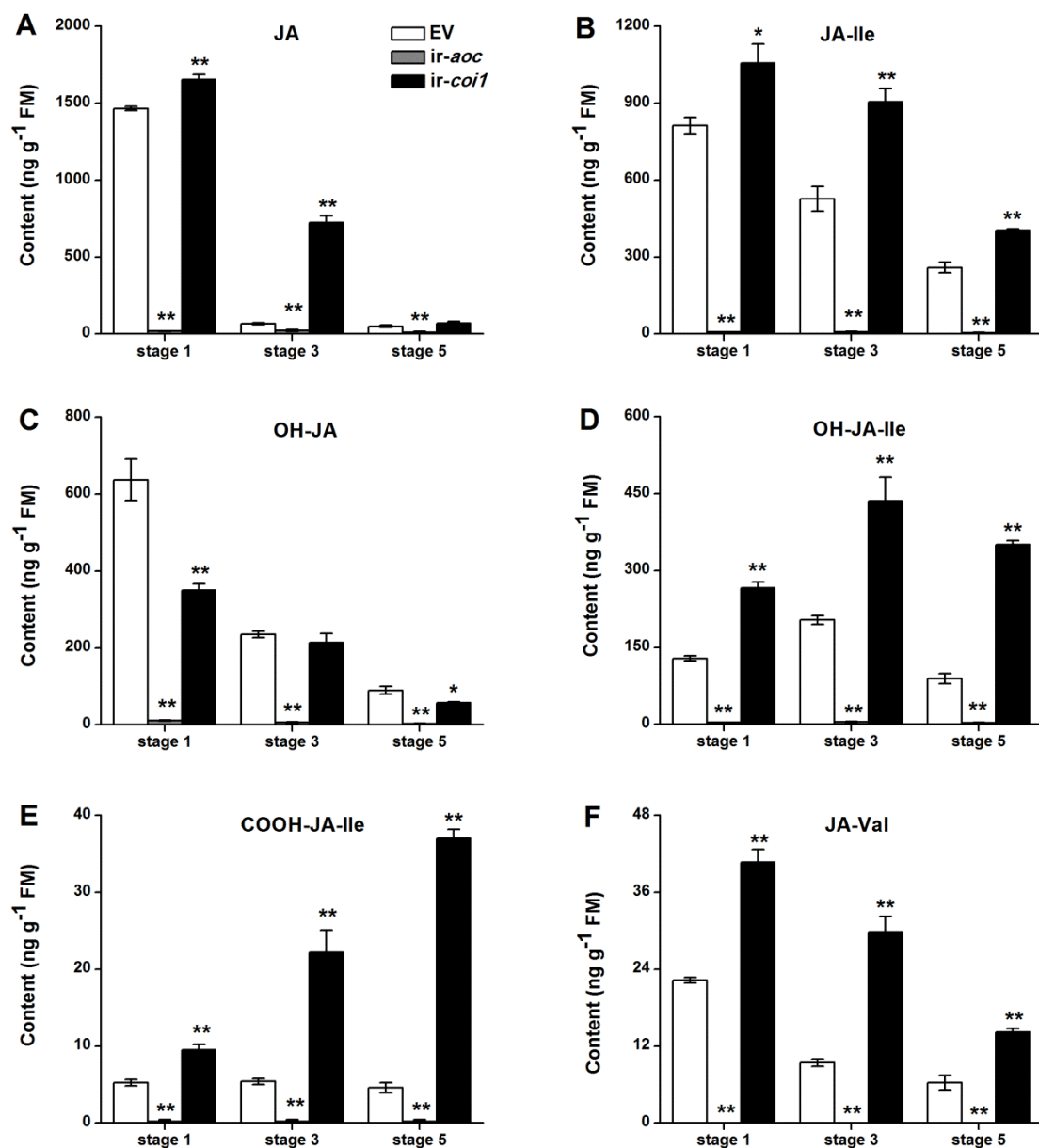
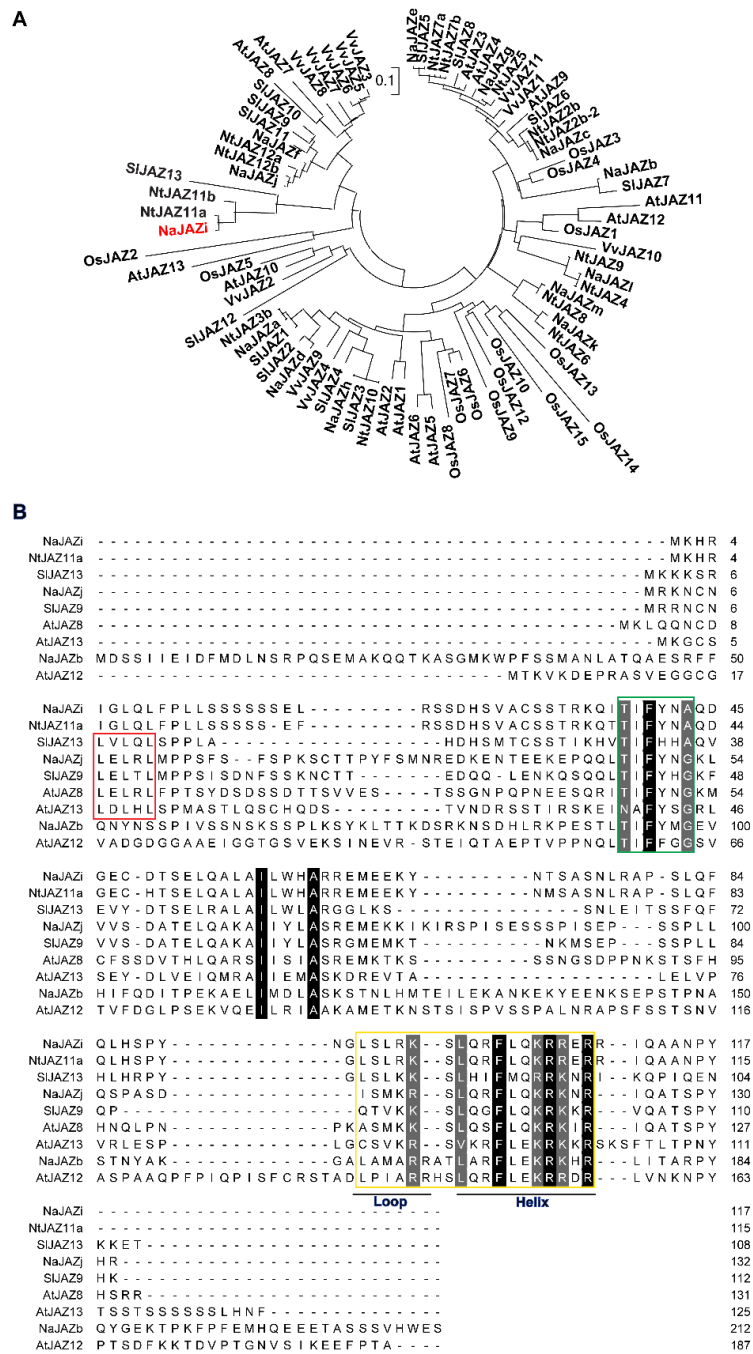


## Supporting Information Appendix



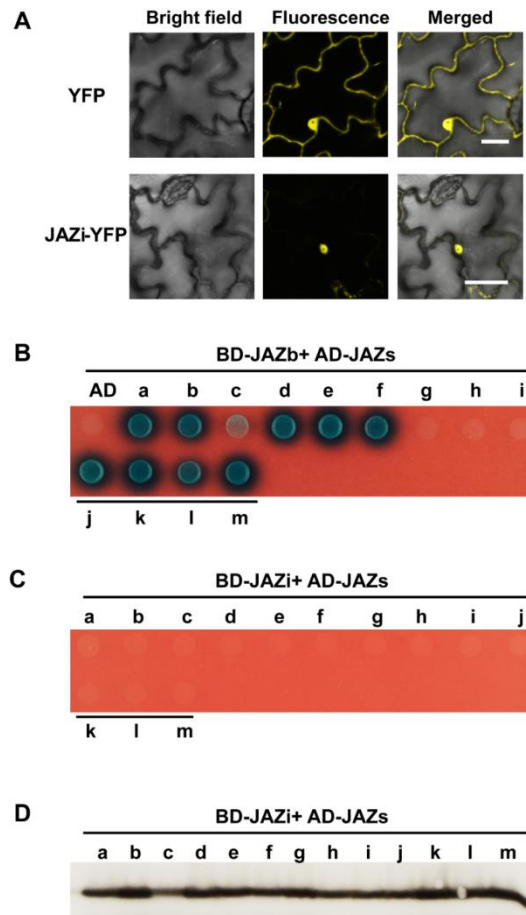
**Fig. S1.** JA levels in lines deficient in JA biosynthesis (*ir-aoc*) and perception (*ir-coi1*).

