

Dental Calculus and the Evolution of the Human Oral Microbiome

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ABSTRACT Characterizing the evolution of the oral microbiome is a challenging, but increasingly feasible, task. Recently, dental calculus has been shown to preserve ancient biomolecules from the oral microbiota, host tissues and diet for tens of thousands of years. As such, it provides a unique window into the ancestral oral microbiome. This article reviews recent advancements in ancient dental calculus research and emerging insights into the evolution and ecology of the human oral microbiome.

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he human oral cavity is home to a diverse ecology of microorganisms, collectively known as the oral microbiome. Composed of more than six hundred taxa,1 the oral microbiota plays a central role in dental health, and increasingly it has been shown to influence extraoral organs and tissues, as well as general health.2 Investigating the evolution of this rich microbial ecology is challenging, but it is critical for understanding the origins and prevalence of oral and systemic disorders as diverse as dental caries, periodontal disease, rheumatoid arthritis, cardiovascular disease, respiratory illness and a range of infectious diseases.

Research on what constitutes a healthy or normal oral microbiome has expanded dramatically over the past decade,³ in large part due to the advent of high-throughput DNA sequencing^{4,5} and

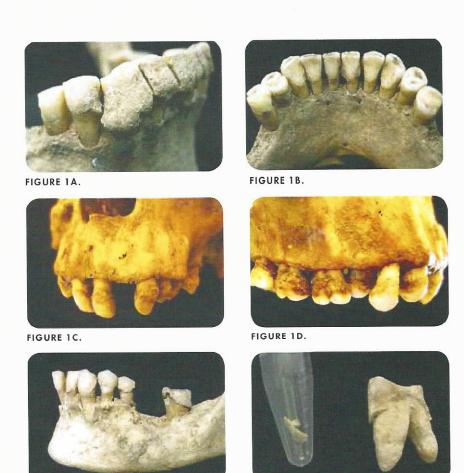
major public funding initiatives, such as the National Institutes of Health's Human Microbiome Project (HMP).⁶ However, the fact that the oral microbiome will cause dental disease in a majority of individuals during their lifetimes has been argued to suggest that even the "healthy" or "normal" oral microbiome today is already in an altered dysbiotic state,⁷⁻⁹ and likely has been for some time.¹⁰⁻¹²

To better understand the oral microbiome and its associations to both dental disease and other so-called "diseases of civilization," it is useful to investigate how our vulnerability to disease is related to human evolutionary history. Evolutionary medicine, a field that integrates both medicine and evolutionary biology, provides a framework for rethinking conventional models of oral health and disease, ¹³ and biological anthropology,

a field that encompasses primatology, paleoanthropology, bioarchaeology and human biology, provides a context in which to characterize the ancestral state of the human microbiome. ¹⁴ Together, these fields provide pathways to better understanding the oral microbiome, and ultimately to restoring and better maintaining oral health in the future.

One of the most exciting recent discoveries in biological anthropology has come from an unlikely quarter – dental calculus (FIGURE 1). Since at least the 1980s, it has been known that archaeological dental calculus contains preserved cellular structures of oral bacteria, 15-17 but it was only recently discovered that it is also a robust and long-term reservoir of well-preserved DNA and proteins. 17-20 Advances in ancient DNA and paleoprotein technologies now allow detailed characterization of these ancient biomolecules, enabling direct comparisons between ancient and modern oral microbial communities. 18,20 Host proteins preserved within ancient dental calculus additionally provide insights into past microbial virulence and host immune response,20 and food particles entrapped within dental calculus offer unprecedented insights into the diets of past populations.²⁰⁻²³

This article provides an overview of recent ancient dental calculus research and discusses four areas in which the investigation of ancient oral microbiota is poised to make significant contributions to an evolutionary perspective on human health and disease: dental caries; periodontal disease; rheumatoid arthritis and cardiovascular disease; respiratory infections and meningitis; and major infectious diseases.



FIGURES 1. Archaeological dental calculus. (A,B) St. Petri cemetery, Dalheim, Germany, ca. 1100 CE. (C,D) Sky cave tomb, Samdzong, Nepal, ca. 500 CE. (E,F) Mortuary cave, Camino del Molino, Spain, ca. 2500 BCE. The amount of dental calculus typically required for analysis is shown in (F). Dental calculus from all three sites has yielded genetic data suitable for ancient microbiome analysis.^{20,77}

FIGURE 1F.

Dental Calculus and Ancient Biomolecules

FIGURE 1E.

Dental calculus is a complex, mineralized bacterial biofilm that forms from dental plaque on the surfaces of teeth. ^{24,25} It is found in all known past and present human populations, and it is nearly ubiquitous among adults without active dental hygiene. ¹⁷ The amount of dental calculus buildup on the dentition varies widely among populations ^{26,27} and may result from a complex combination of factors related to subsistence, dietary abrasiveness, dental hygiene practices and genetic predisposition. However, prior to modern dentistry it is not

uncommon to observe heavy calculus deposits in excess of 100 mg, especially from post-Neolithic periods (FIGURE 1). Importantly for archaeology, dental calculus preserves over time at least as well as bone and dentine, and it has even been found on the teeth of Neanderthals²⁸ and australopithocenes, ²⁹ in addition to humans. Among primates, the oldest known calculus to date was reported on the dentition of a Miocene orangutan ancestor and dates to roughly twelve million years ago³⁰ (FIGURE 2). Dental calculus is thus noteworthy for its availability in the fossil record throughout the entirety of human evolution.

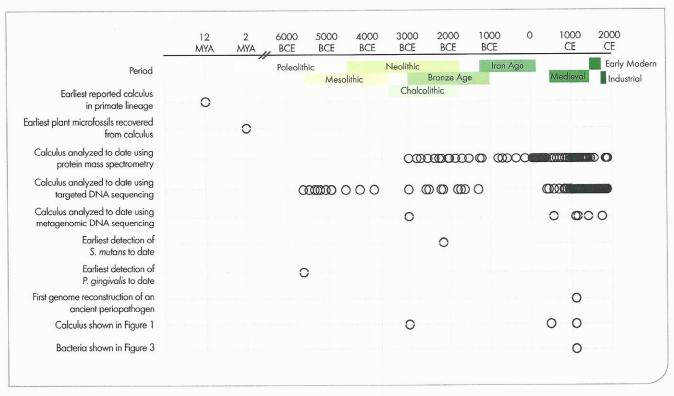


FIGURE 2. Idealized chronology of European prehistory and timeline of major ancient dental calculus findings to date. Includes data from Europe, ^{18,20,23,77} Asia, ^{23,30,77} Africa, ^{23,29,77} North America^{34,77} and South America. ¹⁹ Note that regional European chronologies differ, with major periods typically beginning earlier in southeastern Europe and later in northwestern Europe. African, Asian and New World chronologies are not shown.

