



Dyslexia risk gene relates to representation of sound in the auditory brainstem



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ABSTRACT

Dyslexia is a reading disorder with strong associations with *KIAA0319* and *DCDC2*. Both genes play a functional role in spike time precision of neurons. Strikingly, poor readers show an imprecise encoding of fast transients of speech in the auditory brainstem. Whether dyslexia risk genes are related to the quality of sound encoding in the auditory brainstem remains to be investigated. Here, we quantified the response consistency of speech-evoked brainstem responses to the acoustically presented syllable [da] in 159 genotyped, literate and preliterate children. When controlling for age, sex, familial risk and intelligence, partial correlation analyses associated a higher dyslexia risk loading with *KIAA0319* with noisier responses. In contrast, a higher risk loading with *DCDC2* was associated with a trend towards more stable responses. These results suggest that unstable representation of sound, and thus, reduced neural discrimination ability of stop consonants, occurred in genotypes carrying a higher amount of *KIAA0319* risk alleles. Current data provide the first evidence that the dyslexia-associated gene *KIAA0319* can alter brainstem responses and impair phoneme processing in the auditory brainstem. This brain-gene relationship provides insight into the complex relationships between phenotype and genotype thereby improving the understanding of the dyslexia-inherent complex multifactorial condition.

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1. Introduction

Dyslexia is characterized by poor reading, writing, and spelling skills despite typical intelligence, no visual acuity problems, and appropriate education (ICD-10-CM, <http://www.icd10data.com/ICD10CM/Codes/F01-F99/F80-F89/F81-/F81.0>). Boys are 2–3 times more likely to be affected than girls, and cumulative incidence rates vary from 5–12% (Shaywitz et al., 1990). Dyslexia persists in 4–6% of adults (Schulte-Körne and Remschmidt, 2003) dis-advantaging employment, and compromising participation in public

life. Prevention requires early sensitive screenings and successful remediation, which are both still desirable.

Various cognitive domains support literacy acquisition. Thus, heterogeneous cognitive fingerprints of dyslexia phenotypes exist (Heim and Grande, 2012; Ramus and Ahissar, 2012) and multiple subtypes of dyslexia have been suggested (Bosse et al., 2007), but a *bona fide* theory of the underlying mechanisms has not been established yet. A widely accepted rationale bases dyslexia on an impairment of phonological representations (Snowling, 2001). Others advocate auditory processing deficits such as an impaired oscillatory phase locking for low frequency temporal coding in auditory cortex (Goswami, 2011), or a decreased sensitivity to rapidly changing phonological features (Benasich et al., 2002; Tallal, 1980). Auditory processing deficits might cause an impoverished

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distinction between speech sounds (Vandermosten et al., 2010), a deficient access to otherwise intact phonetic representations (Boets et al., 2013), or a deficient match between memory representations and auditory sensations (Díaz et al., 2012; Jaffe-Dax et al., 2015). Alternatively or additionally, visual attention, visual-magnocellular processing, or visual-auditory integration compose further cognitive problems (Heim et al., 2010; Stein and Walsh, 1997; Valdois et al., 2014; Widmann et al., 2012).

Dyslexia is moderately to highly heritable (Schumacher et al., 2007) with a multifactorial etiology (Fisher and DeFries, 2002) and a complex underlying genetic architecture. Evidence exists for multiple genes to contribute to the phenotype, with considerable genetic heterogeneity across individuals (Carrion-Castillo et al., 2013). Dyslexia is linked to several risk loci including nine so-called *DYX*-regions (*DYX1-DYX9*) (Carrion-Castillo et al., 2013; Giraud and Ramus, 2013; Peterson and Pennington, 2012; Poelmans et al., 2011), but a consistent genome-wide association is still missing. However, *DYX2* on chromosome 6 is the best replicated susceptibility locus (Gabel et al., 2010), with *DCDC2* (Lind et al., 2010; Ludwig et al., 2008; Meng et al., 2005; Newbury et al., 2011; Scerri et al., 2011; Schumacher et al., 2006; Wilcke et al., 2009) as well as *KIAA0319* (Cope et al., 2005; Francks et al., 2004; Harold et al., 2006; Kaplan et al., 2002; Luciano et al., 2007; Meng et al., 2005; Paracchini et al., 2008; Scerri et al., 2011) as strongest candidate genes of this locus. Numerous studies evaluate the genetic origin of dyslexia, excellently compiled in recent reviews (Carrion-Castillo et al., 2013; Giraud and Ramus, 2013).

Despite considerable progress, complex gene-brain relations of *KIAA0319* and *DCDC2* are far from comprehensive, because studies elucidating the genes' impact on cell anatomy and systems physiology are scarce. Animal experiments associate the functional role of both genes with neuronal migration (Burbridge et al., 2008; Meng et al., 2005; Paracchini et al., 2006; Peschansky et al., 2010) and, thus, a role in the formation of the neuronal cell assemblies during brain development. Furthermore, both genes are expressed in mature neurons after migration and contribute to protein binding.