Mean JA (A), JA-Ile (B), OH-JA (C), OH-JA-Ile (D), COOH-JA-Ile (E) and JA-Val (F) levels ( $\pm$ SE,  $n=5$ ) in flowers of EV, *ir-aoc* and *ir-coi1* plants. Samples from three different floral stages were analyzed. Asterisks indicate significant differences in *ir-aoc* or *ir-coi1* compared with control plants (\*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; Student's t-test).



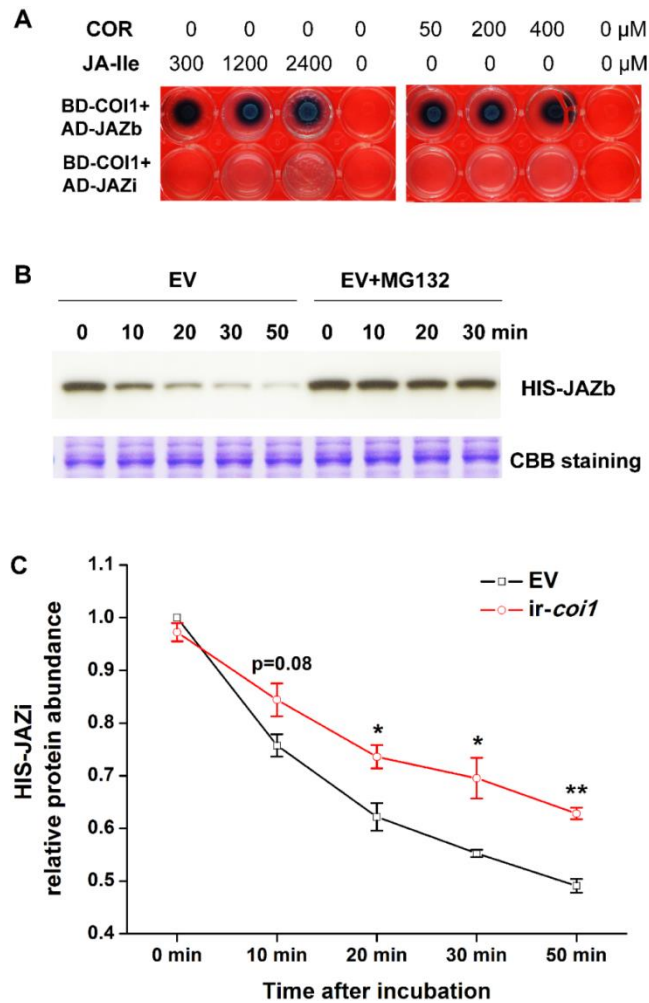
**Fig. S2.** Phylogenetic analysis and sequence alignment of JAZ genes in different species.

(A) Phylogenetic analysis of JAZ gene family from *N. attenuata* (Na), *Oryza sativa*, (*Os*), *N. tabacum* (Nt), *Vitis vinifera* (Vv), *Solanum lycopersicum* (Sl) and Arabidopsis (*At*). (B) Sequence alignment of JAZ genes from different species. EAR motif was highlighted in red box. TIFY domain was highlighted in green box. Jas motif was highlighted in yellow box.



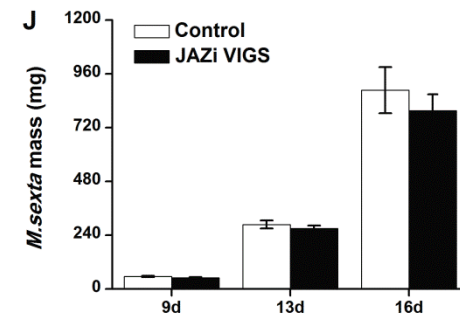
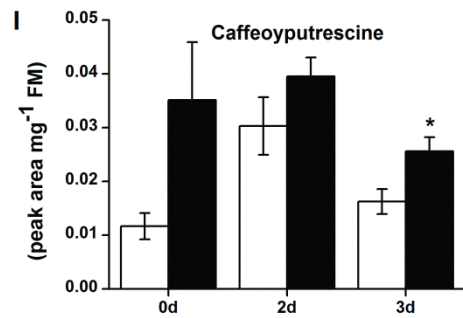
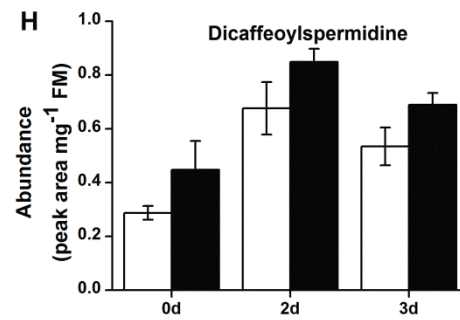
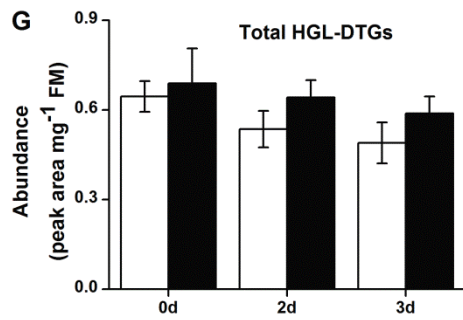
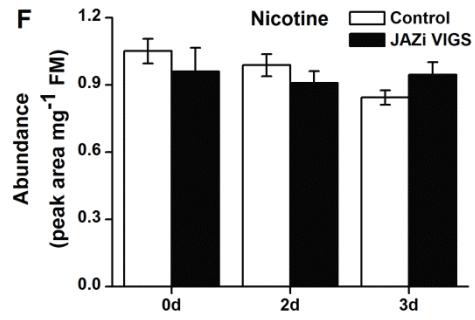
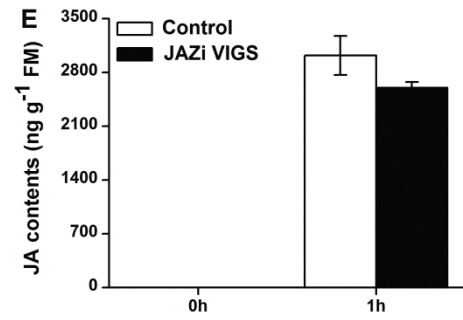
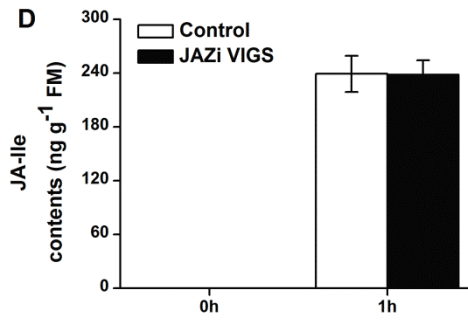
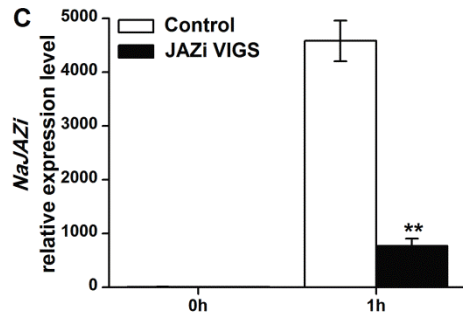
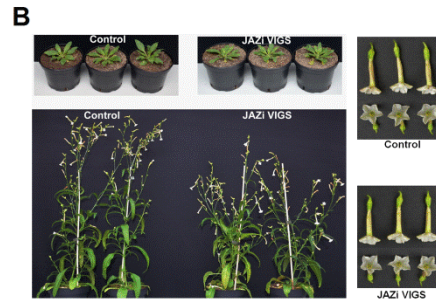
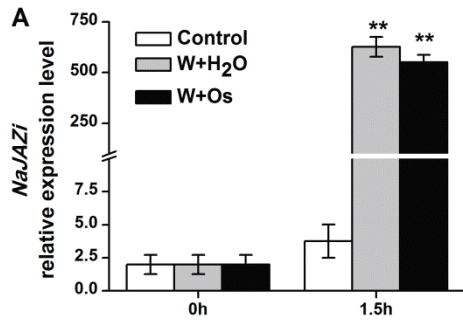
**Fig. S3.** Subcellular localization of JAZi and JAZ homo-/hetero dimerization.

