


Original Article

Release of resource constraints allows greater carbon allocation to secondary metabolites and storage in winter wheat

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ABSTRACT

The atmospheric CO₂ concentration ([CO₂]) is rapidly increasing, and this may have substantial impact on how plants allocate metabolic resources. A thorough understanding of allocation priorities can be achieved by modifying [CO₂] over a large gradient, including low [CO₂], thereby altering plant carbon (C) availability. Such information is of critical importance for understanding plant responses to global environmental change. We quantified the percentage of daytime whole-plant net assimilation (A) allocated to night-time respiration (R), structural growth (SG), nonstructural carbohydrates (NSC) and secondary metabolites (SMs) during 8 weeks of vegetative growth in winter wheat (*Triticum aestivum*) growing at low, ambient and elevated [CO₂] (170, 390 and 680 ppm). R/A remained relatively constant over a large gradient of [CO₂]. However, with increasing C availability, the fraction of assimilation allocated to biomass (SG + NSC + SMs), in particular NSC and SMs, increased. At low [CO₂], biomass and NSC increased in leaves but decreased in stems and roots, which may help plants achieve a functional equilibrium, that is, overcome the most severe resource limitation. These results reveal that increasing C availability from rising [CO₂] releases allocation constraints, thereby allowing greater investment into long-term survival in the form of NSC and SMs.

Key-words: carbon allocation; CO₂; growth; respiration; storage carbohydrates.

INTRODUCTION

For more than 40 years, the control mechanisms of whole-plant carbon (C) allocation have been of central interest to plant scientists (Mooney, 1972). Plants fix CO₂ from the atmosphere and partition the resulting photosynthetic products (carbohydrates) among several uses, including growth of structural biomass, synthesis of secondary metabolites and metabolic processes like respiration and osmoregulation. The atmospheric CO₂ concentration ([CO₂]) has increased from 170 ppm during glacial periods of the past million years to the

current ~400 ppm and is expected to increase up to 430 ppm to 1000 ppm by the year 2100 (Cubasch *et al.*, 2013). However, little is known about how increased [CO₂] alters partitioning of fixed C among different sinks or about the processes regulating allocation (Dietze *et al.*, 2014).

Historically, the regulation of plant C partitioning was thought to be driven by C supply (i.e. source activity), but recent evidence suggests that sink activity may be the main determinant (Fatichi *et al.*, 2014; Korner, 2015). Experiments manipulating source activity via [CO₂] availability can elucidate the contributions of sources and sinks for whole-plant C allocation (Fatichi *et al.*, 2014). However, such studies are still sparse, not only because of the expense of manipulating [CO₂] (Gerhart & Ward, 2010), but also because of challenges in assessing both whole-plant C balance (assimilation minus respiration; A – R) and how non-respired C is distributed among sink components, that is, structural growth (SG), non-structural carbohydrates (NSC), secondary metabolites (SMs) and export (e.g. to rhizosphere symbionts or as volatile organic compounds (VOCs)) (Fig. 1).

In many plant species, short-term stimulation of leaf-level photosynthesis by elevated [CO₂] (Franks *et al.*, 2013; Gerhart & Ward, 2010) is often associated with down-regulation of photosynthesis in the long term (Lee *et al.*, 2011; Long *et al.*, 2004). However, leaf-level net photosynthetic rates cannot be directly scaled up to whole-plant assimilation for a number of reasons. For example, leaf-level assimilation measurements are usually done under ideal light and temperature conditions (Kirschbaum, 2011) and therefore may overestimate actual assimilation rates. Scaling up from leaf to whole-plant gas exchange also requires parameterization of leaf phenology and light distribution. In addition, photosynthesis in non-leaf tissues (bark, stems) may not scale in the same way as in leaves. Therefore, direct whole-plant assessment of gas exchange is a more suitable method for determining whole-plant responses to different [CO₂] (Atkin *et al.*, 2007; Boote *et al.*, 2013).

Respiration (R) is an essential plant functional process providing energy and metabolic intermediates needed for photosynthesis, which in turn provides substrates for R. Some studies have reported that whole-plant R may account for up to 30–50% of the gross photosynthetic CO₂ uptake in herbaceous plants, and due to the interdependency of these processes this proportion remains relatively constant under a gradient of

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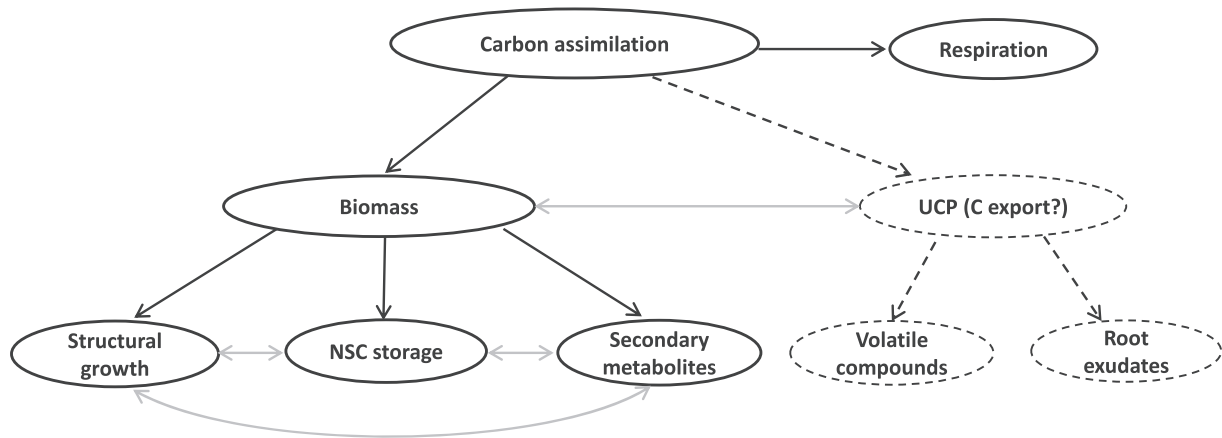


Figure 1. Conceptual model for trade-offs in carbon allocation to functional pools respiration, biomass (structural growth + non-structural carbohydrates (NSC) + secondary metabolites) and an – operationally defined – unaccounted carbon pool (UCP, potentially export to volatile compounds and root exudates). Black solid arrows indicate established (based on literature) carbon flow between pools, while black dashed arrows indicate flows that remain uncertain mainly due to technical difficulties in measuring them. Such flows require further investigation (see Discussion and Outlook sections). Grey solid arrows indicate trade-off situation between different carbon pools, as suggested by growth-defence hypotheses and different carbon storage paradigms.

environmental conditions (Atkin *et al.*, 2007; Gifford, 1995; Pons & Poorter, 2014). In our study, we measured net C assimilation; during the day-time, this is the difference between A and R; at night, as there is no assimilation it represents only R.

Species adapted to low resource availability have intrinsically lower growth rates than species adapted to high resource availability (Endara & Coley, 2011). Therefore, modern plants adapted to current $[\text{CO}_2]$ may not be able to utilize higher $[\text{CO}_2]$ assimilation for growth (Tissue & Lewis, 2012). Under elevated $[\text{CO}_2]$, growth may also be constrained by environment factors such as water and nutrients (Kirschbaum, 2011; Palacio *et al.*, 2014). A 30–40% increase in leaf-level photosynthesis leads only to a 10% increase in relative growth rate (reviewed in Kirschbaum, 2011). By contrast, plants grown at low $[\text{CO}_2]$ may fully utilize all assimilated C for growth while concurrently reduce C loss to exports, particularly in the form of VOCs and rhizodeposits, at least when nutrients are provided and freely accessible (Fig. 1).

Plants maintain large pools of NSC compounds that can enhance plant survival during stresses (Dietze *et al.*, 2014; Hartmann & Trumbore, 2016), and strategies of C allocation to NSC pools have been discussed for more than 25 years (Chapin *et al.*, 1990). NSC pools have often been considered as reservoirs filled with C that is available in excess of demand (accumulation of NSC) (Palacio *et al.*, 2014). However, an alternate model suggests that plants allocate C to NSC to ensure future C availability even at the expense of other C sinks such as SG (reviewed in Dietze *et al.*, 2014; Wiley & Helliker, 2012). Recent studies provide evidence of C allocation to NSC storage even in half defoliated *Quercus velutina* saplings (Wiley *et al.*, 2013) and in *Picea abies* saplings grown under low $[\text{CO}_2]$ (Hartmann *et al.*, 2015).

