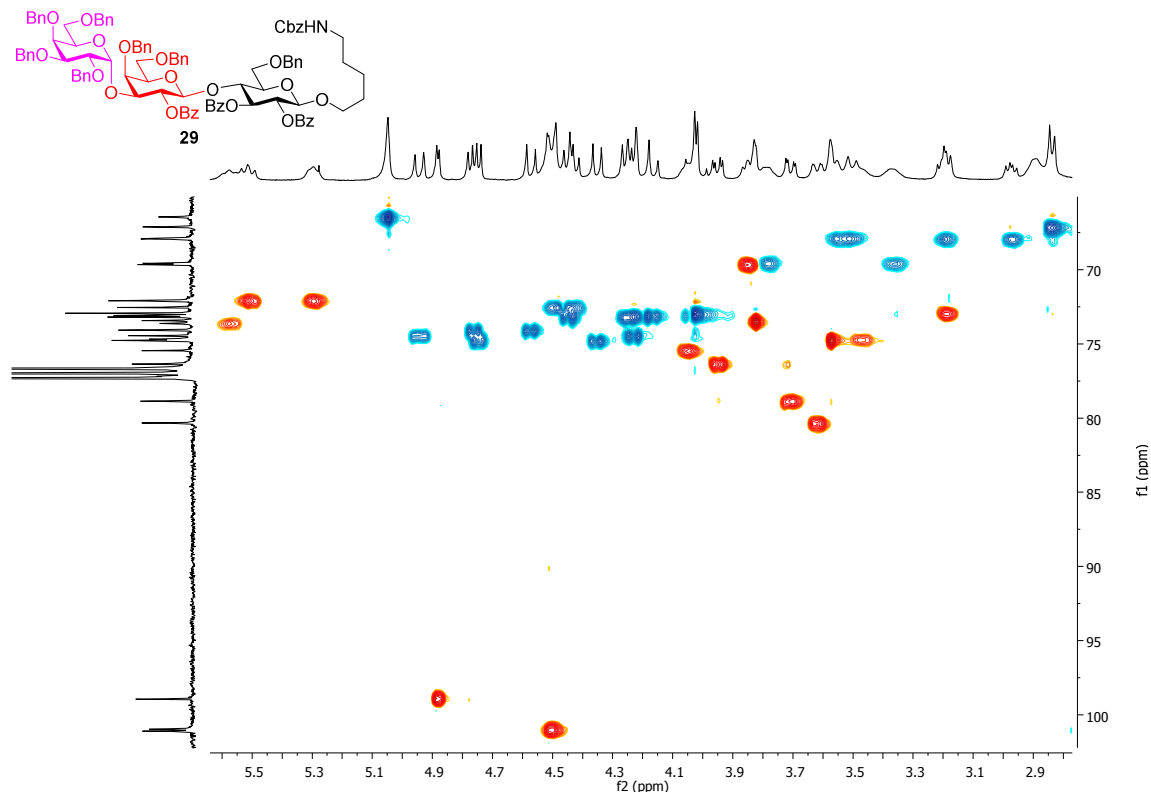


^1H - ^{13}C -coupled-HSQC NMR, 600 MHz, CDCl_3

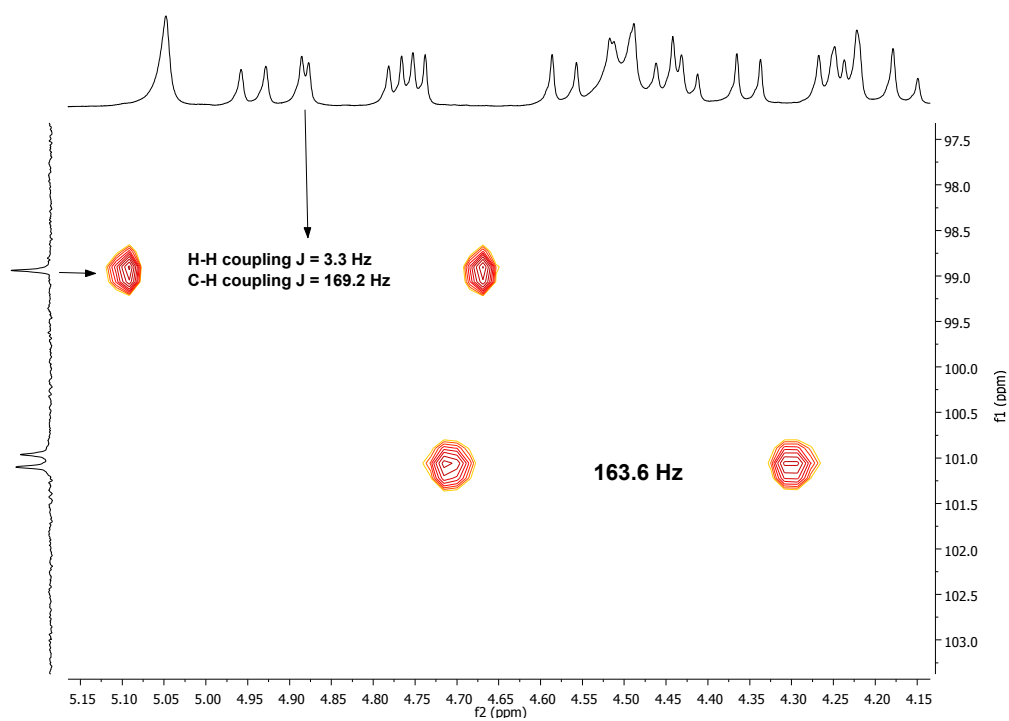


^{13}C NMR, 150 MHz, CDCl_3  ^1H -COSY NMR, 600 MHz, CDCl_3 

^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3



^1H - ^{13}C -coupled-HSQC NMR, 600 MHz, CDCl_3



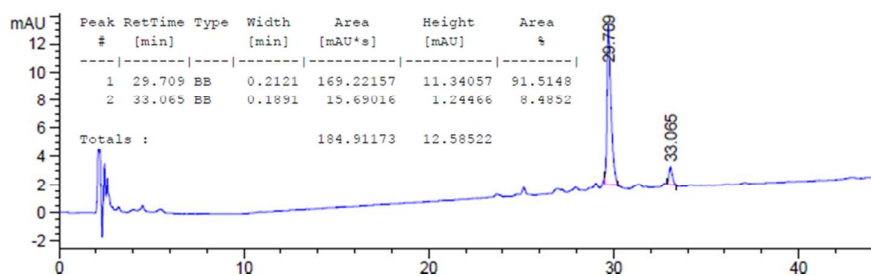
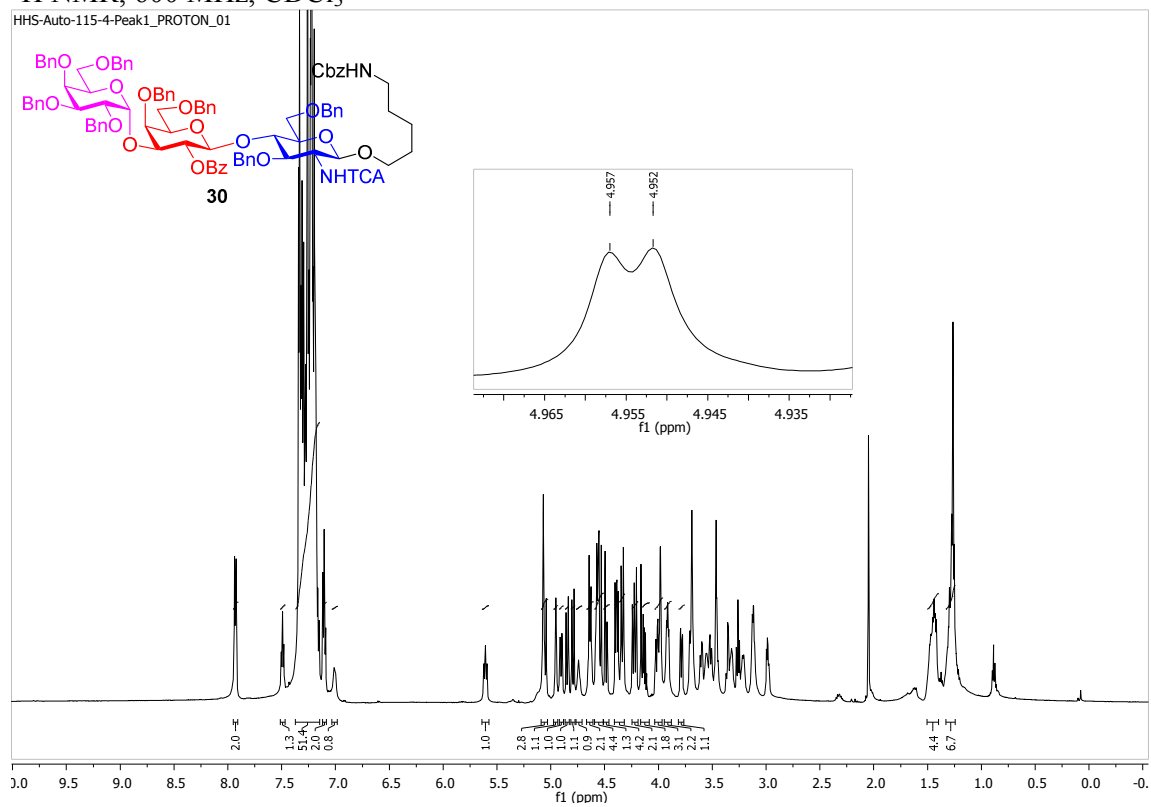
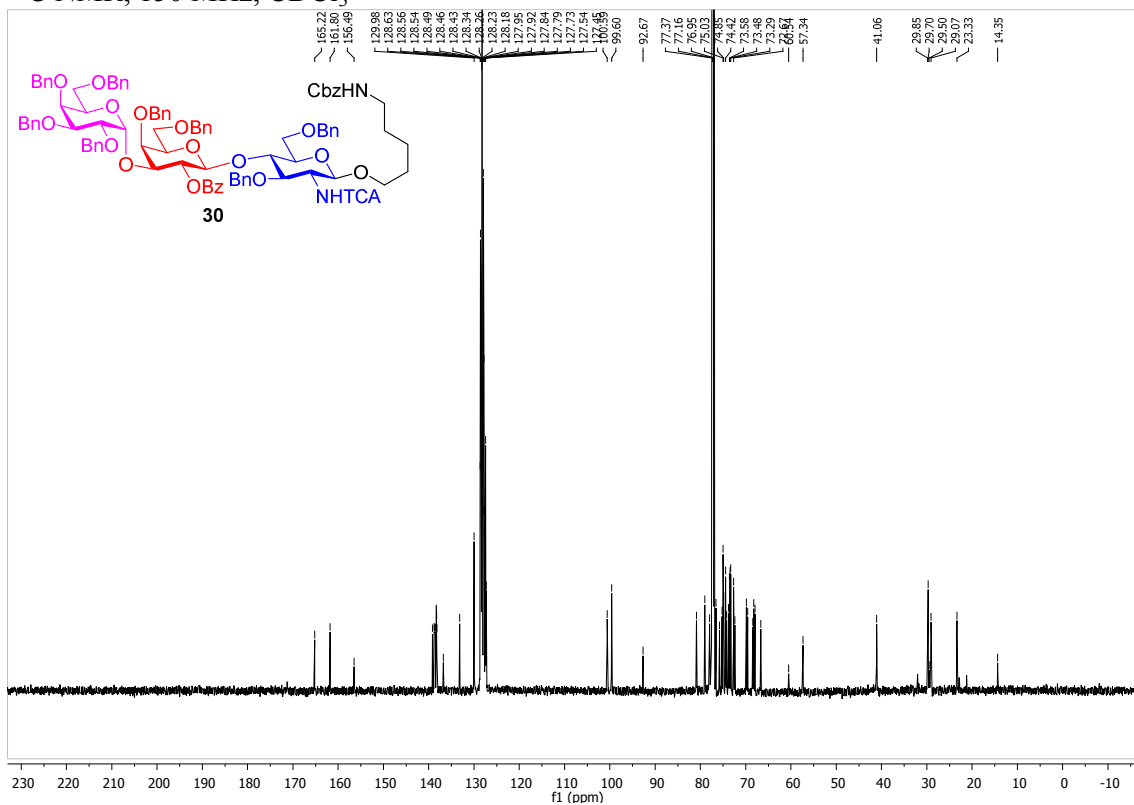
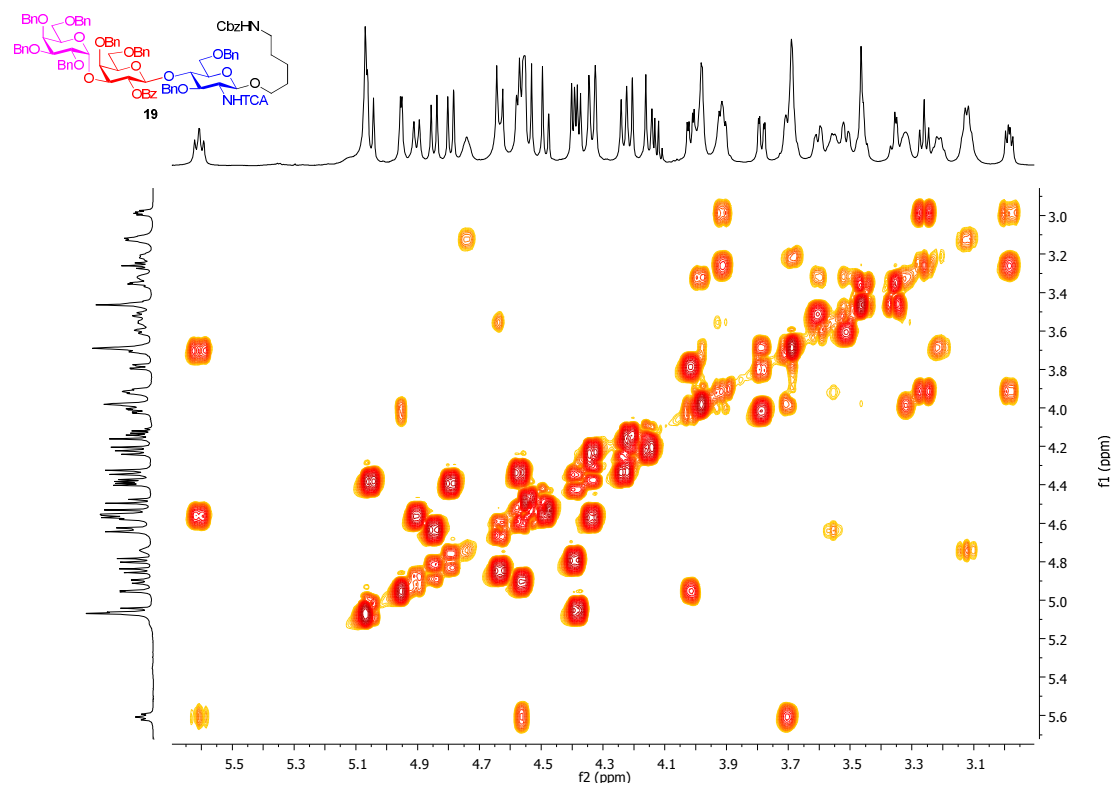
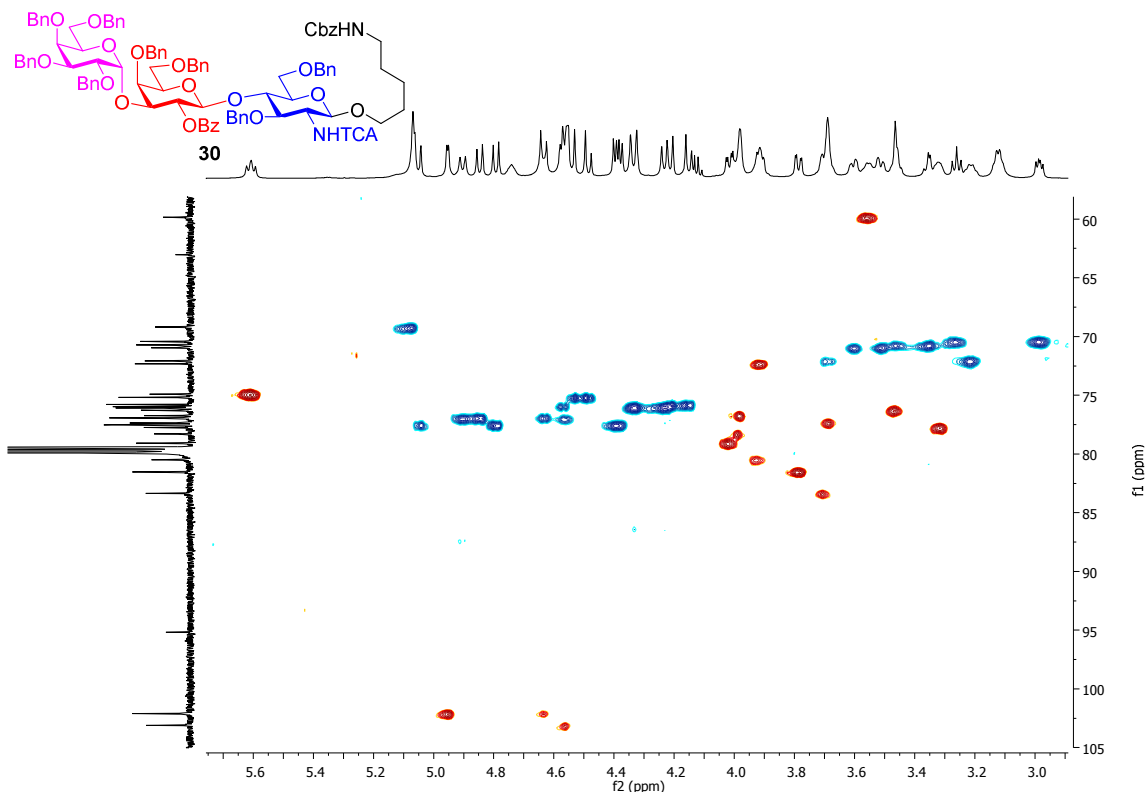
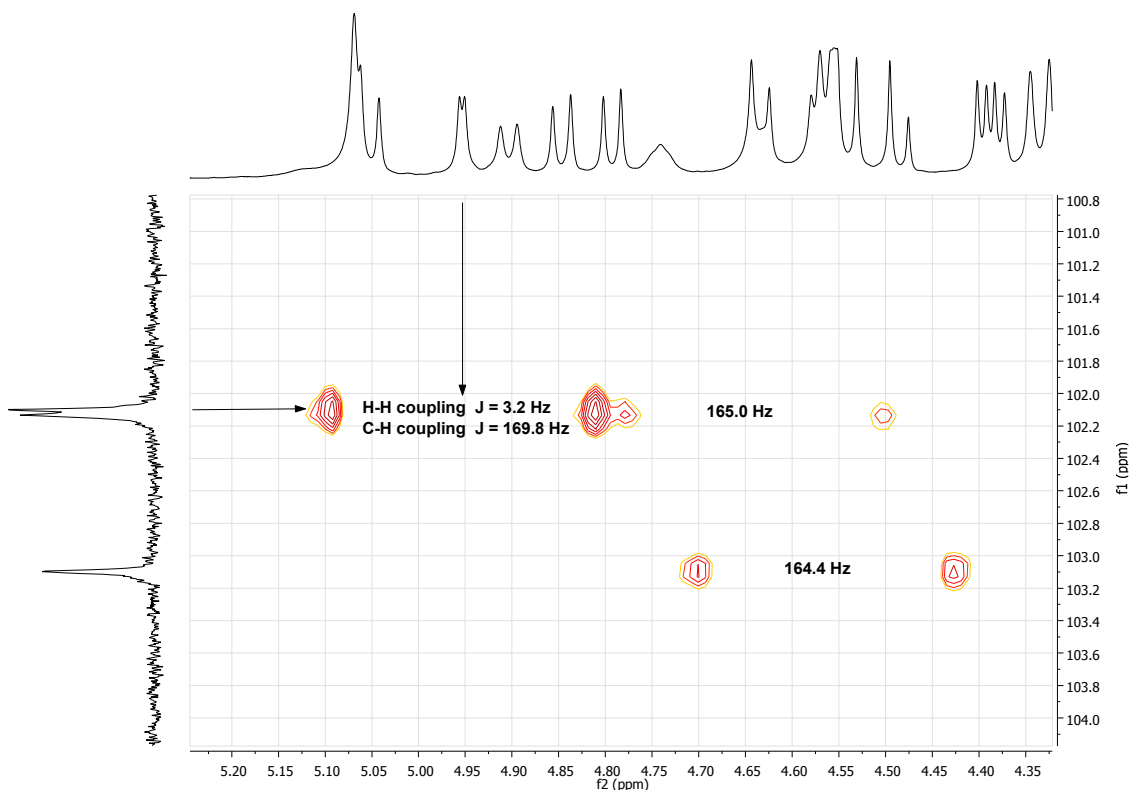


Figure S9. LC-MS of 30.

 ^1H NMR, 600 MHz, CDCl_3 

^{13}C NMR, 150 MHz, CDCl_3  ^1H -COSY NMR, 600 MHz, CDCl_3 

^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3  ^1H - ^{13}C -coupled-HSQC NMR, 600 MHz, CDCl_3 

Entry	Sequence	Ratio (α/β)
1	Gal α 1 \rightarrow 3Gal β 1 \rightarrow 4Glc β 1 \rightarrow linker	13.7
2	Gal α 1 \rightarrow 3Gal β 1 \rightarrow 4GlcNTCA β 1 \rightarrow linker	10.8
3	Gal α 1 \rightarrow 3Gal β 1 \rightarrow 4GlcNTCA β 1 \rightarrow 3Gal β 1 \rightarrow 4Glc β 1 \rightarrow linker	11.8

Table S2. The acceptor dependency of the formation of 1,2-*cis*-galactosidic linkages.