(A) Subcellular localization of NaJAZi. YFP and JAZi-YFP were transiently expressed in *N. attenuata* leaves. After incubation for 48 h, transformed cells were observed under a confocal microscope. The photographs were taken in UV light, visible light, and in combination (merged), respectively. Scale bar, 20  $\mu$ m. (B) NaJAZb homo- and heterodimerization assay by yeast two-hybrid. BD-JAZs and AD-JAZb were co-transformed into yeast strain Y2Hgold. BD and AD-JAZb co-transformed yeast were used as control. (C) NaJAZi homo- and heterodimerization assays were conducted by yeast two-hybrid assays. BD-JAZs and AD-JAZi were co-transformed into yeast strain Y2Hgold. The transformants were grown on QDO (SD/-Ade/-His/-Leu/-Trp/) plate with 40mg/L X- $\alpha$ -gal. (D) Immunoblot analysis of JAZ proteins in yeast strains used for Y2H assays shown in (C). BD-JAZi fusion protein was detected with anti-myc antibody.



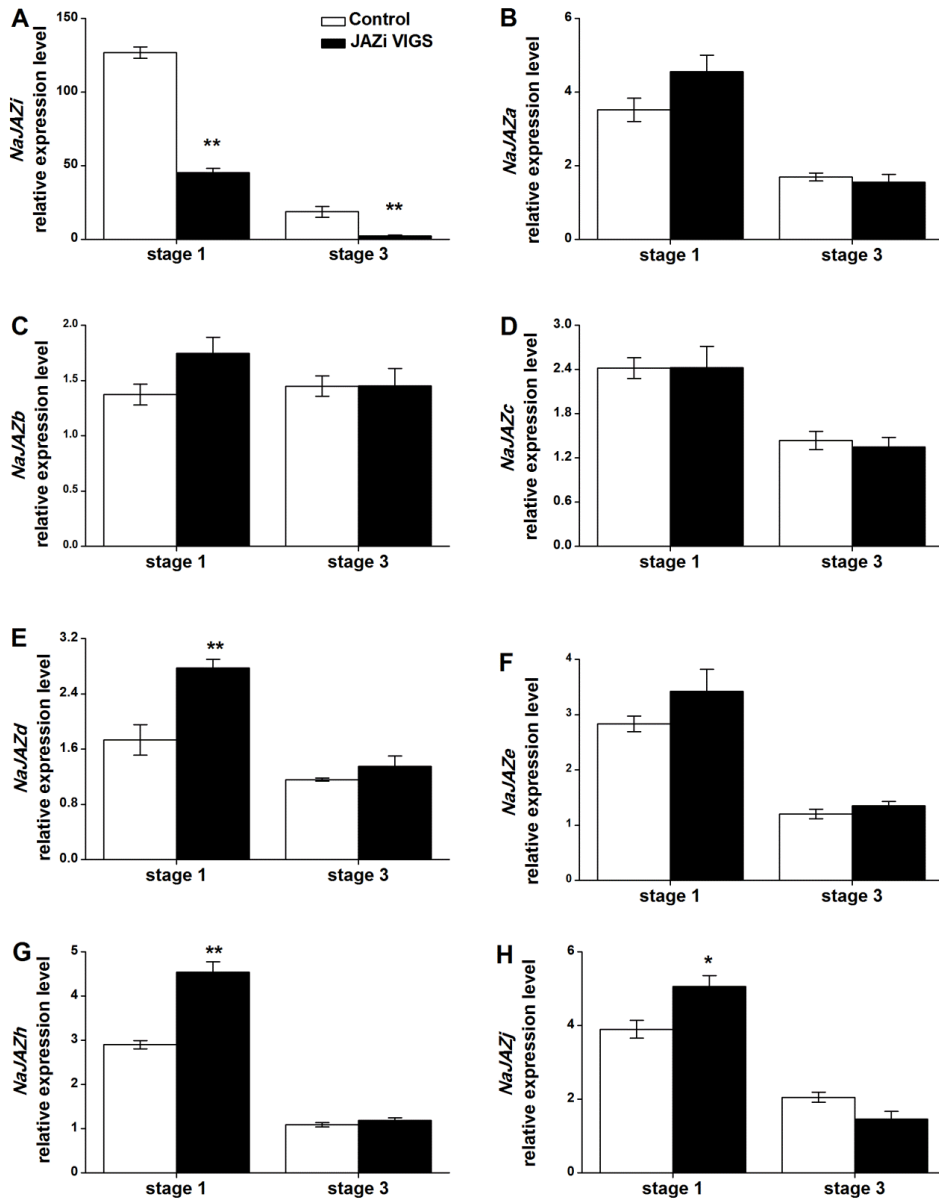
**Fig. S4. Interactions between NaCOI1 and NaJAZs and *in vitro* NaJAZ degradation assay.**

(A) Interactions between NaCOI1 and NaJAZ proteins by yeast two-hybrid assays. GAL4 DNA-binding domain (BD) and activation domain (AD) fusions were co-transformed into yeast strain Y2Hgold. The transformants were grown on QDO (SD/-Ade/-His/-Leu/-Trp/+40mg/L X- $\alpha$ -gal) plates in the presence of coronatine (COR) or JA-Ile or a solvent control. (B) *In vitro* JAZb degradation assays. Purified HIS-JAZb was incubated with total crude extracts from EV and total crude extracts from EV plus MG132. HIS-JAZb was detected using anti-HIS antibody at the indicated incubation time points. The Coomassie Brilliant Blue (CBB) staining is shown as a protein loading control. (C) Mean relative abundance ( $\pm$ SE, n=3) of HIS-JAZi protein levels in different treatments. The relative protein abundance was analyzed by ImageJ software.