Plants may allocate a substantial proportion of available C to SMs that play important roles in biotic and abiotic interactions (Neilson *et al.*, 2013). For example, glucosinolates are one of the major classes of defensive SMs (Halkier & Gershenzon,

2006), and up to 15% of total photosynthetic energy may be required for glucosinolate production in *Arabidopsis thaliana* (Bekaert *et al.*, 2012). This large C investment, however, may divert resources away from other metabolic processes. The growth-differentiation balance hypothesis (GDBH) (Herms & Mattson, 1992) suggests that allocation to SMs is driven by C availability in excess of growth demand. Under ideal conditions, this trade-off may be less constrained, and plants can grow larger leaf canopies with higher SM production than under resource limitation. In support of this hypothesis, C-based SMs increased under elevated $[\text{CO}_2]$ (Robinson *et al.*, 2012), while O'Neill *et al.* (2010) found that only quercetin triglycoside increased but genistein decreased in soybean (*Glycine max*) foliage.

Trade-off situations in allocation occur not only across functional sinks but also across plant tissues and organs. Plants are thought to invest C preferentially into tissues that are responsible for acquiring the most limiting resource; this is termed ‘functional equilibrium’. Such an optimal allocation strategy has been observed in biomass partitioning under low $[\text{CO}_2]$ where plants allocated proportionally more C to above-ground than to belowground tissues (Poorter *et al.*, 2012; Temme *et al.*, 2013; Zhang *et al.*, 2015), but data on the partitioning to different functional sinks, like NSC (Hartmann *et al.*, 2015) and SMs, and across different plant tissues and organs are still very sparse.

The objective of this study was to investigate C allocation tradeoffs in winter wheat (*Triticum aestivum* L.) during the vegetative growth phase. Previous studies have shown that elevated $[\text{CO}_2]$ will cause photosynthetic acclimation of wheat plants (Aranjuelo *et al.*, 2011; Aranjuelo *et al.*, 2015), but much less is known about allocation priorities across functional sinks under these conditions. Such priorities can be deduced from assessments of allocation patterns along a plant C availability gradient, including low $[\text{CO}_2]$ (Hartmann *et al.*, 2015). Although shading may be a much simpler

approach for reducing plant C availability, it may also trigger phytochrome-induced growth stimulation (Casal, 2013) and defence suppression (Ballare, 2014), independent of C availability. Direct manipulations of atmospheric $[\text{CO}_2]$ suffer less from such side effects and allow plants to adjust to C (rather than light) availability.

Because allocation is a continuous process, the whole-plant fluxes ($A - R$ derived from gas exchange) and pools were monitored over the vegetative growth phase using a unique chamber design implemented in a greenhouse. Supplemental nutrients were applied to the substrate in order to minimize nutrient limitations on plants. Allocation to reproduction (seed production) was purposely excluded in this study because pollination has been shown to be problematic and incomplete in these growth chambers, likely due to light limitation. Changes in primary (NSC, including soluble sugars, starch and fructans) and SMs (including flavonoid glycosides and benzoxazinoids) of plant tissues (leaves, stems and roots) were used to partition biomass into SG ($\text{SG} = \text{Biomass} - \text{NSC} - \text{SMs}$) and nonstructural components (NSC and SMs). Specifically, we hypothesized that, with increasing $[\text{CO}_2]$ (170, 390 and 680 ppm): (1) the proportion of C allocated to R remains constant but decreases in biomass ($\text{SG} + \text{NSC} + \text{SMs}$) (relative to A); (2) NSC and SMs increase while SG decreases; and (3) biomass ($\text{SG} + \text{NSC} + \text{SMs}$) increases in stems and roots but decreases in leaves, in accordance with the functional equilibrium hypothesis.

MATERIALS AND METHODS

Plant material

A cultivar of winter wheat from a local seed plantation (Bioland Hof Jeebel Biogartenversand, Salzwedel, Germany) adapted to Central Europe was used in our study. On 7 January 2015, seeds were germinated in plates filled with sand and were watered every day. On 13 and 14 January 2015, seedlings of

similar height were transplanted into pots (diameter 11 cm, height 24 cm) filled with C-free quartz sand. Four pots (12 seedlings per pot) and two pots (10 seedlings per pot) were randomly placed in each plant chamber immediately after transplanting (Fig. 2). We provided the plants with supplemental nutrients by watering pots with a continuous through-flow of a modified Hoagland nutrient solution (Hoagland & Arnon, 1950) to avoid nutrient limitation (other than C).

Growth chambers

Twelve aquarium-style glass chambers (80 cm high \times 75 cm long \times 45 cm wide) were used to isolate the atmosphere around the plants and allowed controlling $[\text{CO}_2]$ (Fig. 2). Chambers were placed on a greenhouse table covered with closed-cell rubber foam mats made of ethylene propylene diene monomer for airtight sealing (as in Hartmann *et al.* (2013)). Air in the chambers was flushed continuously at a flow rate of 14 l min^{-1} through inlet tubing located at the bottom and outlet tubing at the top of the chamber. The $[\text{CO}_2]$ of incoming air was controlled by first scrubbing all CO_2 from atmospheric air using a molecular sieve and then adding pure CO_2 from a gas tank (Schnyder, 1992). The CO_2 -free air was separated into three air circuits (four chambers per circuit); in each circuit, $[\text{CO}_2]$ was measured sequentially with a Vaisala® (*GMP 343*) at intervals of 10 min, using a custom-built switching valve controlled by a micro-logger (Campbell® *CR1000*). Measured $[\text{CO}_2]$ were compared against pre-set values (170, 390 and 680 ppm, respectively), and, if required, CO_2 supply to the airstream was adjusted via mass flow controllers to maintain target $[\text{CO}_2]$ levels. To minimize differences in light levels, treatments were randomly assigned on the table, and three of four chambers per treatment were used to grow plants while the other one was used to monitor $[\text{CO}_2]$ directly in the chamber.

In each chamber, air temperature and photosynthetic photon flux density were monitored continuously (see Hartmann *et al.*, 2013). During the day, average temperatures increased

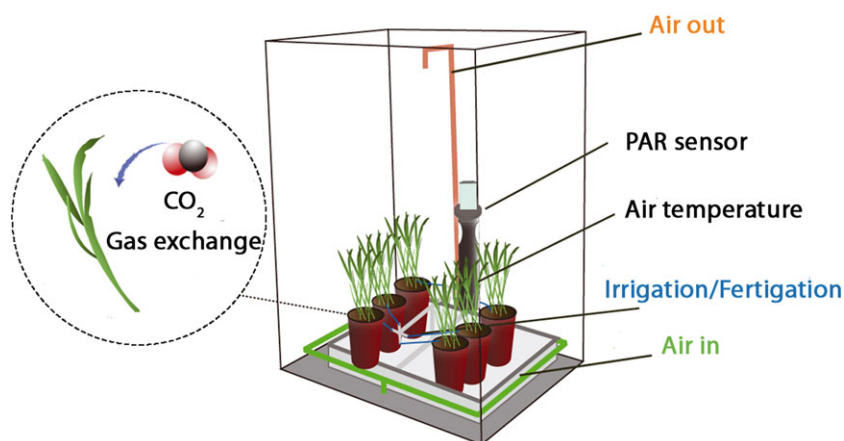


Figure 2. Schematic view of one of the 12 chambers showing the ventilation system and the photosynthetically active radiation (PAR) sensor. Air was flushed continuously through inlet tubing located at the bottom and outlet tubing at the top of the chamber. Four pots (12 seedlings per pot) and two pots (10 seedlings per pot) were randomly placed in each plant chamber. Note that only pots containing 12 seedlings were harvested. Supplemental nutrients were provided by watering the pots with a continuous through-flow of a modified Hoagland nutrient solution.

from ca. 12 °C at 6:30 (local time) to ca. 20–24 °C in the afternoon, and then decreased to ca. 16.5 °C at 22:00. During the night, air temperature declined from ca. 16 to 11 °C. All plants were grown in a 16/8 h light/dark regime using supplemental greenhouse lamps (Son-T Agro® 430 W HPS bulbs, primary light range = 520–610 nm, Philips® Lighting Co., Somerset, NJ, USA). The average daily total photosynthetic photon flux density inside the chambers from 23 January to 20 March was $7.79 \pm 0.92 \text{ mol m}^{-2} \text{ d}^{-1}$.

Destructive harvesting and biomass processing

Allocation priorities likely differ between growth stages of winter wheat (seedling, tillering, stem elongation, flowering and grain filling), and we concentrated on the vegetative growth period (seedling and tillering stage) to rule out the potential confounding effects of developmental stage on plant allocation. According to the growth scale of Zadoks *et al.* (1974), seedling growth of winter wheat is achieved before the occurrence of nine unfolded leaves on the main shoot. Plants were therefore harvested when three, six and eight leaf sheaths were completely developed, denoted as 3 L, 6 L and 8 L periods, respectively. For 390 and 680 ppm treatments, this occurred 3, 7 and 9 weeks after transplanting, respectively, while development in the low [CO₂] treatment was slower, and plants were sampled 3.5, 8 and 10.5 weeks after transplanting. Sampling was always conducted between 16:00 and 21:00. At each sampling, three pots (12 seedlings per pot) from three chambers of each [CO₂] treatment were destructively harvested. Fresh biomass of whole-plant leaves, stems and roots of each pot were determined, and then all tissues were shock-frozen in liquid nitrogen and stored at –80 °C, freeze-dried and weighted for dry biomass. Dry tissues were homogenized and ground to fine powder using a ball mill (Retsch® MM400, Haan, Germany) and stored at –20 °C until analysis.

