

Caterpillars induce jasmonates in flowers and alter plant responses to a second attacker

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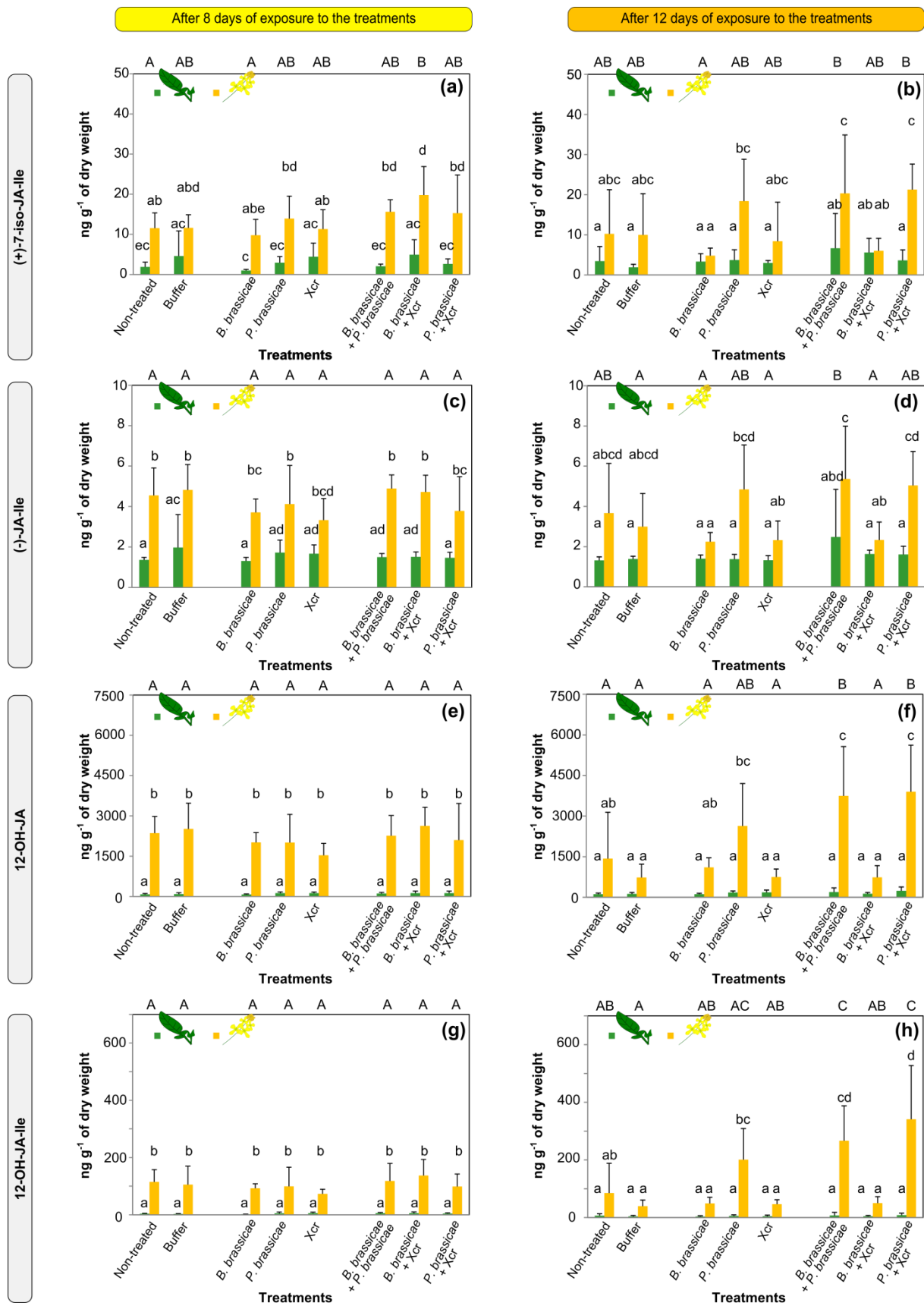
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Methods S1 Protocol for extraction and quantification of the phytohormones and their catabolites