More specifically, *KIAA0319* encodes an integral transmembrane protein (Velayos-Baeza et al., 2010), and is a component in the early endosome, its membrane and the plasma membrane, possibly supporting a broader spectrum of signaling functions. In addition to neuronal migration, *KIAA0319* is associated with a negative regulation of dendrite development. It regulates processes that stop, prevent or reduce the frequency, rate or extent of dendrite development (<http://www.ncbi.nlm.nih.gov/gene/9856>; Gene ID: 9856, updated on 6-Mar-2016). Animal studies indicated that in utero RNA interference (RNAi) targeting *Kiaa0319* in male Wistar rats affected acoustic discrimination abilities of complex stimuli, which was associated with formation of heterotopias in white matter (Szalkowski et al., 2013). Electrophysiologically, a downregulation of *Kiaa0319* expression was followed by a decreased response consistency to sound stimuli as measured from neurons in the primary auditory cortex, resulting in a reduced neuronal discrimination ability (Centanni et al., 2014a,b).

At the cellular level *DCDC2* is involved in processes such as cellular defense response, dendrite morphogenesis, intracellular signal transduction, regulation of smoothed signaling pathway, regulation of Wnt signaling pathway, and regulation of cilium assembly. At the systems level, *DCDC2* is correlated with visual learning and sensory perception of sound. *DCDC2* is a component of axoneme, cytoplasm, cytoskeleton, kinocilium, nucleus and primary cilium. The doublecortin domain, to which *DCDC2* belongs, has been shown to bind tubulin and enhance microtubule polymerization. Its function may affect the signaling of primary cilia (<http://www.ncbi.nlm.nih.gov/gene/51473>; Gene ID: 51473, updated on 6-Mar-2016).

DCDC2 has been reported to be a deafness gene in a Tunisian family motivated by the considerations that hair cell kinocilia and cell primary cilia length regulation is likely influenced by *DCDC2*'s role in microtubule formation and stabilization (Szalkowski et al., 2012). *Dcdc2* knockout mice showed a deficit in rapid auditory processing (Truong et al., 2014), which is consistent with the observation of degraded neural spike timing and, thus, difficulties in the encoding of rapid sequential sensory input as measured in the somatosensory cortex in the same mutants (Che et al., 2014).

Taken together, auditory processing deficits have been linked to gene homologues for both genes (Centanni et al., 2014a,b; Szalkowski et al., 2012; Truong et al., 2014). The physiological consequence of altered functions of *KIAA0319* and *DCDC2* is linked to imprecise neuronal temporal coding. It is plausible to assume that an imprecise encoding of acoustic input leads to processing deficits of ascending speech signals challenging the formation of robust phoneme representations in long-term memory. Thus, a temporal processing deficit might prevent the uncomplicated acquisition and consolidation of literacy skills as suggested by dominating theories (Goswami, 2011; Tallal, 2012).

A huge body of brain-behavior association studies report altered structural and functional correlates pointing to irregular auditory and phonological processes in dyslexia (Banai et al., 2005; Díaz et al., 2012; Hämäläinen et al., 2013; Hornickel and Kraus, 2013; Kujala et al., 2006; Paulesu et al., 2014; Schulte-Körne and Bruder, 2010). Several brain-gene studies considered *KIAA0319* and *DCDC2* in the context of literacy. Late electrophysiological responses to speech sounds (300–700 ms) are affected in rare variants in a region between *KIAA0319* and *DCDC2* (Czamara et al., 2010). A *KIAA0319/TTRAP/THEM2* locus was associated with a reduced left-hemispheric functional asymmetry of posterior superior temporal sulcus during reading (Pinel et al., 2012). The *KIAA0319* single nucleotide polymorphism rs2143340 was related to activation in the bilateral supramarginal gyri during a word rhyming task (Cope et al., 2012). In the same study, alleles of a *DCDC2* complex tandem repeat were related to activation in the right lateral occipital temporal gyrus and the left supramarginal gyrus. These gene-related abnormal functional activations in the parietal lobes are consistent with the *DCDC2*-related reduced white matter volume, and a degraded cortical thickness in the same region (Darki et al., 2014).