Dental calculus formation occurs when dental plaque undergoes periodic mineralization events, although the timing and triggers for this process are not well understood.¹⁷ During mineralization, calcium phosphate ions from saliva and gingival crevicular fluid precipitate within the dental plaque matrix, at once killing the microbiota and calcifying the microbial cells and other debris in situ (FIGURE 3A). Organic structures preserve well in this environment (FIGURE 3B), and bacterial cell walls with functional cell surface protein epitopes, as well as delicate dietary microfossils such as starch granules, have been found to remain intact for more than 10,000 years.^{28,31} Genetic material also survives within dental calculus (FIGURE 3C), in part because the hydroxyapatite mineral within dental calculus strongly binds DNA.17 Genetic

analyses of archaeological dental calculus have found that it typically contains tento thousandfold more DNA than bone or dentine from the same individual, making it the richest source of ancient DNA yet identified in the archaeological record.¹⁷

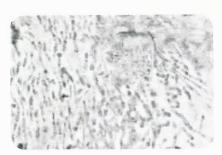
A number of recent studies have explored the potential of dental calculus to reconstruct aspects of ancient health and disease (FIGURE 2) and have shown it to be an area of great potential. Made possible by dramatic advances in ancient DNA³² and paleoprotein³³ technologies over the past decade (FIGURE 4), these studies have uncovered specific aspects of individuals' health states, ¹⁸⁻²⁰ diets, ^{20,23,28} ancestry³⁴ and even craft activities³⁵ using a combination of genetic, proteomic and microscopic analyses of dental calculus. This type of specificity and the ability to address issues that typically leave no

lasting mark in the macroarcheological record provide opportunities to address questions that were previously thought to be unanswerable. By using these new and emerging techniques, dental calculus can be used to address fundamental questions about the evolution of human oral health.

Oral Microbiota and Disease — Targets for Discovery

Dental Caries

Dental diseases, such as caries and periodontal disease, are among the most prevalent diseases affecting industrialized societies.⁷ In the 1960s, the average number of decayed, missing and filled teeth (DMFT index) among 12-year-old children in Western Europe was greater than 5, and by age 15 the average DMFT exceeded 10.³⁶ Thus, a European child reaching



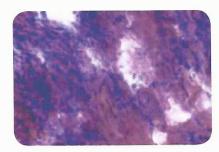




FIGURE 3A.

FIGURE 3B.

FIGURE 3C.

FIGURES 3. Micrographs of ancient bacteria embedded within archaeological dental calculus dating to the medieval period, ca. 1100 CE.²⁰ (A) Numerous calcified bacterial cells are visible within dental calculus using scanning electron microscopy (SEM) with backscattered electron imaging. (B) Staining of decalcified dental calculus thin sections reveals the presence of both Gram-positive and Gram-negative bacterial cells with relatively intact cell walls. (C) Hoechst stain, a blue fluorescent dye that binds double-stranded DNA, reveals the presence of abundant genetic material.

adulthood in the mid-20th century could expect one-third of his or her dentition to be compromised by dental decay. From the 19th century to the mid-20th century, dental decay was perceived as so inevitable that complete dental extraction and replacement with dentures became a popular gift for young women and brides in Western Europe and parts of North America.37-39 Extensive dental public health interventions and the introduction of fluoridated dentifrices over the past 50 years have significantly reduced caries rates,36 but even today dental caries affect more than 40 percent of children and 90 percent of adults in the United States.⁴⁰

Dental caries are easily observed in the archaeological record, and they have been systematically studied for more than a century.41 Extensive data have now been collected on paleoanthropological and archaeological populations around the world, spanning time periods from the Pleistocene⁴² to the 19th century.⁴³ It is clear from these studies that diet is the major driver of caries frequency.¹² Although caries are observed during all time periods, and are, in fact, also present in nonhuman primates, 44,45 caries frequencies vary remarkably through time and space. The most salient patterns relate to the transition between foraging (hunting and gathering) and agriculture,46 and later to the widespread availability of refined flour and sugar, especially sucrose, during the 18th century.41

Numerous studies have documented increases in caries frequency with the onset of agriculture. 12,41 The Eastern Woodlands of North America is perhaps the most intensively studied region, and analysis of caries frequency at 180 sites reveals that during the 2,000 years preceding the introduction of maize agriculture the percentage of teeth affected by dental caries was approximately 2 to 5 percent. After the introduction of maize ca. 500 CE, caries frequencies increased steadily from 14 percent during the Late Woodland period to 18 percent during the late Prehistoric period to 22 percent during the early Contact period¹² (FIGURE 5). Although not all regions saw such sharp increases, a general trend toward increased caries frequency in agricultural populations is observed globally, 12,41 with relatively few exceptions. 47-50

The origin of today's extreme levels of dental decay, however, lies not with agriculture, but with the introduction of refined flours and sugars during the Early Modern period (ca. 1450-1800 CE; FIGURE 2). In Western societies, tooth decay was so severe during this time that smiling is all but absent from European and American portraiture of the period. 51 Since then, consumption of refined carbohydrates has been met with increasingly aggressive oral hygiene regiments and prophylactic dental care as necessary measures to prevent premature tooth loss. 24 The need

to continuously engage in such behavior has been argued to be a sign that even the "healthy" oral microbiome today is in an altered state of dysbiosis.^{7,11}

One question that emerges with these observations is to what degree the oral microbiota of these populations changed in step with subsistence practices. Recent research on the gut microbiome has identified major changes in microbial ecology associated with foraging, agricultural and industrial lifestyles,52 but less is known about the oral microbiome of traditional societies.53,54 A recent study of ancient oral microbial communities spanning the past 8,000 years reported evidence for slight, phylum-level microbial shifts correlated with the onset of agriculture and industrialization;18 deeper sequencing and a larger sample size in future studies is anticipated to clarify these associations and provide greater taxonomic resolution during these transitions.