Extraction of phytohormones was done by stirring 20 mg of ground freeze-dried plant material in 1.5 ml of methanol for 30 min, and then centrifuging it twice (at 14,000 rpm, for 10 min at 4 °C) and combining the supernatants. The final methanolic crude extract was then evaporated (speed-vac at 30 °C) and re-dissolved in 500 µl methanol. The following internal standards were added to the methanolic extract: 60 ng D₆-abscisic acid (D₆-ABA) (Santa Cruz Biotechnology, Santa Cruz, U.S.A.), 60 ng of D₆-jasmonic acid (D₆-JA) (HPC Standards GmbH, Cunnorsdorf, Germany), 60 ng D₄-salicylic acid (D₆-SA) (Sigma-Aldrich, Merck KGaA, Darmstadt, Germany), and 12 ng of JA-¹³C₆-isoleucine conjugate [JA-¹³C₆-Ile]. To obtain JA-¹³C₆-Ile, JA was conjugated to ¹³C₆-Ile (Sigma-Aldrich, Merck KGaA, Darmstadt, Germany) as described by Kramell *et al.* (Kramell *et al.*, 1988).

Resulting extracts were analysed by high performance liquid chromatography (Agilent 1200 HPLC system, Agilent technologies, Santa Clara, USA) coupled with a mass spectrometer (MS) (API 5000, Applied Biosystem, Foster city, USA) and equipped with a Turbospray ion source. Two µl of extracts was separated on a Zorbax Eclipse XDB-C18 column (50 x 4.6 mm, 1.8 µm, Agilent technologies, Santa Clara, USA). Two solvents formed the mobile phase: formic acid (0.05 %) in ultrapure water as solvent A, and acetonitrile as solvent B. The following gradient was used: 0-0.5 min, 5 % B; 0.5-9.5 min, 5-42 % B; 9.5-9.51 min, 42-100 % B; 9.51-12 min, 100 % B and 12.1-15 min, 5 % B. The flow rate was 1.1 ml min⁻¹ and the column was kept at 25 °C. In the MS, the liquid effluent was ionized by electrospray ionisation in a negative mode (-4500 eV). The turbo gas temperature was set at 700 °C. Nebulizing gas was set at 60 psi, curtain gas at 25 psi, heating gas at 60 psi, and collision gas at 7 psi. The MS was run in multiple reaction monitoring (MRM) mode at *m/z* 263.0 to 153.2 (collision energy (CE) -22 V; declustering potential (DP) -35 V) for ABA; at *m/z* 269.0 to 159.2 (CE -22 V; DP -35 V) for D₆-ABA; at *m/z* 209.1 to 59.0 (CE -24 V; DP -35 V) for JA; at *m/z* 215.1 to 59.0 (CE -24 V; DP -35 V) for D₆-JA; at *m/z* 136.9 to 93.0 (CE -22 V; DP -35 V) for SA; at *m/z* 140.9 to 97.0 (CE -22 V; DP -35 V) for D₄-SA; at *m/z* 290.9 to 165.1 (CE -24 V; DP -45 V) for cis-OPDA, at *m/z* 322.2 to 130.1 (CE -30V; DP -50V) for JA-Ile conjugate; at *m/z* 328.2 to 136.1 (CE -30V; DP -50V) for JA-¹³C₆-Ile conjugate; at *m/z* 338.2 to 130.1 (CE -

30V; DP -50V) for 12-hydroxy-jasmonoyl-isoleucine [12-OH-JA-Ile] conjugate; at m/z 352.2 to 130.1 (CE -30V; DP -50V) for 12-carboxyjasmonoyl-isoleucine [12-COOH-JA-Ile] conjugate; and at m/z 225.1 to 59.0 (CE -24V; DP -35V) for 12-hydroxy-jasmonate [12-OH-JA]. Phytohormones were quantified in ng g^{-1} of dry biomass (Analyst 1.5, Applied Biosystems, Foster city, USA) using their respective internal standards. The D_6JA was used for the quantification of *cis*-OPDA with a response factor of 0.5, and for 12-OH-JA with a response factor of 1.0. 12-OH-JA-Ile conjugate and 12-COOH-JA-Ile conjugate were quantified using $\text{JA-}^{13}\text{C}_6\text{-Ile}$ conjugate as internal standard applying a response factor of 1.0.