The impact of dyslexia risk genes on early auditory processing is currently unknown. Interestingly, the sensation related processing of speech sounds has been found to be noisy at a very early stage in the auditory pathway of poor readers. Speech evoked brainstem responses (cABRs) were unstable and indistinctive in poor readers and in children with poor phonological skills (Banai et al., 2005; Chandrasekaran et al., 2009; Hornickel et al., 2009; Hornickel and Kraus, 2013; Strait et al., 2011; White-Schwoch and Kraus, 2013). Particularly striking is the sensitivity of cABRs in the phase of the formant transition of a given stimulus (Hornickel and Kraus, 2013). Formant transitions are fast changes of frequency bands that constitute important phonetic features because a correct encoding of formant transitions enables us to distinguish between stop consonants. Ultimately, we investigated how the two prominent dyslexia susceptible genes *DCDC2* and *KIAA0319* relate to the stability of speech-evoked brainstem responses in the phase of the formant transition of the syllable [da], which has been reported to be an electrophysiological marker of dyslexia in the early auditory pathway (Hornickel and Kraus, 2013). Here, we provide the first evidence that the dyslexia-associated gene *KIAA0319* affects response consistency in the auditory brainstem and, thus, impairs phoneme encoding at a very early stage in the auditory pathway.

2. Materials and methods

2.1. Participants

One hundred fifty nine native German-speaking children were enrolled, consisting of 95 preliterate children, aged 4–7, participating in the Legascreen project (www.legascreen.de), along with 64 literate children, aged 11–13, who were former participants of the German Language Developmental Study (GLAD-Study) e.g. (Friedrich and Friederici, 2004). A summary of demographic, psychometric and electrophysiological data is given in Table 1. All participants had normal hearing, passing a hearing screening at a 25 dB hearing level (air conduction) for octaves from 250 to 4000 Hz. Click-evoked brainstem responses were normal. No neurological diseases were known or evident. All parents gave written informed consent, while children gave additional documented verbal assent to participate in the study. Experimental procedures were approved by the University of Leipzig Ethical Review Board.

2.2. Psychometrics

Literate children were tested for reading comprehension and reading speed (*Lesegeschwindigkeits- und -verständnistest für die Klassen 6–12*, LGVT, Schneider et al., 2007) as well as for performance in spelling and writing (*Deutscher Rechtschreibtest*, DERET, Stock and Schneider, 2008). To the best of our knowledge, none of the participants were formally diagnosed with a reading disorder. Non-verbal intelligence of literate children was determined by the Kaufman Assessment Battery for Children (K-ABC, Kaufman et al., 2009) and was missing in one participant. Preliterate children were tested with the Wechsler Preschool and Primary Scale of Intelligence (WPPSI-III, Wechsler et al., 2009). In the group of preliterate children 12 individuals yielded a non-verbal IQ <85. Because the brainstem measure varies with intelligence as reported in previous studies (Hornickel and Kraus, 2013; White-Schwoch and Kraus, 2013), we controlled for intelligence in all statistical tests across all participants.

2.3. Acoustic stimulus

The Klatt-synthesized syllable [da] was provided by Nina Kraus (Hornickel et al., 2009; Hornickel and Kraus, 2013). The syllable was 170 ms long with a pitch onset (100 Hz) at 10 ms. The formant transition duration was 50 ms composed of a linear rising F1 (400–720 Hz), a linear falling F2 (1700–1240 Hz), F3 (2580–2500 Hz), and flat F4 (3300 Hz), F5 (3750 Hz), and F6 (4900 Hz). The steady-state vowel lasted 110 ms (Fig. 1a).

2.4. Neurophysiological data recording and reduction

Children were seated comfortably in a reclining chair in an electrically shielded, soundproof booth and were allowed to watch a movie of their choice (SPL <45 dB). Throughout the recording session, the children were listening to the movie in the left ear and to the speech stimuli in the right ear, which is common practice (e.g. Hornickel and Kraus, 2013). Before and after stimulation with the target syllable a train of 2000 clicks was presented to test the integrity of the auditory pathway and to ensure stable recording conditions throughout the experiment. The syllable was presented to the right ear through Etymotic ER-3 insert earphones (Etymotic Research, Elk Grove Village, IL) at an intensity level of 80 dB SPL, at a rate of 4.35 Hz, and with both polarities (condensation and rarefaction). Brainstem responses were collected using BrainVision V-Amp in combination with an EP-PreAmp, an extremely low-level noise bipolar amplifier (BrainVision) at 20 kHz sampling rate. Three single multitrode Ag/AgCl electrodes were attached to the scalp from Cz to

the ipsilateral earlobe, with the forehead as the ground. Impedances were down-regulated (<5 k Ω) and the inter-electrode impedance difference was not higher than 1.5 k Ω . The continuous signal was filtered off-line with the firfilt EEGLAB plugin (Windowed Sinc FIR-filter, bandpass 70–2000 Hz, Kaiser window, beta = 7.8572, filter order = 100300, fs = 20 kHz), epoched from –40 to 190 ms, and baseline corrected to a 40 ms interval preceding sound onset. Epochs with any activity exceeding the range of 35 μ V were rejected and a total of 6000 epochs were considered for further analyses. A comparison of click-evoked wave V latencies across session and participants showed that recording parameters were comparable throughout the session and across participants ($p > 0.4$).