Direct investigation of ancient carious lesions using ancient DNA techniques has identified the presence of the cariopathogen *Streptococcus mutans*,⁵⁵ but reconstructing the full polymicrobial community contributing to such lesions⁵⁶ is challenging because dentine is highly susceptible to postmortem alteration by environmental microbes.²⁰ By contrast, dental calculus is much better preserved.²⁰ Genetic sequences from *S. mutans* have been detected in the dental calculus of diverse populations,¹⁸⁻²⁰ but interestingly



FIGURE 4. Extraction of genetic material from archaeological dental calculus requires highly specialized ancient DNA laboratories to prevent contamination from modern sources.

it has not been detected in samples prior to the Bronze Age (ca. 2200-1000 BCE) 18 (FIGURE 2). Recent phylogenetic analysis of modern S. mutans strains has estimated that S. mutans underwent an exponential expansion ca. 10,000 years ago (95 percent confidence interval: 3,268-14,344 years ago), suggesting an association with the onset of agriculture in the Near East.57 Future ancient DNA investigations of dental calculus using whole-genome capture enrichment technologies32 may be able to reveal the natural history of S. mutans and determine the major drivers of its evolution and functional role within the oral cavity of humans.

Periodontal Disease, Rheumatoid Arthritis and Cardiovascular Disease

Periodontal diseases affect up to 90 percent of the worldwide population,9 and moderate-to-severe chronic periodontitis is estimated to affect 13 percent of U.S. adults older than age 30,58 although new diagnostic criteria suggest that this may be a gross underestimate.⁵⁹ In addition to dental morbidities, periodontitis is also associated with increased risk of a wide range of so-called "diseases of civilization," including type II diabetes, obesity, rheumatoid arthritis, cardiovascular disease, stroke and pulmonary disease. 9,60-65 This association may reflect a partially causal relationship. For example, Porphyromonas gingivalis, an oral bacterium strongly associated

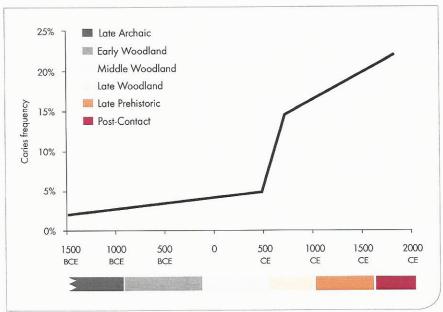


FIGURE 5. Dental caries frequency in prehistoric Eastern Woodland populations of North America. A dramatic increase in caries frequency is associated with the adoption of maize agriculture ca. 500 CE.¹²

with periodontitis, ⁶⁶ has recently been implicated in the development and progression of rheumatoid arthritis, ^{67,68} and treatment of periodontitis has been found to alleviate rheumatoid arthritis symptoms. ^{69,70} Periodontitis is also associated with a 19 percent increase in the risk of cardiovascular disease, ⁷¹ and a recent study of cardiovascular specimens found that >80 percent of diseased heart valves and >90 percent of aortic aneurisms have been infected with cariogenic or pathogenic periodontal bacteria. ⁶²

Although there is ample evidence for periodontitis in the archaeological record, periodontal diseases are very poorly studied. The most widely used laboratory guide for osteological analysis, *Standards for Data Collection From Human Skeletal Remains*, ⁷² contains recording metrics for age estimation from dental development and wear, enamel hypoplasia measurement, caries scoring and dental calculus quantification, but no information on how to record or measure periodontal disease in archaeological dentitions. As a result, it is rarely noted in archaeological reports. When periodontal disease is mentioned,

the descriptions are usually qualitative and limited to small case reports. ¹² At present there have been no large-scale, systematic studies of periodontal disease prevalence in the past, leaving many gaps in our understanding of the antiquity, prevalence and significance of periodontal disease in human evolutionary history.

Although clinical metrics⁷³ are difficult to apply to archaeological specimens, molecular characterization of periodontal disease is increasingly feasible. Recent conceptual changes in periodontology over the past 50 years now point to disrupted microbial communities and host-mediated inflammatory tissue destruction as the proximate causes of periodontal disease,74,75 and new molecular approaches, such as metagenomics and metaproteomics, allow culture-free investigation and comparison of oral microbiota and host immune response from both clinical and archaeological dental samples (FIGURE 2). For example, a recent study of medieval dental calculus found that periopathogens common today were also associated with suspected periodontal disease cases nearly a thousand years ago.20 Frequencies of

the periodontal pathogens P. gingivalis, Tannerella forsythia, Treponema denticola and Filifactor alocis were found to be highly elevated in dental calculus collected from archaeological dentitions with generalized moderate or severe attachment loss. Additionally, virulence factors expressed by these taxa and host innate immune proteins were found at high abundance among the proteins recovered from these samples, strongly suggesting periodontitisassociated inflammation. In dental calculus from one individual, genetic sequences for T. forsythia were so abundant that a near complete ancient genome for this pathogen could be reconstructed. The ancient strain was found to harbor the same 14 virulence proteins found in modern T. forsythia, but it lacked several mobile elements, including a putative antibiotic resistance gene, tetQ, found today in the T. forsythia reference strain.

Two additional studies have begun to look at temporal changes in periodontal pathogens. In a study of ancient Chilean and Argentinian dental calculus, P. gingivalis was detected in most time periods from 2500 BCE to the present, 19 and in a study of ancient Europeans, P. gingivalis was detected in all major periods dating back to the Mesolithic, ca. 5550 BCE-3450 BCE18 (FIGURE 2). Interestingly, the latter study found that P. gingivalis frequencies were lower in Mesolithic hunter-gatherers than in post-Neolithic farmers, a finding consistent with observations that huntergatherer societies in the recent past and today appear to have lower rates of periodontal disease than agricultural and industrialized societies. 11,76 However, further research is necessary to ensure that this finding is not simply an artifact of more advanced DNA decay in older samples.77

The investigation of *P. gingivalis* in archaeological dental calculus may also have implications for the study of rheumatoid arthritis. In 1990,

rheumatologist Bruce Rothschild, MD, proposed a radical hypothesis — that rheumatoid arthritis is a vector-transmitted infectious disease that originated in central North America and spread to the Old World after European contact in the region ca. 1750.^{78,79} Among the evidence he assembled, he noted that the first documented case of rheumatoid arthritis in Europe dates to 1800, while cases in North America are known from archaeological remains up to 6,500 years old, and that Native Americans today have unusually high rates of the

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autoimmune disease (more than fivefold higher than the U.S. general population 80). Although aspects of this hypothesis are now quite dated, 81 the recent link made between rheumatoid arthritis and *P. gingivalis* 67,68 has rekindled interest in a possible microbial origin for this disease. By using ancient DNA from dental calculus along with osteological data, it may be possible to unravel the historic biogeography of virulent *P. gingivalis* strains and their relationship to autoimmunity.