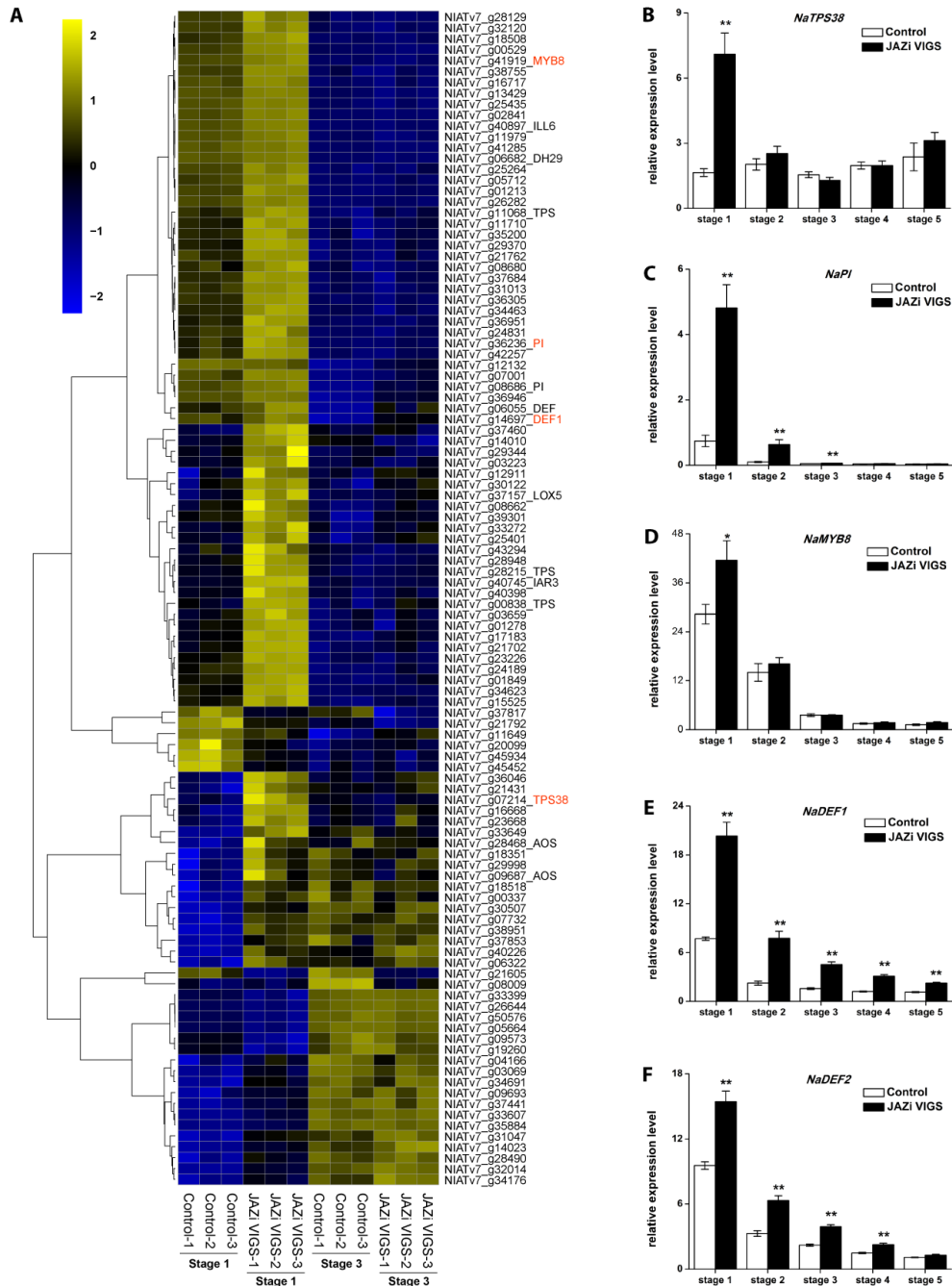
**Fig. S5.** Silencing of *NaJAZi* does not affect leaf resistance to *M. sexta* larvae.

(A) Mean transcripts levels ( $\pm$ SE, n=5) of *NaJAZi* in *N. attenuata* leaves after W+R or wounding control (W+H<sub>2</sub>O). Transcripts levels were analyzed by RT-qPCR. (B) Growth phenotype of *NaJAZi*-silencing plants. (C) Mean transcript levels ( $\pm$ SE, n=7) of *NaJAZi* in leaves of VIGS control and *JAZi* VIGS plants after W+R treatment. Transcripts levels were analyzed by RT-qPCR. Mean JA (D) and JA-Ile (E) levels ( $\pm$ SE, n=7) in leaves of VIGS control and *JAZi* VIGS plants after W+R treatments. Samples were harvested before or 1 h after treatments. Mean nicotine (F), 17-hydroxygeranylinalool diterpene glycosides (DTGs) (G), caffeoyputrescine (H) and dicaffeoylspermidine (I) levels ( $\pm$ SE, n=7) in leaves of VIGS control and *JAZi* VIGS plants after W+R treatments. (J) *M. sexta* performance on VIGS control and *JAZi* VIGS plants. *M. sexta* larva mass ( $\pm$ SE, n=30) was measured at 9, 13 and 16 d after feeding on indicated plants. Asterisks indicate significant differences in *JAZi* VIGS plants compared with VIGS EV control plants (\*\*,  $p < 0.01$ ; Student's t-test).



**Fig. S6.** Transcript abundance of JAZ genes in *NaJAZi*-silenced plants.

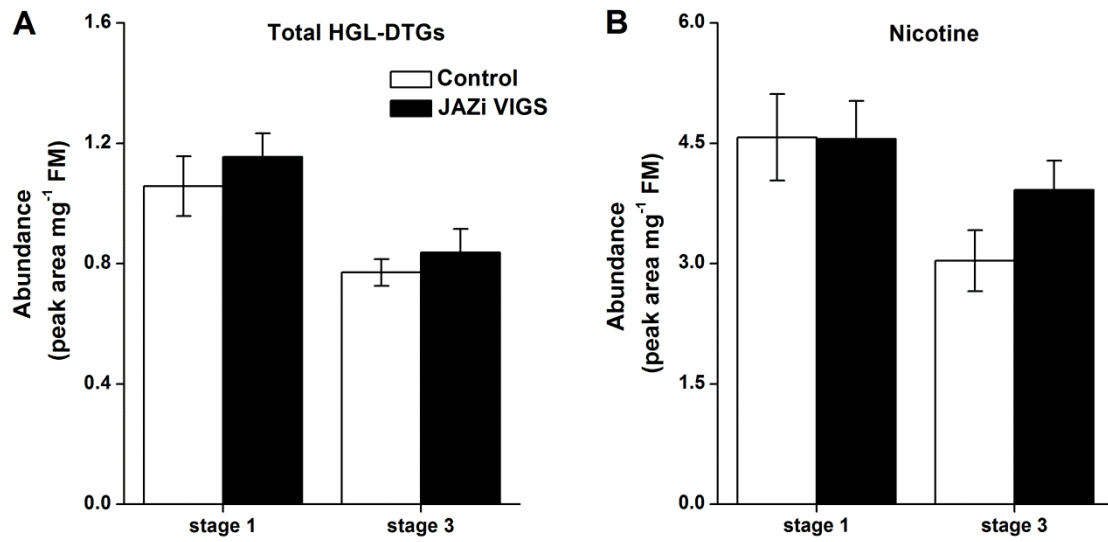
Mean transcript levels ( $\pm$ SE, n=5) of *JAZi* (A), *JAZa* (B), *JAZb* (C), *JAZc* (D), *JAZd* (E), *JAZe* (F), *JAZh* (G) and *JAZj* (H) in the flowers of VIGS control and JAZi VIGS plants. Transcripts levels were analyzed by RT-qPCR. Asterisks indicate significant differences in JAZi VIGS plants compared with control plants (\*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; Student's t-test).