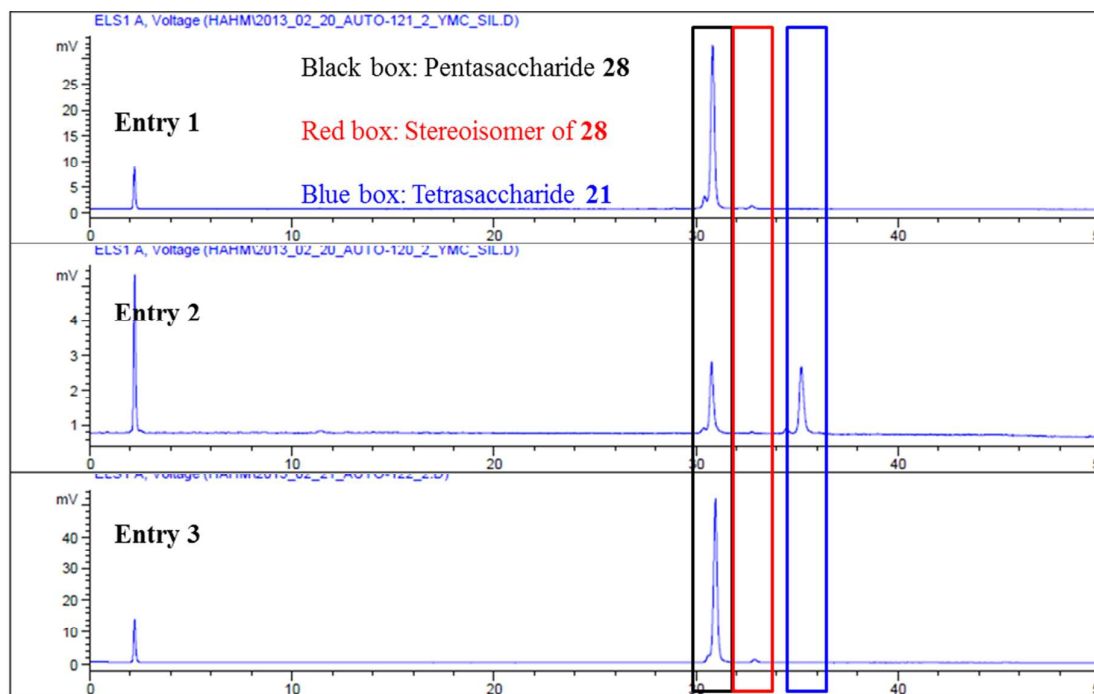


Figure S10. LC-MS of 28.

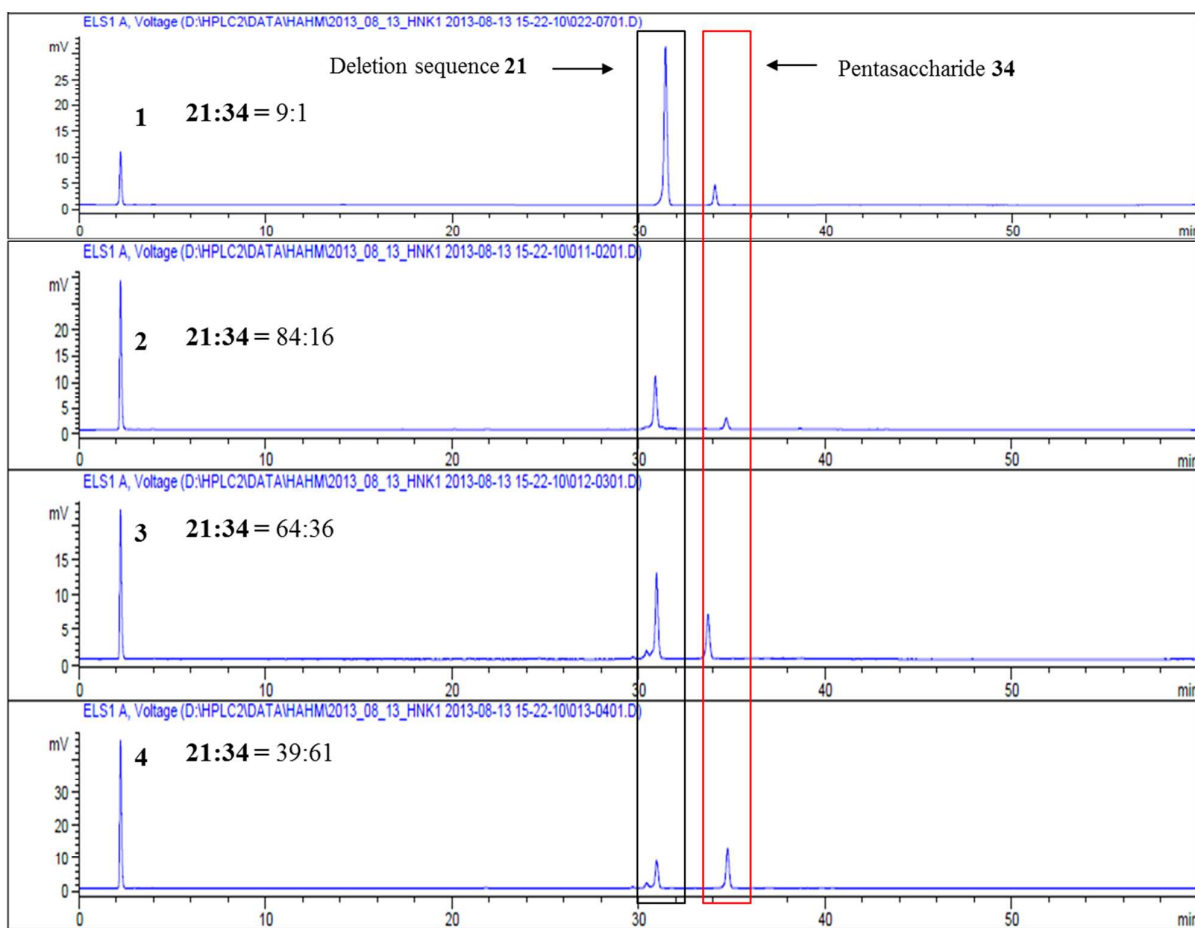


Figure S11. Optimization of automated synthesis of 34.

