

CARSTEN KETTNER

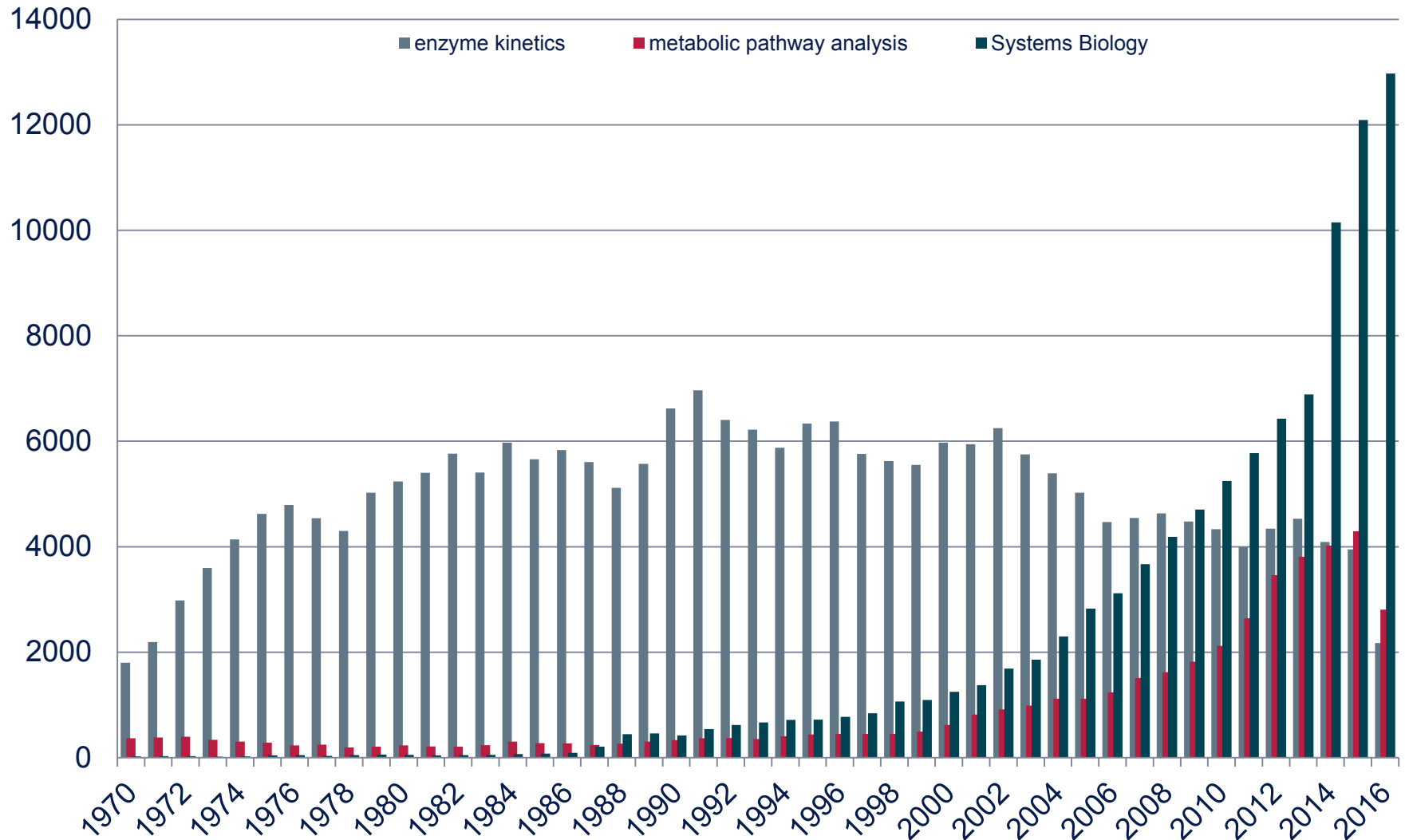
STRENDA DB: THE PDB FOR ENZYME FUNCTION DATA?

**OPEN SCIENCE DAYS 2017
BERLIN
16 & 17 OCTOBER 2017**



BEILSTEIN INSTITUT

Publications in enzymology



Source: PubMed (<http://www.ncbi.nlm.nih.gov/pubmed>), March 2017

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PLOS MEDICINE

Essay

October 2014 | Volume 11 | Issue 10 | e1001747

How to Make More Published Research True

John P. A. Ioannidis^{1,2,3,4*}

¹Meta-Research Innovation Center at Stanford (METRICS), Stanford University, Stanford, California, United States of America, ²Department of Medicine, Stanford Prevention Research Center, Stanford, California, United States of America, ³Department of Health Research and Promotion, Stanford University School of Medicine, Stanford, California, United States of America, ⁴Department of Statistics, Stanford University School of Humanities and

NATURE CELL BIOLOGY VOLUME 10 | NUMBER 10 | OCTOBER 2008

commentary

The challenges of integrating multi-omic data sets

Bernhard Palsson & Karsten Zengler

The capability to generate multi-omic data sets raises the issue of resource allocation for data generation versus data curation and integration. The initial experience of researchers shows that the effort required for the latter can be much greater than that for the former.

PeerJ

On the reproducibility of science: unique identification of research resources in the biomedical literature

Nicole A. Vasilevsky¹, Matthew H. Brush¹, Holly Paddock², Laura Ponting³, Shreejoy J. Tripathy⁴, Gregory M. LaRocca⁴ and Melissa A. Haendel¹

PeerJ 1:e148; DOI 10.7717/peerj.148

Missing and imprecise information ...on the assay conditions...

no indication of UniProtKB AC	85%
Missing enzyme concentration	63%
no indication of temperature	12%
"room temperature"	6%
incomplete biochemical reactions (missing substrates/ products)	14%
no standard units for concentrations of compounds	20%
experimental conditions in references	10%
inconsistent experimental conditions within the publication	6%

Survey in SABIO-RK, as for September 2017

Journals unite for reproducibility

Science, Nov. 2014, 346(6210)

PERSPECTIVE

doi:10.1038/nature11556

A call for transparency and optimization of preclinical research

Story C. Landis¹,
Robert B. Darnell⁸,
Robert M. Golub¹³,
John Huguenard¹⁸,
Malcolm R. Macleod¹,
John D. Porter¹, Osvaldo

Meeting in 2014:
“...scientific journals are standing together in their conviction that reproducibility and transparency are important issues...”
Marcia McNutt, *Science*, Nov. 2014 346(6210)

The signatories represent journals that publish preclinical biological research — an area of research that encompasses both exploratory studies and hypothesis-testing studies, with many different designs. The reproducibility of these studies is expected to vary. The journals agree to adhere to the following principles with the aim of facilitating the interpretation and repetition of experiments as they have been conducted in the published study. These measures and principles do not obviate the need for replication and reproduction in subsequent investigations to establish the robustness of published results across multiple biological systems.