**Fig. S7.** Up- and down-regulated genes in *NaJAZi*-silenced plants.

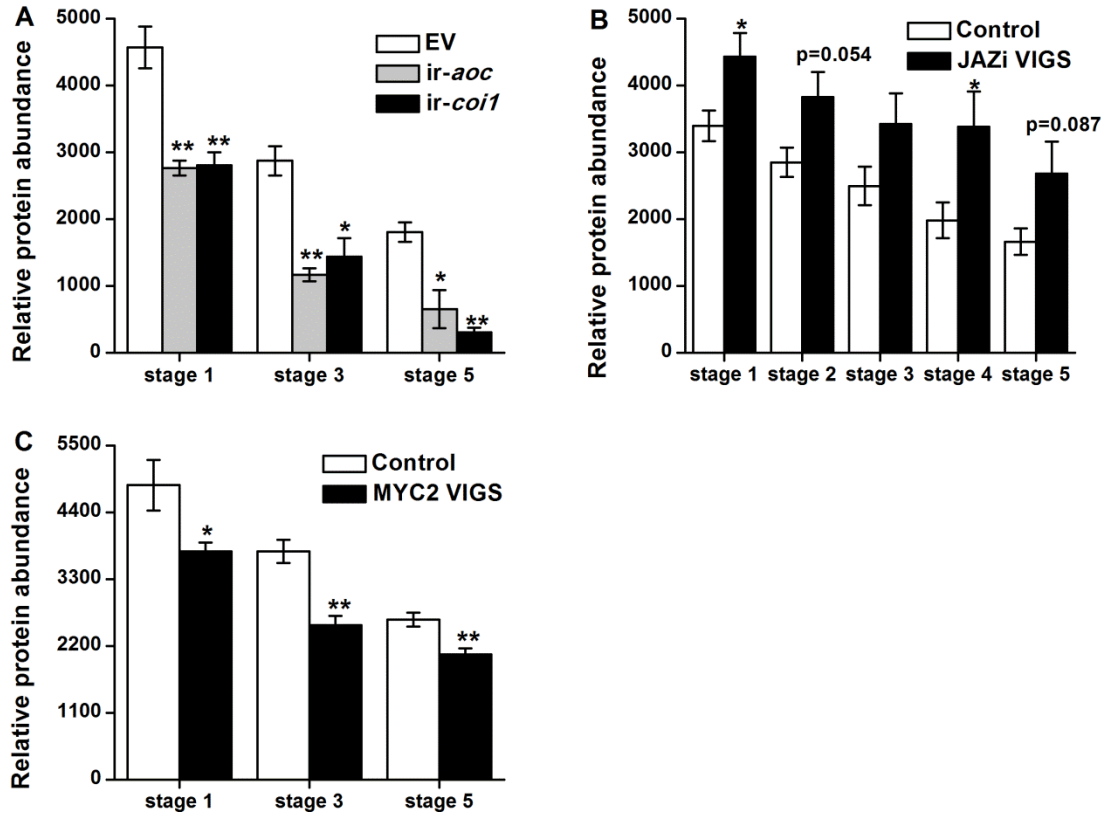
Heatmap shows up- and down-regulated genes in *JAZi*-silencing flowers. The color gradient refers to relative signal abundance. Mean transcript levels ( $\pm$ SE,  $n=5$ ) of *NaTPS38* (B), *NaPI* (C), *NaMYB8* (D), *NaDEF1* (E) and *NaDEF2* (F) in flowers of VIGS control and *JAZi* VIGS plants. Transcripts levels were analyzed by RT-qPCR. Asterisks indicate significant differences in *JAZi* VIGS plants compared with control plants (\*\*,  $p < 0.01$ ; Student's t-test).





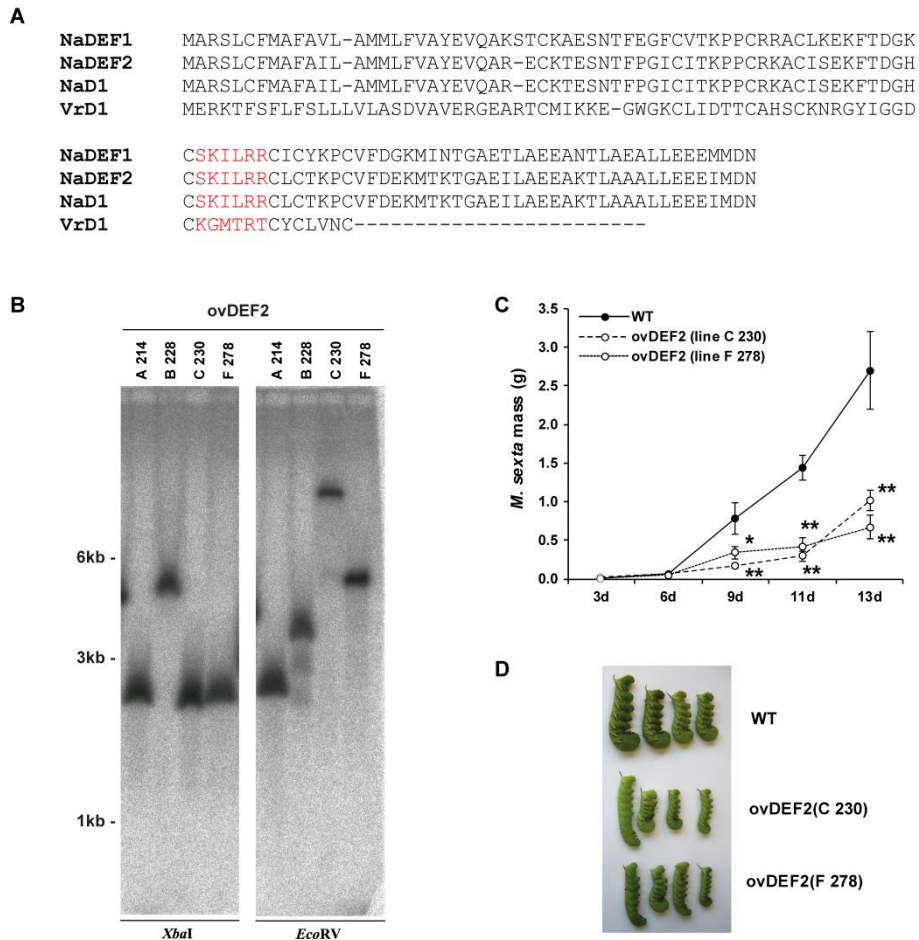
**Fig. S8.** Silencing of *NaJAZi* does not affect nicotine and DTG levels in developing flowers.

Mean 17-hydroxygeranylinalool diterpene glycosides (DTGs) (A) and nicotine (B) relative abundance ( $\pm$ SE, n=5) in the flowers of VIGS control and JAZi VIGS plants.



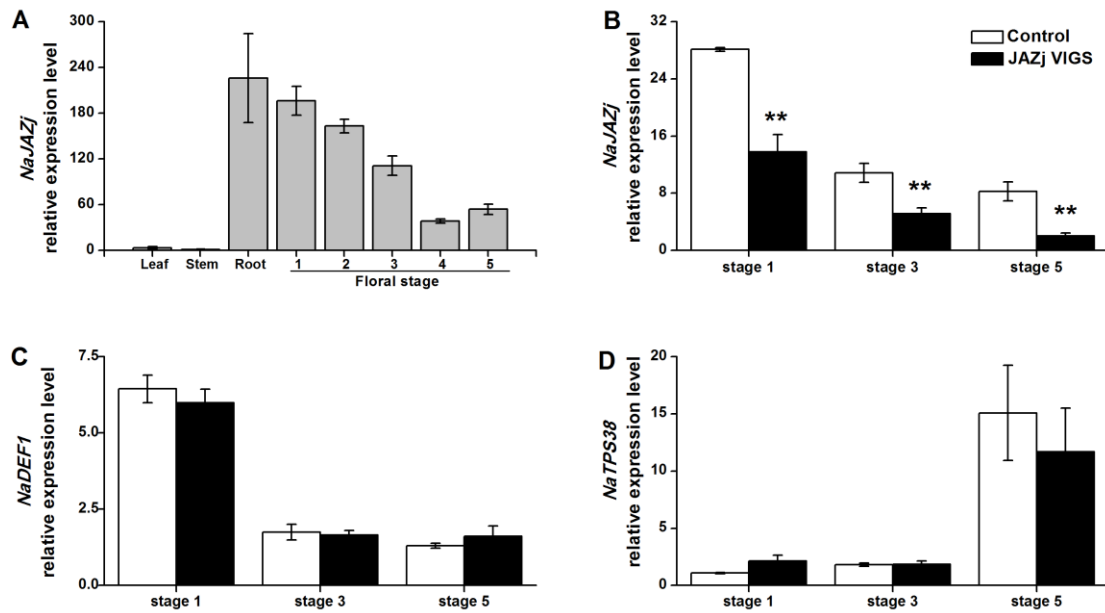
**Fig. S9.** Relative defensin protein levels in different genotypes.

(A) Mean defensin protein levels ( $\pm$ SE, n=3) in flowers of *ir-aoc*, *ir-coi1* and EV plants (\*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; Student's t-test). (B) Mean defensin protein levels ( $\pm$ SE, n=5) in flowers of VIGS control and JAZi VIGS plants (\*,  $p < 0.05$ ; Student's t-test). (C) Mean defensin protein levels ( $\pm$ SE, n=5) in flowers of VIGS control and MYC2 VIGS plants (\*,  $p < 0.05$ ; Student's t-test). The relative protein abundance was analyzed by ImageJ software.