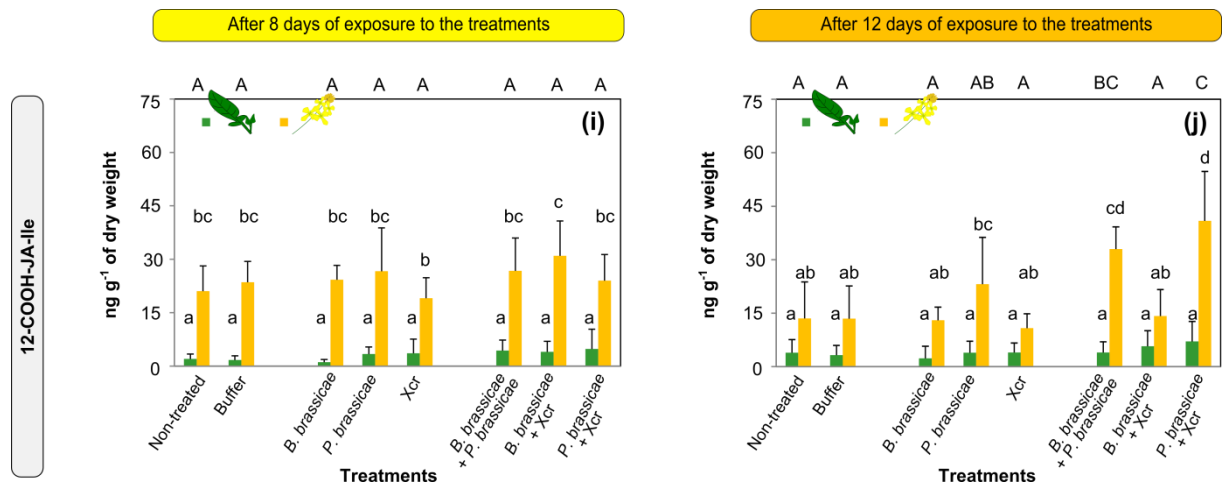


Fig. S1 Concentration of active jasmonates and their catabolites (mean + SD) quantified in leaves and inflorescences of *Brassica nigra* plants exposed to single or dual attack for 8 or 12 d. Quantities (ng g⁻¹ of plant dry weight) in leaves (green) and inflorescences (yellow) of the jasmonate-derived phytohormones: (+)-7-iso-jasmonoyl-L-isoleucine [(+)-7-iso-JA-Ile] at (a) day 8 and (b) day 12, and of the catabolic forms: 12-hydroxy-jasmonate [12-OH-JA] at (e) day 8 and (f) day 12, 12-hydroxy-jasmonoyl-isoleucine [12-OH-JA-Ile] at (g) day 8 and (h) day 12, 12-carboxy-jasmonoyl-isoleucine [12-COOH-JA-Ile] at (i) day 8 and (j) day 12, in plants that were nontreated, exposed to buffer, or exposed to single or dual attack by *Brevicoryne brassicae*, *Pieris brassicae*, and/or *Xanthomonas campestris* pv. *raphani* (Xcr). We had 6 replicates per treatment and time point. Uppercase letters indicate overall significant differences between treatments; lowercase letters indicate significant differences between each treatment for leaves and inflorescences at the 0.05 level.