<http://www.nih.gov/about/reporting-preclinical-research.htm>

Reporting Preclinical

Background

NIH held a joint workshop in June 2014 with the Nature Publishing Group and Science on the issue of reproducibility and rigor of research findings, with journal editors representing over 30 basic/preclinical science journals in which NIH-funded investigators have most often published. The workshop focused on the common opportunities in the scientific publishing arena to

STandards for Reporting ENzymology DAta



founded in 2003 and supported by the Beilstein-Institut
www.beilstein-strenda.org

Barbara M. Bakker,
Antonio Baici,
Athel Cornish-Bowden,
Paul F. Fitzpatrick,
Peter Halling,
Thomas S. Leyh,
Claire O'Donovan,
Frank M. Raushel



Johann M. Rohwer,
Santiago Schnell
Dietmar Schomburg,
Neil Swainston,
Ming-Daw Tsai,
Roland Wohlgemuth,
Ulrike Wittig,
Carsten Kettner



Standards for Reporting ENzymology DAta



Measuring enzyme activities under standardized *in vivo*-like conditions for systems biology

Karen van Eunen^{1,2}, Jildau Bouwman^{1,2}, Pascale Daran-Lapujade^{2,3}, Jarne Postmus⁴, André B. Canelas^{2,3}, Femke I. C. Menzonides^{1,2}, Rick Orij⁴, Isil Tuzun⁵, Joost van den Brink^{2,3}, Gertien J. Smits⁴, Walter M. van Gulik^{2,3}, Stanley Brul⁴, Joseph J. Heijnen^{2,3}, Johannes H. de Winde^{2,3}, M. Joost Teixeira de Mattos⁵, Carsten Kettner⁵, Jens Nielsen⁷, Hans V. Westerhoff^{1,2,8} and Barbara M. Bakker^{1,2,9}

¹ Department of Molecular Cell Physiology, Vrije Universiteit Amsterdam, The Netherlands

² Kluyver Centre for Genomics of Industrial Fermentation, Delft, The Netherlands

³ Department of Biotechnology, Delft University of Technology, The Netherlands

⁴ Department of Molecular Biology and Microbial Food Safety, Swammerdam Institute for Life Sciences, University of Amsterdam, The Netherlands

⁵ Department of Molecular Microbial Physiology, Swammerdam Institute for Life Sciences, University of Amsterdam, The Netherlands

⁶ Leibniz Institute for Food Biotechnology, Dortmund, Germany

⁷ University of Technology, Gøteborg, Sweden

⁸ Interdisciplinary Biocentre, The University of Manchester, UK

⁹ Infectious Diseases, University Medical Centre Groningen, University of Groningen, The Netherlands

Mission:

1. Development of experimental standard conditions;

actively engaged in ongoing efforts of the international scientific community to define standards for yeast and other organisms and to get them widely adopted. Hence, the authors would specifically welcome responses from readers who would like to be involved in such efforts and/or have specific comments on the proposed standards or the scientific strategy to define them.

(Received 7 October 2009, revised 20 November 2009, accepted 27 November 2009)

doi:10.1111/j.1742-4658.2009.07524.x

titative models require data from many laboratories. Therization of experimental systems and assay conditions is crucial. Standards should be representative of the *in vivo* conditions. However, enzyme-kinetic parameters are measured under assay conditions that do not yield the maximum activity of each enzyme. In practice, these kinetic parameters of different enzymes are measured in different media, at different pH values, with different ionic strengths, etc. In a previous paper (Bouwman et al., 2009), the Dutch Vertical Genomics Consortium, the European Yeast Network and the Standards for Reporting Enzymology Network (SREN) have developed a single assay medium for determining kinetic parameters in yeast. The medium is as close as possible to the conditions for the yeast *Saccharomyces cerevisiae*, and at the same time is experimentally feasible. The *in vivo* conditions were estimated for the yeast strain CEN.PK.113-7D grown in aerobic glucose-limited chemostat cultures at an extracellular pH of 5.0 and a specific growth rate of 0.1 h^{-1} . The cytosolic pH and concentrations of calcium, sodium, potassium, phosphorus, sulfur and magnesium were determined. On the basis of these data and literature data, we propose a defined *in vivo*-like medium containing 300 mM potassium, 50 mM phosphate, 245 mM glutamate, 20 mM sodium, 2 mM free magnesium and 0.5 mM calcium, at a pH of 6.8. The V_{max} values of the glycolytic and fermentative enzymes of *S. cerevisiae* were measured in the new medium. For some enzymes, the results deviated conspicuously from those of assays done under enzyme-specific, optimal conditions.



Standards for Reporting ENzymology DAta

Mission:

1. Development of experimental standard conditions;
2. **Definition of minimum information for reporting enzyme function data (STRENDA Guidelines);**

Checklist level 1A	
Data	Comments
Identity of the enzyme	
Name of reaction catalyst	name, preferably the accepted name from the IUBMB Enzyme list
EC number	
Sequence accession number	
Organism/species & strain	NCBI Taxonomy ID
Additional information on the enzyme	
Isoenzyme	naturally occurring variant
	that localization is based on
	determined
	source, procedure used or reference along with
	tagged, fusion protein, lacking native
	selection criteria. Specify whether protein or enzyme
	factors
Substrate purity	Origin of substrate
Measured reaction	as a stoichiometrically balanced equation
Assay temperature	
Assay pressure	if it is not atmospheric; indicate if not aerobic
Atmosphere if not air	
Metal salt(s) & concentrations	e.g., 10 mM KCl, 1.0 mM MgSO ₄
Other assay components	e.g., 1.0 mM EDTA, 1.0 mM dithiothreitol
Coupled assay components	if relevant
Enzyme/protein concentration	Molar concentration if known, otherwise mass concentration e.g., mg ml ⁻¹ or better μM