**Fig. S10.** Ectopic expression of *NaDEF2* in leaves enhances resistance to *M. sexta* larvae.

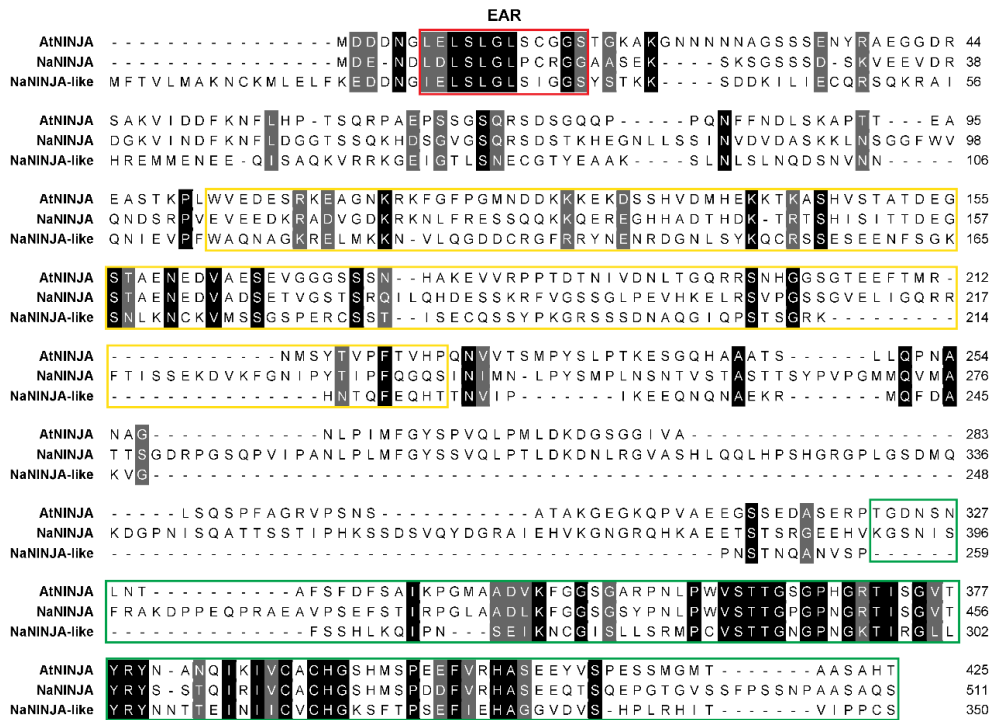
(A) Alignment analysis of *N. attenuata* NaDEF1, NaDEF2, *N. alata* NaD1 and *V. radiata* VrD1. The putative  $\alpha$ -amylase inhibitory activity region is highlighted in red. (B) Southern blot analysis for the determination of T-DNA copy numbers of four independent *ovDEF2* lines. Genomic DNA was isolated from seedlings homozygous to the hygromycin resistance marker and digested in separate reactions with *Xba*I and *Eco*RV. A radiolabeled fragment of the hygromycin resistance marker (*hpt*II) served as probe. (C) *M. sexta* performance on leaves of rosette-stage wild type (WT) and *ovDEF2* plants in the glasshouse. *M. sexta* larval mass was determined 3, 6, 9, 11 and 13 days after infestation ( $\pm$ SE, n = 12 plants). Asterisks indicate statistically significant differences between control and transgenic plants (\*, p < 0.05; \*\*, p < 0.01; Student's t-test). (D) Pictures depict *M. sexta* caterpillars after 13 days of feeding from WT and *ovDEF2* plants.



**Fig. S11.** Silencing of *NaJAZj* does not affect the expression of floral defense-related genes.

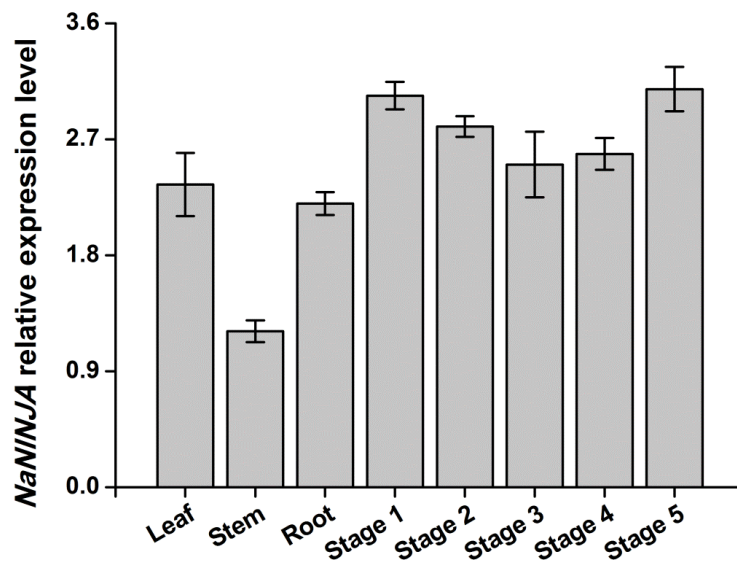
Mean transcript levels ( $\pm$ SE,  $n=5$ ) of *NaJAZj* in different tissues of *N. attenuata*.

Transcripts levels were analyzed by RT-qPCR. (B) Mean transcripts levels ( $\pm$ SE,  $n=4-5$ ) of *NaJAZj* (B), *NaDEF1* (C) and *NaTPS38* (D) in the flowers of VIGS control and JAZj VIGS plants. Transcripts levels were analyzed by RT-qPCR. Asterisks indicate significant differences in NINJA-like VIGS plants compared with control plants (\*\*,  $p < 0.01$ ; Student's t-test).



**Fig. S12.** Alignment analysis of NINJA and NINJA-like.

EAR motif is highlighted in the red box. B domain is highlighted in the yellow box and the C domain is highlighted in the green box.



**Fig. S13.** Transcript accumulations of the *NaNINJA* gene.

Mean transcripts levels ( $\pm$ SE, n=5) of *NaNINJA* in different tissues of *N. attenuata*.

Transcripts levels were analyzed by RT-qPCR.

**Table S1.** The transcripts per million (TPM) values of JAZs from the RNA-seq analysis.

Gene name	Leaf control	Leaf treated	Stem treated	Root treated	Flower bud	Corolla early	Corolla late	Opening flower	Pedicel	Stigma	Nectary	Anther	Ovary
NaJAZa	43.95	1634.56	800.62	344.79	216.39	274.45	104.7	70.56	152.9	138.37	105.83	114.17	108.3
NaJAZb	71.67	431.55	251.72	5.02	24.27	43.09	43.13	20.06	15.24	19.48	2.2	12.91	13.61
NaJAZc	24.4	114.45	69.67	78.99	89.16	67.87	63.8	51.38	72.59	102.1	66.49	21.04	84.32
NaJAZd	7.74	736.5	359.93	78.29	37.15	38.95	47.38	34.15	17.65	47.27	67.33	50.82	53.3
NaJAZe	4.47	9.56	8.46	77.91	23.47	14.52	5.9	8.06	8.31	10.99	12.37	20.86	17.33
NaJAZf	0	151.31	3.5	1.09	1.11	2.59	0.3	0	0	0	0	0.75	0
NaJAZg	0	0	0	51.52	0	0.08	0	0	0.1	0.12	0	0	0
NaJAZh	42.29	740.37	551.18	48.82	74.2	54.96	28.27	17.65	33.64	47.41	30.33	31.28	33.9
NaJAZi	0	0.53	0.29	0.07	157.91	248.72	0.57	1.08	0.25	151.92	8.93	61.48	16.27
NaJAZj	0.59	690.1	219.95	11.69	35.08	35.65	20.26	12.43	3.14	12.08	13.81	40.8	7.78
NaJAZk	1.1	1.74	3.6	2.56	2.21	2.52	4.22	1.72	3.35	2.21	2.63	0.84	3.21
NaJAZl	0	0.17	0.2	0.23	0	0.18	0	0.24	0	0	0	0	0.24
NaJAm	3.71	2.73	9.33	10.93	9.01	7.67	8.58	15.1	6.58	10.5	16.14	6.4	26.73