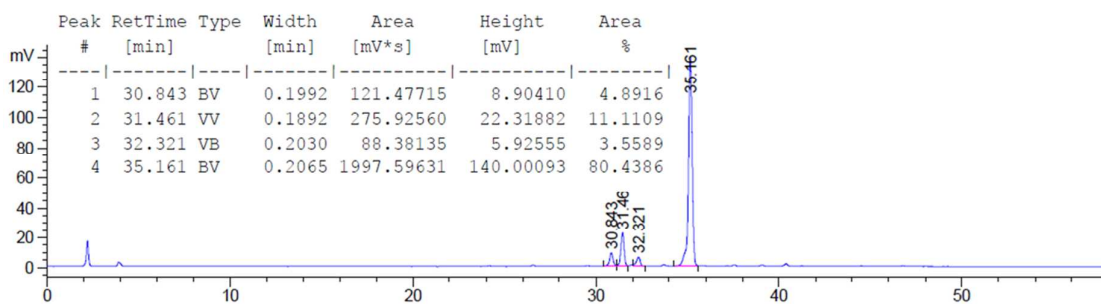
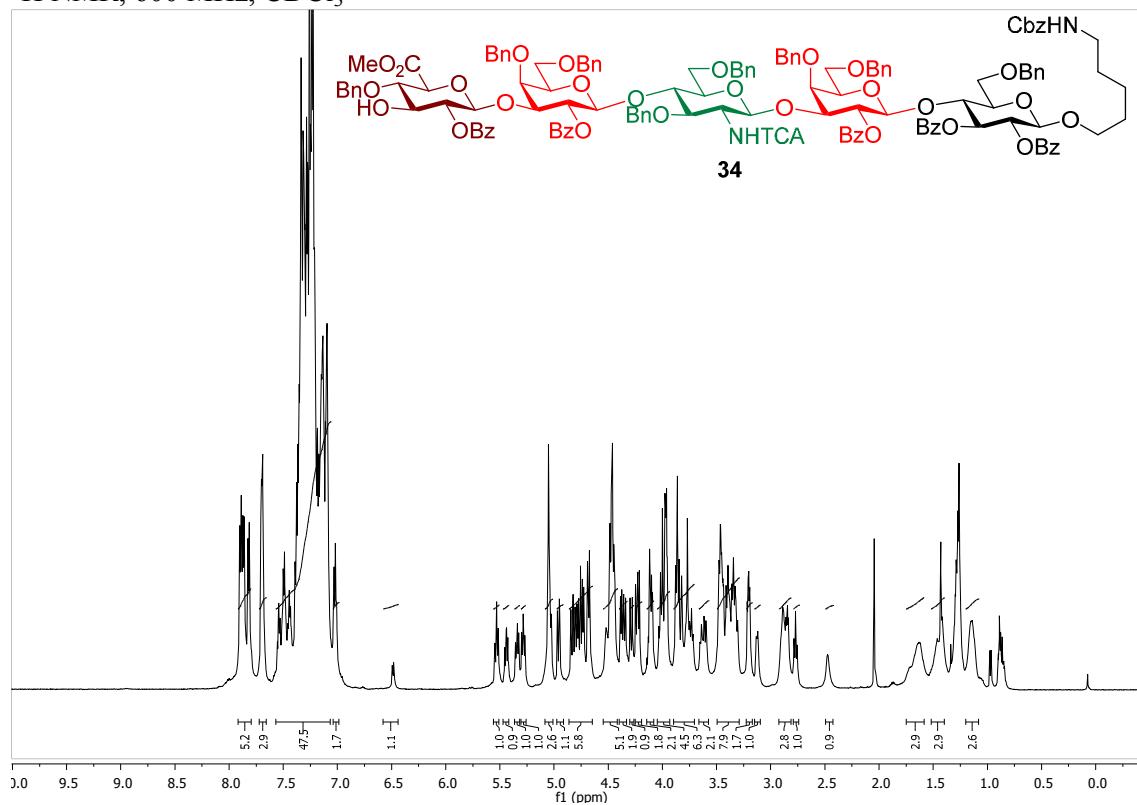
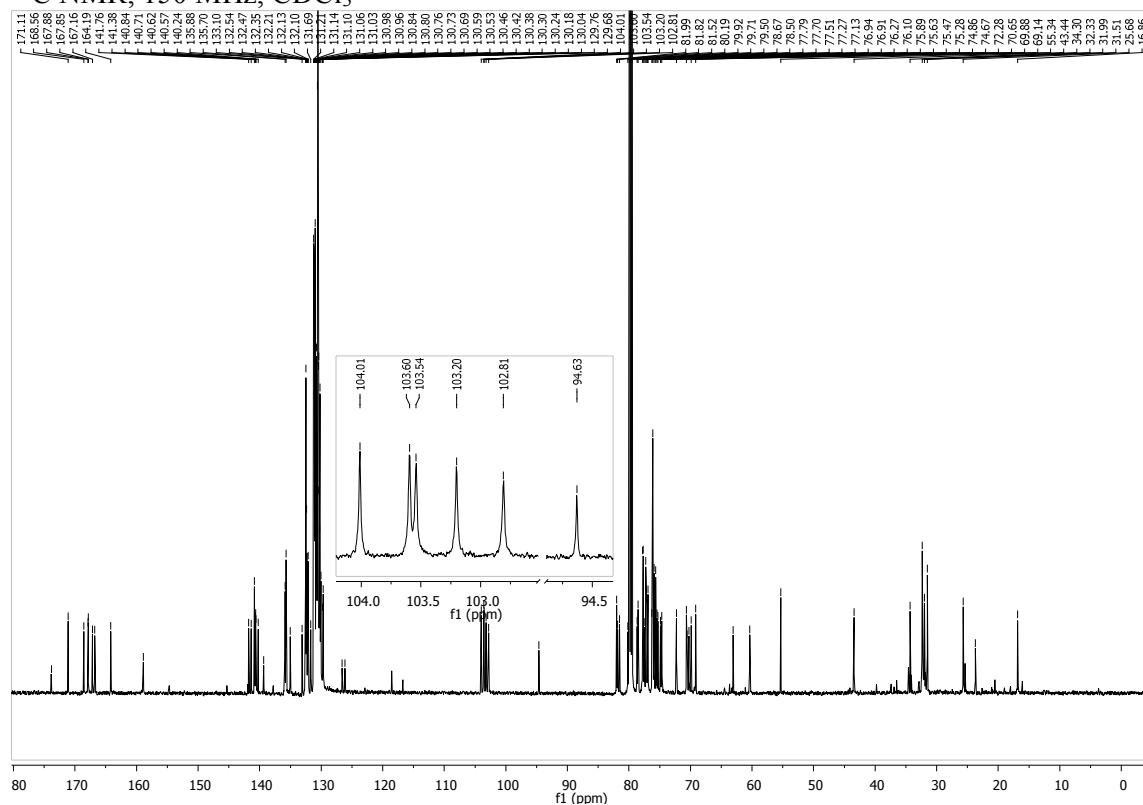
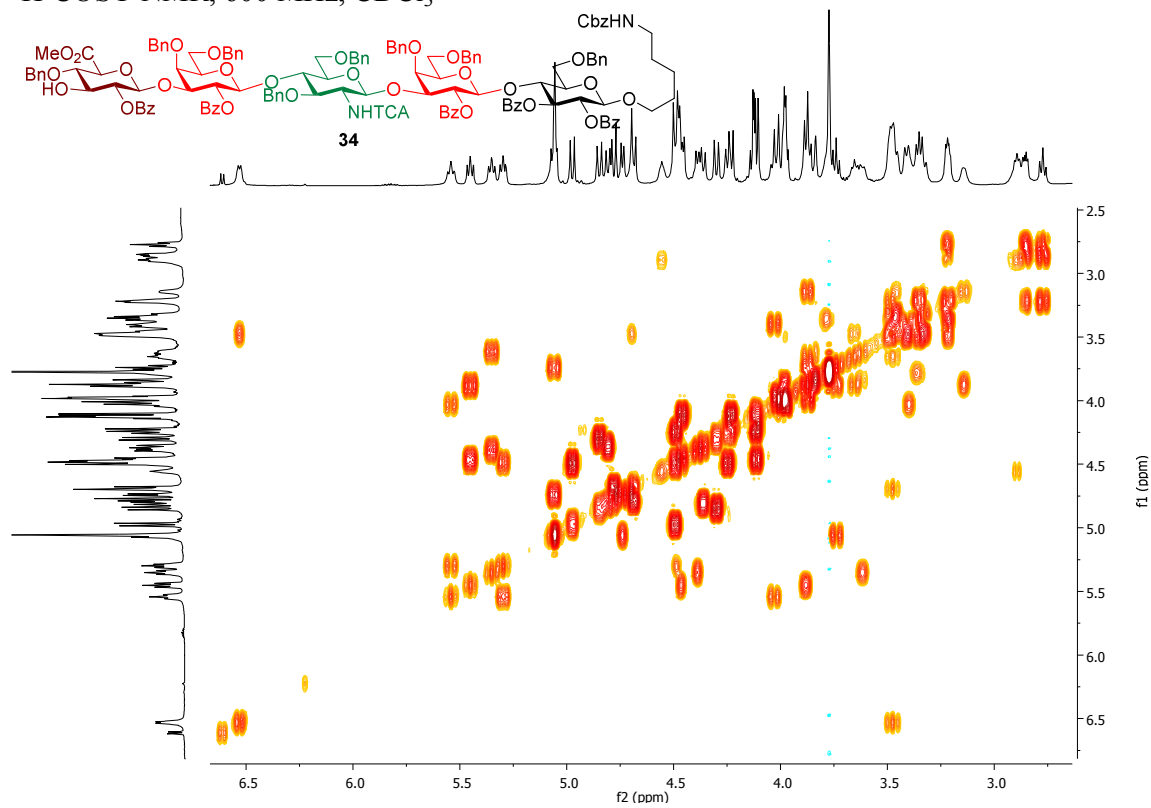
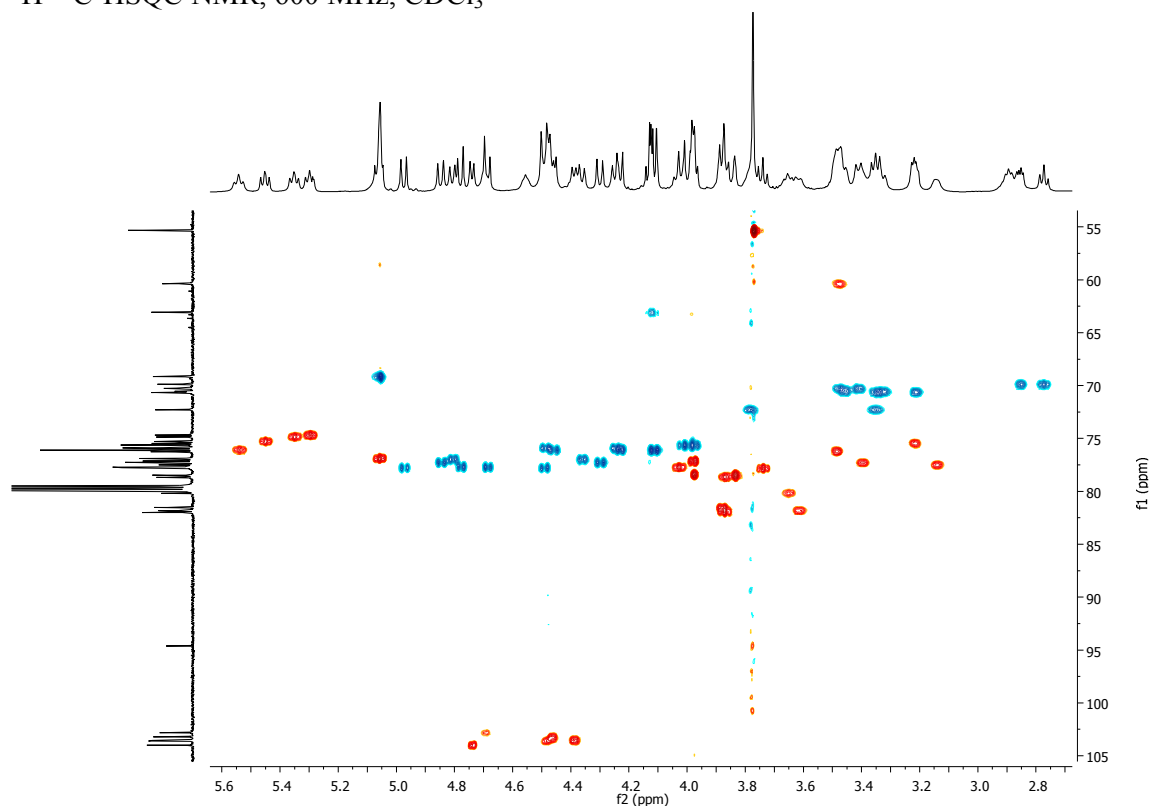


Figure S12. LC-MS of 34.