<http://www.beilstein-institut.de/en/projects/strenda/guidelines/>

STRENDA Guidelines highly recommended by:



ACS Catalysis
Archives in Biochemistry and Biophysics
Antimicrobial Agents and Chemotherapy
BBA (all nine sections)
Biochem. and Biophys. Res. Communications
Biochemical Journal
Biochemistry
Biophysical Chemistry
Clinical and Vaccine Immunology
eLife
FEBS Journal
Free Radical Research
Infection and Immunity
Journal of the American Chemical Society
mBio
Molecular and Cellular Biology
Proceedings of the National Academy of Sciences
The Journal of Bacteriology
The Journal of Biological Chemistry
The Journal of Virology
Trends in Biotechnology

FAIRsharing.org

PLoS Journals
BioMedCentral Journals
Beilstein J. Org. Chem.
eLife
FEBS Lett.
J. Biomed. Sci.
Nature
OMICS
Science

Standards are...

Standards are...

... demanded and appreciated by the community

... created by the community after internal and external consultation

... recommended by journals

... included in instructions for authors

Standards are...

Standards are...

... demanded and appreciated by the community

... created by the community after internal and external consultation

... recommended by journals

... included in instructions for authors

In addition, they are...

... often (comprehensively) long

... too complicated

... for experts only

Standards are useful...



Bosch Fawstin, <http://fawstin.blogspot.de>

Standards are useful...

...BUT...

...they are disregarded since

- “nobody” reads instructions
- not consistently enforced by editors

Some prerequisites for acceptance

- | | |
|---------------------------|-------------------------------|
| 1. Sufficiency. | Comprehensive and relevant |
| 2. Practicability. | Useful but not too burdensome |
| 3. Stability. | Long-term access and support |



Making standards useful



Home Manage Manuscripts Manage Users Query Guidelines References Help

Welcome admin | Logout

Experiment Overview ?

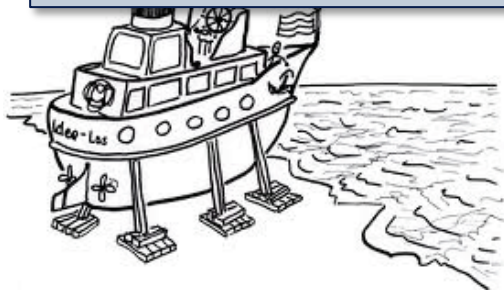
Manuscript Data	
Title	Description of methods to characterize the yeast hexokinase in detail
Author Names	Kettner, C., Doe, J., Hewitt, P.
Status	finalized
User	ckettner
PMID	25297050

Experiment	
Experiment	
Description	General characterization of hexokinase
Methodology	here the details of the experimental procedure with regards to methodology used and techniques ...

Missions:

1. Development of experimental standard conditions;
2. Definition of minimum information for reporting enzyme functional data (STRENDA Guidelines);
3. **Generation of a comprehensive data validation system (STRENDA DB).**

Adenosine triphosphate	Substrate	1	5 mM
pH: 5	pD:	Temperature: 25 °C	Protein Concentration: 86 nM





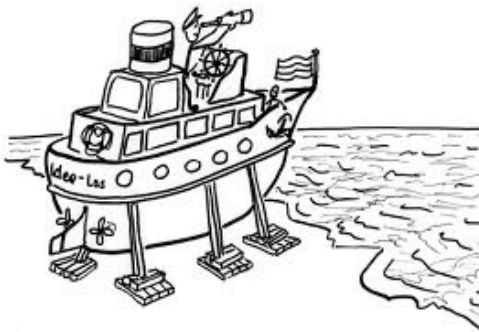
Making standards applicable

Mission:

1. Development of experimental standard conditions;
2. Definition of minimum information for reporting enzyme function data (STRENDA Guidelines);
3. **Generation of a comprehensive data validation system (STRENDA DB).**

EC Number	2.7.1.1
Sequence modifications	no
PTM	no
Organism	Saccharomyces cerevisiae (strain ATCC 204508 / S288c) (Baker's yeast)

- Customize, transform and update guidelines
- Assessment tool for authors and journals
- Direct data submission by authors
- Storage of data in a freely accessible database
- STRENDA DB: the “PDB for enzyme function”



Overview



IMPORTANT:

- not a substitute for the review process!
- emphasis on monitoring information rather than defining acceptance criteria

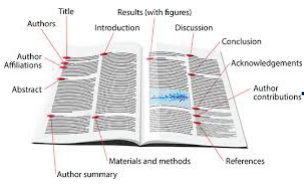
Review
correctio

Check for compliance with STRENDA guidelines



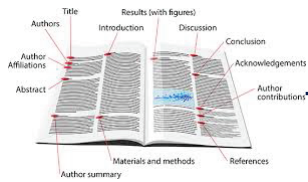
¹ PDB: 1BDG, hexokinase

² Acker & Auld (2014) *Perspectives in Science* 1(1-6):56-73.



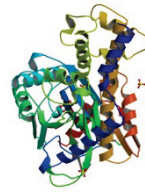
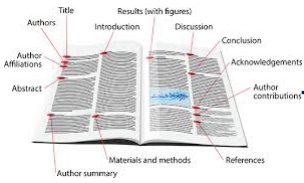
Definition of

- Type of Experiment
- Methodology
- Techniques



Definition of

- Type of Experiment
- Methodology
- Techniques



Protein data:

- Identifier
- Source
- Modifications
- Reaction



Add Protein ?

Manuscript Data Experiment

Is the protein data registered in UniProtKB? Yes No

Search for Proteins ?

Search for Proteins in UniProtKB Search

Protein Description ?