2.5. Neurophysiological data analysis

Response consistency was calculated for the portion of the response reflecting the formant transition, 7–60 ms post stimulus onset. Response consistency was the result of a correlation of the subaverages of an equal number of odd and even responses (Fig. 1c). Odd and even responses were determined after the summation of a single response to the original stimulus and a single response to the inverted stimulus resulting in the operationalization of the envelope FFR (Aiken and Picton, 2008; Hornickel and Kraus, 2013). Metrics were quantified with custom-written MATLAB scripts. For statistical analyses, a Fisher transformation was applied to the correlation coefficients by calculating the inverse of the hyperbolic tangent function to linearize the distribution of these coefficients (Fisher, 1921).

2.6. DNA extraction and genotyping

We selected single nucleotide polymorphisms (SNPs) of *KIAA0319* (rs761100, rs2179515, rs2143340, rs3212236, rs9461045, and rs6935076) and *DCDC2* (rs807724, rs1087266, rs807701, rs793842, rs1091047, rs6922023) because they were repeatedly associated with dyslexia in previous studies (Cope et al., 2005; Harold et al., 2006; Meng et al., 2005; Schumacher et al., 2006; Wilcke et al., 2009).

DNA was extracted from saliva samples using standard procedures (Quinque et al., 2006) or using Oragene DNA Genotek Kits (Kanata, Ontario, Canada). Genotyping was performed first with the matrix-assisted laser desorption/ionization time-of-flight mass spectrometry system iPLEX (Agena, Hamburg, Germany). Genotyping data had to fulfill the following quality measures: SNP-wise Hardy-Weinberg-Equilibrium (HWE; $p > 0.05$ Bonferroni corrected), SNP-wise call rate >97%, individual-wise: call-rate >90% and minor allele frequency (MAF) >0.05. We excluded SNPs if any genotype comprised less than 5% of the cohort (see Table 2). Therefore, rs2143340, rs3212236, and rs9461045 were excluded from analyses.

2.7. Statistical analyses

To test the reliability of the recordings throughout the session a repeated-measures mixed model ANCOVA was employed to the peak-V-latencies to clicks collected at the start and the end of the recording session. Across all participants, a principal component analysis (PCA) with a varimax rotation was employed considering the 9 selected *DCDC2/KIAA0319* SNPs to account for the linkage disequilibrium between neighboring SNPs (Meng et al., 2005; Zondervan and Cardon, 2007). Any component with an eigenvalue greater than 1 was retained for further analyses (Kaiser, 1960). The extraction of the factor loadings of the main principal components allocated a weighting factor to every participant. This weighting factor compounds the burden of the genetic risk for each participant. This method has been adopted from Ge et al. (2012).

Table 1
Participant characteristics.

	Preliterater		Literate	
<i>N</i>	95	M	64	M
Demographics				
Age (years)	5.8 (0.9, 4.3–7.5)	–	12.7 (0.6, 11.4–13.8)	–
Sex (male/female)	48/47	–	39/25	–
Familial risk (no risk/risk)	49/44	2	46/18	–
Profession mother (median)	3	8	2	–
Profession father (median)	3	8	2	–
Brainstem measures				
<i>Peak-V-latency</i>				
Pre (ms)	5.40 (0.23, 4.65–6.05)	3	5.48 (0.20, 5.15–6.05)	1
Post (ms)	5.42 (0.24, 4.55–6.05)	3	5.51 (0.19, 5.15–5.95)	1
<i>Response consistency (r)</i>	0.72 (0.13, 0.31–0.91)	–	0.67 (0.22, –0.04–0.97)	–
Psychometrics				
<i>Literacy</i>				
DERET (mean PR)	–	–	43.6 (29.5, 0–99)	1
LGVT speed (mean PR)	–	–	37.1 (22.9, 5–98)	–
LGVT comp (mean PR)	–	–	40.8 (24.7, 1–94)	–
WR (mean%correct)	–	–	94.4 (11.5, 30–100)	–
WR speed (s)	–	–	36.4 (27.3, 15–210)	–
NWR (mean%correct)	–	–	79.2 (15.9, 23–100)	–
NWR reading speed (s)	–	–	59.1 (36.9, 25–292)	–
<i>Intelligence</i>				
Intelligence (mean IQ)	99 (12, 73–127)	–	111 (10, 86–126)	1
<i>Handedness</i>				
(right/left/ambidexterity)	83/9/-	3	59/5/-	–

M, missing data; DERET, standardized test writing skills; LGVT speed, standardized test reading speed; LGVT comp, standardized test reading comprehension; WR, word reading; NWR, nonword reading; professional education was operationalized with an ordinal scale with 1, without professional education; 2, Professional School, Vocational School; 3, Master Craftsman, Technical College, Bachelor, University of Cooperative Education; 4, Higher classes of civil service; 5, University of Applied Sciences; 6, University Degree; State Examination; values represent group averages (\pm SD, range) unless otherwise indicated.