¹H NMR, 600 MHz, CDCl₃¹³C NMR, 150 MHz, CDCl₃

^1H -COSY NMR, 600 MHz, CDCl_3  ^1H - ^{13}C -HSQC NMR, 600 MHz, CDCl_3 

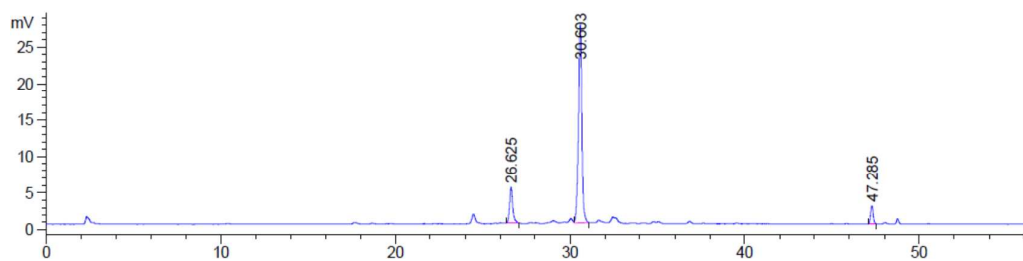
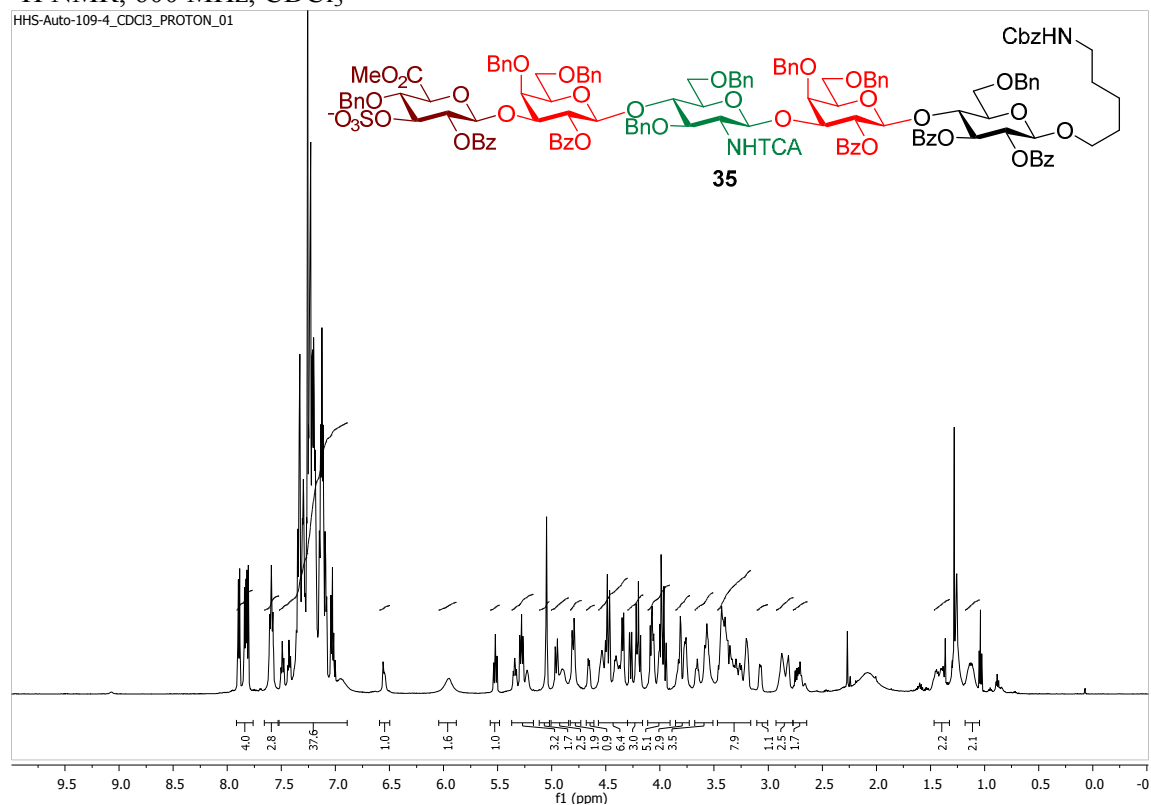
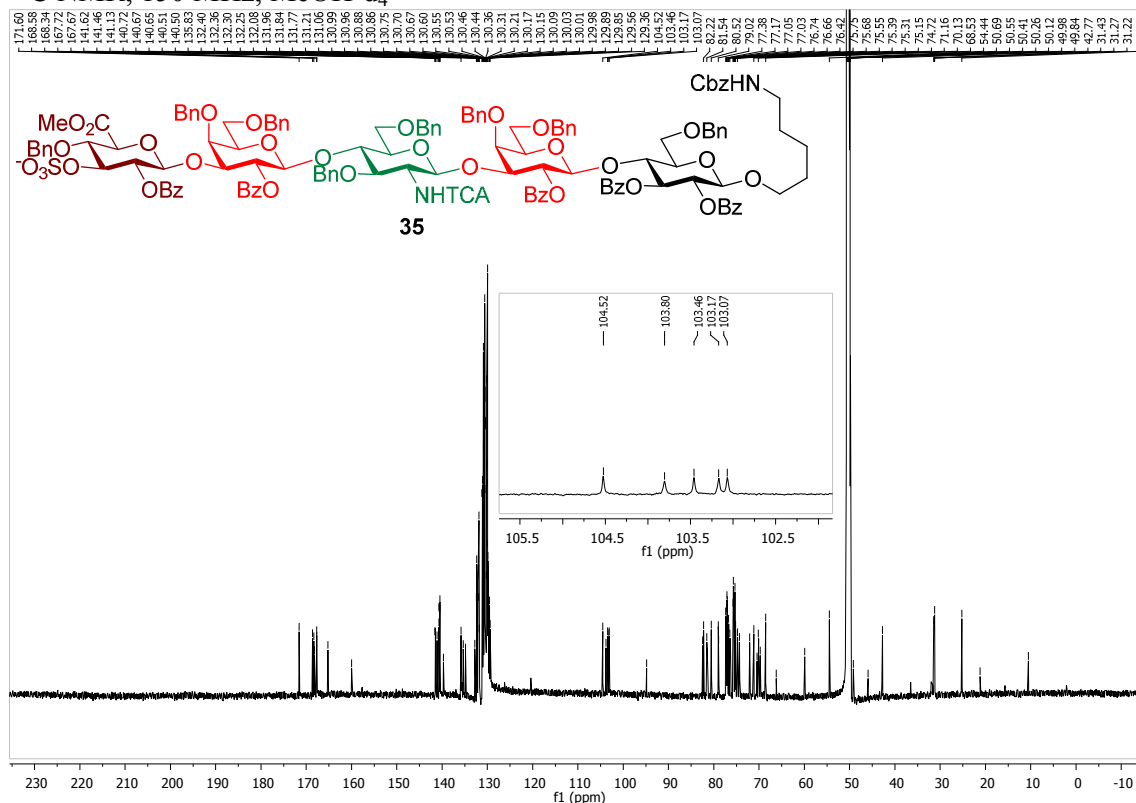
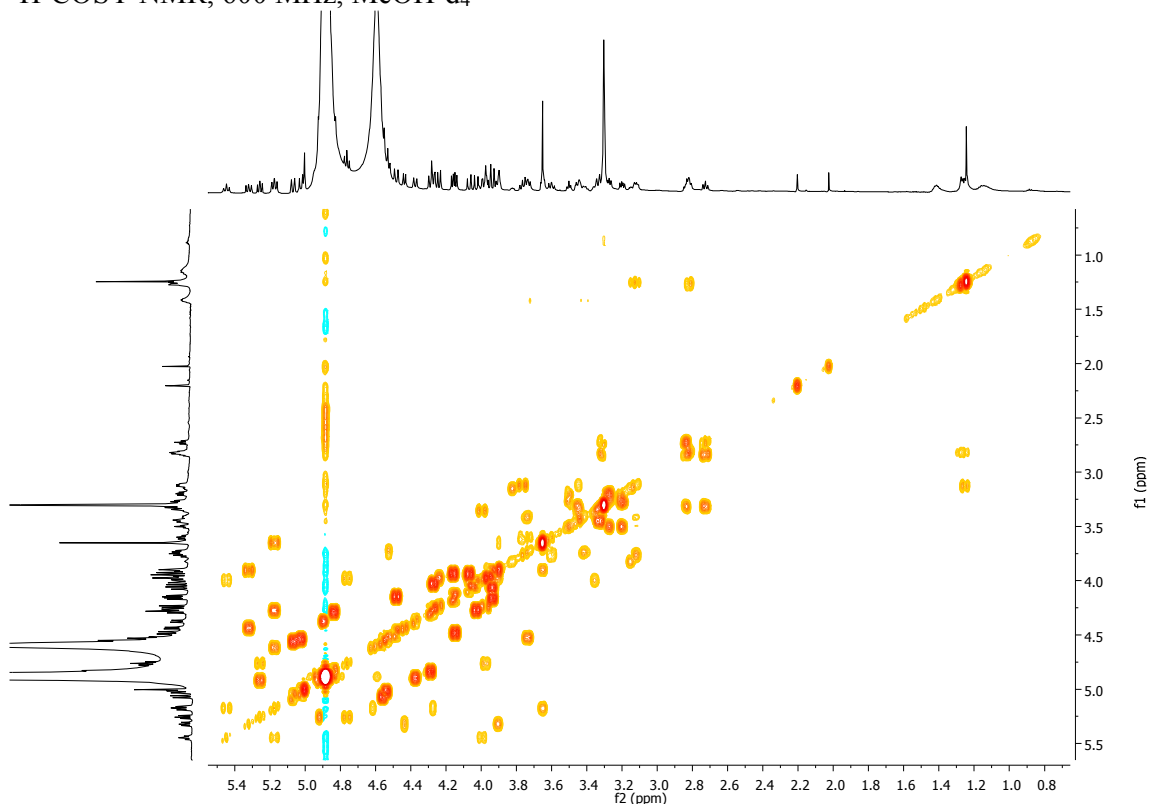
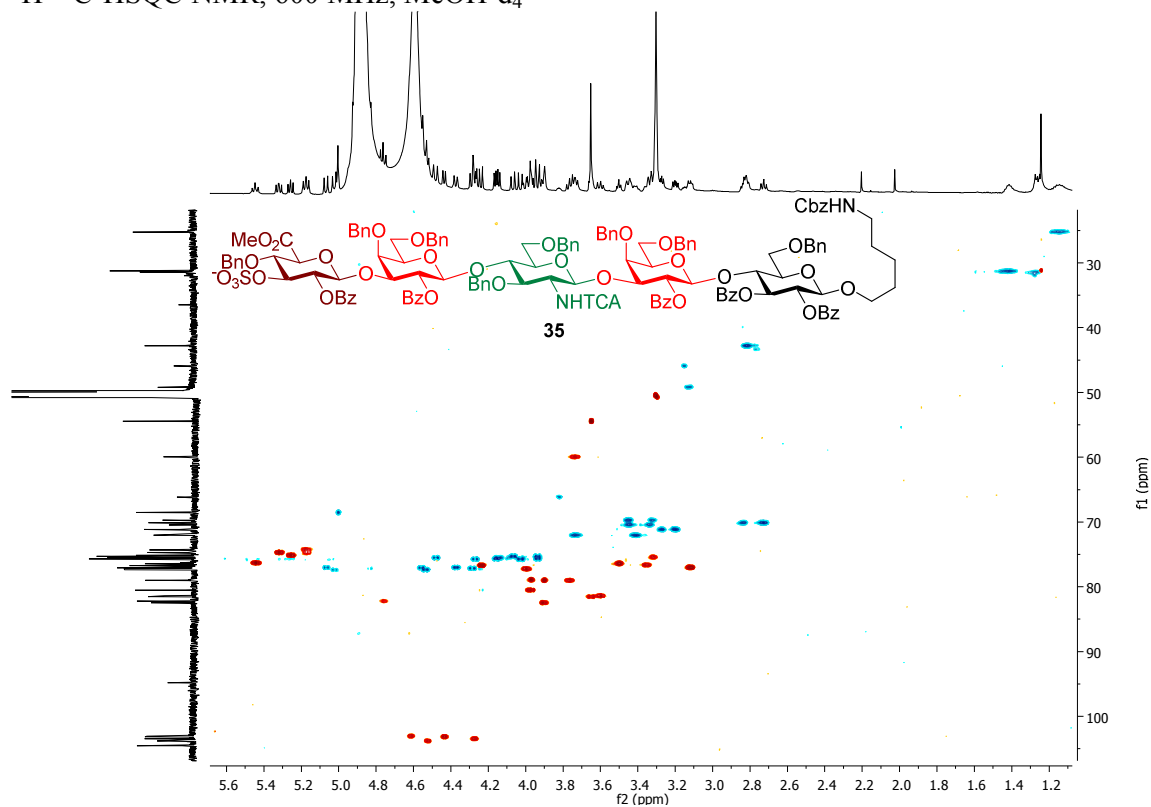


Figure S13. Purification of α -Gal epitope **35**. Conditions: column: C18-Nucleodur (21 \times 250 mm; 5 μ m); flow rate: 10 mL \cdot min $^{-1}$; eluents: 0.01 M NH_4HCO_3 in water/MeCN; gradient: 45% (5 min) \rightarrow 55% (in 40 min) \rightarrow 100% (in 5 min); detection: ELSD.

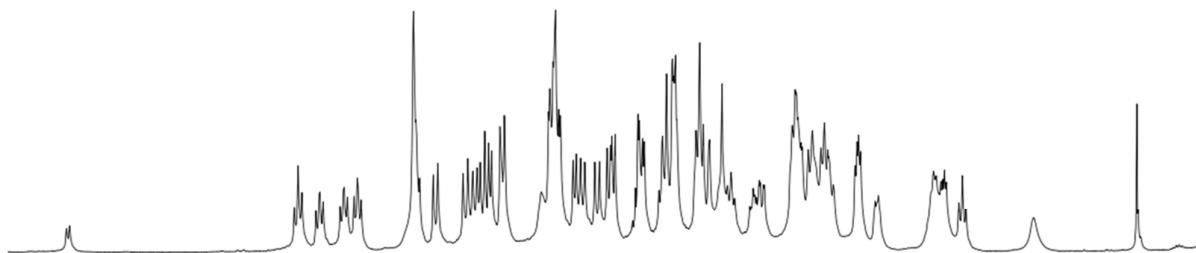
^1H NMR, 600 MHz, CDCl_3



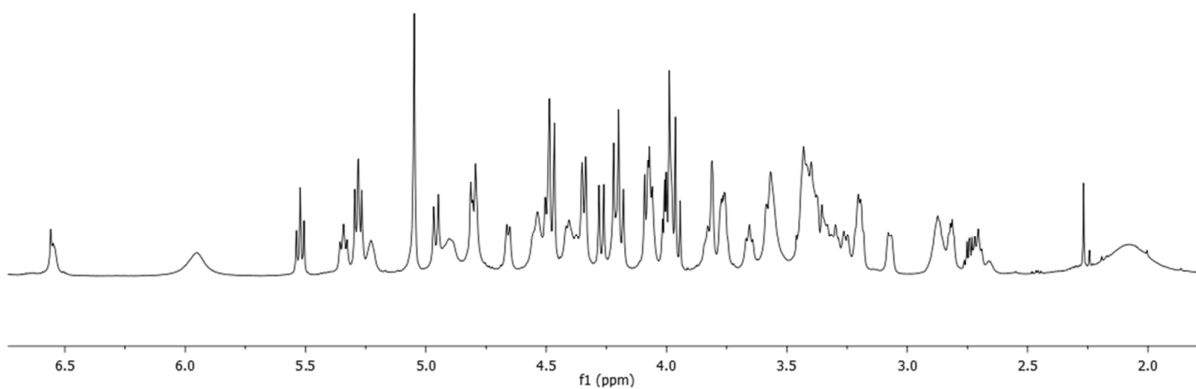
^{13}C NMR, 150 MHz, MeOH- d_4  ^1H -COSY NMR, 600 MHz, MeOH- d_4 