Finally, the recent link made between cardiovascular disease and periodontal and cariogenic taxa⁸² raises questions as to the antiquity of oral involvement in atherosclerotic plaques. Cardiovascular disease is conventionally viewed as a disease of modernity, but archaeological evidence now confirms its presence in diverse ancient

cultures, from the elites of ancient empires to prehistoric hunter-gatherers. Calcific atherosclerosis has been identified, first by autopsy and later by X-ray and computed tomography (CT), in the coronary and peripheral arteries of mummies originating from ancient cultures in Egypt, the Alps, Peru, the American Southwest, the Aleutian Islands and Korea. 83,84 Moreover, radiological scans suggest that the prevalence of atherosclerosis was relatively high in prehistory, and, in the case of Egypt, no significant difference was found in the incidence or prevalence of atherosclerotic plaques between today and 3,500 years ago.85

A collaboration of cardiologists, radiologists, molecular biologists and archaeologists known as the Horus study team has been at the forefront of ancient atherosclerosis research since 2008.83 Because many of the risk factors associated with cardiovascular disease today do not apply to ancient populations, they have proposed that the high rates of atherosclerosis in the past may have resulted from chronic systemic inflammation caused by long-term infection by gastrointestinal parasites or repeated exposure to microbial or viral pathogens;83,86 however, in the absence of direct evidence for such infections, they note that other yet-to-be discovered risk factors may also play a role. Poor oral health may be just such a risk factor. Untreated caries, heavy calculus deposits and alveolar recession suggest both dysbiotic oral microbial communities and sustained inflammation were prevalent in many ancient populations. A promising future direction in this research would be to genetically test ancient calcific atherosclerotic plaques directly for the presence of oral taxa. Systematic CT scanning has identified atherosclerotic plagues in more than 50 ancient mummies from around the world, and biopsies of these plaques could be analyzed and

compared to microbial profiles generated from dental calculus collected from the same individuals. Taxonomic matches would be strong evidence for a long-term role of oral involvement in the initiation of cardiovascular disease.

Respiratory Infections and Meningitis

The oral microbiome is the natural reservoir for a large number of pathobionts (endogenous potential pathogens), including Streptococcus pneumoniae, Haemophilus influenzae, Moraxella catarrhalis, Streptococcus pyogenes, Corynebacterium diphtheriae, Bordetella pertussis and Neisseria meningitidis,1 which cause a wide range of acute and potentially life-threatening respiratory and meningeal infections, especially in children. Importantly, asymptomatic carriage of these microbes is relatively prevalent, although it varies widely among populations. For example, reported carriage rates of S. pneumonia range from 1 percent in parts of Scandinavia to more than 80 percent in parts of France and Gambia, while H. influenzae carriage rates range from 3 percent in Sweden to 88 percent in Costa Rica, and carriage rates for M. catarrhalis, a common cause of middle ear infections, range from 2 percent in parts of Sweden to 82 percent in parts of the Netherlands.⁸⁷ In Europe, N. meningitidis is typically carried by an average of 24 percent of teenagers and 8 to 13 percent of adults,88 and nearly onefifth of American schoolchildren carry S. pyogenes at any given time.89 For pathogens with widespread vaccine programs, such as Corynebacterium diphtheriae and Bordetella pertussis, carriage rates are generally low, 90-94 but were presumably once much higher. In addition to these pathobionts, even commensal oral taxa pose serious health risks among the elderly and immunocompromised. Aspiration of common oropharyngeal taxa can result

in aspiration pneumonia, 95,96 the leading cause of death and the second most common cause of hospitalization among nursing home patients. 96 Additionally, it is common for the elderly to acquire extraoral pathobionts, such as *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Escherichia coli*, in their dental plaque, which also contribute to aspiration pneumonia. 2

To date, multiple pathobionts have been identified in archaeological dental calculus, including *S. pneumoniae*, *H. influenzae*, *S. pyogenes*, *C. diphtheriae* and *N. meningitidis*. ²⁰ Additionally, genetic

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sequences consistent with *Bordetella* parapertussis, an organism responsible for persistent cough in children that is similar to pertussis, ⁹⁷ and several species of *Moraxella* were also identified. These pathobionts were detected in the dental calculus of two individuals excavated from the medieval St. Petri cemetery in Dalheim, Germany (ca. 1100 CE). Although there is no evidence that these organisms were causing disease in these individuals, they nevertheless provide the first direct evidence of respiratory pathobiont carriage in the oral cavity before the 20th century.²⁰

Only a century ago, respiratory infections were the leading causes of death in the U.S., 98 and studies of periosteal rib lesions in skeletal collections 99 and soft tissue changes in South American mummies 100-102 suggest that pneumonia

and/or other respiratory infections were a major cause of human mortality in prehistory. The discovery that dental calculus is a reliable source of genetic material from common respiratory pathobionts opens up the possibility of conducting epidemiological studies of past carriage rates, and also presents the opportunity to investigate the evolution of these taxa through time.

Dental calculus may also help to date the origin of infectious diseases that have recently emerged from the oral cavity, such as bacterial meningitis and gonorrhea. Phylogenetic studies of Neisseria, a genus of bacteria that colonize mucosal surfaces in animals, indicate that N. meningitidis and N. gonorrhoeae belong to a recently diverged pathogenic clade in humans that is most closely related to the common nasopharynx commensal Neisseria lactamica; 103-105 however, the timing and context of this divergence are not well understood. Both N. meningitidis and N. gonorrhoeae are obligate human taxa, indicating that this divergence must have occurred since the chimp-human split approximately 6 million years ago, and whole-genome comparisons with other Neisseria species suggest that N. meningitidis may have undergone a population bottleneck and acquired its virulence genes for polysaccharide capsule synthesis very recently, perhaps only a few centuries ago. 105 Nonspecific endocranial meningeal reactions are often found in skeletons, 106 and with the recent identification of putative N. meningitidis genetic sequences in archaeological dental calculus, this hypothesis has become testable. Future ancient DNA investigations using whole-genome capture enrichment technologies³² hold great promise for resolving the origins and evolution of N. meningitidis, a pathobiont whose mortality rate from bacterial meningitis and septicemia continues to exceed 10 percent even in developed nations. 104