Whole-plant gas exchange

Gas exchange data were measured with a Picarro® 2101-i (precision 0.01–0.4%, Picarro Inc. Santa Clara, CA, USA) for the first 7 weeks (extending beyond the 6 L period) under 680, 390 and 170 ppm [CO₂]. However, because the Picarro failed after week 7, gas exchange for week 8, 9, 10 and 11 were measured with a Spectronus® FTIR (Fourier transform infrared spectroscopy, Ecotech Pty Ltd., Knoxfield, VIC, Australia) using otherwise the same hardware and the same sequence protocol as mentioned above. Reference gases were used to verify and ensure comparability across devices.

Within 2 h, the [CO₂] of air leaving the 12 chambers and 6 reference air samples (taken from the air circuit) were measured sequentially at intervals of 6 min 40 s, and the sequence was controlled by a micro-logger (Campbell® CR1000) and a custom-built switching valve unit. Transition periods after valve switching were excluded from analysis, and a core period of 3 min 20 s was used to estimate instantaneous whole-plant gas exchange, defined as:

$$\Delta[\text{CO}_2] = \frac{[\text{CO}_2]_{\text{non-plant}} - [\text{CO}_2]_{\text{plant}}}{M} \quad (1)$$

where [CO₂]_{non-plant} is the [CO₂] of outgoing air from the chamber without plants, and [CO₂]_{plant} is the [CO₂] of outgoing air from chambers with plants. *M* is the number of plants grown in the chamber.

The whole-plant gas exchange rate was assumed to be constant within the 2 h cycle and whole-plant C flux at hour *j* was calculated as:

$$C_j \text{ (g h}^{-1}\text{)} = \frac{\Delta[\text{CO}_2] (\mu\text{mol mol}^{-1}) * \text{VFR (l min}^{-1}\text{)} * 60 \text{ min} * 12 \text{ (g mol}^{-1}\text{)}}{22.4 \text{ (l mol}^{-1}\text{)} * 1000000} \quad (2)$$

where VFR is the volumetric flow rates of air going through the chambers, that is, 14 l min^{–1}, and 22.4 l mol^{–1} is the molar volume of gas at normal conditions. The cumulative daily whole-plant net C assimilation (*A*_{cum,*n*}) and night-time respiration (*R*_{cum,*n*}) over the duration of the experiment (*n* = number of days *d*, *j* = hour of day) were then computed using the following equation:

$$A_{\text{cum},n} = \sum_{i=1}^n \sum_{j=7}^{22} C_j \quad (3)$$

$$R_{\text{cum},n} = \sum_{i=1}^n \sum_{j=23}^{24} C_j + \sum_{i=1}^n \sum_{j=1}^{06} C_j \quad (4)$$

*A*_{cum,*n*} and *R*_{cum,*n*} between 3 L and 6 L were used to calculate the fraction of C allocated to R. Whole-plant A and R collected over the last two or three days prior to biomass sampling were averaged for calculations of mass-based (i.e. specific) rates of daily net assimilation and respiration.

While C-free quartz sand was used to avoid any C input into the system other than from plant photosynthesis, it was technically impossible to sterilize the all components of such a greenhouse facility (air, water, plant material) to a degree that would completely avoid microbial infestation. Hence, microbial respiration of root exudates or litter may be included in the gas exchange measurements. However, given that root exudation (Grayston *et al.*, 1997; Phillips *et al.*, 2009) represents a much smaller C flux than respiration (<10% versus 30–50%) (Atkin *et al.*, 2007; Gifford, 1995; Pons & Poorter, 2014), gas exchange data represent mainly plant assimilation and respiration (Hartmann *et al.*, 2015; Hartmann *et al.*, 2013).

NSC analysis

Concentrations of soluble sugars and starch in plant tissues were determined using methods described in Raessler *et al.* (2010). For soluble sugars, usually 1 ml (or 0.5 ml for small samples) of sterilized water was added to 50 mg (or 10 mg for small samples) dry biomass. The mixture was vortexed, incubated at 65 °C for 10 min and then centrifuged at 12 000 *g* for 10 min. The supernatant was collected and stored on ice, and the remaining pellet was re-extracted twice using the same procedure. The supernatants were pooled and then diluted at a ratio of 1:20 (or 1:8 for small samples) and stored at –20 °C. For starch, 50 mg (or 10 mg for small samples) dry biomass

were extracted with 0.35 ml (or 0.175 ml for small samples) of water and 0.5 ml (or 0.25 ml for small samples) of perchloric acid (52%). The mixture was vortexed, incubated at 65 °C for 10 min and then centrifuged at 12 000 *g* for 10 min. The supernatant was collected, and the remaining pellet was re-extracted. The supernatants were pooled and then diluted at a ratio of 1:55, stored at –20 °C.

Sucrose, glucose and fructose of both soluble sugars and starch extractions were determined by High-Performance Liquid Chromatography coupled with Pulsed Amperometric Detection (HPLC-PAD), using a Dionex® ICS 3000 ion chromatography system equipped with an autosampler (Thermo Fisher GmbH, Idstein, Germany). Starch concentrations were calculated by subtracting glucose and half of the sucrose concentrations in water-soluble extract from glucose concentrations in the hydrolyzed extract then multiplying a conversion factor of 0.9 (Sullivan, 1935). Fructan concentrations were calculated using the same method but with fructose instead of glucose concentrations, given that fructan is one of the most important C storage compounds in wheat (Pollock & Cairns, 1991). Total NSC concentrations reported are the sum of soluble sugars, starch and fructans.

SM analysis

Samples from 680 ppm at 8 L were used for the identification of major SMs in leaf, stem and root tissues; 800 µl methanol and two glass beads were added to 30 mg of dry biomass. The mixture was bead-beaten for 40 s at 6.0 m s⁻¹ with a FastPrep Instrument (MP Biomedicals, Santa Ana, USA), then vortexed and centrifuged at 13 000 *g* for 5 min. The supernatant was collected and then analysed by HPLC coupled to a mass spectrometer. Compounds were separated on a Nucleodur Sphinx RP18ec column with dimensions of 250 × 4.6 mm and a particle size of 5 µm (Macherey Nagel, Dueren, Germany) using an Agilent 1100 series HPLC with a flow rate of 1.0 ml min⁻¹. The column temperature was maintained at 20 °C. Phenolic compounds were separated using 0.2% (v/v) formic acid and acetonitrile as mobile phases A and B, respectively, with the following elution profile: 0–28 min, 5–61% B in A; 28–30 min 100% B; and 30–35 min 5% B.

Compound detection and quantification were accomplished with an Esquire 6000 ESI ion-trap mass spectrometer (Bruker Daltonics, Bremen, Germany). Flow coming from the column was diverted in a ratio of 4:1 before entering the mass spectrometer electrospray chamber. ESI-MS was operated in negative mode scanning *m z*⁻¹ between 50 and 1600 with an optimal target mass of 400 *m z*⁻¹. The mass spectrometer was operated using the following specifications: skimmer voltage, 60 V; capillary voltage, 4200 V; nebulizer pressure, 35 psi; drying gas, 11 l min⁻¹; gas temperature, 330 °C. Capillary exit potential was kept at –121 V. Compounds of leaves and stems were identified by comparing the fragmentation patterns with previously reported wheat phenolic profiles (Moheb *et al.*, 2011; Wojakowska *et al.*, 2013). Root compounds were identified based on profiles of benzoxazinoids in grasses (Wouters *et al.*, 2014).

For quantification, 500 (300) µl 95% methanol and two glass beads were added into 50 (30) mg samples, the mixture extracted using the same procedure described above but the remaining pellet was re-extracted again. Supernatants were pooled and then analysed by HPLC-UV using the same chromatographic conditions as above. The UV wavelengths 240, 260, 280 and 330 nm were monitored. All compounds were quantified using external standard curves. Feruoylputrescine (HPC Standards GmbH, Cunnorsdorf, Germany) was used to quantify ferulic acid-based compounds in leaves and stems, and also putrescine-containing compounds in roots. Luteolin 6-C-glucoside and apigenin 6-C-glucoside (Sigma-Aldrich) were used to quantify luteolin-based and apigenin-based compounds in leaves, respectively. Chrysoeriol (LGC standards, Middlesex, UK) was used to quantify chrysoeriol-based and tricrin-based compounds in leaves. DIMBOA-Glc and HDMBOA-Glc in roots were quantified by standards DIMBOA-Glc and HDMBOA-Glc purified from maize roots (Wouters *et al.*, 2014). Leaf and stem compounds were quantified at a UV absorption spectrum of 330 nm, and root compounds at 280 nm.