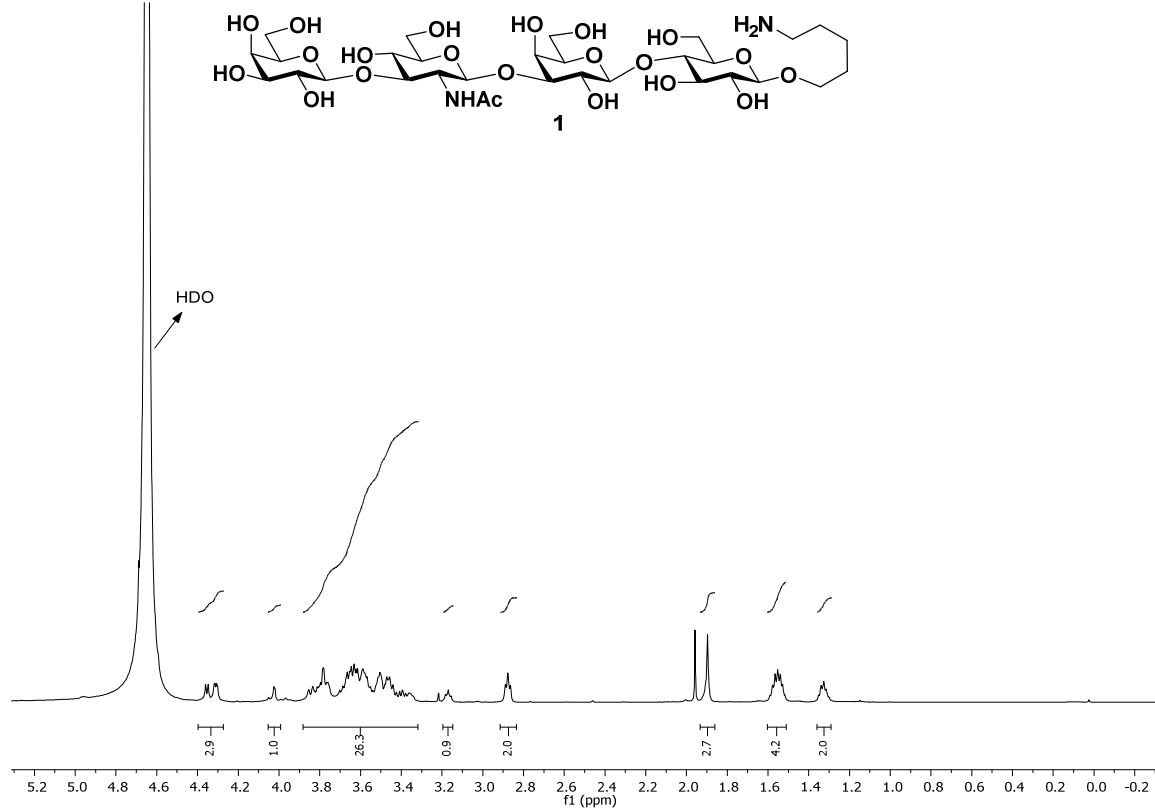
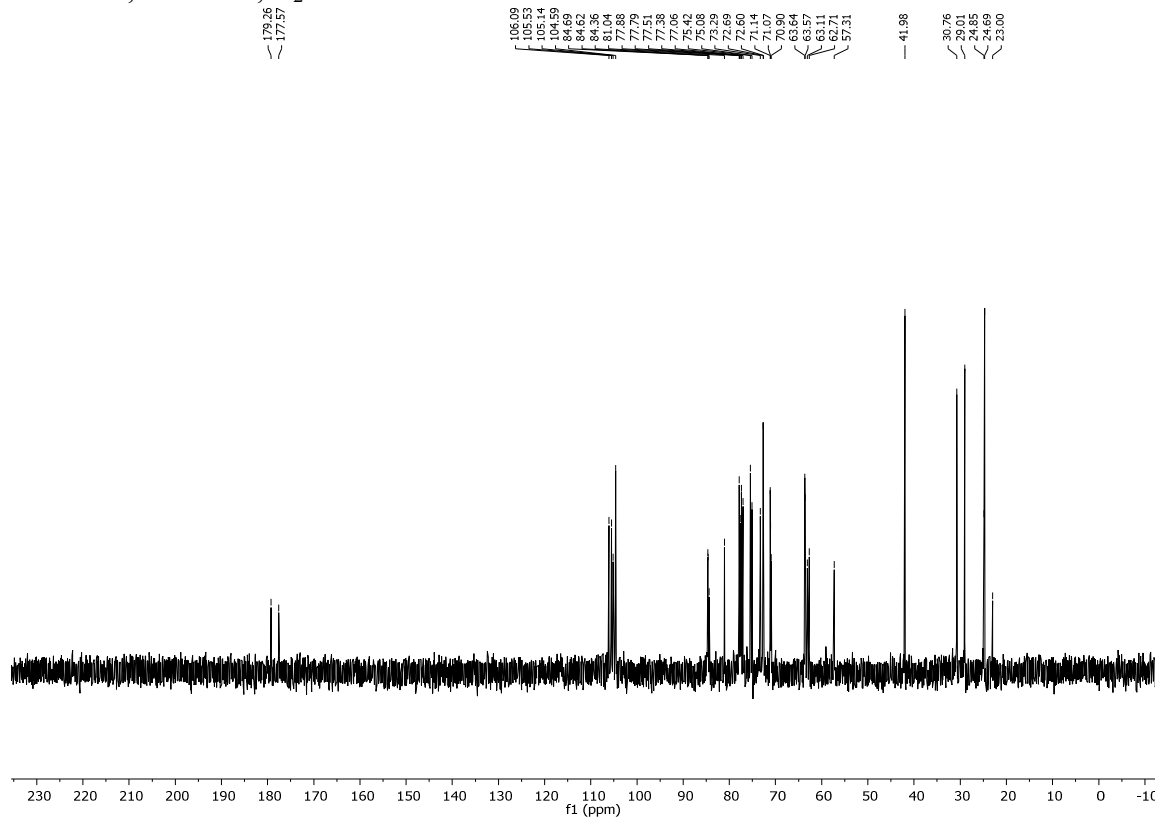
^1H - ^{13}C -HSQC NMR, 600 MHz, MeOH- d_4 

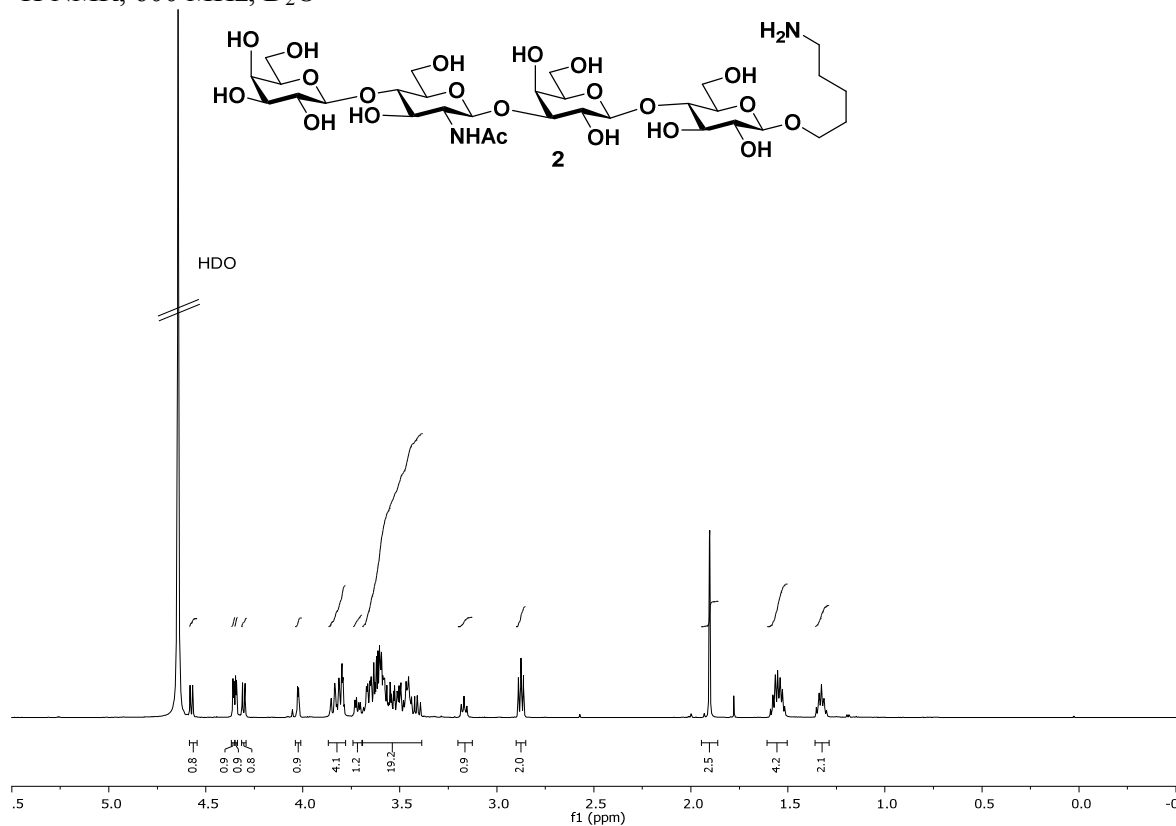
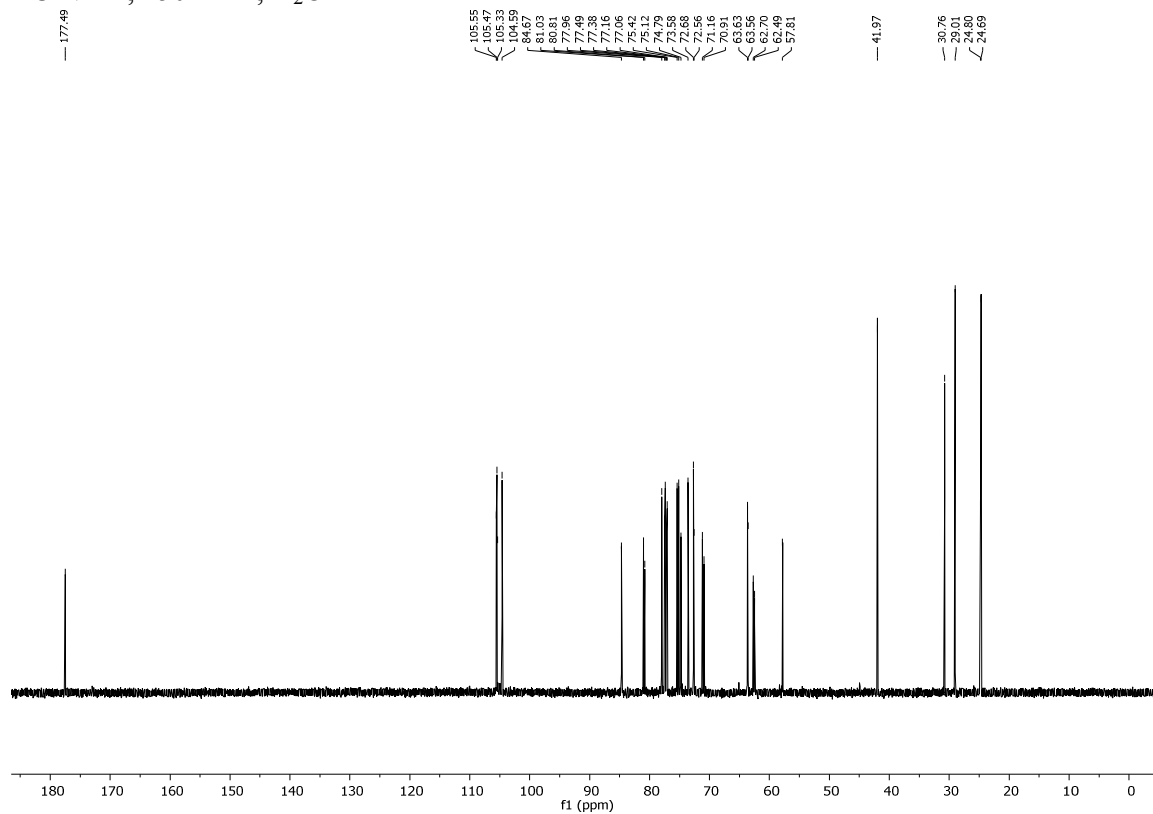
non-sulfated pentasaccharide 34