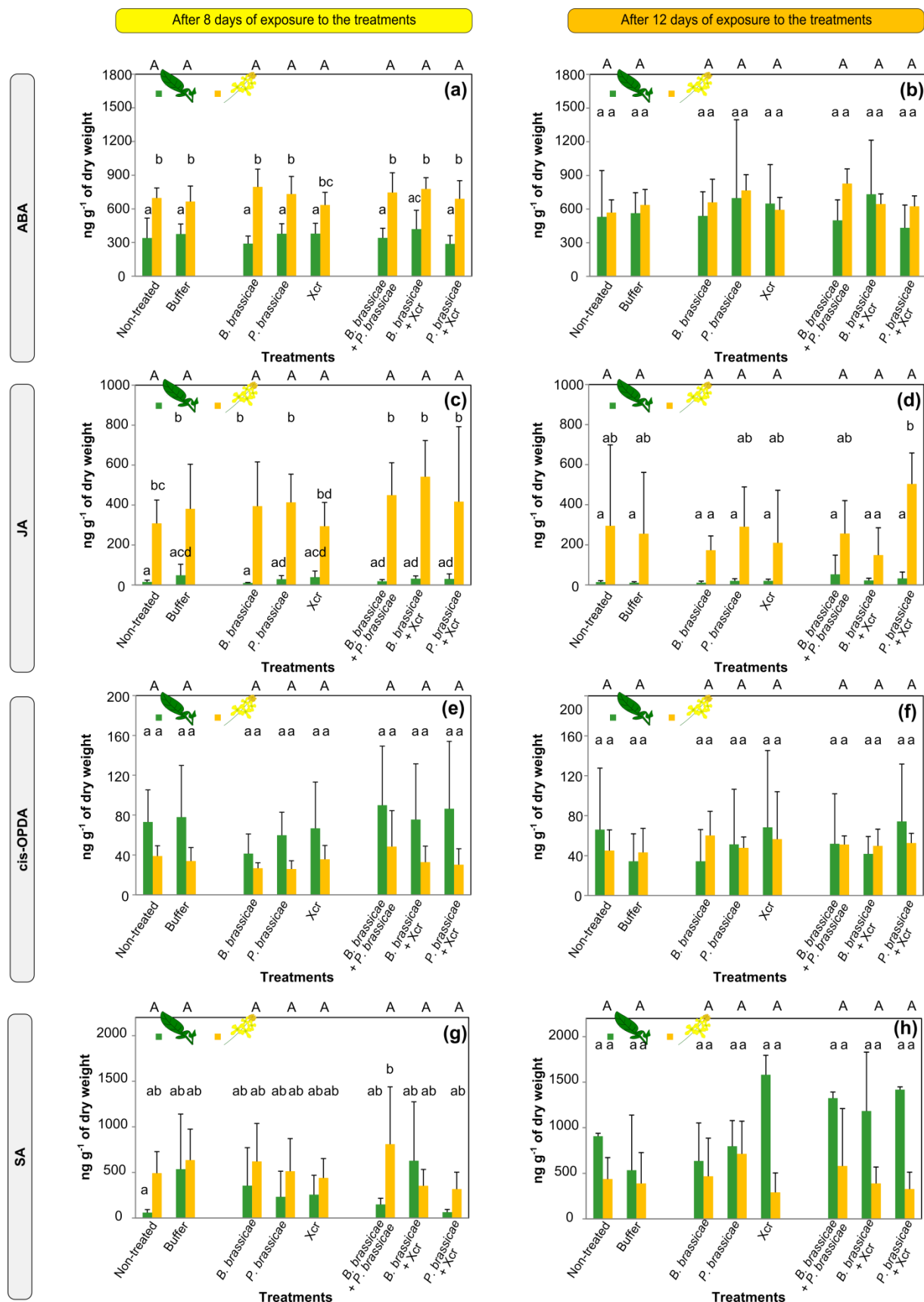


Fig. S2 Concentration of abscisic acid (ABA), jasmonic acid (JA), *cis*-(+)-12-oxophytodienoic acid (*cis*-OPDA) and salicylic acid (SA) quantified in leaves and inflorescences (mean + SD) of *Brassica nigra* plants exposed to single and dual attack for 8 and 12 d. Quantities (ng g^{-1} of dry weight) in leaves (green) and inflorescences (yellow) of the phytohormones: ABA at (a) day 8 and (b) day 12; JA at (c) day 8 and (d) day 12, *cis*-OPDA at (e) day 8 and (f) day 12; SA at (g) day 8 and (h) day 12, in plants that were nontreated, exposed to buffer, or exposed to single or dual attack by *Brevicoryne brassicae*, *Pieris brassicae*, and/or *Xanthomonas campestris* pv. *raphani* (Xcr). We had 6 replicates per treatment and time point. Uppercase letters indicate overall significant differences between treatments; lowercase letters indicate significant differences between each treatment for leaves and inflorescences at the 0.05 level.