Elemental analysis

C and N concentrations of tissues were determined by an elemental analyser (varioEL II). To do so, 15 mg dry samples were weighed into tin foil, combusted and separated by specific columns, and then assessed with a thermal conductivity detector.

C balance

Tissue-specific C, NSC and SM content was calculated by multiplying tissue-specific C (Supporting Information Table S1), NSC and SM concentrations by tissue dry mass. C content in tissue-specific NSC and SM was then calculated by multiplying tissue-specific NSC and SM content by their mass proportion of C (0.4 for NSC and specific fraction to each compound for the different SMs (Supporting Information Table S2)). C allocation to biomass, NSC and SMs was defined as the difference in C content of whole-plant biomass, NSC and SMs between 6 L and 3 L divided by the difference in $A_{cum,n}$ between 6 L and 3 L. Note that quantifying allocation to biomass requires relatively large increase in biomass; therefore, C allocation between 8 L and 6 L is not reported, given that average biomass increment of low [CO₂] plants was smaller than its standard deviation within and across chambers.

When net C gain estimated from gas exchange exceeded the C that accumulated in biomass, the difference was attributed to an ‘unaccounted carbon pool’ (UCP). UCP was calculated by mass balance using the following equation:

$$UCP = A_{cum,n} - R_{cum,n} - SG - NSC - SMs. \quad (5)$$

Data analysis

All of the data were analysed using Levene tests and Tukey’s HSD tests that treated each growth chamber as a biological

replicate ($n=3$), to check homogeneity of variances and detect significant differences between treatments, respectively. Data were log-transformed when variance was not homoscedastic. All statistical analysis was conducted in R, version 3.23 (R Development Core Team, 2014).

RESULTS

Whole-plant gas exchange and tissue biomass

Whole-plant daily assimilation, respiration and net carbon gain increased with increasing $[\text{CO}_2]$, but the increase was much stronger at 170 ppm (compared to 390 ppm) than at 390 ppm (compared to 680 ppm) (Fig. 3a–c). However, after 9 weeks, plants grown at 390 and 680 ppm exhibited no significant difference in day-time assimilation (A), night-time respiration (R) and net carbon gain ($A - R$; Fig. 3a–c). Initially, R/A in plants grown at 170 and 390 ppm $[\text{CO}_2]$ were higher than that in plants grown at 680 ppm $[\text{CO}_2]$, but the difference was not statistically significant and disappeared over time (Fig. 3d). At 3 L, mass-based A increased with increasing $[\text{CO}_2]$ (Fig. 4a). Mass-based R was higher at 390 and 680 ppm $[\text{CO}_2]$ than at 170 ppm $[\text{CO}_2]$, but there was no significant difference between the two higher $[\text{CO}_2]$ treatments (Fig. 4b). Both mass-based A and R declined strongly over time at 390 and 680 ppm $[\text{CO}_2]$ but much less at 170 ppm $[\text{CO}_2]$ (Fig. 4a,b).

Biomass varied with $[\text{CO}_2]$ in a way similar to whole-plant assimilation and respiration. At 3 L and 6 L, the dry mass of all tissues increased with increasing $[\text{CO}_2]$, and the increase was much greater comparing plants grown between 170 and

390 ppm than between 390 and 680 ppm (Fig. 5a–d). However, at 8 L, there was no significant difference in all tissues between 390 and 680 ppm $[\text{CO}_2]$ treatments (Fig. 5a–d).

NSC and SMs

The response of soluble sugars and total NSC (soluble sugars + starch + fructans) to $[\text{CO}_2]$ differed between tissues and developmental stages. At 3 L, plant leaves grown at 170 ppm $[\text{CO}_2]$ had slightly higher concentrations of soluble sugars and total NSC than plant leaves grown at 390 ppm $[\text{CO}_2]$, but the opposite was observed in stems and roots (Fig. 6a–f). However, while the concentrations of soluble sugars and total NSC (soluble sugars + starch + fructans) remained relatively constant at 170 ppm $[\text{CO}_2]$ between 3 L and 6 L, they doubled at 390 ppm $[\text{CO}_2]$ and increased by more than three times at 680 ppm $[\text{CO}_2]$ over the same period (Fig. 6a–f). Whole-plant NSC content increased in proportions that were much higher than increases in total plant biomass (Fig. 7). At 6 L, concentrations of soluble sugars and total NSC (soluble sugars + starch + fructans) were significantly higher at 680 ppm than at 390 ppm $[\text{CO}_2]$ while the difference between 390 and 170 ppm $[\text{CO}_2]$ was much smaller. Between 6 L and 8 L, the concentrations of soluble sugars and total NSC (soluble sugars + starch + fructans) increased at 390 ppm $[\text{CO}_2]$ but showed a declining trend at 680 ppm $[\text{CO}_2]$ (Fig. 6a–f).

SM concentrations were much higher in leaves and roots than that in stems (Fig. 6g–i). Leaf SM concentrations significantly increased with increasing $[\text{CO}_2]$ over all developmental stages (Fig. 6g). At 3 L, stem SM concentrations significantly

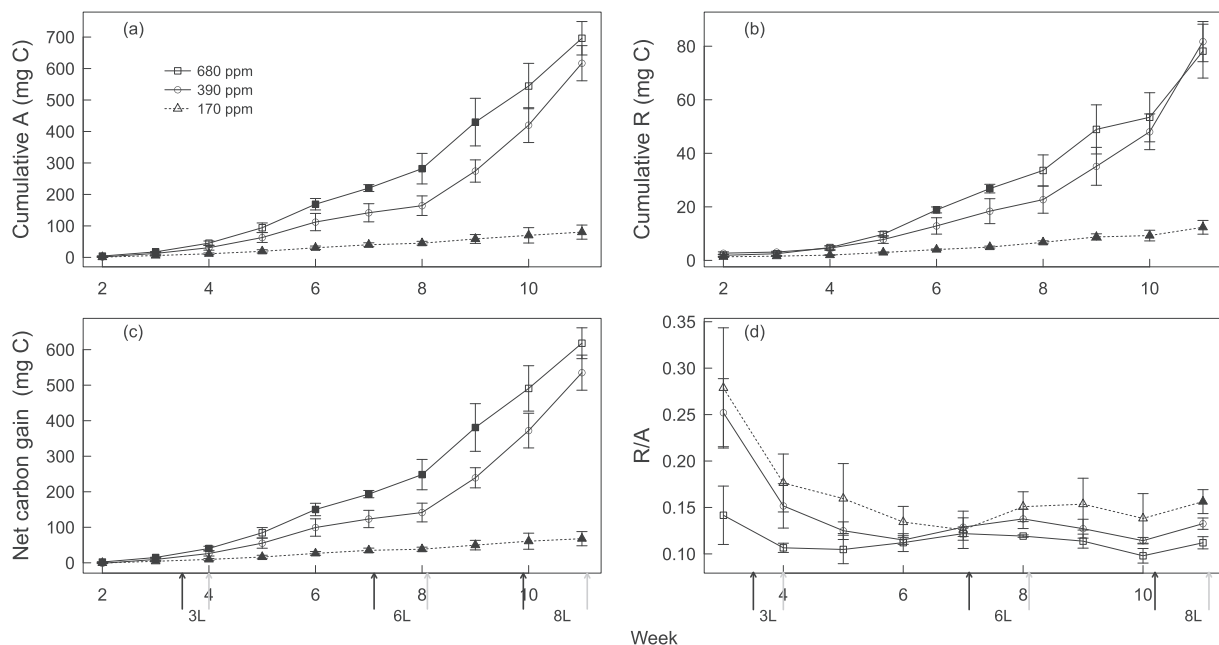


Figure 3. Weekly cumulative whole-plant net carbon assimilation A (a), respiration R (b), net carbon gain (c) and R/A (d) of winter wheat (*Triticum aestivum* cv. Genius) for the three $[\text{CO}_2]$ treatments: 680 ppm $[\text{CO}_2]$ (squares, black line); 390 ppm $[\text{CO}_2]$ (circles, black line); 170 ppm $[\text{CO}_2]$ (triangles, dashed line). Values are the means (mg C) of three individual chambers; error bars represent ± 1 SD. Black arrows indicate sampling dates for 680 and 390 ppm $[\text{CO}_2]$ treatments; grey arrows indicate sampling dates for 170 ppm $[\text{CO}_2]$ treatment. Significant differences between 680 and 170 ppm $[\text{CO}_2]$ treatments compared to ambient $[\text{CO}_2]$ (390 ppm) are indicated by filling of symbols ($P < 0.05$, Tukey's HSD). Harvests were carried out after emergence of three, six and eight leaf sheaths; denoted as 3 L, 6 L and 8 L periods, respectively.