UniProtKB AC *

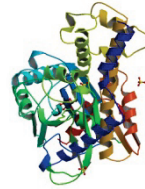
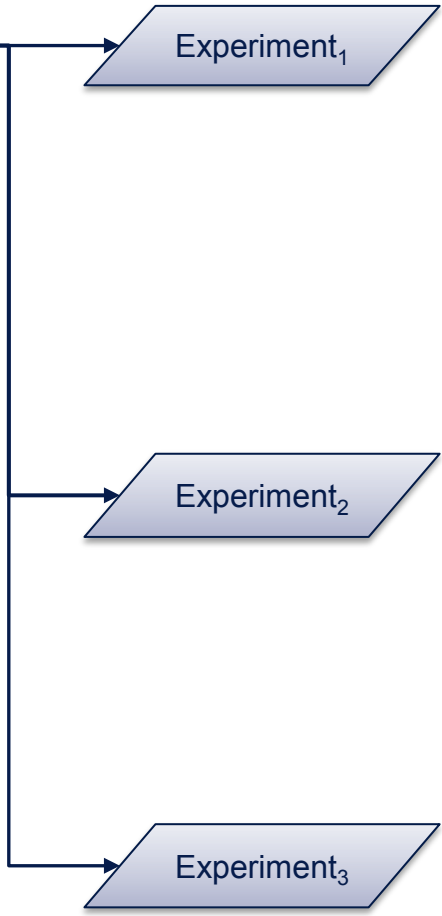
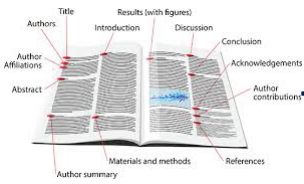
Protein Name *

Sequence *

```
MIVITTFVNGTSYCTVIASVQSYKAAIDFYTKFLSLENRSPDENSTLLSNDISLSKILLRPDEKINKNVEAHLKELNSITKIQDWRSHATQSLVFNTS
DILAVKDTLNAMNAPLQGYPTLFPMLQYTLDFLGNVVGVI STKNAVSTKPTPPAPEASAE SGLSSKVVSYTDLAYRMKTTDITYSLPKPLNRPQKAIA
VMTSGGDAPGMNSNVRVSAIFKGCRAFVVMGEYGLVRGGPEYIKE FHWEDVRGWSAEGGTNI GTARCMEFKKREGRLGAQHLI EAGVDALIVCGG
DGSLTGADLFRSEWPSLIEELLKTNRISNEQYERMKHLNICGTVGSIDNDMSTT DATIGAYSALDRICKAIDYVEATANSHSRAFVVEVMGRNCWLLALL
AGIATSADYIFIPEKPATSEWQDQMCDIVSKHRSRGRKRTIIVVVAEGAIADLTPI SPDVHKVLDVRLGLDTRITTLGHVQRGGTAVAYDRILATLQG
LEAVNAVLESTPDTFPLIAVNNKIVRKLPMESVKLTQVAEAIQAKDFKRAMSLRDTFIEHLNFMAINSADHNEPKLPKDKRKLKIAIVNVGAPAGG
INSAVYSMATYCMSQGHRPVAIYNGWSGLARHESVRLNWKMDLQWQRGGSEIGTNRVTFEEADLGMIAIYFQRYEFDGLIIVGGFEAFESLHQLERAR
ESYPAFRI PMVLI PATLSNNVPGTEYSLGSDTALNLM EYCDVVKQASSTRGRAFVVDCCGGNSGLATYASLAVGAQVSYVPEEGISLEQLSEDI EYL
AQSFKEAERGRFGKLLKSTNASKALSA TKLAEVITAEADGRFDKAPYPGHVQQGLSPFIDRTRATRMAIKAVGFIKDNQAIAEARRAAEENFNADD
KTI SDTAAVVGKGSVVVNSIRQLYDYETEVSRRMFKVIHWQAIRLIADHLVGRKRV D
```

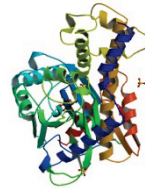
Protein Sequence Modifications ?

Does the protein contain any sequence modification(s) in comparison to that of the UniProtKB entry? Yes No Unknown



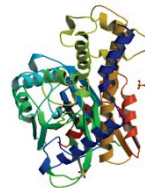
Protein data:

- Identifier
- Source
- Modifications
- Reaction



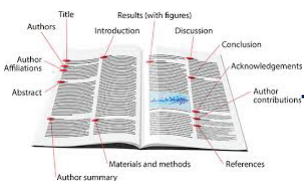
Protein data:

- Identifier
- Source
- Modifications
- Reaction



Protein data:

- Identifier
- Source
- Modifications
- Reaction



- Assay components
- Temperature
- pH



Assay Conditions ?

Manuscript Data +

Experiment +

Assay Conditions for Experimental Subset: 2

Small Assay Components

Role	Compound Name	ChEBI ID	PubChem CID	Stoich.	Concentration(s)	Actions
Substrate	Fructose 6-Phosphate	15946	69507	1	0.0 - 100 mM	Edit Delete
Salt	MAGNESIUM CHLORIDE	6636	5360315		20.0 mM	Edit Delete
Substrate	Adenosine triphosphate	30616	5957	1	10.0 mM	Edit Delete
Buffer	HEPES sodium salt set to pH 7.5 with HCl	46758	2724248		75.0 mM	Edit Delete
Salt	potassium chloride	32588	4873		150.0 mM	Edit Delete

Please create an entry in this table for every compound added to the assay mixture (except water, which may be taken to be the solvent unless shown otherwise).

[Add component](#)

Macromolecular Components

Role	Class	Compound Name	Database used	Identifier	Stoich.	Concentration(s)	Actions
No records found.							

Please create an entry in this table for every compound added to the assay mixture (except water, which may be taken to be the solvent unless shown otherwise).

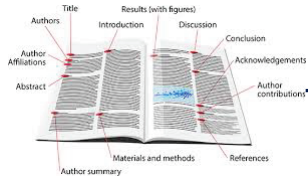
[Add component](#)

Concentration of the assayed protein

ATP-dependent 6-phosphofructokinase subunit beta (ATP-dependent 6-phosphofructokinase) (ATP-PFK) (Phosphofructokinase 2) (Phosphohexokinase) *

How was the protein concentration measured? *

optical study according to ABC



- Kinetic params
- Inhibition
- Activation



Edit Kinetic Parameter ?