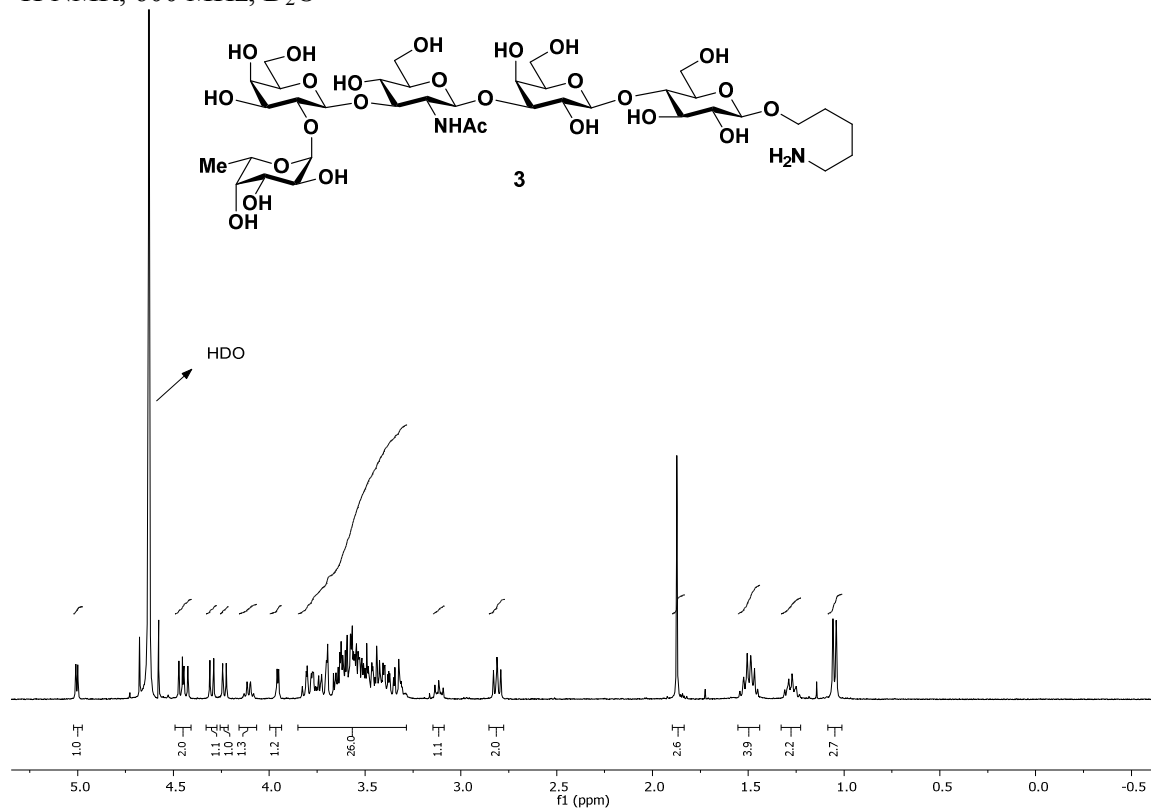
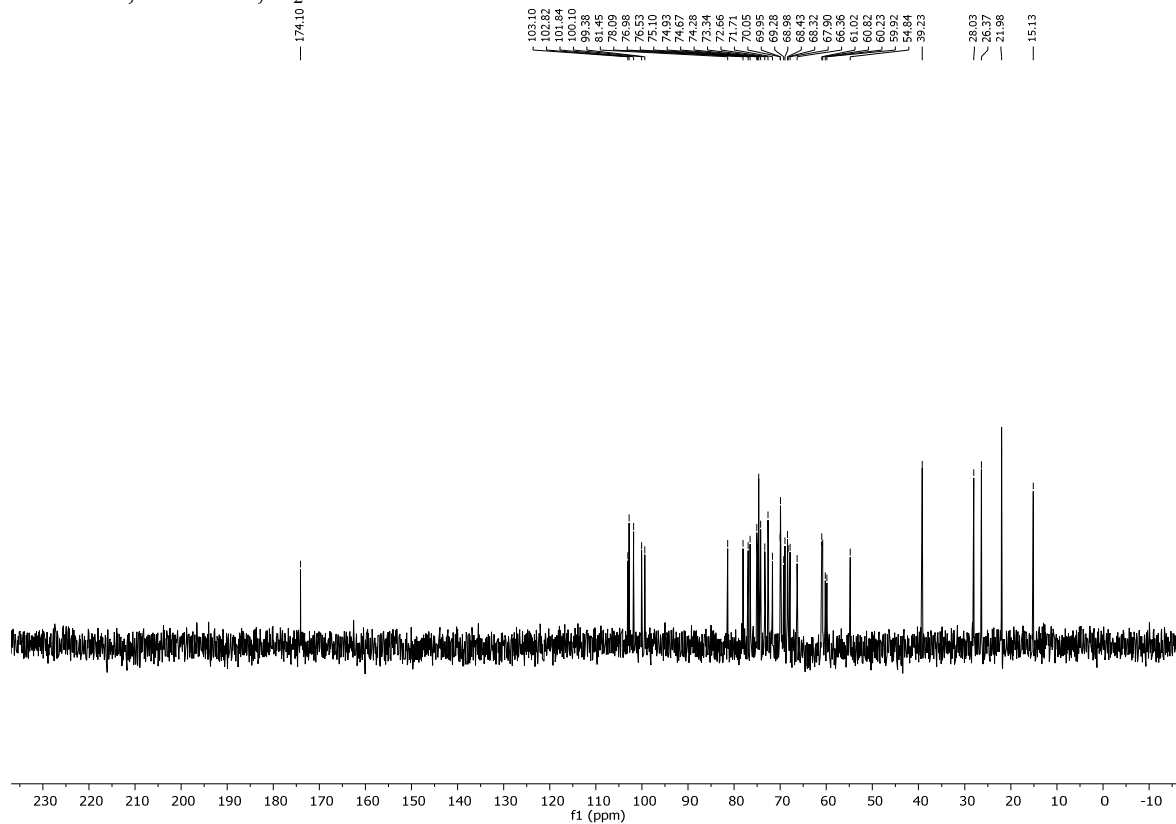


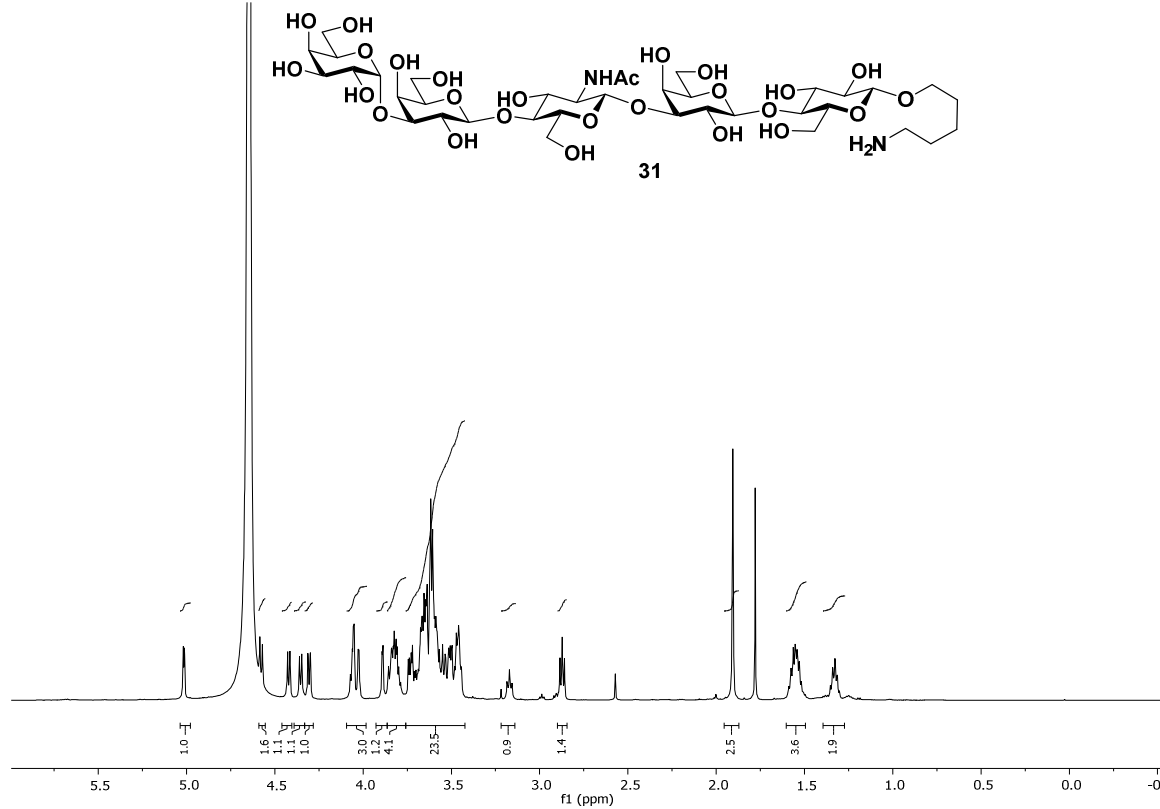
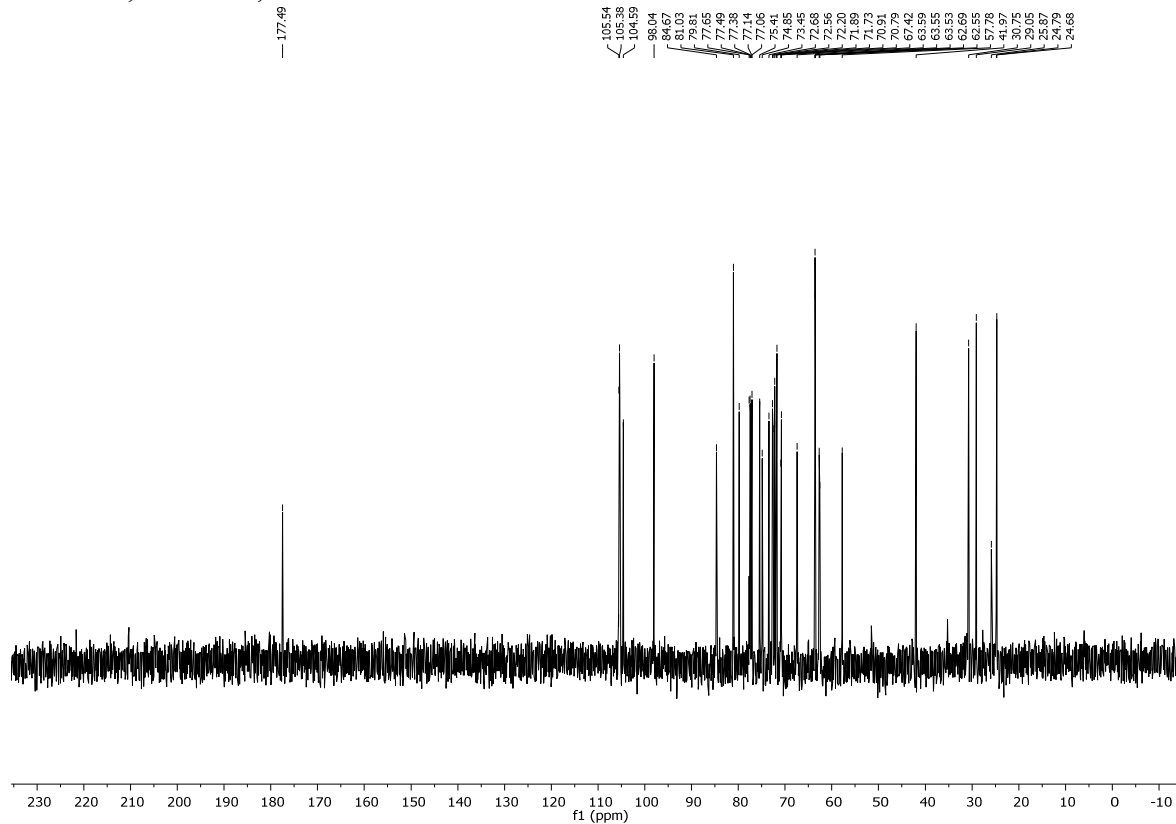
sulfated pentasaccharide 35