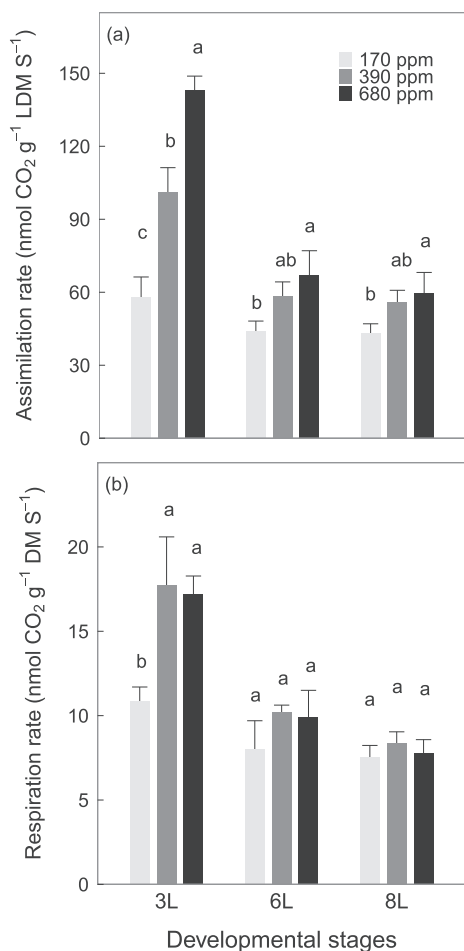


Figure 4. Leaf mass-based net assimilation rate (a) and whole-plant mass-based night-time respiration rate (b) of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (black bars); 390 ppm [CO₂] (dark grey bars); 170 ppm [CO₂] (light grey bars). Values are the means (nmol CO₂ g⁻¹ s⁻¹) of three individual chambers; error bars represent ±1 SD. Different letters indicate significant differences between [CO₂] treatments ($P < 0.05$, Tukey's HSD). Harvests were carried out after emergence of three, six and eight leaf sheaths; denoted as 3 L, 6 L and 8 L periods, respectively.

increased with increasing [CO₂], but at 6 L and 8 L there was no significant difference across [CO₂] treatments (Fig. 6h). At 6 L and 8 L, root SM concentrations were significantly lower at 170 ppm than at 390 ppm [CO₂]. By contrast, root SM concentrations were slightly higher at 390 ppm than at 680 ppm [CO₂], although this difference was statistically not significant (Fig. 6i). Note that most leaf SMs are C-based compounds whereas most root SMs contain N (Supporting Information Figs S1 & S2). Within leaves, low [CO₂] tended to decrease the concentrations of Luteolin-based, chrysoeriol-based, tricrin-based and apigenin-based SMs, rather than ferulic acid-based SMs (Supporting Information Fig. S1). In addition, the concentrations of ferulic acid-based, luteolin-based and chrysoeriol-based SMs decreased from 3 L to 8 L across all [CO₂] treatments, whereas tricrin-based and apigenin-based SMs increased at 680 ppm [CO₂] but remained relatively constant

at 390 and 170 ppm [CO₂] (Supporting Information Fig. S1). Within roots, low [CO₂] tended to decrease DIMBOA-Glc (38%) more than HDMBOA-Glc (25%) (Supporting Information Fig. S2).

The response of SMs/NSC to [CO₂] differed between tissues and developmental stages (Fig. 6j–l). At 3 L and 6 L, leaf SMs/NSC was lower at 170 ppm [CO₂] than at 390 ppm [CO₂], but at 8 L there was no difference in leaf SMs/NSC across [CO₂] treatments (Fig. 6j). By contrast, stem and root SMs/NSC was higher at 170 ppm than at 390 and 680 ppm [CO₂] (Fig. 6k,l).

Proportional C allocation to respiration, structural growth, NSC and SMs

Proportional C allocation to R (R/A) remained relatively constant across [CO₂] treatments (Fig. 8). The proportion of C allocation to biomass (SG+NSC+SMs) was lower in plants grown at 170 ppm [CO₂] than at 680 ppm [CO₂], although this was not statistically significant (Fig. 8). Proportional C allocation to NSC was significantly higher in plants grown at 680 ppm than at 390 and 170 ppm [CO₂], and proportional C allocation to SMs significantly increased with increasing [CO₂] (Fig. 8). The unaccounted C pool (A – R – SG – NSC – SMs) showed large variations and no discernible trend with [CO₂].

Within-plant partitioning of biomass, NSC and SMs

Allocation of total plant biomass and NSC to leaves, stems and roots differed with [CO₂] (Fig. 9a–f). With increasing [CO₂], allocation of biomass and NSC to leaves decreased but allocation to stems and roots increased (Fig. 9a–f). However, allocation of SMs was not affected by [CO₂]; plants in all treatments showed the highest proportions in leaves and the lowest proportions in stems (Fig. 9g–i).

DISCUSSION

Continuous measurements of gas exchange combined with the recurrent quantification of whole-plant NSC and SMs allowed us to track C fluxes to different plant organs and functional pools. Manipulations of the whole-plant C balance provided insights into the response of plant C allocation to changes in C availability. Our study revealed that C allocation to respiration is maintained as a fixed proportion of net plant C gain, even over a large gradient of [CO₂]. With increasing C availability, the fraction of assimilation allocated to biomass (SG+NSC+SMs), in particular NSC and SMs, increased. Moreover, biomass and NSC increased more in stems and roots than in leaves with increasing C availability. However, under low C availability resource constraints favoured the establishment of a functional equilibrium, that is, biomass and NSC increased in leaves, the tissue responsible for the acquisition of the most limiting resource, CO₂. Increasing C availability released this constraint and allowed plants to invest proportionally more C into NSC and SMs.

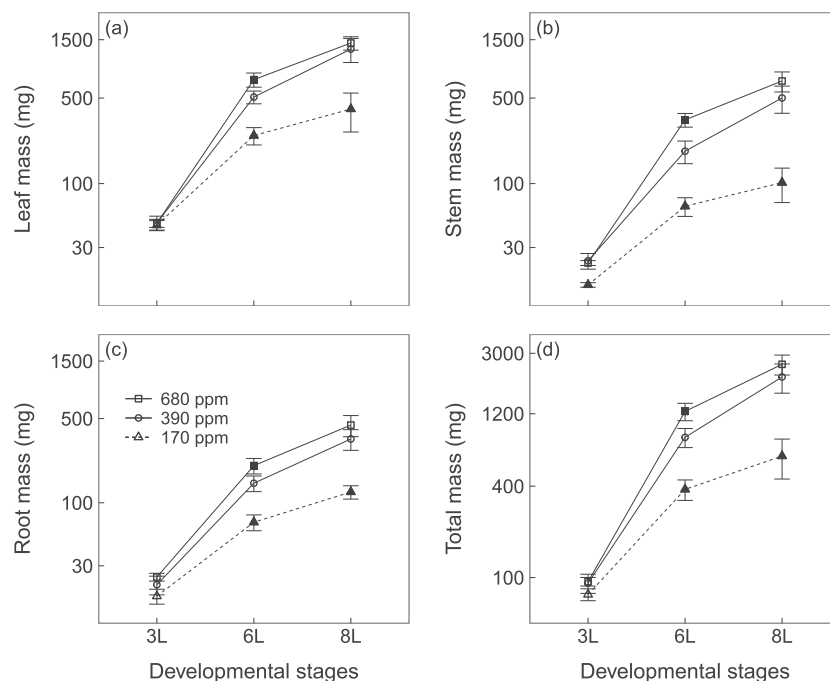


Figure 5. The dry mass of leaf (a), stem (b), root (c) and total mass (d) of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, black line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, dashed line). Values are the means (mg) of three individual chambers; error bars represent ± 1 SD. Significant differences between 680 and 170 ppm [CO₂] treatments compared to ambient [CO₂] (390 ppm) are indicated by filling of symbols ($P < 0.05$, Tukey's HSD). Note the dry mass is plotted on a log scale. Harvests were carried out after emergence of three, six and eight leaf sheaths; denoted as 3 L, 6 L and 8 L periods, respectively.