Major Infectious Diseases

Finally, although not true members of the oral microbiome, several opportunistic and obligate pathogens can be found transiently within dental plaque, buccal mucosa and saliva.1 These include the causative agents of tuberculosis (Mycobacterium tuberculosis), leprosy (Mycobacterium leprae), plague (Yersinia pestis), syphilis (Treponema pallidum), gastritis (Helicobacter pylori) and smallpox (Variola virus), among others. M. tuberculosis, for example, is present in sputum and regularly comes into contact with the oral cavity throughout the entire course of the disease. 107 In addition to sputum, M. tuberculosis has also been detected in 92 percent of dental plaque samples from infected patients using polymerase chain reaction (PCR)-based techniques. 108 Leprosy involves the oral cavity in up to 60 percent of cases, and multiple oral structures may develop lesions and ulcers, including the hard and soft palate, the gingiva, tongue, lips and buccal mucosa. 109,110 During outbreaks of bubonic plague, Y. pestis that escapes the lymphatic system and infects the lungs causes pneumonic plague, a highly infectious form of the disease that results in lethal fulminant pneumonia. 111 T. pallidum is known to cause oral lesions during the secondary phase of syphilis infection, which may last for many years, 112 and this provides ample opportunity for passive or active incorporation and preservation in calcifying dental plaque biofilms. Finally, H. pylori is readily found in the saliva and dental plaque of infected individuals,113 and smallpox causes oropharyngeal lesions. 114

Infectious diseases have played a major role in shaping human history, and many pathogens continue to present serious challenges to public health. Little is known, however, about the origins or evolutionary history of most human infectious agents. Ancient DNA research has contributed

greatly to what is known about the origins of a handful of pathogens, including M. tuberculosis, ^{115,116} Yersinia pestis, ¹¹⁷⁻¹¹⁹ and H. pylori, ¹²⁰ and it has been used to confirm the presence of several additional pathogens in ancient infections, including M. leprosy, ¹²¹ T. pallidum ¹²² and Variola virus. ¹²³

However, genetic detection rates for most ancient pathogens are low, even in remains with overt and relatively diagnostic paleopathology indicators. Antemortem tissue destruction likely enhances postmortem decay, which may contribute to poor preservation of pathogen DNA

Because dental calculus calcifies during life, it does not undergo the same decomposition processes as the rest of the body, and it is nearly ubiquitous in skeletal collections.

within infected bone, and examples of well-preserved soft tissue are relatively rare outside of a few geographic regions. Dental calculus presents a promising alternative for screening ancient skeletons for infectious pathogens. Because dental calculus calcifies during life, it does not undergo the same decomposition processes as the rest of the body, and it is nearly ubiquitous in skeletal collections. 124 DNA within dental calculus has been shown to preserve well over long time scales, and because it forms incrementally, serially entrapping microbes and debris from discrete periods of time, 17 it may even be possible to retrieve pathogen DNA from individuals who survived disease events and were no longer infected at the time of death. This may make it possible to trace the evolutionary history of several diseases that have proven difficult to study

using bone samples, including treponemal diseases such as venereal syphilis, ¹²⁵ which is of particular importance given its historical and clinical significance, ¹²⁵ as well as its past intractability to ancient DNA analysis. ^{126,127}

Although none of the above pathogens has yet been identified from archaeological dental calculus, the fact that so many infectious agents transiently inhabit the oral cavity during disease progression makes future detection of pathogens from dental calculus at least plausible. If successful, such analyses could greatly expand our understanding of human pathogen evolution.

Conclusion

The incorporation of genetic material from commensal, pathobiont and pathogenic microorganisms into dental plaque, and later dental calculus, presents a rare opportunity to study the evolution of the human oral microbiome and associated diseases in archaeological skeletal collections spanning thousands of years. Great progress has already been made in developing the tools and technologies necessary to extract genomic and proteomic information from ancient dental calculus, and clinical research on the oral microbiome is laying the theoretical foundations for making this information relevant in today's dental practices and hospitals. Through collaborations between oral health science and ancient dental calculus research, we can leverage knowledge of the ancestral oral microbiome to improve human health today.

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REFERENCES

- 1. Dewhirst FE, Chen T, Izard J, et al. The human oral microbiome. J Bacteriol 2010;192(19):5002-17.
- 2. Scannapieco FA. The oral microbiome: Its role in health

- and in oral and systemic infections. Clin Microbiol Newsl 2013;35(20):163-69.
- 3. Kumar PS, Mason MR. Mouthguards: Does the indigenous microbiome play a role in maintaining oral health? Front Cell Infect Microbiol 2015;5:35.
- 4. Chen H, Jiang W. Application of high-throughput sequencing in understanding human oral microbiome related with health and disease. Front Microbiol 2014;5:508.
- 5. Cox MJ, Cookson WO, Moffatt MF. Sequencing the human microbiome in health and disease. *Hum Mol Genet* 2013;22{R1}:R88-94.
- 6. Group NHW, Peterson J, Garges S, et al. The NIH Human Microbiome Project. Genome Res 2009;19(12):2317-23. 7. Marsh PD. Are dental diseases examples of ecological
- catastrophes? Microbiology 2003;149(Pt 2):279-94.

 8. Marsh PD. Dental diseases are these examples of ecological catastrophes? Int J Dent Hyg 2006;4 Suppl 1:3-10; discussion 50-2.

 9. Pihlstrom BL, Michalowicz BS, Johnson NW. Periodontal diseases. Lancet 2005;366(9499):1809-20.
- 10. Hillson S. Recording dental caries in archaeological human remains. Int J Osteoarchaeol 2001;11(4):249-89.
- 11. Hujoel P. Dietary carbohydrates and dental-systemic diseases. J Dent Res 2009;88(6):490-502.
- 12. Larsen CS. Bioarchaeology: Interpreting behavior from the human skeleton. Cambridge University Press; 2015.
- 13. Trevathan WR. Evolutionary medicine. A Rev Anthropol 2007;36(1):139.
- Warinner C, Lewis CM Jr. Microbiome and health in past and present human populations. Am Anthropol 2015;in press.
 Dobney K, Brothwell D. Dental calculus: Its relevance to ancient diet and oral ecology. Teeth Anthropol BAR Int Ser 1986;291:55-81.
- Dobney K, Brothwell D. A scanning electron microscope study of archaeological dental calculus. In: Olsen S, editor. Scanning Electron Microscopy in Archaeology. BAR International Series, vol. 452. Oxford: BAR; 1988. p. 372-85.
- 17. Warinner C, Speller C, Collins MJ. A new era in palaeomicrobiology: Prospects for ancient dental calculus as a long-term record of the human oral microbiome. *Philos Trans R Soc Lond B Biol Sci* 2015;370(1660):20130376.
- 18. Adler CJ, Dobney K, Weyrich LS, et al. Sequencing ancient calcified dental plaque shows changes in oral microbiota with dietary shifts of the Neolithic and Industrial revolutions. *Nat Genet* 2013.
- 19. De La Fuente CP, Flores SV, Moraga ML. Human bacterial DNA from dental calculus: A new source of genetic material. Am J Phys Anthropol 2012;147:127-27.
- 20. Warinner C, Rodrigues JF, Vyas R, et al. Pathogens and host immunity in the ancient human oral cavity. Nat Genet 2014;46(4):336-44.
- Fox CL, Juan J, Albert RM. Phytollith analysis on dental calculus, enamel surface and burial soil: Information about diet and paleoenvironment. Am J Phys Anthropol 1996;101(1):101-12
- 22. Hardy K, Blakeney T, Copeland L, et al. Starch granules, dental calculus and new perspectives on ancient diet. J Archaeol Sci 2009;36(2):248-55.
- 23. Warinner C, Hendy J, Speller C, et al. Direct evidence of milk consumption from ancient human dental calculus. Sci Rep 2014;4:7104.
- 24. Schroeder HE. Formation and Inhibition of Dental Calculus. Bern: Hans Huber Publishers; 1969.

