Electronic Supplementary Information

Determining Substrate Specificities of β1,4-Endo-Galactanases Using Plant Arabinogalactan Oligosaccharides Synthesized by Automated Glycan Assembly

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Building Block Synthesis

Synthesis of BB1 (in analogy to a previously reported procedure)¹

Reaction Scheme:



NMR Spectra:

¹H NMR (400 MHz, CDCl₃) of **11**







¹H NMR (400 MHz, CDCl₃) of **12**



 $^{\rm 13}{\rm C}$ NMR (101 MHz, CDCl₃) of ${\rm 12}$



¹HNMR (400 MHz, CDCl₃) of **13**



$^{\rm 13}{\rm C}$ NMR (101 MHz, CDCl_3) of ${\bf 13}$



^1H NMR(400 MHz, CDCl₃) of $\beta\text{-anomer}$ of **BB1**



 ^{13}C NMR (101 MHz, CDCl₃) of $\beta\text{-anomer of }\textbf{BB1}$



Synthesis of BB2

Reaction Scheme:



NMR Spectra:

¹H NMR (400 MHz, CDCl₃) of **14**:



¹³C NMR (101 MHz, CDCl₃) of **14**:



¹H NMR (400 MHz, CDCl₃) of **15**:



¹³C NMR (101 MHz, CDCl₃) of **15**:



 ^1H NMR (400 MHz, CDCl3) of $\beta\text{-anomer of}$ BB2:



 ^{13}C NMR (101 MHz, CDCl3) of $\beta\text{-anomer of }\text{\textbf{BB2}}\text{:}$



Synthesis of Arabinose BB5

Reaction Scheme:



NMR Spectra:

¹H NMR (400 MHz, CDCl₃) of **16**:



¹³C NMR (101 MHz, CDCl₃) of **16**:



¹H NMR (400 MHz, CDCl₃) of **17**:



¹³C NMR (101 MHz, CDCl₃) of **17**:



¹H NMR (400 MHz, CDCl₃) of **18**:



¹³C NMR (101 MHz, CDCl₃) of **18**:



¹H NMR (400 MHz, CDCl₃) of **BB5**:



¹³C NMR (101 MHz, CDCl₃) of **BB5**:



Automated Glycan Assembly

 $\label{eq:Benzyloxycarbonylaminopentyl 2-O-benzoyl-3, 6-O-dibenzyl-\beta-D-galactopyranosyl-(1\rightarrow 4)-2-O-benzoyl-3, 6-O-dibenzyl-4-O-fluorenylcarboxymethyl-\beta-D-galactopyranoside$



HPLC was performed using a YMC Diol column and linear gradients from 90% to 40% hexane in ethyl acetate (35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranoside (1)



Crude RP-HPLC of the semi-protected disaccharide (ELSD trace):



HPLC was performed using a C5 column and a linear gradient from 80% to 0% H₂O (containing 0.1% of formic acid) in MeCN (50 min, flow rate 1.0 mL/min).

RP-HPLC of deprotected disaccharide 1 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

 ^1H NMR (600 MHz, D2O) of disaccharide 1







HSQC (D₂O) of disaccharide 1



 $\label{eq:Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranoside}$



Crude NP-HPLC (ELSD trace):



(35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranoside (2)



Crude RP-HPLC of the semi-protected tetrasaccharide (ELSD trace):



RP-HPLC of deprotected tetrasaccharide 2 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of tetrasaccharide **2**:







 $\label{eq:sphere:eq:sphe$



Crude NP-HPLC (ELSD trace):



HPLC was performed using a YMC Diol column and linear gradients from 90% to 30% hexane in ethyl acetate (35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranos





Crude RP-HPLC of thesemi-protected hexasaccharide (ELSD trace):

HPLC was performed using a C5 column and a linear gradient from 80% to 0% H₂O (containing 0.1% of formic acid) in MeCN (50 min, flow rate 1.0 mL/min).

RP-HPLC of deprotected hexasaccharide 3 (ELSD trace):



RP-HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (700 MHz, D₂O) of hexasaccharide **3**:



 ^{13}C NMR (176 MHz, D2O) of hexasaccharide **3**:



HMQC (D₂O) of hexasaccharide **3**



 $\label{eq:Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,4-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranoside}$



HPLC was performed using a YMC Diol column and linear gradients from 90% to 40% hexane in ethyl acetate (35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranosyl- $(1 \rightarrow 3)$ - β -D-galactopyranosyl- $(1 \rightarrow 4)$ - β -D-galactopyranoside (4)



Crude RP-HPLC of the semi-protected tetrasaccharide (ELSD trace):



RP-HPLC of deprotected tetrasaccharide 4 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).









HSQC (D₂O) of tetrasaccharide 4:



Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-[2,3,5-O-tribenzoyl- α -L-arabinofuranosyl]-6-O-benzyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3,6-O-dibenzyl- β -D-galactopyranoside



Crude NP-HPLC (ELSD trace):



(35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl β -D-galactopyranosyl-(1 \rightarrow 4)-3-*O*-[α -L-arabinofuranosyl]- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-galactopyranoside (5)





Crude RP-HPLC of the semi-protected tetrasaccharide (ELSD trace):

HPLC was performed using a C5 column and a linear gradient from 80% to 0% H₂O (containing 0.1% of formic acid) in MeCN (50 min, flow rate 1.0 mL/min).