Minimizing carbon loss – with increasing C availability the proportion of C in biomass (in particular nonstructural components) increases but respiration does not

The higher relative allocation to respiration in the first weeks of the experiment (until 3L) in the two lower [CO₂] treatments may be due to relatively greater carbon supply from seed storage than from photosynthesis. Because seed storage (~46 mg seed mass on average) was depleted rapidly over time, as indicated by strong increases in plant dry biomass (ca. 230 and 1150 mg at 3L and 6L at low [CO₂]), R/A decreased dramatically at low and ambient [CO₂] while it remained quite constant at elevated [CO₂]. Whole-plant R/A remained relatively constant across [CO₂] treatments afterwards, corroborating previously observed homeostasis of leaf-level R/A in response to changing [CO₂] in *Eucalyptus saligna* (Ayub *et al.*, 2011), *G. max* (Ayub *et al.*, 2014) and wheat (Gifford, 1995). The homeostasis of whole-plant R/A was also observed across a range of temperatures in two *Plantago* species (Atkin *et al.*, 2007) and at different irradiance levels in five herbaceous species (Pons & Poorter, 2014). However, all these studies assessed gas exchange during several hours or up to few days only, and hence our study, spanning many weeks, provides further insights into long-term responses of R/A to changes in [CO₂]. Note that our estimates of the R/A ratio (ca. 10–14%) were based on nighttime respiration (R) and daytime net photosynthesis (i.e. A = gross photosynthesis – daytime respiration), not on total respiration and gross photosynthesis as in other studies.

Assuming daytime respiration rates equal those measured at night, the ratio of total respiration to gross photosynthesis would be ~30% and within the range of proportions previously reported (ca. 30–50%) (Atkin *et al.*, 2007; Gifford, 1995; Pons & Poorter, 2014).

Whole-plant assimilation and respiration are largely dependent on plant size and, therefore, do not represent photosynthetic or respiratory efficiency. By contrast, mass-based rates can indicate whether other factors than CO₂ availability, like leaf N or sugar concentrations, may constrain assimilation or respiration. Leaf mass-based net day-time assimilation declined over time and correlated negatively with leaf NSC concentrations but positively with leaf N concentrations in all [CO₂] treatments (Supporting Information Figs S3 & S4). Moreover, it decreased to a greater extent at elevated [CO₂] than at low [CO₂] as leaf NSC increased and N concentrations decreased to a greater extent at elevated [CO₂]. These findings support previously reported sugar- (reviewed in Long *et al.*, 2004) and Rubisco-mediated (Aranjuelo *et al.*, 2011; Pinto *et al.*, 2014) photosynthetic regulation under elevated [CO₂] but may also indicate proportionally greater light limitation at elevated than at low [CO₂] in the growth chambers.

The tight coupling between assimilation and respiration across [CO₂] treatments suggests that a large proportion of respiration serves to provide energy required during assimilation, for example, for processes like sucrose synthesis and phloem loading (Crous *et al.*, 2011). However, assimilation was not the only factor influencing respiration. In absence of

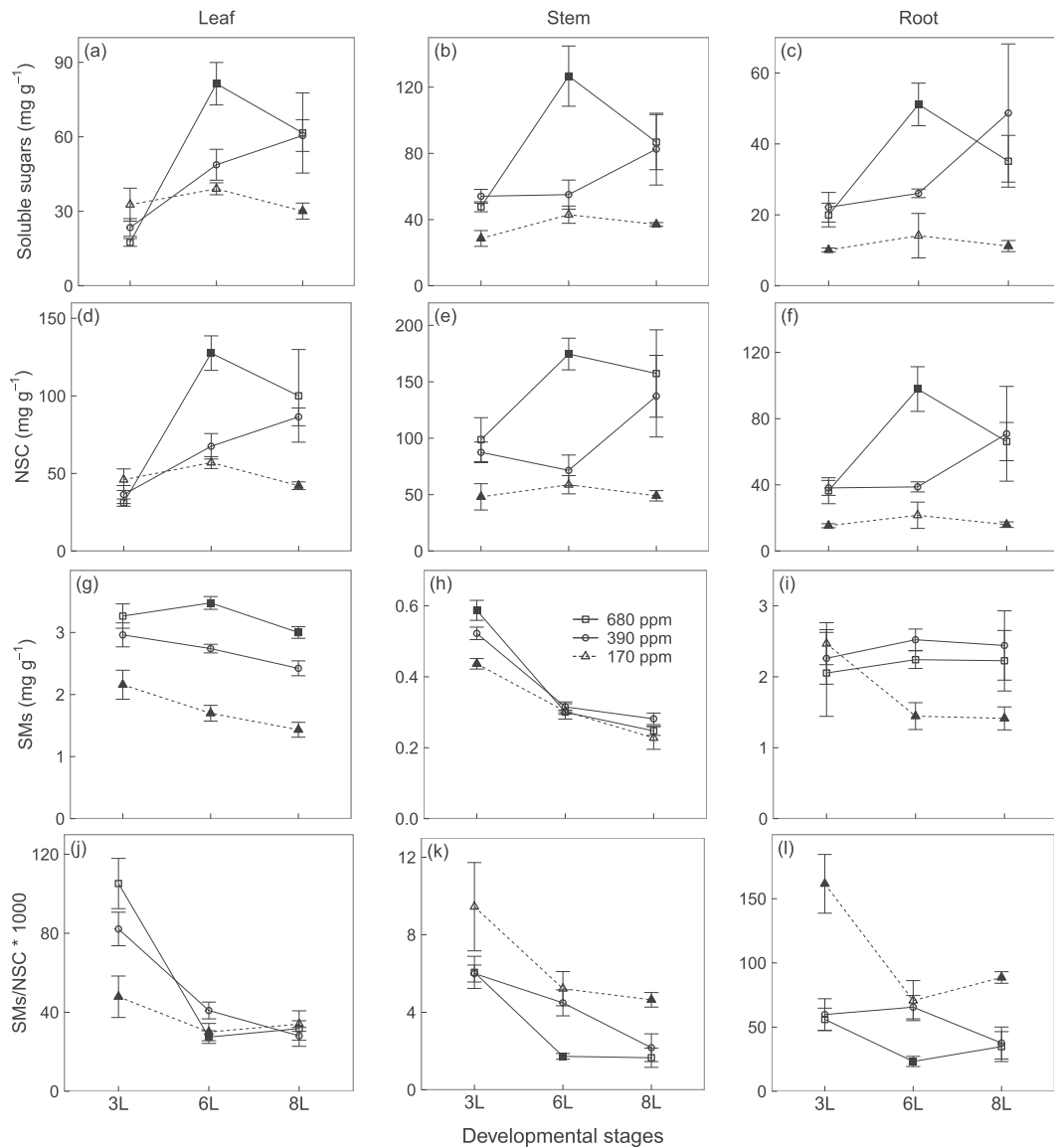


Figure 6. Concentrations of soluble sugars (mg g^{-1} dry weight, a–c), total non-structural carbohydrates (NSC, mg g^{-1} dry weight, d–e), secondary metabolites (SMs, mg g^{-1} dry weight, g–i) and ratios of SMs to NSC (j–l) in leaves, stems and roots of winter wheat (*Triticum aestivum* cv. Genius) for the three $[\text{CO}_2]$ treatments: 680 ppm $[\text{CO}_2]$ (squares, black line); 390 ppm $[\text{CO}_2]$ (circles, black line); 170 ppm $[\text{CO}_2]$ (triangles, dashed line). Total NSC concentrations were calculated as the sum of concentrations of soluble sugars, starch and fructans. Values are the means of three individual chambers; error bars represent ± 1 SD. Significant differences between 680 and 170 ppm $[\text{CO}_2]$ treatments compared to ambient $[\text{CO}_2]$ (390 ppm) are indicated by filling of symbols ($P < 0.05$, Tukey's HSD). Note the different scales on y-axes. Harvests were carried out after emergence of three, six and eight leaf sheaths; denoted as 3 L, 6 L and 8 L periods, respectively.

assimilation, that is, during nights, respiration rates were quite low, but these may have been the result of low night-time temperatures in the greenhouse. The negative correlation between mass-based night-time respiration and whole-plant NSC concentrations (Supporting Information Fig. S3) indicates that substrate supply, at least at ambient and elevated $[\text{CO}_2]$, was not a limiting factor for respiration.

Interestingly and contrary to our expectations, plants grown at high $[\text{CO}_2]$ invested relatively more C in biomass (in particular in NSC and SMs) than plants grown at low $[\text{CO}_2]$. In these plants, a down-regulation of growth via modulation of phytohormones under stress (Achard *et al.*, 2006; Park *et al.*, 2007; Zhang *et al.*, 2009) could provide an explanation for the

different allocation patterns. By contrast, at elevated $[\text{CO}_2]$, photosynthesis was apparently light-, not C-limited, and plants tended to utilize more C for biomass. This is consistent with the growth rate hypothesis (Coley *et al.*, 1985; Endara & Coley, 2011) stating that plants in resource-rich environments generally grow faster in order to be able to compete for other resources such as water, nutrients or, as in our study, for light. Greenhouse light intensities were much lower than those occurring in the field. While this may put an upper limit on absolute resource fluxes, the general agreement of allocation patterns in our study with those observed in field-grown plants downplays the impact of greenhouse light limitation on allocation patterns.