- 25. Jin Y, Yip HK. Supragingival calculus: Formation and control. Crit Rev Oral Biol Med 2002;13(5):426-41.
- 26. Lieverse AR. Diet and the aetiology of dental calculus. Int J Osteographical 1999:9(4):219-32.
- 27. White DJ. Dental calculus: Recent insights into occurrence, formation, prevention, removal and oral health effects of supragingival and subgingival deposits. Eur J Oral Sci 1997:105(5):508-22.
- 28. Henry AG, Brooks AS, Piperno DR. Microfossils in calculus demonstrate consumption of plants and cooked foods in Neanderthal diets (Shanidar III, Iraq; Spy I and II, Belgium). Proc Natl Acad Sci U S A 2011;108(2):486-91.
- 29. Henry AG, Ungar PS, Passey BH, et al. The diet of Australopithecus sediba. Nature 2012;487(7405):90-3. 30. Hershkovitz I, Kelly J, Latimer B, et al. Oral bacteria in Miocene Sivapithecus. J Hum Evol 1997;33(4):507-12.
- 31. Linossier A, Gajardo M, Olavarria J. Paleomicrobiological study in dental calculus: Streptococcus mutans. Scanning Microsc 1996;10(4):1005-13; discussion 14.
- Shapiro B, Hofreiter M. A paleogenomic perspective on evolution and gene function: New insights from ancient DNA. Science 2014;343(6169):1236573.
- 33. Cappellini E, Collins MJ, Gilbert MT. Biochemistry. Unlocking ancient protein palimpsests. Science 2014;343(6177):1320-2. 34. Ozga A, Nieves-Colón M, Honap T, et al. Successful enrichment and recovery of whole mitochondrial genomes from ancient human dental calculus. Am J Phys Anthropol 2015;in press. 35. Blatt SH, Redmond BG, Cassman V, Sciulli PW. Dirty teeth and ancient trade: Evidence of cotton fibres in human dental calculus from Late Woodland, Ohio. Int J Osteoarchaeol
- 36. Marthaler TM. Changes in dental caries 1953-2003. Caries Res 2004;38(3):173-81.

2011;21(6):669-78.

- 37. Gordon SC, Kaste LM, Barasch A, et al. Prenuptial dental extractions in Acadian women: First report of a cultural tradition. J Womens Health (Larchmt) 2011;20(12):1813-8.
- 38. Nitschke I, Muller F, Hopfenmuller W. The uptake of dental services by elderly Germans. Gerodontology 2001;18(2):114-20.
- 39. Waldman HB. Dentistry within the British National Health Service. J Am Dent Assoc 1979;99(3):439-47.
- 40. Beltran-Aguilar ED, Barker LK, Canto MT, et al. Surveillance for dental cories, dental sealants, tooth retention, edentulism and enamel fluorosis United States, 1988-1994 and 1999-2002.

 MMWR Surveill Summ 2005;54(3):1-43.
- Lanfranco LP, Eggers S. Caries through time: An anthropological overview. InTech Open Access Publisher; 2012.
 Grine FE, Gwinnett AJ, Oaks JH. Early hominid dental pathology: Interproximal caries in 1.5 million-year-old Paranthropus robustus from Swartkrans. Arch Oral Biol 1990;35(5):381-6.
- 43. Mant M, Roberts C. Diet and Dental Caries in Post-Medieval London. Int J Hist Archaeol 2015;19(1):188-207.
- 44. Gilmore CC. A comparison of antemortem tooth loss in human hunter-gatherers and nonhuman catarrhines: Implications for the identification of behavioral evolution in the human fossil record. Am J Phys Anthropol 2013;151(2):252-64.
- 45. Miyanohara M, Imai S, Okamoto M, et al. Distribution of Streptococcus traglodytae and Streptococcus dentirousetti in chimpanzee oral cavifies. Microbiol Immunol 2013;57(5):359-65. 46. Larsen CS. The agricultural revolution as environmental catastrophe: Implications for health and lifestyle in the Holocene. Quart Int 2006;150(1):12-20.

- 47. Eshed V, Gopher A, Hershkovitz I. Tooth wear and dental pathology at the advent of agriculture: New evidence from the Levant. Am J Phys Anthropol 2006;130(2):145.
- Domett KM. Health in late prehistoric Thailand. BAR; 2001.
 Lubell D, Jackes M, Schwarcz H, Knyf M, Meiklejohn C. The Mesolithic-Neolithic transition in Portugal: Isotopic and dental evidence of diet. J Archaeol Sci 1994;21(2):201-16.
- 50. Humphrey LT, De Groote I, Morales J, et al. Earliest evidence for caries and exploitation of starchy plant foods in Pleistocene hunter-gatherers from Morocco. Proc Natl Acad Sci U S A 2014;111(3):954-9.
- 51. Jones C. The Smile Revolution: In Eighteenth Century Paris: Oxford University Press; 2014.
- 52. Obregon-Tio AJ, Tito RY, Metcalf J, et al. Subsistence strategies in traditional societies distinguish gut microbiomes. *Nat Commun* 2015;6:6505.
- 53. Clemente JC, Pehrsson EC, Blaser MJ, et al. The microbiome of uncontacted Amerindians. Sci Adv 2015;1(3).
- 54. Contreras M, Costello EK, Hidalgo G, et al. The bacterial microbiota in the oral mucosa of rural Amerindians. Microbiology 2010:156(Pt 11):3282-7.
- 55. Simon M, Montiel R, Smerling A, et al. Molecular analysis of ancient caries. *Proc Biol Sci* 2014;281(1790).
- 56. Gross El, Beall CJ, Kutsch SR, et al. Beyond Streptococcus mutans: Ddental caries onset linked to multiple species by 16S rRNA community analysis. PLoS One 2012;7(10):e47722.
 57. Cornejo OE, Lefebure T, Bitar PD, et al. Evolutionary and population genomics of the cavity causing bacteria Streptococcus
- 58. Albandar JM, Brunelle JA, Kingman A. Destructive periodontal disease in adults 30 years of age and older in the United States, 1988-1994. J Periodontal 1999;70(1):13-29. 59. Eke PI, Dye BA, Wei L, et al. Prevalence of periodontitis in adults in the United States: 2009 and 2010. J Dent Res 2012;91(10):914-20.

mutans. Mol Biol Evol 2013;30(4):881-93.