Manuscript Data	
Title	ATP-dependent 6-phosphofructokinase subunit beta of yeast
Author Names	Alpha B, Gamma D and Doe J
Status	open
User	ckttner
Creation Date	Feb 22, 2016
Last Work Date	Mar 7, 2016

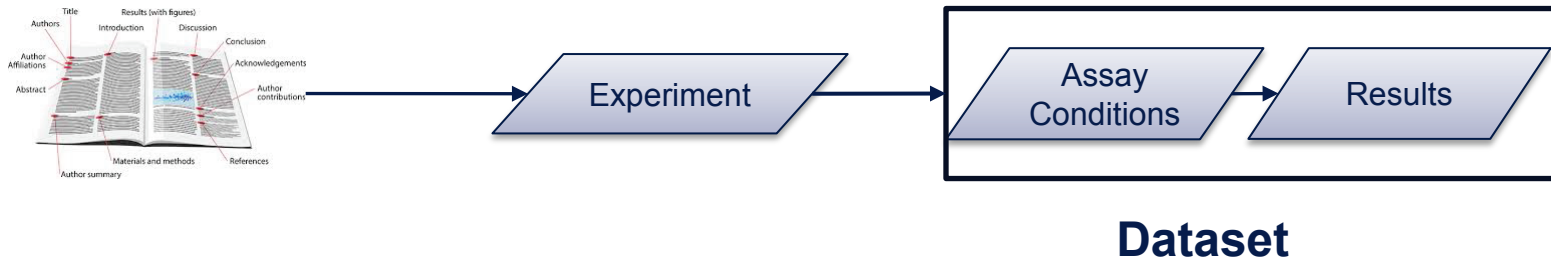
Experiment	
Experiment	
Description	Native PFK
Methodology	activity studied according to the methodology published by Duden K et al. (1995), J. Biochemistry.
Protein	
Protein Name	ATP-dependent 6-phosphofructokinase subunit beta (ATP-dependent 6-phosphofructokinase) (ATP-PFK) (Phosphofructokinase 2) (Phosphohexokinase)
UniProtKB AC	P16862
EC Number	2.7.1.11
Sequence modifications	no
PTM	unknown
Organism	Saccharomyces cerevisiae (strain ATCC 204508 / S288c) (Baker's yeast)

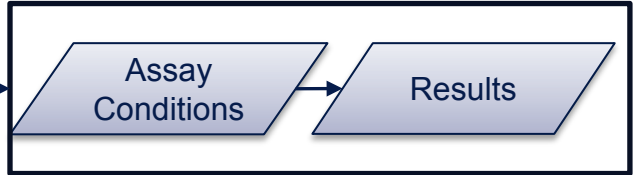
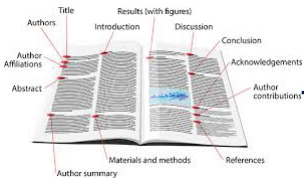
Kinetic parameters for Substrate: Fructose 6-Phosphate

Fill in only those parameters you have obtained. Please do not enter values of those you are uncertain. You need to enter at least one value.

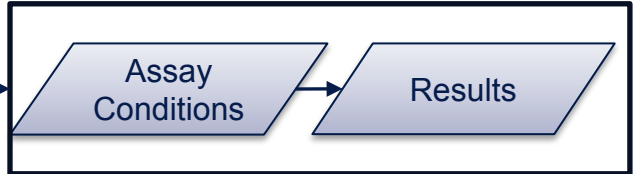
K_m	<input type="text" value="3e-7"/>	(+/-)	<input type="text" value="2.5e-8"/>	<input type="text" value="M"/>
k_{cat}	<input type="text" value="1500"/>	(+/-)	<input type="text" value="32"/>	<input type="text" value="s^-1"/>
V	<input type="text"/>	(+/-)	<input type="text"/>	<input type="text" value="mM min^-1"/>
k_{cat}/K_m	<input type="text"/>	(+/-)	<input type="text"/>	<input type="text" value="M^-1 s^-1"/>
V/K_m	<input type="text"/>	(+/-)	<input type="text"/>	<input type="text" value="s^-1"/>

Cancel Save

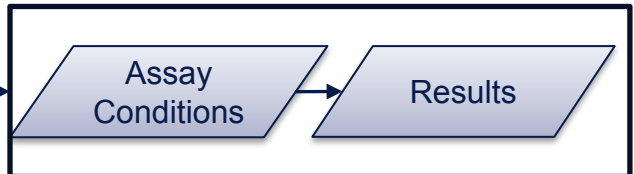




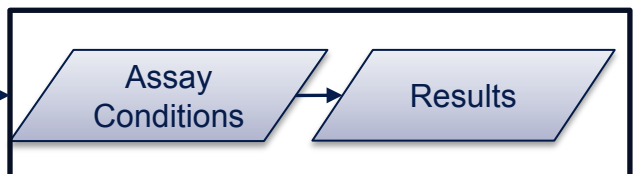
Dataset₁



Dataset₂



Dataset₃

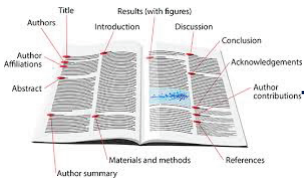


Dataset_n

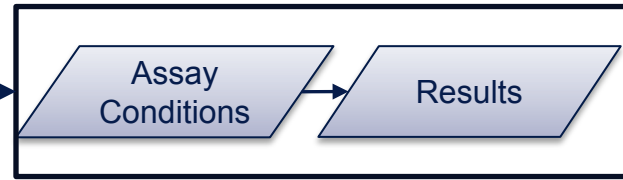
Examples

e.g. pH profile of protein X:
Exp₁: pH 5
Exp₂: pH 6
...

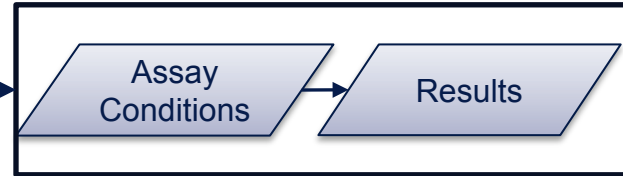
e.g. Temp. profile of protein Y:
Exp₁ @ 15 °C
Exp₂ @ 20 °C
...