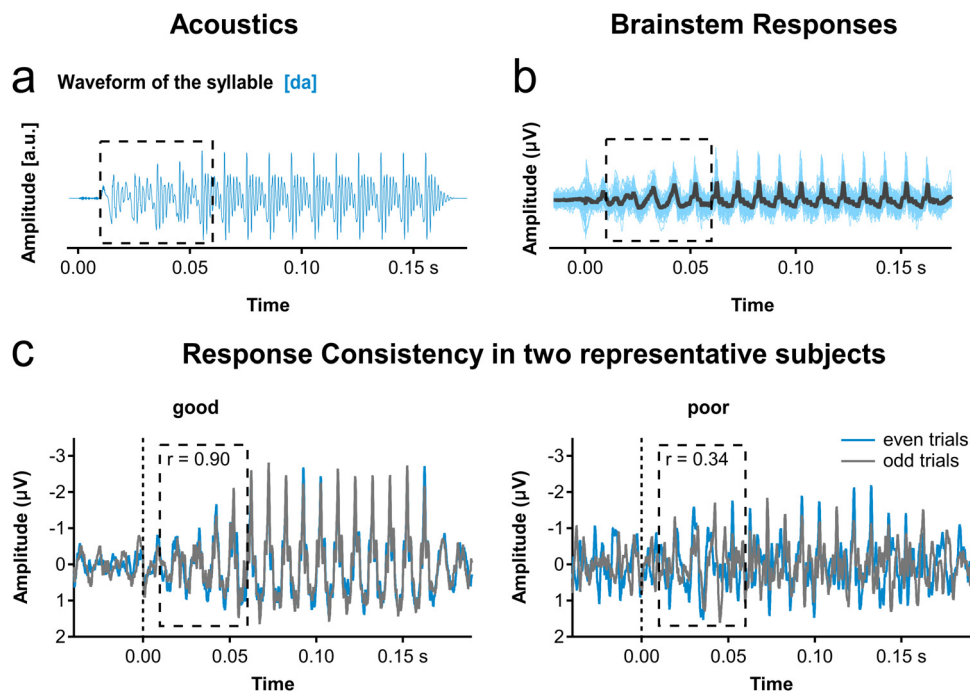


Fig. 1. Acoustic stimulus and speech evoked brainstem responses. (a) Stimulus was a Klatt-synthesized [da] syllable of 170 ms duration with a 50 ms formant transition (dotted square) and a steady-state vowel of 110 ms duration. (b) The grand average of complex auditory brainstem responses (cABR, black lines) was calculated from 6000 stimulations per syllable and subject. Individual average waveforms are depicted in blue. (c) cABRs are shown for two representative subjects. High correlation coefficients between even and odd trials in the period of the formant transition (dotted squares) illustrate good response consistency in the subject depicted on the left. Small correlation coefficients illustrate a poor response consistency in the subject on the right.

To determine the association between the genetic risk and the electrophysiological measure a multiple regression was calculated with a repeated-measures ANCOVA considering PCA-components

as within-subjects factor (*DCDC2* and *KIAA0319*), and response consistency of the brainstem as well as age, gender, familial risk and intelligence as covariates. Additional partial correlations were cal-

Table 2
Characteristics of the investigated set of single nucleotide polymorphisms.

Nearest Gene	SNP	MAF	N participants in Genotypic Groups			Genotypic Groups			Risk Allele
			Major	Hetero	Minor	Major	Hetero	Minor	
DCDC2	rs807724	0.248	89	61	9	AA	GA	GG	minor
DCDC2	rs1087266	0.409	55	78	26	CC	CT	TT	minor
DCDC2	rs807701	0.399	59	73	27	TT	TC	CC	minor
DCDC2	rs793842	0.428	54	74	31	GG	GA	AA	minor
DCDC2	rs1091047	0.192	107	43	9	GG	CG	CC	minor
DCDC2	rs6922023	0.189	108	42	9	AA	GA	GG	major
KIAA0319	rs761100	0.428	50	82	27	GG	GT	TT	minor
KIAA0319	rs2179515	0.349	66	75	18	AA	GA	GG	major
KIAA0319	rs6935076	0.371	65	70	24	CC	CT	TT	minor
KIAA0319	rs2143340	0.151	117	36	6	TT	TC	CC	minor
KIAA0319	rs3212236	0.157	115	38	6	AA	GA	GG	minor
KIAA0319	rs9461045	0.157	115	38	6	CC	TC	TT	minor

SNP, Single Nucleotide Polymorphism; MAF, Minor Allele Frequency; Major, Homozygous Major Allele; Hetero, Heterozygous; Minor, Homozygous Minor Allele.

culated to determine the univariate amount of variance of response consistency explained by the risk genes; age, sex, familial risk, and intelligence were variables of no interest. We hypothesized that a higher genetic risk loading (meaning a larger sum of risk alleles across considered SNPs within an individual participant) would be associated with a worsened electrophysiological signature. Two-tailed *p*-values were reported. Statistical analyses were performed in SPSS (IBM).