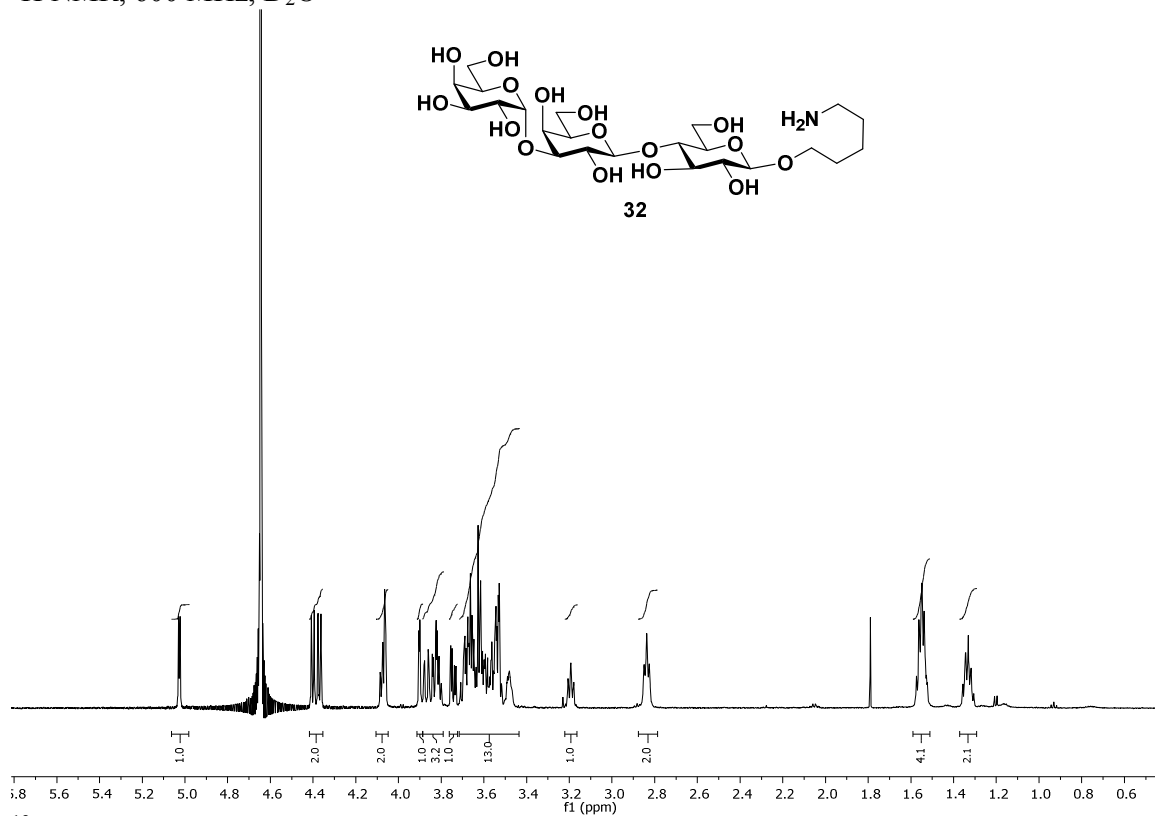
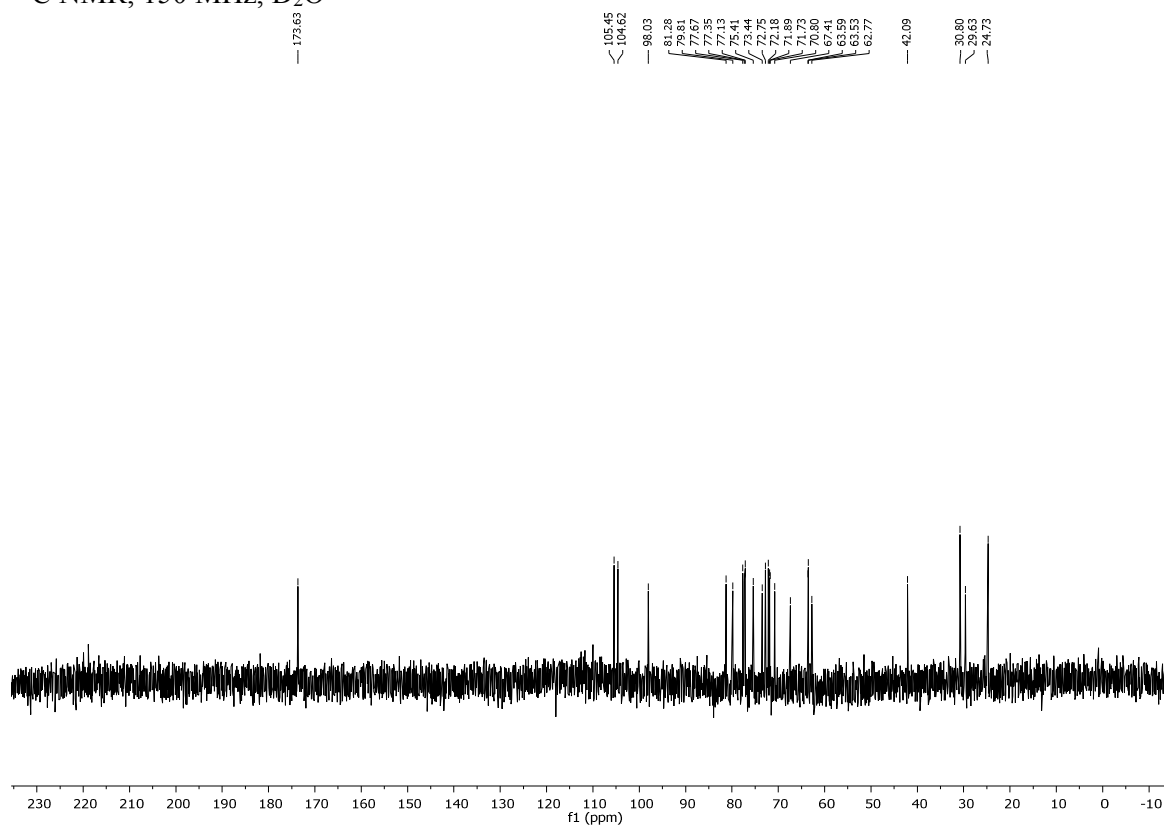


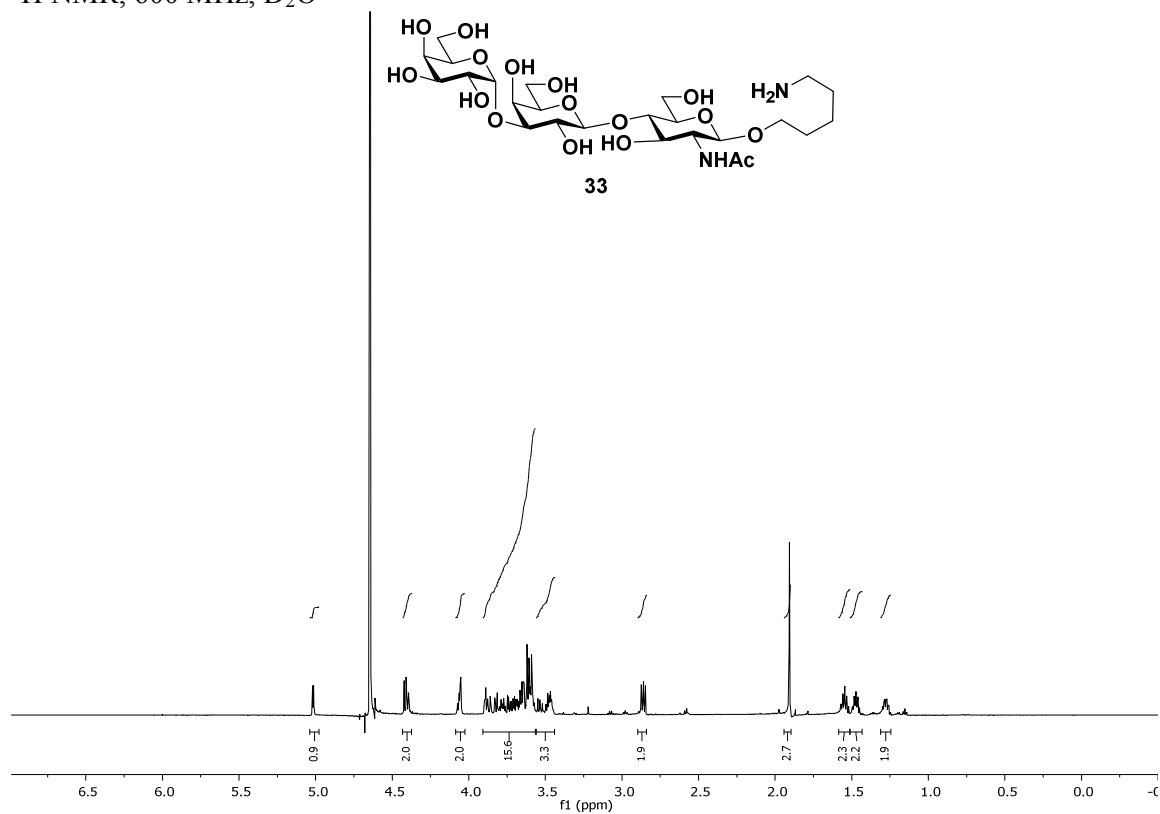
^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 

^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 

^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 

^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 

^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 

^1H NMR, 600 MHz, D_2O  ^{13}C NMR, 150 MHz, D_2O 