Table S1 Output of the generalized linear model for the effects of treatment, plant part and day (duration of exposure to the treatments) on the concentration of the jasmonic acid (JA)-related phytohormones: the active forms (+)-7-iso-jasmonoyl-L-isoleucine [(+)-7-iso-JA-Ile] and (-)-jasmonoyl-L-isoleucine [(-)-JA-Ile], and of their catabolic forms 12-hydroxy-jasmonate [12-OH-JA], 12-hydroxy-jasmonoyl-isoleucine [12-OH-JA-Ile], 12-carboxy-jasmonoyl-isoleucine [12-COOH-JA-Ile]. We assessed compound concentration in leaves and inflorescences of flowering *Brassica nigra* plants that were exposed to single or dual attack for 8 or 12 d. Output of the analyses including both time points in the statistical model is shown on the left side. On the right side, the output for each of the time points is shown.

	Factors	Wald Chi-Square	df	P		Factors	Wald Chi-Square	df	P	
(+)-7-iso-JA-Ile	Overall				Day 8	Treatment	20.787	7	0.004	
	Treatment	29.113	7	< 0.001		Plant part	163.381	1	< 0.001	
	Plant part	148.560	1	< 0.001		Treatment*Plant part	11.333	7	0.125	
	Day	0.046	1	0.830						
	Treatment*Plant part	20.323	7	0.005		Day 12	Treatment	25.611	7	< 0.001
	Treatment*Day	19.421	7	0.007			Plant part	41.058	1	< 0.001
	Plant part*Day	1.699	1	0.192			Treatment*Plant part	19.592	7	0.007
Treatment*Plant part*Day	14.259	7	0.047							
(-)-JA-Ile	Overall				Day 8	Treatment	11.116	7	0.134	
	Treatment	29.34	7	< 0.001		Plant part	206.233	1	< 0.001	
	Plant part	216.308	1	< 0.001		Treatment*Plant part	8.615	7	0.281	
	Day	3.908	1	0.048	Day 12	Treatment	31.726	7	< 0.001	
	Treatment*Plant part	15.373	7	0.032		Plant part	60.949	1	< 0.001	
	Treatment*Day	20.146	7	0.005		Treatment*Plant part	17.110	7	0.017	
	Plant part*Day	4.046	1	0.044						
Treatment*Plant part*Day	13.090	7	0.070							
12-OH-JA	Overall				Day 8	Treatment	8.202	7	0.315	
	Treatment	44.288	7	< 0.001		Plant part	349.145	1	< 0.001	
	Plant part	368.557	1	< 0.001		Treatment*Plant part	8.756	7	0.271	
	Day	1.550	1	0.213	Day 12	Treatment	63.125	7	< 0.001	
	Treatment*Plant part	40.043	7	< 0.001		Plant part	110.69	1	< 0.001	
	Treatment*Day	47.403	7	< 0.001		Treatment*Plant part	56.934	7	< 0.001	
	Plant part*Day	3.110	1	0.078						
Treatment*Plant part*Day	43.510	7	< 0.001							
12-OH-JA-Ile	Overall				Day 8	Treatment	7.713	7	0.359	
	Treatment	64.975	7	< 0.001		Plant part	234.939	1	< 0.001	
	Plant part	260.717	1	< 0.001		Treatment*Plant part	7.258	7	0.403	
	Day	4.996	1	0.025	Day 12	Treatment	78.722	7	< 0.001	
	Treatment*Plant part	61.011	7	< 0.001		Plant part	104.735	1	< 0.001	
	Treatment*Day	62.301	7	< 0.001		Treatment*Plant part	74.892	7	< 0.001	
	Plant part*Day	4.071	1	0.044						
Treatment*Plant part*Day	60.039	7	< 0.001							
12-COOH-JA-Ile	Overall				Day 8	Treatment	12.679	7	0.080	
	Treatment	61.905	7	< 0.001		Plant part	359.996	1	< 0.001	
	Plant part	469.932	1	< 0.001		Treatment*Plant part	8.225	7	0.313	
	Day	3.259	1	0.071	Day 12	Treatment	74.745	7	< 0.001	
	Treatment*Plant part	34.191	7	< 0.001		Plant part	150.127	1	< 0.001	
	Treatment*Day	34.489	7	< 0.001		Treatment*Plant part	54.262	7	< 0.001	
	Plant part*Day	9.861	1	0.002						
Treatment*Plant part*Day	34.950	7	< 0.001							