Joint effects on literacy skills and dyslexia status for the SNPs of *KIAA0319* and *DCDC2* were calculated by comparing one model including the intercept and the co-variables (age, sex, familial risk and intelligence) against a second model including the intercept, the same co-variables and the SNPs of the gene. The comparison was carried out by a likelihood ratio test (R package 'lm test', Zeileis and Hothorn, 2002) to investigate the additional information provided by the SNPs.

3. Results

We measured and analyzed cABRs to the [da] syllable in 64 literate children and in 95 preliterate children. Fig. 1b shows the averaged cABRs across participants for the [da] syllable. Fig. 1c exemplifies individual brainstem responses of two single participants. One participant showed a high response consistency with a Pearson correlation coefficient of $r=0.900$ while the other participant showed a reduced response consistency with a Pearson correlation coefficient of $r=0.335$. The supplementary material provides an alternative account of quantifying the physiological representation of sound.

For literate children we calculated the joint associations between the two genes *KIAA0319* and *DCDC2* and literacy skills. For *KIAA0319*, we observed significant association with writing and spelling performance ($\chi(3)=13.37$, $p=0.004$) and no association with reading speed test ($\chi(3)=6.88$, $p=0.076$) or reading comprehension test ($\chi(3)=2.14$, $p=0.544$). For *DCDC2*, associations were observed for reading speed ($\chi(6)=19.37$, $p=0.004$) as well as reading comprehension ($\chi(6)=17.29$, $p=0.008$). No significant relations occurred between *DCDC2* and performance in spelling and writing ($\chi(6)=3.69$, $p=0.718$).

Fig. 2a shows how much of the variance of the three measures of literacy are explained by age, sex, familial risk, and intelligence alone or when additionally including genetic variance.

We performed a PCA to account for the linkage disequilibrium of SNPs located near each other (Fig. 2b). Three components displayed an eigenvalue greater than one. The first component had an eigenvalue of 3.2 and explained 26.8% of the allelic variance in the population with a factor loading allocating four *DCDC2* SNPs (rs807724=0.925, rs1087266=0.747,

rs80771=0.716, rs793842=0.678). The second component had an eigenvalue of 2.3 and explained 26.4 % of variance loading on all *KIAA0319* SNPs (rs761100=−0.943, rs2179515=0.898, rs6935076=0.793). The third component had an eigenvalue of 1.6 and explained 26.4 % of variance loading on the remaining two *DCDC2* SNPs (rs1091047=0.923, rs6922023=−0.807). The *DCDC2* SNPs rs793842 and rs807701 did also load on component three (0.660, 0.631). Total variance explained by these three principal components was 79.6 %.

The repeated-measures ANCOVA yielded a Gene × Brainstem interaction ($F(2153)=4.35$, $p=0.014$). Tests of within-subjects contrasts indicated a quadratic relationship between genotypes and response consistency of the auditory brainstem ($F(1153)=6.7$, $p=0.011$). To further disentangle this gene-brain association we calculated post-hoc partial correlations between the response consistency at the level of the auditory brainstem and the factor loadings of the principal components extracted. Age, gender, familial risk, and intelligence were variables of control. Response consistency showed a trending positive correlation with the first principal component ($r=0.144$, $p=0.075$) allocating genetic risk of *DCDC2*, a significant negative correlation with the second principal component ($r=−0.190$, $p=0.018$) allocating genetic risk of *KIAA0319* (Fig. 2c), and no correlation with the third principal component ($r=−0.02$, $p=0.801$).

4. Discussion

A PCA separated the genetic risk of dyslexia in our sample into three principal components. Two components explained the variance of risk loading associated with *DCDC2* and one component explained the variance of risk loading of *KIAA0319*. Most interesting was the partial correlation showing that children with a higher amount of risk alleles of *KIAA0319* have less stable speech evoked brainstem responses whereas children with lower *KIAA0319* risk loading have more stable responses. Surprisingly, an opposite trend emerged for the partial correlation between *DCDC2* and electrophysiology. A higher number of *DCDC2* risk alleles was associated with more stable cABRs. This electrophysiology by gene interaction suggests that especially the *KIAA0319* risk gene carriers are prone to noisy processing of speech stimuli at a very early stage of the auditory pathway. Conversely, the data suggest that *DCDC2* risk variants might have a protective function for early auditory sensations. The different contribution of the risk factors to auditory sensations is unexpected, but most interesting. It supports reports of disparate cognitive profiles of dyslexia, when distinctively clustering the cognitive measures phonological, auditory, visual attention, and automatic skills, as first described by Heim and Grande (2012). It is highly plausible that this heterogeneity of