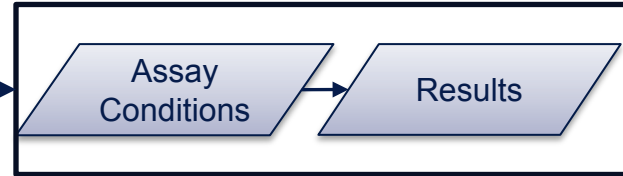
Experiment



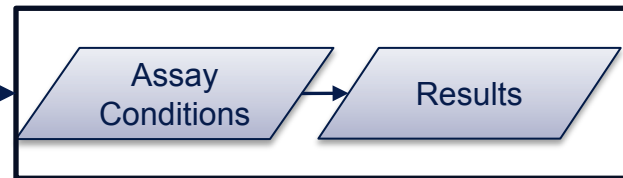
Dataset₁



Dataset₂



Dataset₃

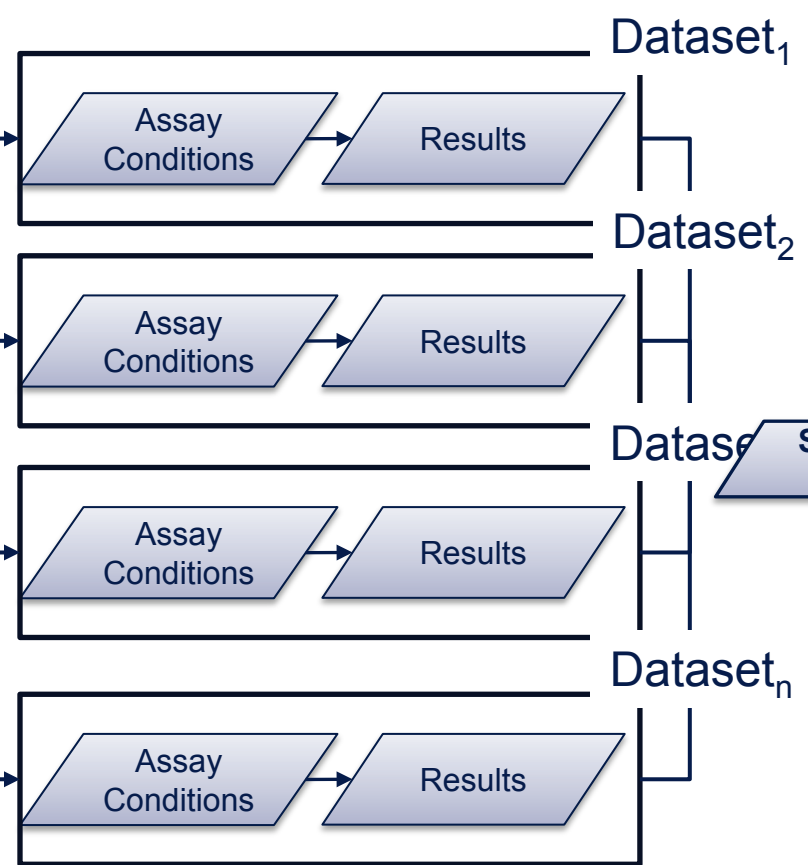
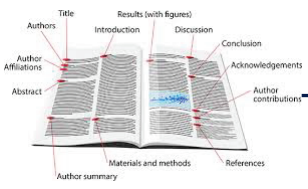


Dataset_n

Examples

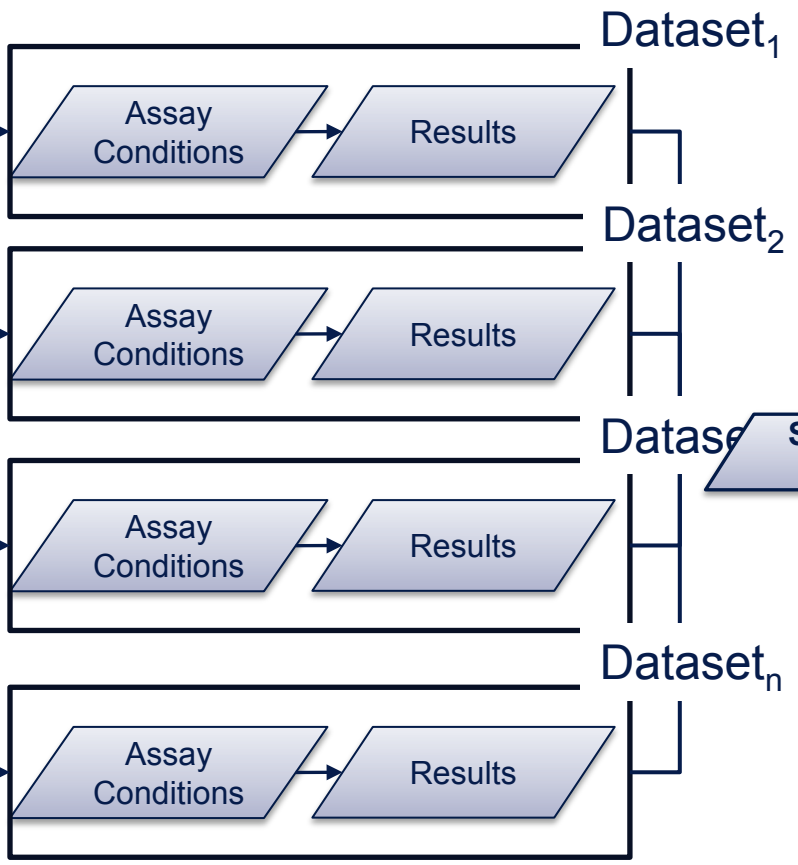
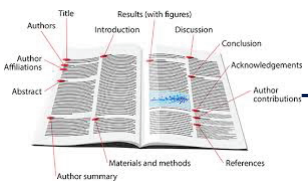
e.g. pH profile of protein X:
Exp₁: pH 5
Exp₂: pH 6
...

e.g. Temp. profile of protein Y:
Exp₁ @ 15 °C
Exp₂ @ 20 °C
...