Table S2 Output of the generalized linear model for the effects of treatment, plant part and day (duration of exposure to the treatments) on the concentration of the phytohormones: salicylic acid (SA), abscisic acid (ABA), jasmonic acid (JA), and *cis*-(+)-12-oxophytodienoic acid (*cis*-OPDA). We assessed compound concentration in leaves and inflorescences of flowering *Brassica nigra* plants that were exposed to single or dual attack for 8 or 12 d. Output of the analyses including both time points in the statistical model is shown on the left side. On the right side, the output for each of the time points is shown.

ABA	Overall	Factors	Wald Chi-Square	df	P	Day 8	Factors	Wald Chi-Square	df	P
		Treatment	9.813	7	0.199		Treatment	7.205	7	0.408
Plant part	59.929	1	< 0.001	Plant part	240.084	1	< 0.001			
Day	9.267	1	0.002	Treatment*Plant part	9.121	7	0.244			
Treatment*Plant part	8.787	7	0.268							
Treatment*Day	2.392	7	0.935	Day 12	Factors	Wald Chi-Square	df	P		
Plant part*Day	23.332	1	< 0.001		Treatment	5.885	7	0.553		
Treatment*Plant part*Day	3.357	7	0.850	Plant part	2.536	1	0.111			
				Treatment*Plant part	5.471	7	0.603			

JA	Overall	Factors	Wald Chi-Square	df	P	Day 8	Factors	Wald Chi-Square	df	P
		Treatment	8.948	7	0.256		Treatment	7.184	7	0.410
Plant part	218.377	1	< 0.001	Plant part	181.623	1	< 0.001			
Day	10.985	1	0.001	Treatment*Plant part	7.311	7	0.397			
Treatment*Plant part	10.470	7	0.163							
Treatment*Day	7.075	7	0.421	Day 12	Factors	Wald Chi-Square	df	P		
Plant part*Day	9.512	1	0.002		Treatment	11.687	7	0.111		
Treatment*Plant part*Day	10.687	7	0.153	Plant part	60.951	1	< 0.001			
				Treatment*Plant part	10.111	7	0.182			

cis-OPDA	Overall	Factors	Wald Chi-Square	df	P	Day 8	Factors	Wald Chi-Square	df	P
		Treatment	7.594	7	0.370		Treatment	8.761	7	0.270
Plant part	15.660	1	< 0.001	Plant part	31.468	1	< 0.001			
Day	0.033	1	0.856	Treatment*Plant part	2.903	7	0.894			
Treatment*Plant part	5.519	7	0.597							
Treatment*Day	5.322	7	0.621	Day 12	Factors	Wald Chi-Square	df	P		
Plant part*Day	12.673	1	< 0.001		Treatment	4.583	7	0.711		
Treatment*Plant part*Day	1.749	7	0.972	Plant part	0.072	1	0.789			
				Treatment*Plant part	4.230	7	0.753			

SA	Overall	Factors	Wald Chi-Square	df	P	Day 8	Factors	Wald Chi-Square	df	P
		Treatment	3.885	7	0.793		Treatment	13.316	7	0.065
Plant part	5.431	1	0.020	Plant part	12.605	1	< 0.001			
Day	19.907	1	< 0.001	Treatment*Plant part	14.090	7	0.05			
Treatment*Plant part	10.932	7	0.142							
Treatment*Day	10.344	7	0.170	Day 12	Factors	Wald Chi-Square	df	P		
Plant part*Day	29.203	1	< 0.001		Treatment	5.685	7	0.577		
Treatment*Plant part*Day	9.550	7	0.216	Plant part	18.404	1	< 0.001			
				Treatment*Plant part	9.354	7	0.228			