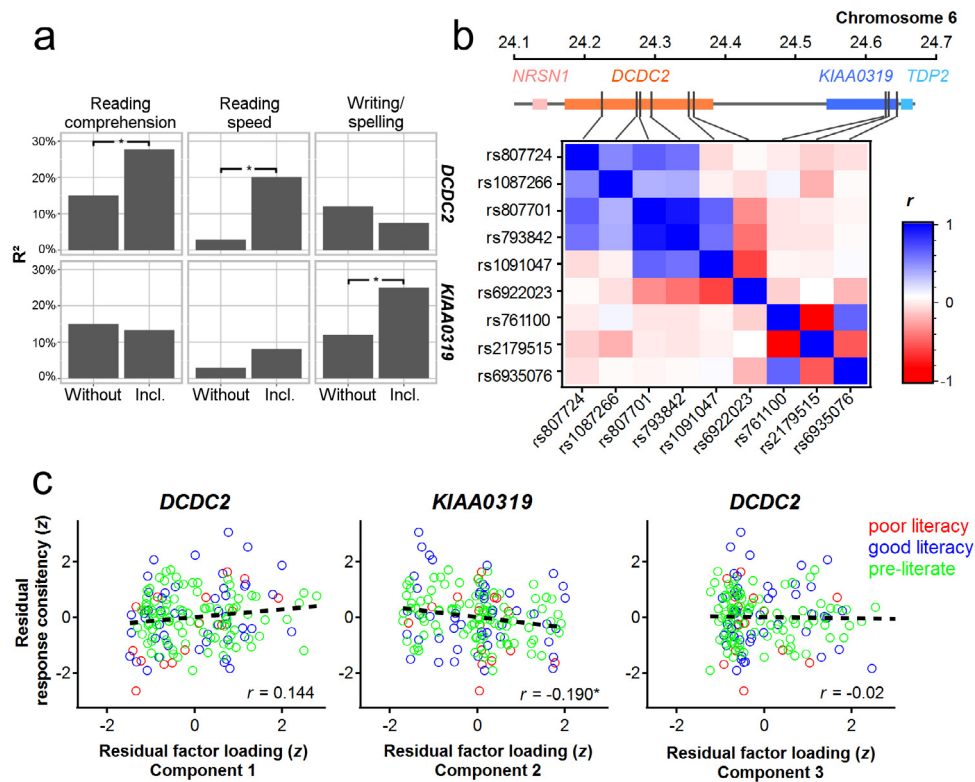


Fig. 2. Genetic risk of dyslexia relates to speech evoked brainstem responses. (a) Shown is how much of the variance (adjusted R^2) of three measures of literacy skills (reading comprehension, reading speed and writing/spelling) are explained by age, sex, familial risk and intelligence alone (left bars 'without genetics') or when additionally including variants of *DCDC2* (right bars 'incl. genetics', upper panel) or variants of *KIAA0319* (right bars 'incl. genetics', lower panel). Statistical significance of differences between the models without and including genetics were assessed using the likelihood ratio test. Significant differences between both models at the level of $p \leq 0.05$ are marked with an asterisk. (b) Gene and marker locations on the top of the correlation matrix are proportional to physical distances on chromosome 6. The correlation matrix demonstrates the linkage disequilibrium of neighboring SNPs. (c) Response consistency showed a trend for a positive correlation with the individual factor load of the first principal component allocating the genetic risk loading of *DCDC2*. Response consistency was negatively correlated with the individual factor load of the second principal component allocating the genetic risk loading of *KIAA0319*. No correlation was evident between response consistency and the factor loadings of the third principal component. To illustrate the distribution of children with poor literacy skills compared to children with good literacy skills poor performers are depicted in red in case mean literacy across reading speed, reading comprehension and spelling performance was under the 25th percentile. Blue indicates good performers (≥ 25 th percentile) and green indicates preliterary children. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

underlying cognitive profiles could be associated with heterogeneous neurobiological mechanisms caused by specific genotypes (Cicchini et al., 2015).