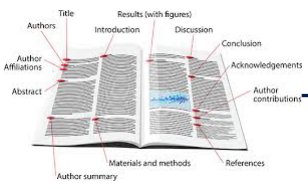
non-public



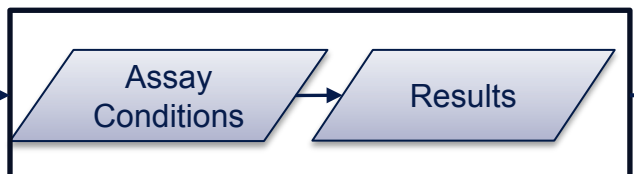
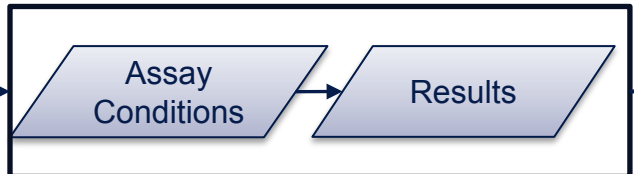
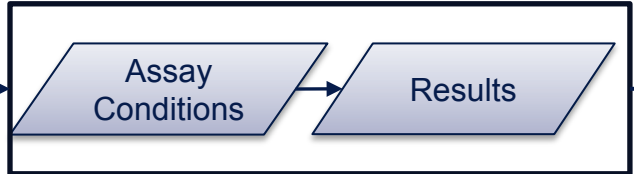
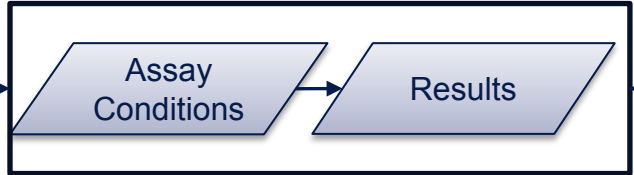


non-public

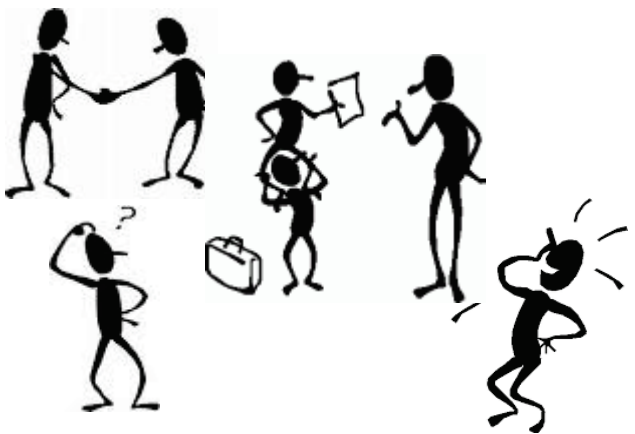




Enzyme

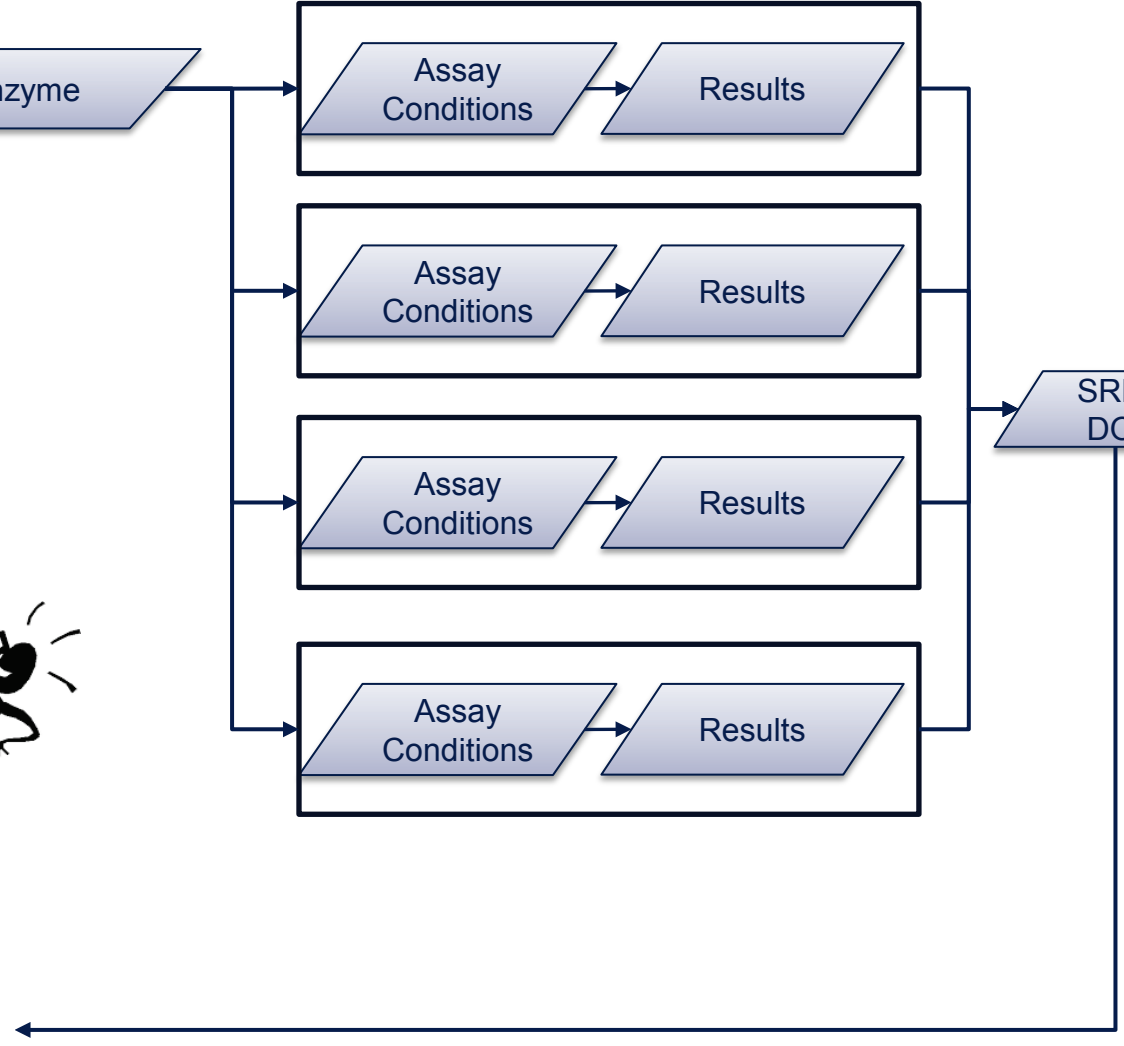


SRN/
DOI



public

PMID



STRENDA DB: Data Overview



Home Data Submission Query Guidelines References Help

Experiment Overview

Manuscript Data	
Title	Mechanistic Studies of the Flavoprotein Tryptophan 2-Monooxygenase 1. Kinetic Mechanism
Author Names	Emanuele JJ, Fitzpatrick PF
Status	published
User	fitzpatrickp
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Publication Date	Nov 16, 2016

Experiment
Experiment Title
Strenda ID
DOI
Manuscript Title
Authors
Methodology
Protein
Protein Name
UniProt
EC Num
Sequen
PTM
Organis

kinetic mechanism with phenylalanine

Assay Conditions		Results
Small Assay Components		kinetic mechanism with methionine
Name		
Dithiothreitol		
1185-53-1		
Edta disodium		
L-phenylalanine		
oxygen		
Physical Properties		
pH	pD	Temperature
8.3		25.0 °C

STRENDA DB

Experimental data fact sheet

This document provides all functional enzyme data that were obtained under the given experimental conditions, entered into STRENDA DB and assigned to the unambiguous SRN shown in the first line. This document can be submitted together with the corresponding manuscript to a journal at one's own option.