**Table S2.** DNA primers used in this study.

Gene	Sequence (5'-3')	Purpose
JAZi-RT-F	TCATTCTGTGGCATGTTTCGT	RT-qPCR
JAZi-RT-R	TGAAACTGCAGAGATGGTGC	RT-qPCR
AD/BD-NaJAZi-F	GCCAGCATATGATGAAGCACAGAATTGGCCT	Y2H
AD/BD-NaJAZi-R	TCCATCCCGGGTGGTATGGATTAGCTGCTTGAA	Y2H
AD/BD-NaJAZb-F	GCCAGCATATGATGGATTCAAGTATTATTGA	Y2H
AD/BD-NaJAZb-R	TCAATCCCGGGTGGCTTTCCCAATGAACGCTTG	Y2H
BD-COI1-F	TTCATCATATGATGGAGGAGCGTAGCTCCAC	Y2H
BD-COI1-R	TCCATCCCGGGAATTCAGCGAGAAGGGAATTTG	Y2H
AD-MYC2a-F	CCCAGCATATGATGACGGACTATAGAATACC	Y2H
AD-MYC2a-R	TACAGATCGATGTCGCGATTCAGCAATTCTGG	Y2H
AD-MYC2b-F	CCCAGCATATGATGAATTTGTGGAATACTAG	Y2H
AD-MYC2b-R	TACAGATCGATGGCGTGTTCAGCAACTCTGG	Y2H
AD/BD-NINJA-F	GCGCGCATATGATGGATGAAAACGATCTTGA	Y2H
AD/BD-NINJA-R	TAAATCCCGGGAAGCTTTGGGCAGAGGCAGCCG	Y2H
AD-NaJAZa-F	GCCAGCATATGATGGCATCATCGGAGATTGT	Y2H
AD-NaJAZa-R	TCCATCCCGGGTGGACGAATTGAATACCTACAC	Y2H
AD-NaJAZc-F	TCAGTCCCGGGTATGGAGAGAGATTTTATGGG	Y2H
AD-NaJAZc-R	TCAATCTCGAGCGGTCTCCTTACCGGCTATCA	Y2H

BD-NaJAZc-F	TCAGTCCATGGAGATGGAGAGAGATTTTATGGG	Y2H
BD-NaJAZc-R	TCAATGTTCGAC G GGTCTCCTTACCGGCTATCA	Y2H
AD/BD-NaJAZd-F	GCCAGCATATGATGGGGTTATCGGAGATTGT	Y2H
AD/BD-NaJAZd-R	TCGATCCCGGGTGAAAGAACTGCTCAGTTTTCA	Y2H
AD/BD-NaJAZe-F	GCCAGCATATGATGGGGTTGACTCATCATGT	Y2H
AD/BD-NaJAZe-R	TCAATCCCGGGTGCGTCTCCTTGACCAAATTGA	Y2H
AD/BD-NaJAZf-F	GCCAGCATATGATGAGAAGAACTGTAACCTG	Y2H
AD/BD-NaJAZf-R	TCGATCCCGGGTGGTGTATGATATGGAGAAGTTT	Y2H
AD/BD-NaJAZg-F	GCCAGCATATGATGGAGAGAGATTTTCATGGG	Y2H
AD/BD-NaJAZg-R	TCGATCCCGGGTGTATAGTAGCAGGAAGAACAG	Y2H
AD/BD-NaJAZh-F	GCCCGCATATGATGTCAAATTCGCAAATTC	Y2H
AD/BD-NaJAZh-R	TCGGTCCCGGGTGTAACCTGAAATTGAGATCGA	Y2H
AD/BD-NaJAZj-F	GCCCGCATATGATGAGAAAAAACTGTAACCT	Y2H
AD/BD-NaJAZj-R	TCATTCCCGGGTGGCGATGATAAGGAGAAGTTG	Y2H
AD/BD-NaJAZk-F	GACAGCATATGATGCCGCCGGAAGAATCAGT	Y2H
AD/BD-NaJAZk-R	TAGGTCCCGGGTGCCTGTCTTTTCGCTTCTCAA	Y2H
AD/BD-NaJAZl-F	GCCCGCATATGATGTATTGCAGCTCCAAAGT	Y2H
AD/BD-NaJAZl-R	CAGGTCCCGGGTACTATTCTTTTCCTTCAAAC	Y2H
AD/BD-NaJAZm-F	GCCAGCATATGATGGCGCCGGAAGAAACAGT	Y2H
AD/BD-NaJAZm-R	TCCATCCCGGGTGCTCTTTGGCATCTTTGTCAT	Y2H
AD-TOPLESS-F	CCGCAGAATTCATGTCATCTCTCAGCAGAGA	Y2H
AD-TOPLESS-R	AATCAGGATCCCTCTTGGTGCTTGTTCGGAGC	Y2H
AD/BD-NINJA-like-F	CCGCAGAATTCATGTTTACTGTGTTAATGGC	Y2H
AD/BD-NINJA-like-R	AATCAGGATCCAGAACAAAGGGGGGATTACAG	Y2H
TOPO-NINJA-like-F	CACCATGTTTACTGTGTTAATGGC	pENTR
TOPO-NINJA-like-R	AGAACAAGGGGGGATTACAG	pENTR
TOPO-JAZi-F	CACCATGGGCTGTCCCTAAGAAAA	pENTR
TOPO-JAZi-R	ACAGGTGATCCAACCTTCCA	pENTR
NINJA-like-RT-F	GCTCAGAATGCTGGGAAAAG	RT-qPCR
NINJA-like-RT-R	CATCTTCCGGTGAACCACT	RT-qPCR
TPS38-RT-F	ATGGGCTGTTGGTTTCACT	RT-qPCR
TPS38-RT-R	TGCATTGATGTCCCATCTGT	RT-qPCR
PI-RT-F	ACACGAGACTTGGGAAATGG	RT-qPCR
PI-RT-R	GTGTCCCTGGAAAACCTTCA	RT-qPCR
MYB8-RT-F	ACCGGGACGAACAGATAATG	RT-qPCR
MYB8-RT-R	CGACGAAGAATTTGGGTGTT	RT-qPCR
DEF2-RT-F	TGCATTACCAAACCACTTG	RT-qPCR
DEF2-RT-R	CAGCCAAAGTTTTTGTCTCC	RT-qPCR
DEF1-RT-F	ACATTCGAGGGATTCTGCGT	RT-qPCR