Risk variants in *KIAA0319* and *DCDC2* have recently been shown to interact with each other and to influence dyslexia phenotypes in a non-additive manner (Powers et al., 2013). This trans-genetic interaction between *DCDC2* risk haplotypes and *KIAA0319* risk haplotypes is based on the strong linkage disequilibrium of two *DCDC2* risk haplotypes to *READ1* (regulatory element associated with dyslexia; GenBank accession No BV677278). *READ1* regulates *KIAA0319* expression through a *KIAA0319* risk haplotype (Powers et al., 2016). Interestingly, Powers and colleagues suggest that *READ1* alleles act in both ways, either deleterious or protective depending on length or structure of the allele. Long *READ1* alleles with insertions in repeat unit 2 show significant associations with severe reading disability. Conversely, short *READ1* alleles with a deletion of one copy of repeat 1 showed nominal associations not surviving a Bonferroni correction (Powers et al., 2016). Further combined analyses testing different combinations of *READ1* and a *KIAA0319* risk haplotype confirm the deleterious effect of *READ1* allele 5 and *READ1* allele 6 in synergy with the *KIAA0319* risk haplotype. In contrast, *READ1* allele 3 in combination with the same *KIAA0319* risk haplotype was related to unaffected phenotypes. This gene-behaviour association demonstrates opposing effects of dyslexia risk genes on behaviour. In the light of these opposing effects it occurs likely that *DCDC2* and *KIAA0319* risk alle-

les could conversely act on brain-gene relationships. The present data suggest such a *KIAA0319-DCDC2* interaction to have an impact on the response consistency of the auditory brainstem.

The neurobiological substrates dominantly contributing to the complex auditory brainstem potentials are the nuclei of the auditory midbrain including the inferior colliculus and the lateral lemniscus. However, it cannot be excluded that preceding central auditory processes e.g. in the auditory nerve, cochlear nucleus or the superior olive could contribute to irregular patterns in scalp-recorded responses, which aggregate phase-locked activity of the auditory midbrain (Bidelman, 2015; Glaser et al., 1976; Skoe and Kraus, 2010; Smith et al., 1975; Sohmer et al., 1977). Information about gene expression in the auditory pathway and especially in the inferior colliculus and the lateral lemniscus is fragmentary, and gene expression across mammals and life span is diverse. The Allen Brain Atlas (www.brain-map.org) provides the first neuroanatomical precise, genome-wide maps of transcript distributions. *KIAA0319* and *DCDC2* are highly expressed in the central nucleus of the inferior colliculus of the prenatal human brain relative to other tissues (Miller et al., 2014; Shen et al., 2012). In the Adult Human Brain Tissue Gene Expression Profiles such information is still missing (Hawrylycz et al., 2012).

How these genes eventually interact with neurophysiology especially in the region of the inferior colliculus and the lateral lemniscus and with behavior is largely unknown. Lately, animal studies started elucidating electrophysiological correlates of *KIAA0319* and

DCDC2 knockdown in cortical regions of rodents (Centanni et al., 2016, 2014a,b; Che et al., 2014; Truong et al., 2014). In the context of auditory processing it is of particular interest that in utero RNAi of *Kiaa0319* knockdown in rats introduced a decreased consistency of responses to speech combined with a reduced neural discrimination ability of speech sounds (Centanni et al., 2014a,b). The authors report that a reduced expression of *Kiaa0319* increased neural excitability and input resistance when recording from affected neurons in the auditory cortex. These observations are in line with the current finding that a decreased response consistency was associated with an increased genetic burden of *KIAA0319* risk alleles. A most recent electrophysiological study of the same group reports that suppression of the dyslexia risk variant of *Dcdc2* in rats does not affect spike timing at the level of the auditory cortex, although animals lost the ability to identify speech sounds from a continuous stream (Centanni et al., 2016). This is reminiscent of the current findings. The present data show that *DCDC2* risk alleles have a trending association with an increased response consistency. This indicates better phase-locking capabilities of neurons in the auditory midbrain in carriers of *DCDC2* risk alleles. However, a different animal study showed that a *Dcdc2* knockout impaired temporal encoding as evident in degraded neural spike timing during the encoding of rapid sequential sensory inputs to the somatosensory cortex (Che et al., 2014). This discrepancy might be disentangled on the background of above elaborated *trans-gene* interactions suggesting that certain *DCDC2* haplotypes regulate *KIAA0319* expression resulting in opposing behavioral effects.

Here, we present the first observation of an interaction between dyslexia risk genes and the encoding of sounds in the auditory brainstem in humans. The small number of participants makes it necessary to consider these observations as preliminary with a need for replication. To date, no studies exist elucidating the physiology of *Kiaa0319* or *Dcdc2* knockout or knockdown in auditory brainstem regions. Hence, future studies are necessary to further elucidate the here observed brain-gene interactions. The specification of such brain-gene interactions will ultimately lead to a better understanding of the origin of diverse dyslexia phenotypes.

Authors contributions

N.E.N. and A.F. conceived and designed the experiments, N.E.N., B.M., J.L., and G.S., performed the experiments; N.E.N., B.M., J.L. analyzed the data; N.E.N., M.G., I.K., N.K., H.K., A.W., M.A.S., J.B., F.E., and T.G., contributed unpublished analytic tools, N.E.N. wrote the paper. All authors commented on the paper.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.dcn.2017.01.008>.

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