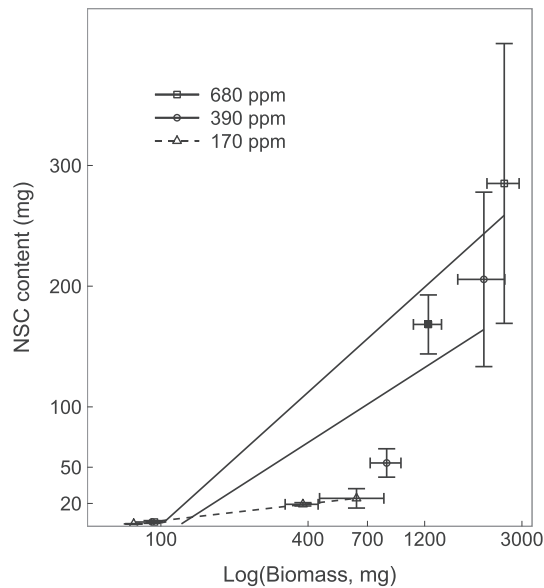


Figure 7. Correlations between whole-plant nonstructural carbohydrates (NSC) content and biomass in winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, black line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, dashed line). Values are the means (mg) of three individual chambers; error bars represent ± 1 SD. Note the dry mass is plotted on a log scale. Significant differences in NSC content and biomass between 680 and 170 ppm [CO₂] treatments compared to ambient [CO₂] (390 ppm) are indicated by filling of symbols ($P < 0.05$, Tukey's HSD).

The unaccounted C pool showed relatively large variations across treatments, which may be attributed, at least partially, to changes in C export, like root exudation or the emission of

VOCs. Plants release root exudates as a means to facilitate organic matter decomposition presumably for nutrient acquisition, and several studies found that root exudation was much lower in plants grown at ambient than at elevated [CO₂] (Johansson *et al.*, 2009; Phillips *et al.*, 2009). In our experiment, nutrients were provided and freely accessible (sand and nutrient solution) thus making nutrient uptake facilitation unnecessary. However, other studies showed that plants subjected to low [CO₂] always exhibited higher emissions of VOCs (Possell & Hewitt, 2011; Wilkinson *et al.*, 2009), likely for quenching reactive oxygen species (Harrison *et al.*, 2013; Loreto & Schnitzler, 2010) that are increasingly produced during inhibition of the photosynthetic electron transfer chain under limited CO₂ availability to the Calvin cycle (Brosché *et al.*, 2010).

Ensuring long-term survival – larger increases in NSC and SMs than structural growth with increasing C availability

How plants build up NSC remains a conceptual paradigm with potentially far-reaching implications for both understanding and predicting plant responses to environmental changes (Dietze *et al.*, 2014; Hartmann & Trumbore, 2016). There is substantial evidence suggesting that environmental stress limits growth faster than photosynthesis thus leading to an initial excess C accumulating as NSC (Palacio *et al.*, 2014). By contrast, trade-off allocation to NSC, that is, allocation of C to storage at the cost of growth, was directly assessed only in few recent studies (Gibon *et al.*, 2009; Hartmann *et al.*, 2015; Wiley *et al.*, 2013) but may represent an important insurance strategy against potential future threats like herbivory and thus promote long-term survival (Wiley & Helliker, 2012).

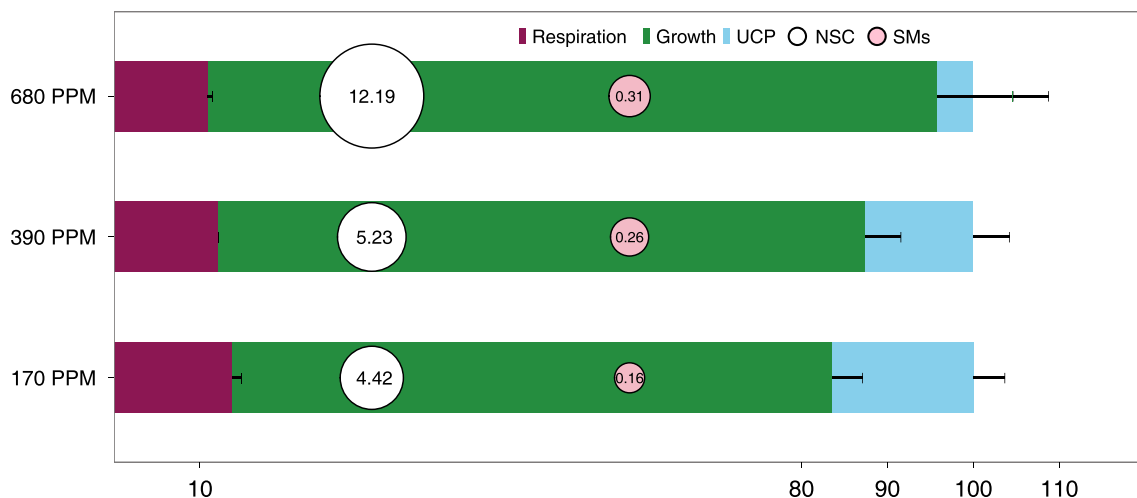


Figure 8. Percentage proportions of cumulative net carbon assimilation between 3 L and 6 L allocated to respiration (violet bars), growth (green bars), unaccounted carbon pool (UCP, blue bars), total nonstructural carbohydrates (NSC, white bubbles) and secondary metabolites (SMs, pink bubbles) of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂]; 390 ppm [CO₂]; 170 ppm [CO₂]. All concentration data were scaled up to whole-plant content by multiplying them with tissue dry mass, and then converted to carbon content before calculating proportions. The numbers in the white and pink bubbles indicate the percentage of net carbon assimilation allocated to NSC and SMs, respectively. Note, however, that the size of these bubbles has been adjusted to allow numbers to be printed and is not linearly related to the pool sizes. Values are means of three individual chambers; error bars represent ± 1 SE.

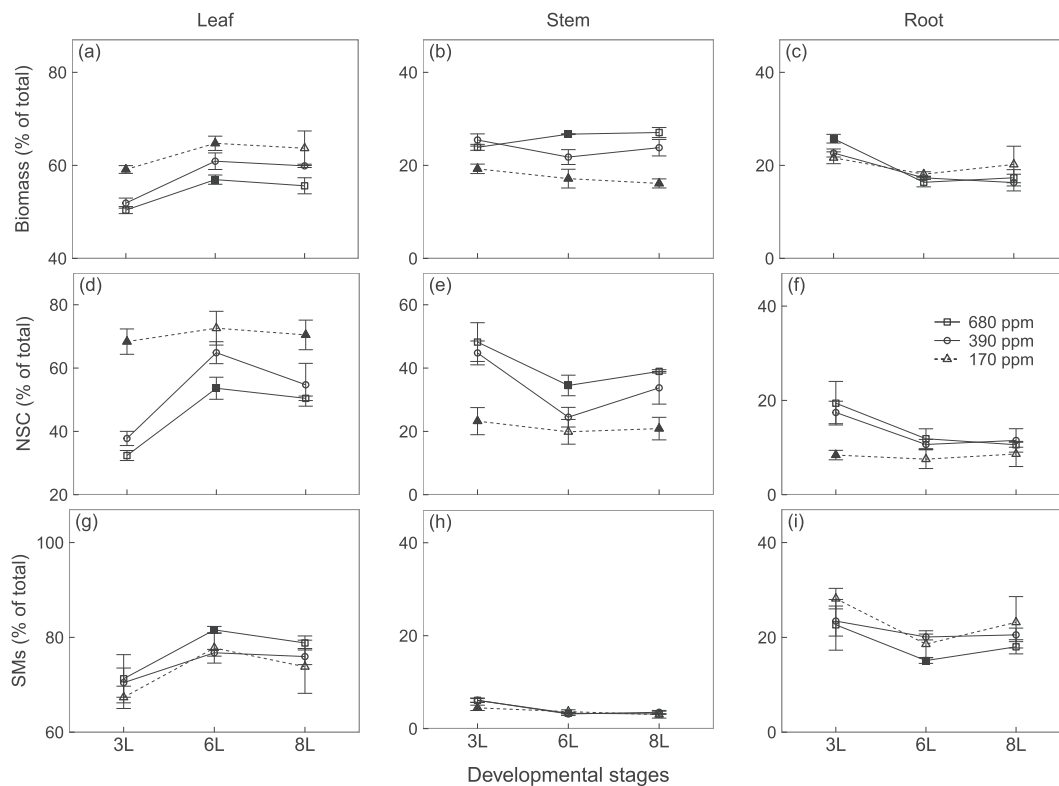


Figure 9. Percentage proportion of biomass (a–c), non-structural carbohydrates (NSC, d–e) and secondary metabolites (SMs, g–i) allocated to leaves, stems and roots of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, black line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, dashed line). Values are the means of three individual chambers; error bars represent ±1 SD. Significant differences between 680 and 170 ppm [CO₂] treatments compared to ambient [CO₂] (390 ppm) are indicated by filled symbols ($P < 0.05$, Tukey's HSD). Note the different scales on y-axes. Harvests were carried out after emergence of three, six and eight leaf sheaths; denoted as 3 L, 6 L and 8 L periods, respectively.