- 60. Kuo LC, Polson AM, Kang T. Associations between periodontal diseases and systemic diseases: A review of the interrelationships and interactions with diabetes, respiratory diseases, cardiovascular diseases and osteoporosis. *Public Health* 2008;122(4):417-33.
- 61. Liao F, Li ZB, Wang YN, et al. Porphyromonas gingivalis may play an important role in the pathogenesis of periodonititis-associated rheumatoid arthritis. Med Hypotheses 2009;72(6):732-35.
- 62. Nakano K, Nemoto H, Nomura R, et al. Detection of oral bacteria in cardiovascular specimens. Oral Microbiol Immunol 2009;24(1):64-68.
- 63. Nakano K, Nomura R, Matsumoto M, Ooshima T. Roles of Oral Bacteria in Cardiovascular Diseases From Molecular Mechanisms to Clinical Cases: Cell-Surface Structures of Novel Serotype k Streptococcus mutans Strains and Their Correlation to Virulence. J Pharmacol Sci 2010;113(2):120-25.
- 64. Scannapieco FA, Bush RB, Paju S. Associations between periodontal disease and risk for atherosclerosis, cardiovascular disease and stroke. A systematic review. *Ann Periodontal* 2003;8(1):38-53.
- 65. Han Y, Wang X. Mobile Microbiome Oral Bacteria in Extraoral Infections and Inflammation. J Dent Res 2013:0022034513487559.
- 66. Socransky SS, Haffajee AD. Periodontal microbial ecology. Periodontal 2000 2005;38:135-87.
- 67. Leech MT, Bartold P. The association between rheumatoid

arthritis and periodontitis. Best Prac Res Clin Rheumatol 2015. 68. Kaur S, White S, Bartold PM. Periodontal disease and rheumatoid arthritis: A systematic review. J Dent Res 2013;92(5):399-408.

69. Al-Katma MK, Bissada NF, Bordeaux JM, Sue J, Askari AD. Control of periodontal infection reduces the severity of active rheumatoid arthritis. J Clin Rheumatol 2007;13(3):134-7.

70. Erciyas K, Sezer U, Ustun K, et al. Effects of periodontal therapy on disease activity and systemic inflammation in rheumatoid arthritis patients. Oral Dis 2013;19(4):394-400.

71. Janket SJ, Baird AE, Chuang SK, Jones JA. Meta-analysis of periodontal disease and risk of coronary heart disease and stroke. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003:95(5):559-69.

72. Buikstra JE, Ubelaker DH. Standards for data collection from human skeletal remains: Proceedings of a seminar at the Field Museum of Natural History (Arkansas Archaeology Research Series 44). Fayetteville Arkansas Archaeological Survey 1994. 73. Care AHCotPo. Parameters of Care. J Periodontol 2000;71(5):847-83.

74. Armitage GC. Learned and unlearned concepts in periodontal diagnostics: A 50-year perspective. *Periodontal* 2000 2013;62(1):20-36.

75. Bartold PM, Van Dyke TE. Periodonitiis: A host-mediated disruption of microbial homeostasis. Unlearning learned concepts. Periodontol 2000 2013;62(1):203-17.

76. Hillson SW. Diet and dental disease. World Archaeol 1979;11(2):147-62.

77. Ziesemer KA, Mann AE, Sankaranarayanan K, et al. Intrinsic challenges in ancient microbiome reconstruction using 16S rRNA gene amplification. Sci Rep 2015;5:16498.

78. Peschken CA, Esdaile JM. Rheumatic diseases in North America's indigenous peoples. Semin Arthritis Rheum 1999;28(6):368-91.

79. Rothschild BM, Woods RJ, Rothschild C, Sebes JI.
Geographic distribution of rheumatoid arthritis in ancient North
America: Implications for pathogenesis. Semin Arthritis Rheum
1992;22(3):181-7.

80. Alamanos Y, Drosos AA. Epidemiology of adult rheumatoid arthritis. *Autoimmun Rev* 2005;4(3):130-36.

81. Kacki S. Erosive polyarthropathy in a Late Roman skeleton from northern France: A new case of rheumatoid arthritis from the pre-Columbian Old Word? IJPP 2013;3(1):59-63.

82. Koren O, Spor A, Felin J, et al. Human oral, gut, and plaque microbiota in patients with atherosclerosis. *Proc Natl Acad Sci U* S A 2011;108 Suppl 1:4592-8.

83. Thomas GS, Wann LS, Allam AH, et al. Why Did Ancient People Have Atherosclerosis[®] From Autopsies to Computed Tomography to Potential Causes. Glob Heart 20 14;9(2):229:37. 84. Kim MJ, Kim YS, Oh CS, et al. Anatomical confirmation of computed tomography-based diagnosis of the atherosclerosis discovered in 17th century Korean mummy. PLoS One 2015;10(3):e0119474.

 Allam AH, Mandour Ali MA, Wann LS, et al. Atherosclerosis in ancient and modern Egyptians: The Horus study. Glob Heart 2014;9(2):197-202.

86. Finch CE, Crimmins EM. Inflammatory exposure and historical changes in human life spans. Science 2004;305(5691):1736-9.
87. Garcia-Rodriguez JA, Fresnadillo Martinez MJ. Dynamics of nasopharyngeal colonization by potential respiratory pathogens.
J Antimicrob Chemother 2002;50 Suppl \$2:59-73.

88. Christensen H, May M, Bowen L, Hickman M, Trotter CL. Meningococcal carriage by age: A systematic review and meta-

analysis, Lancet Infect Dis 2010;10(12):853-61.

89. DeMuri GP, Wald ER. The Group A Streptococcal Carrier State Reviewed: Still an Enigma, J Pediatric Infect Dis Soc 2014;3(4):336-42.

90. Farizo KM, Strebel PM, Chen RT, et al. Fatal respiratory disease due to Corynebacterium diphtheriae: Case report and review of guidelines for management, investigation and control. Clin Infect Dis 1993;16(1):59-68.

