AP-1/σ1A and AP-1/σ1B adaptor-proteins differentially regulate neuronal early endosome maturation via the Rab5/Vps34-pathway

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Supplementary Information

Supplemental Information



Figure S1: AP-1/Vps34p direct interactions: Determination of σ 1-adaptin binding of Vps34p domains by the yeast-3-hybrid assay. The C-terminal, regulatory Vps34p domain (aa 809-887) showed weak, unspecific binding to the N-terminal core γ 1 adaptin domain, but no binding to either σ 1A or σ 1B. This weak interaction is not due to an autoactivation, because without γ 1, the weak binding activity was lost. We did not analyze this weak binding in more detail, because it can not explain σ 1 isoform specific functions. This binding could also be mediated by a γ 1 domain occupied or sterically blocked by β 1 and μ 1 adaptins. The core domain of Vps34p (aa 257-869), which contains the catalytic center, did not bind to any of the adaptins. In the presence of the auxotrophic marker histidine (+His) all yeast clones grow, excluding a toxic effect of the non-yeast proteins and protein fragments. Addition of the histidine-synthesis blocker 3-AT to 2.5 mM inhibits growth of the yeast clones, confirming that they are indeed auxotrophic for histidine.



Figure S2: Rabex-5 sequence motif for σ 1B binding: Sequence alignment of Rabex-5 and RabGAP5 and the effects of amino acid exchanges on the σ 1B binding. Rabex-5 and RabGAP5 bound σ 1B, but not σ 1A adaptin, indicating that both proteins might use the same sequence motif for σ 1B binding. Aligning there sequences revealed two homologous motifs in both proteins: **P_E_**A:E_C:L_**L** and **P_L_**Q:K_P:E_Q:G_**V**. Homologous sequence motifs are boxed. Exchange of the E and L residues following the P's in both sequences by A in Rabex-5, indeed abolished σ 1B binding in yeast-3-hybrid assays.









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Fig. S3: AP-1/ σ 1A and AP-1/ σ 1B dependency of ArfGAP1 distribution on membranes: A We expressed GFP-tagged versions of the ubiquitously expressed ArfGAP1 and of the brain specific ArfGAP1 isoform in mouse embryonic fibroblast cell lines from wt, o1B -/- and mice deficient in any functional AP-1 complex (µ1A -/-, ΔAP-1) and determined their distribution on peripheral endosomes as well as on the peri-nuclear trans-Golgi network. Over 85% of the ArfGAP1 proteins bound to endosomes and this fraction increased in σ 1B -/cells. Fraction sizes of endosomal GFP-ArfGAP1 proteins in the mouse embryonic fibroblast (MEF) cell lines. **B** Confocal microscopy images showing the localization of AP-1 complexes, labelled by anti-y1 antibodies and Alexa-633 secondary antibodies (red), and of GFP-ArfGAP1 proteins at the peri-nuclear trans-Golgi network. AP-1 and ArfGAP1 proteins formed neighboring domains with limited colocalization, best visible on the larger trans-Golgi network, in line with transient interactions. C Distribution of both ArfGAP1 proteins on endosomes and the trans-Golgi network (TGN) in the various MEF cell lines. Numbers are the signal intensities, expressed as mean (m), highest (h) and lowest (l) intensity ranges of the respective organelles within a cell. There was no qualitative difference in their distribution of both proteins on endosomes and on the trans-Golgi network in the various cell lines. The range of concentrations was much higher in the trans-Golgi network than on endosomes. The trans-Golgi network membrane is larger than the area of the labelled endosomes allowing for a wider distribution of the proteins and indicating the concentration of the proteins in smaller subdomains.

The quantifications shown in A and C were determined in $n \ge 10$ cells and the box-plot diagrams depict the statistics of the cohort.