As a result of positive C balance between 3 L and 6 L, total NSC (soluble sugars + starch + fructans) concentrations doubled at 390 ppm [CO₂] and increased by more than three times at 680 ppm [CO₂]. Therefore, greater C availability increased total NSC concentrations and whole-plant NSC content in proportions that were much higher than increases in total plant biomass. At ambient and elevated [CO₂], SG may be limited by environmental factors such as nutrients and water (Kirschbaum, 2011; Palacio *et al.*, 2014). The much higher C/N ratio at elevated than at ambient [CO₂] also suggests that SG was likely N-, rather than C-limited (Supporting Information Table S1). At 8 L, the subsequent decline in NSC concentrations at elevated [CO₂] may be due to photosynthetic acclimation of wheat plants to elevated [CO₂] (Aranjuelo *et al.*, 2011; Aranjuelo *et al.*, 2015). In addition, under elevated [CO₂], the size of the C pool in whole-plant NSC is equivalent to twice the whole-plant daily net assimilation at 6 L (1.5 times at 8 L). Thus, NSC at elevated [CO₂] may represent a multiple-day buffer pool for supporting metabolic activities.

Many manipulative studies have investigated the effects of elevated [CO₂] or of shading (to reduce C availability) on SMs (Lindroth, 2012), but little is known about the responses of whole-plant SMs over a large gradient of C availability. Leaf SM concentrations increased with increasing C availability,

which is in accord with the GDBH and suggests that the SM pool is filled with C that is available in excess of growth demand (Hermes & Mattson, 1992). Plants grown under low C availability may gain immediate benefits by reducing C investments into SMs. However, SMs can serve multiple functions, such as detoxification, primary metabolism and chemical defence (Neilson *et al.*, 2013). Large costs needed to repair and regrow plant tissues following stress conditions such as herbivory could reduce the long-term fitness of low [CO₂] plants with low SMs, as suggested by the growth rate hypothesis (Coley *et al.*, 1985). In addition, plants at 3 L had relatively more young leaves and higher leaf and stem SMs than plants at 8 L. Greater leaf and stem SMs in young vulnerable leaves that also have greater photosynthetic capacity may represent an investment insurance strategy.

N-rich SMs play important roles in ecological interactions, but very few studies investigated how they may respond to elevated [CO₂] (Lindroth, 2012). As expected, low [CO₂] decreased N-containing SMs (benzoxazinoid derivatives) in roots. This is consistent with the GDBH and suggests that limited C resources are preferentially invested in SG rather than SMs. Furthermore, low [CO₂] tended to decrease DIMBOA-Glc more than HDMBOA-Glc (Supporting Information Fig. S2), possibly because HDMBOA-Glc is more efficient against pathogens and insect herbivory (Glaser *et al.*, 2011) and represents

the better investment under C limitation. By contrast, concentrations of these compounds were slightly lower at elevated [CO₂] than at ambient [CO₂], likely because the production of these N-rich SMs (putrescine-based and benzoxazinoids) may have been limited by N supply rather than C supply (Supporting Information Table S1). This interpretation is in agreement with results from other studies showing that elevated [CO₂] inhibited nitrate assimilation in wheat (Bloom *et al.*, 2010). Given the defensive role of benzoxazinoid derivatives (Ahmad *et al.*, 2011), the responses observed in our study support previously reported increases in egg densities of western corn rootworm (*Diabrotica virgifera virgifera*) in the soil of soybean fields under elevated [CO₂] (Schroeder *et al.*, 2006).

The trade-off between allocation to NSC versus SMs was tissue specific and depended on the developmental stage of the plants. At 3 L, leaf SMs/NSC increased with increasing C availability, indicating that leaves tend to invest more NSC into SM production. By contrast, root SMs/NSC declined with increasing C availability, suggesting that plant roots grown at ambient and elevated [CO₂] increase NSC more than SMs, likely due to a potential N limitation on root SM production, as discussed above.

Achieving a functional equilibrium – with increasing C availability wheat plants prioritize allocation of biomass and NSC to stems and roots over leaves

Allocation patterns of biomass and NSC are consistent with the 'functional equilibrium' hypothesis (Poorter *et al.*, 2012). At low [CO₂], plants were apparently constrained to invest proportionally more biomass and NSC in leaves, while with increasing C availability biomass and NSC increased in stems and roots. Under low [CO₂], this optimization strategy allows plants to counteract C limitation, while a more balanced resource partitioning was required to support taller plants under elevated [CO₂]. Moreover, export of NSC from leaves to stems and roots in plants at elevated [CO₂] may help plants to build storage and avoid high NSC-induced negative feedback on assimilation (Long *et al.*, 2004). By contrast, proportional allocation to SMs across plant organs did not vary between treatments, likely because leaf SMs and root SMs differed in C and N cost.

OUTLOOK

While our experimental design allowed us to quantify several functional sinks of the plant C balance, it did not allow direct assessments of root exudates and VOCs and thereby prevented differentiating among C exports. The high variability in the unaccounted C pool in our experiment is likely due to the accumulation of uncertainties across different measurements and devices (Vaisala, Picarro, FTIR, elemental analyser, HPLC-PAD and HPLC-UV). Thus, future studies would benefit from direct assessment of 'export' C fluxes, including root exudates and VOCs, while keeping in mind that very short-lived or rapidly decomposed export may be detected in experimental set-ups as respiration (i.e. CO₂).

Given that SMs can serve multiple functions, the role of C allocation to SMs as a strategic investment for defence can only

be assessed by simultaneously evaluating responses of herbivores and/or pathogens. Such studies have been conducted on soybean under elevated [CO₂] (reviewed in Zavala *et al.*, 2013) and can guide future studies with low C availability ([CO₂]). Because our knowledge about the mechanisms controlling plant allocation is still very limited, future studies should also focus on biochemical and molecular regulation of growth (Claeys *et al.*, 2014) and explore regulatory mechanisms of other allocation sinks like storage and SMs.

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SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

Table S1. Carbon (C) and nitrogen (N) concentrations and C/N ratios in leaves, stems and roots of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] 390 ppm [CO₂] and 170 ppm [CO₂]. Values represent the means (± 1 SD) of three individual chambers. Different letters indicate significant differences between [CO₂] treatments ($P < 0.05$ Tukey's HSD)

Table S2. Molecular formula, molecular weight and C fraction of standards used for the quantification of secondary metabolites in leaves, stems and roots of winter wheat (*Triticum aestivum* cv. Genius).

Figure S1. Concentrations of ferulic acid-based (a), luteolin-based (b), chrysoeriol-based (c) and triclin-based (d) and apigenin-based (e) secondary metabolites in leaves of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, blue line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, red line). Values are the means (mg g^{-1}) of three individual chambers; error bars represent ± 1 SD. Significant differences between 680 and 170 ppm [CO₂] treatments compared to ambient [CO₂] (390 ppm) are indicated by filled symbols ($P < 0.05$, Tukey's HSD).

Figure S2. Concentrations of Putrescine-based (a), DIMBOA-Glc-based (b) and HDMBOA-Glc-based (c) secondary metabolites in roots of winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, blue line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, red line). Values are the means (mg g^{-1}) of three individual chambers; error bars represent ± 1 SD. Significant differences between 680 and 170 ppm [CO₂] treatments compared to ambient [CO₂] (390 ppm) are indicated by filled symbols ($P < 0.05$, Tukey's HSD).

Figure S3. Correlations between leaf nonstructural carbohydrate (NSC) concentrations (mg g^{-1}) and leaf assimilation rate ($\text{nmol CO}_2 \text{ g}^{-1} \text{ s}^{-1}$) (a), and correlations between whole-plant NSC concentrations (mg g^{-1}) and respiration rate ($\text{nmol CO}_2 \text{ g}^{-1} \text{ s}^{-1}$) (b) in winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, blue line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, red line). Values are the means of three individual chambers; error bars are ± 1 SD.

Figure S4. Correlations between leaf nitrogen (N) concentrations (mg g^{-1}) and assimilation rate ($\text{nmol CO}_2 \text{ g}^{-1} \text{ s}^{-1}$) in winter wheat (*Triticum aestivum* cv. Genius) for the three [CO₂] treatments: 680 ppm [CO₂] (squares, blue line); 390 ppm [CO₂] (circles, black line); 170 ppm [CO₂] (triangles, red line). Values are the means of three individual chambers; error bars are ± 1 SD.