DEF1-RT-R	TCCTCGGCTAAAGTTTCAGCT	RT-qPCR
NINJA-RT-F	ATCTACCGTGGGTCTCAACG	RT-qPCR
NINJA-RT-R	AGTTTGTCTTTCGCTTGCAT	RT-qPCR
JAZa-RT-F	ATGACGATATTCTACGGCGG	RT-qPCR
JAZa-RT-R	TAAGTGAAGCTCGTCTCGCA	RT-qPCR
JAZb-RT-F	ACACCAAATGCATCCACAAA	RT-qPCR
JAZb-RT-R	GACGCCGTTTCTTCTTCTTG	RT-qPCR
JAZc-RT-F3	TACCTGCCTCAGGTCATTCC	RT-qPCR
JAZc-RT-R3	GGAACCGCTGCTGACATTAT	RT-qPCR
JAZd-RT-F	ACCGCAGTTTTGAACCAACT	RT-qPCR
JAZd-RT-R	ATTTGCCTTAGCTGCTGGAA	RT-qPCR
JAZe-RT-F	CGCACTACACGTCGACAACCT	RT-qPCR
JAZe-RT-R	CAGCGCTGTTAGTTGGAACA	RT-qPCR
JAZh-RT-F	TCGAATTTTCGTGCAGACTTG	RT-qPCR
JAZh-RT-R	TACAGCACTCTGACGAACGG	RT-qPCR
JAZj-RT-F	AGCTCAGGCTTATGCCTCCT	RT-qPCR
JAZj-RT-R	TCTGAAATTGGTGACCGGAT	RT-qPCR
MYC2a-RT-F	GGCCCCGAACAACACTACTACA	RT-qPCR
MYC2a-RT-R	CCCCGTTCGATTAAAGTCTGA	RT-qPCR
MYC2b-RT-F	TCTGGTTCGATGAAGTCAAG	RT-qPCR
MYC2b-RT-R	CTGCTTCGACGTGATTCAAA	RT-qPCR
JAZj-VIGS-F	TCATCACTAGTCCACAGCAGCTAACAATATT	VIGS
JAZj-VIGS-R	ATCTCCCCGGGCAAAATATATGTACAAATGG	VIGS
MYC2a-VIGS-F	CCAGCACTAGTATCAAGAGGTAGCAACGATG	VIGS
MYC2a-VIGS-R	ACTTTCCCGGGGACACATTTGGTACAACAGC	VIGS
MYC2b-VIGS-F	CCAGCACTAGTATGAATTTGTGGAATACTAG	VIGS
MYC2b-VIGS-R	ACTTTCCCGGGGGCATAGGTCCATGTCTCGC	VIGS
JAZi-VIGS-F	TCATCACTAGTATCACAATTTTCTACAACG	VIGS
JAZi-VIGS-R	ATCTCCCCGGGGAAGTATCCGCATATCGCAA	VIGS
NINJA-like-VIGS-F	CACGCGGATCCATGTTTACTGTGTTAATGGC	VIGS
NINJA-like-VIGS-R	CCGCATCGATTTGATTCAAAGAAAGATTCA	VIGS

**Table S3.** Gene accession numbers used in this study

Name	Species	Accession number
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<b>AtAFP1</b>	<i>Arabidopsis</i>	AT1G69260
<b>AtAFP2</b>	<i>Arabidopsis</i>	AT1G13740
<b>AtAFP3</b>	<i>Arabidopsis</i>	AT3G29575
<b>AtAFP4</b>	<i>Arabidopsis</i>	AT3G02140
<b>AtNINJA</b>	<i>Arabidopsis</i>	AT4G28910
<b>AtJAZ1</b>	<i>Arabidopsis</i>	AT1G19180
<b>AtJAZ2</b>	<i>Arabidopsis</i>	AT1G74950
<b>AtJAZ3</b>	<i>Arabidopsis</i>	AT3G17860
<b>AtJAZ4</b>	<i>Arabidopsis</i>	AT1G48500
<b>AtJAZ5</b>	<i>Arabidopsis</i>	AT1G17380
<b>AtJAZ6</b>	<i>Arabidopsis</i>	AT1G72450
<b>AtJAZ7</b>	<i>Arabidopsis</i>	AT2G34600
<b>AtJAZ8</b>	<i>Arabidopsis</i>	AT1G30135
<b>AtJAZ9</b>	<i>Arabidopsis</i>	AT1G70700
<b>AtJAZ10</b>	<i>Arabidopsis</i>	AT5G13220
<b>AtJAZ11</b>	<i>Arabidopsis</i>	AT3G43440
<b>AtJAZ12</b>	<i>Arabidopsis</i>	AT5G20900
<b>AtJAZ13</b>	<i>Arabidopsis</i>	AT3G22275
<b>OsJAZ1</b>	<i>Oryza sativa</i>	Os04g55920
<b>OsJAZ2</b>	<i>Oryza sativa</i>	Os07g05830
<b>OsJAZ3</b>	<i>Oryza sativa</i>	Os08g33160
<b>OsJAZ4</b>	<i>Oryza sativa</i>	Os09g23660
<b>OsJAZ5</b>	<i>Oryza sativa</i>	Os04g32480
<b>OsJAZ6</b>	<i>Oryza sativa</i>	Os03g28940
<b>OsJAZ7</b>	<i>Oryza sativa</i>	Os07g42370
<b>OsJAZ8</b>	<i>Oryza sativa</i>	Os09g26780
<b>OsJAZ9</b>	<i>Oryza sativa</i>	Os03g08310
<b>OsJAZ10</b>	<i>Oryza sativa</i>	Os03g08330
<b>OsJAZ11</b>	<i>Oryza sativa</i>	Os03g08320
<b>OsJAZ12</b>	<i>Oryza sativa</i>	Os10g25290
<b>OsJAZ13</b>	<i>Oryza sativa</i>	Os10g25230
<b>OsJAZ14</b>	<i>Oryza sativa</i>	Os10g25250
<b>OsJAZ15</b>	<i>Oryza sativa</i>	Os03g27900
<b>NtJAZ2b</b>	<i>N. tabacum</i>	KC246550
<b>NtJAZ2b-2</b>	<i>N. tabacum</i>	KC246551
<b>NtJAZ3b</b>	<i>N. tabacum</i>	KC246552
<b>NtJAZ4</b>	<i>N. tabacum</i>	KC246553
<b>NtJAZ5</b>	<i>N. tabacum</i>	KC246554
<b>NtJAZ6</b>	<i>N. tabacum</i>	KC246555
<b>NtJAZ7a</b>	<i>N. tabacum</i>	KC246556

<b>NtJAZ7b</b>	<i>N. tabacum</i>	KC246557
<b>NtJAZ8</b>	<i>N. tabacum</i>	KC246558
<b>NtJAZ9</b>	<i>N. tabacum</i>	KC246559
<b>NtJAZ10</b>	<i>N. tabacum</i>	KC246560
<b>NtJAZ11a</b>	<i>N. tabacum</i>	KC246561
<b>NtJAZ11b</b>	<i>N. tabacum</i>	KC246562
<b>NtJAZ12a</b>	<i>N. tabacum</i>	KC246563
<b>NtJAZ12b</b>	<i>N. tabacum</i>	KC246564
<b>VvJAZ1</b>	<i>Vitis vinifera</i>	XM-002284819
<b>VvJAZ2</b>	<i>Vitis vinifera</i>	XM-002262714
<b>VvJAZ3</b>	<i>Vitis vinifera</i>	XM-003634778
<b>VvJAZ4</b>	<i>Vitis vinifera</i>	XM-002272327
<b>VvJAZ5</b>	<i>Vitis vinifera</i>	XM-002277733
<b>VvJAZ6</b>	<i>Vitis vinifera</i>	XM-002277769
<b>VvJAZ7</b>	<i>Vitis vinifera</i>	XM-002277916
<b>VvJAZ8</b>	<i>Vitis vinifera</i>	CBI30922
<b>VvJAZ9</b>	<i>Vitis vinifera</i>	XM-002277121
<b>VvJAZ10</b>	<i>Vitis vinifera</i>	XM-002263220
<b>VvJAZ11</b>	<i>Vitis vinifera</i>	XM-002282652
<b>SIJAZ1</b>	<i>Solanum lycopersicum</i>	Solyc07g042170
<b>SIJAZ2</b>	<i>Solanum lycopersicum</i>	Solyc12g009220
<b>SIJAZ3</b>	<i>Solanum lycopersicum</i>	Solyc03g122190
<b>SIJAZ4</b>	<i>Solanum lycopersicum</i>	Solyc12g049400
<b>SIJAZ5</b>	<i>Solanum lycopersicum</i>	Solyc03g118540
<b>SIJAZ6</b>	<i>Solanum lycopersicum</i>	Solyc01g005440
<b>SIJAZ7</b>	<i>Solanum lycopersicum</i>	Solyc11g011030
<b>SIJAZ8</b>	<i>Solanum lycopersicum</i>	Solyc06g068930
<b>SIJAZ9</b>	<i>Solanum lycopersicum</i>	Solyc08g036640
<b>SIJAZ10</b>	<i>Solanum lycopersicum</i>	Solyc08g036620
<b>SIJAZ11</b>	<i>Solanum lycopersicum</i>	Solyc08g036660
<b>SIJAZ12</b>	<i>Solanum lycopersicum</i>	Solyc01g009740
<b>SIJAZ13</b>	<i>Solanum lycopersicum</i>	LOC104649733