91. Tam TW, Bentsi-Enchill A. The return of the 100-day cough:
Resurgence of pertussis in the 1990s. CMAJ 1998;159(6):695-6.
92. Wagner KS, White JM, Neal S, et al. Screening for
Corynebacterium diphtheriae and Corynebacterium ulcerans
in patients with upper respiratory tract infections 20072008: A multicentre European study. Clin Microbiol Infect
2011;17(4):519-25.

93. Bhatta DR, Gokhale S, Sharma AL, et al. Carrier state of Haemophilus influenzae type b (Hib), Streptococcus pneumoniae, Streptococcus pyogenes, Neisseria meningitidis and Corynebacterium diphtheriae among school children in Pokhara, Nepal. Asian Pac J Trop Dis 2014;4(1):45-49.

94. Klement E, Uliel L, Engel I, et al. An outbreak of pertussis among young Israeli soldiers. Epidemiol Infect 2003;131(03):1049-54.

95. Mojon P. Oral health and respiratory infection. J Can Dent Assoc 2002;68(6):340-5.

96, Shay K. Infectious complications of dental and periodontal diseases in the elderly population. *Clin Infect Dis* 2002;34(9):1215-23.

97. Heininger U, Stehr K, Schmitt-Grohe S, et al. Clinical characteristics of illness caused by Bordetella parapertussis compared with illness caused by Bordetella pertussis. Pediatr Infect Dis J 1994:13(4):306-9.

98. Jones DS, Podolsky SH, Greene JA. The burden of disease and the changing task of medicine. N Engl J Med 2012;366(25):2333-8.

99. Lambert PM. Rib lesions in a prehistoric Puebloan sample from southwestern Colorado. Am J Phys Anthropol 2002;117(4):281-92. 100. Aufderheide A, Wittmers L, Arriaza B. Pneumonia in antiquity: A comparison between two preantibiotic population samples from northern Chile and the United States. Chungará, Revista de Antropología Chilena 2008;40(2):173-80. 101. Aufderheide AC. Progress in soft tissue paleopathology. JAMA 2000:284(20):2571-3.

102. Aufderheide AC, Aturaliya S, Focacci G. Pulmonary disease in a sample of mummies from the AZ-75 cemetery in northern Chile's Azapa Valley. Chungará 2002:253-63.

103. Marri PR, Paniscus M, Weyand NJ, et al. Genome sequencing reveals widespread virulence gene exchange among human Neisseria species. PLoS One 2010;5(7):e11835.

104. Stephens DS. Biology and pathogenesis of the evolutionarily successful, obligate human bacterium Neisseria meningitidis. Vaccine 2009;27 Suppl 2:B71-7.

105. Schoen C, Blom J, Claus H, et al. Whole-genome comparison of disease and carriage strains provides insights into virulence evolution in Neisseria meningitidis. Proc Natl Acad Sci U S A 2008;105(9):3473-8.

106. Ortner DJ. Identification of pathological conditions in human skeletal remains. Academic Press; 2003.

107. Cheung MK, Lam WY, Fung WY, et al. Sputum microbiota in tuberculosis as revealed by 16S rRNA pyrosequencing. PLoS One 2013;8(1):e54574.

108. Eguchi J, Ishihara K, Watanabe A, Fukumoto Y, Okuda K. PCR method is essential for detecting Mycobacterium tuberculosis in oral cavity samples. Oral Microbiol Immunol 2003;18(3):156-9.

109. Pallagatti S, Sheikh S, Kaur A, Aggarwal A, Singh R. Oral cavity and leprosy. *Indian Dermatol Online J* 2012;3(2):101-4. 110. Dave B, Bedi R. Leprosy and its dental management guidelines. *Int Dent J* 2013;63(2):65-71.

111. Lathem WW, Price PA, Miller VL, Goldman WE. A plasminogen-activating protease specifically controls the development of primary pneumonic plague. Science 2007;315(5811):509-13.

112. Scott CM, Flint SR. Oral syphilis – re-emergence of an old disease with oral manifestations. *Int J Oral Maxillofac Surg* 2005;34(1):58-63.

113. Gebara EC, Pannuti C, Faria CM, et al. Prevalence of Helicobacter pylori detected by polymerase chain reaction in the oral cavity of periodontitis patients. Oral Microbiol Immunol 2004;19(4):277-80.

114. Breman JG, Henderson DA. Diagnosis and management of smallpox. N Engl J Med 2002;346(17):1300-8.

115. Bos KI, Harkins KM, Herbig A, et al. Pre-Columbian mycobacterial genomes reveal seals as a source of New World human tuberculosis. *Nature* 2014;514(7523):494-7.

116. Bouwman AS, Kennedy SL, Muller R, et al. Genotype of a historic strain of Mycobacterium tuberculosis. Proc Natl Acad Sci U S A 2012;109(45):18511-6.

117. Bos KI, Schuenemann VJ, Golding GB, et al. A draft genome of Yersinia pestis from victims of the Black Death. Nature 2011;478(7370):506-10.

118. Rasmussen S, Allentoff ME, Nielsen K, et al. Early divergent strains of Yersinia pestis in Eurasia 5,000 years ago. Cell 2015;163(3):571-82.

119. Wagner DM, Klunk J, Harbeck M, et al. Yersinia pestis and the plague of Justinian 541-543 AD: a genomic analysis. Lancet Infect Dis 2014;14(4):319-26.

120. Maixner F, Krause-Kyora B, Turaev D, et al. The 5300-yearold Helicobacter pylori genome of the Iceman. Science 2016;351(6269):162-5.

121. Schuenemann VJ, Singh P, Mendum TA, et al. Genome-wide comparison of medieval and modern Mycobacterium leprae. Science 2013;341(6142):179-83.

122. Kolman CJ, Centurion-Lara A, Lukehart SA, Owsley DW, Tuross N. Identification of Treponema pallidum subspecies pallidum in a 200-year-old skeletal specimen. J Infect Dis 1999;180(6):2060-3.

123. Biagini P, Theves C, Balaresque P, et al. Variola virus in a 300-year-old Siberian mummy. N Engl J Med 2012;367(21):2057-9.

124. Warinner C, Speller C, Collins MJ, Lewis CM Jr. Ancient human microbiomes. *J Hum Evol* 2015;79:125-36.

125, Powell ML, Cook DC. The myth of syphilis: The natural history of treponematosis in North America. University Press of Florida; 2005.

126. Bouwman AS, Brown TA. The limits of biomolecular palaeopathology: Ancient DNA cannot be used to study venereal syphilis. J Archaeol Sci 2005;32(5):703-13.

127. von Hunnius TE, Yang D, Eng B, Waye JS, Saunders SR. Digging deeper into the limits of ancient DNA research on syphilis. J Archaeol Sci 2007;34(12):2091-100.

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