RP-HPLC of the deprotected tetrasaccharide 5 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of tetrasaccharide **5**:





 $\label{eq:Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3,6-O-dibenzyl-$-D-galactopyranosyl-(1$-$-2-O-benzoyl-3-O-[2,3,5-O-tribenzoyl-$-C-benzoyl-$-2-O-benzoyl-$$





Aminopentyl β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-galactopyranosyl-(1 \rightarrow 4)-3-*O*-[α -L-arabinofuranosyl]- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-galactopyran

Crude RP-HPLC of the semi-protected hexasaccharide (ELSD trace):



acid) in MeCN (50 min, flow rate 1.0 mL/min).



RP-HPLC of deprotected hexasaccharide **6** (ELSD trace):

¹H NMR (700 MHz, D₂O) of hexaasaccharide **6**:



¹³C NMR (151 MHz, D₂O) of hexasaccharide **6**:







Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3,6-O-dibenzyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3,6-O-dibenzyl- β -D-galactopyranosyl- $(1 \rightarrow 4)$ 2-O-benzoyl-3-O-[2,3,5-O-tribenzoyl- α -L-arabinofuranosyl]-6-O-benzyl- β -D-galactopyranoside



Crude NP-HPLC (ELSD trace):



(35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-galactopyranosyl-(1 \rightarrow 4)- β -D-galactopyranosyl-(1 \rightarrow 4)-3-O-[α -L-arabinofuranosyl]- β -D-galactopyranoside (7)





Crude RP-HPLC of the semi-protected pentasaccharide (ELSD trace):

RP-HPLC of deprotected tetrasaccharide 7 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of pentasaccharide **7**:





 ^{13}C NMR (151 MHz, D2O) of pentasaccharide **7**:

Benzyloxycarbonylaminopentyl 2,3,5-*O*-tribenzoyl-a-L-arabinofuranosyl- $(1\rightarrow 3)$ -2,5-*O*-dibenzoyl- α -L-arabinofuranosyl- $(1\rightarrow 3)$ -2-*O*-benzoyl-4,6-*O*-dibenzyl- β -D-galactopyranoside



HPLC was performed using a YMC Diol column and linear gradients from 90% to 40% hexane in ethyl acetate (35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl α -L-arabinofuranosyl-(1 \rightarrow 3)- α -L-arabinofuranosyl-(1 \rightarrow 3)- β -D-galactopyranoside (8)







acid) in MeCN (50 min, flow rate 1.0 mL/min).

RP-HPLC of deprotected trisaccharide 8 (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of trisaccharide **8**:



¹³C NMR (151 MHz, D₂O) of trisaccharide 8:



HSQC (D₂O) of trisaccharide 8



 $\label{eq:Benzyloxycarbonylaminopentyl 2-O-benzoyl-3,6-O-dibenzyl-\beta-D-galactopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3-O-[2,5-O-dibenzoyl-3-O-[2,3,5-O-tribenzoyl-\alpha-L-arabinofuranosyl]-\alpha-L-arabinofuranosyl]-6-O-benzyl-\beta-D-galactopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-O-dibenzyl-\beta-D-galactopyranoside$





HPLC was performed using a YMC Diol column and linear gradients from 90% to 40% hexane in ethyl acetate (35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

$\label{eq:amplitude} \begin{array}{ll} \mbox{Aminopentyl} & \beta\mbox{-D-galactopyranosyl-(1 \rightarrow 4)-3-$O-[3-$O-[α-L-arabinofuranosyl]-α-L$







HPLC was performed using a C5 column and a linear gradient from 80% to 0% H₂O (containing 0.1% of formic acid) in MeCN (50 min, flow rate 1.0 mL/min).





HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of pentasaccharide **9**:



 ^{13}C NMR (151 MHz, D2O) of pentasaccharide **9**:



Benzyloxycarbonylaminopentyl 2,3,5-O-tribenzoyl-a-L-arabinofuranosyl- $(1 \rightarrow 5)$ -2,3-O-dibenzoyl- α -L-arabinofuranosyl- $(1 \rightarrow 5)$ -2,3-O-dibenzoyl- α -L-arabinofuranosyl-2-O-benzoyl-4,6-O-dibenzyl- β -D-galactopyranoside



(35 min, flow rate 1 mL/min) and from 40% to 0% hexane in ethyl acetate (10 min, flow rate 1 mL/min).

Aminopentyl α -L-arabinofuranosyl-(1 \rightarrow 5)- α -L-arabinofuranosyl-(1 \rightarrow 5)- α -L-arabinofuranosyl-(1 \rightarrow 3)- β -D-galactopyranoside (10)



Crude RP-HPLC of the semi-protected tetrasaccharide (ELSD trace):



HPLC was performed using a C5 column and a linear gradient from 80% to 0% H_2O (containing 0.1% of formic acid) in MeCN (50 min, flow rate 1.0 mL/min).

RP-HPLC of deprotected tetrasaccharide **10** (ELSD trace):



HPLC was performed using a Hypercarb column and a linear gradient from 97.5% to 30% H_2O (containing 0.1% of formic acid) in MeCN (45 min, flow rate 0.7 mL/min) and from 30% to 0% H_2O in MeCN (10 min, flow rate 0.7 mL/min).

¹H NMR (600 MHz, D₂O) of tetrasaccharide **10**:



¹³C NMR (600 MHz, D₂O) of tetrasaccharide **10**:



HSQC (D_2O) of tetrasaccharide **10**



Supplementary Figure



Supplementary Figure 1. Investigation of the substrate specifities of E-GALCJ, E-GALN and E-GALCT endo-galactanases using synthetic type-I arabinogalactan oligosaccharides. HPLC analyses of reactions of the galactanases with different substrates (indicated by boxes). Peaks are annotated with the corresonding AG fragments containing an aminopentenyl linker or with free reducing end (with or without red bar). Note that the α - and the β -form of the fragments with free reducing end elute separately or as double peak.

References

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