Experiment Title	Kinetic mechanism of tryptophan 2-monooxygenase with phenylalanine
Strenda ID	WZOV5O
DOI	10.22011/strenda_db.WZOV5O
Manuscript Title	Mechanistic Studies of the Flavoprotein Tryptophan 2-Monooxygenase 1. Kinetic Mechanism
Authors	Emanuele JJ, Fitzpatrick PF
Methodology	Continuous assay with oxygen electrode

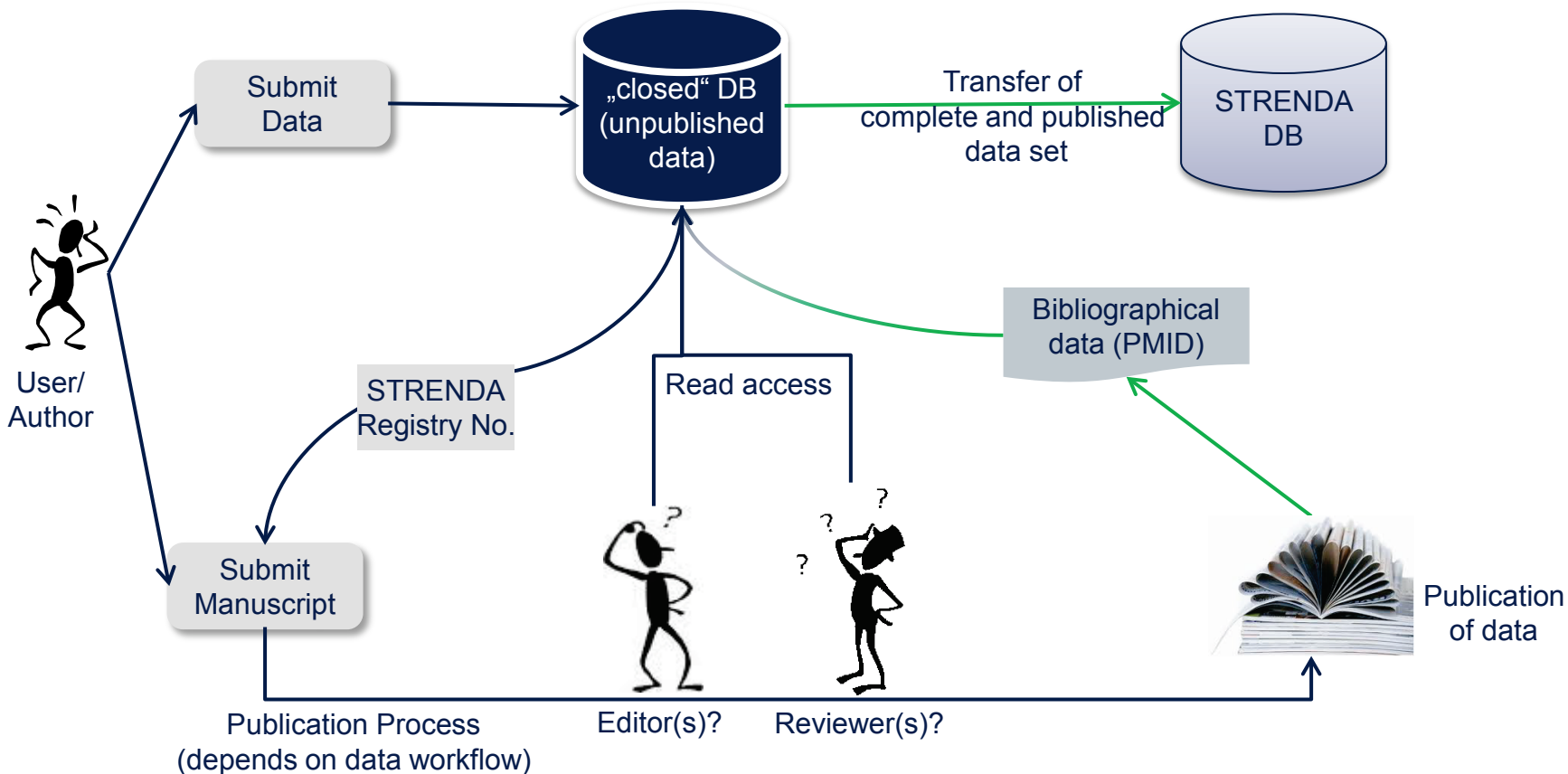
Protein

Protein Description

Is the protein data registered in UniProtKB?	yes
UniProtKB AC	P06617
Protein Name	Tryptophan 2-monooxygenase
Sequence	MYDHFNSPSIDILYDYGPFLLKCEMTGGIGSYSAGTPTPRVAIVGAGISGLVAATELLR AGVKDVLVYESRDRIGGRVWSQVPDQTRPRYIAEMGAMRPPSATGLPHYLKPKGISTG TTPDPGVVDTELHYRGKRYHWPAGKKPELFRRVYEGWQSLLEGGYLLVAPLD ITAMLKSGRLEEAAIAWQGLNVFRDCSFYNAIVCIFTGRHPPGGDRWARPEDFELFGS LGIGSGGFLPVFQAGFTEILRMVINGYQSDQRLIPDGISSLAARLADQSPDGKALDRVR

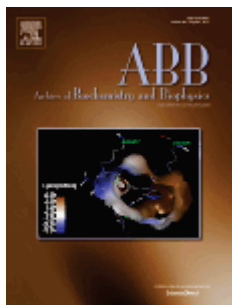
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First data coming in ...



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