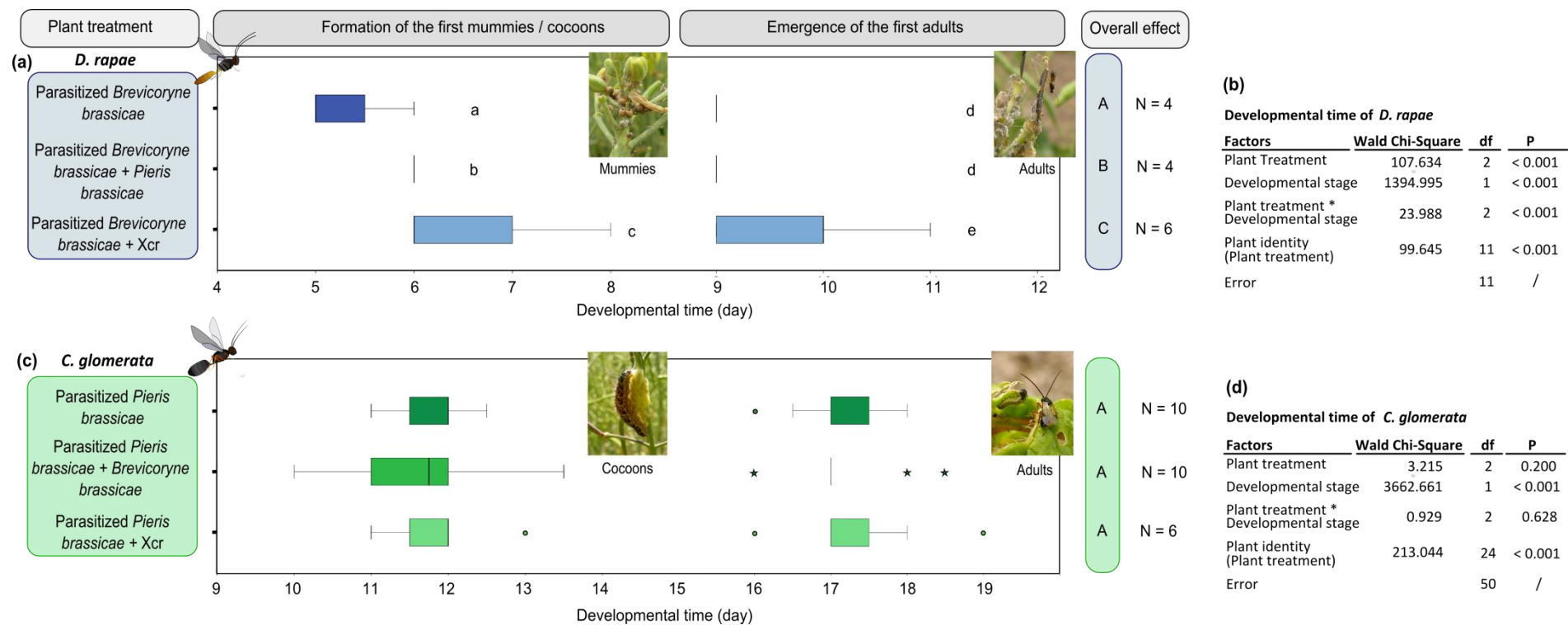


Fig. S3 Developmental time of the parasitoid *Diaeretiella rapae* and of the parasitoid *Cotesia glomerata* developing in *Brevicoryne brassicae* aphids and *Pieris brassicae* caterpillars, respectively, reared on flowering *Brassica nigra* plants exposed to single or dual attack.

(a) Developmental time of males and females *D. rapae* (median, 1st and 3rd quartiles, \pm SD) and **(c)** of males and females *C. glomerata* (median, 1st and 3rd quartiles, \pm SD) that developed in and emerged from their respective herbivorous hosts. Hosts of the parasitic wasps were reared on plants exposed to single or dual attack by *B. brassicae*, *P. brassicae*, and/or *Xanthomonas campestris* pv. *raphani* (Xcr). **(b, d)** Statistics; overall effects of the treatment were tested with a general linear model with a normal distribution using likelihood function and chi-square test. Interaction between plant treatment and developmental stage was included in the model. The Bonferroni *post-hoc* test was used for pairwise comparisons at the 0.05 significance level. Uppercase letters indicate overall significant differences between treatments, lowercase letters indicate significant differences between each treatment for males and females at the 0.05 level. N, Number of plant replicates. Outliers are represented by circles (out) and stars (far out).

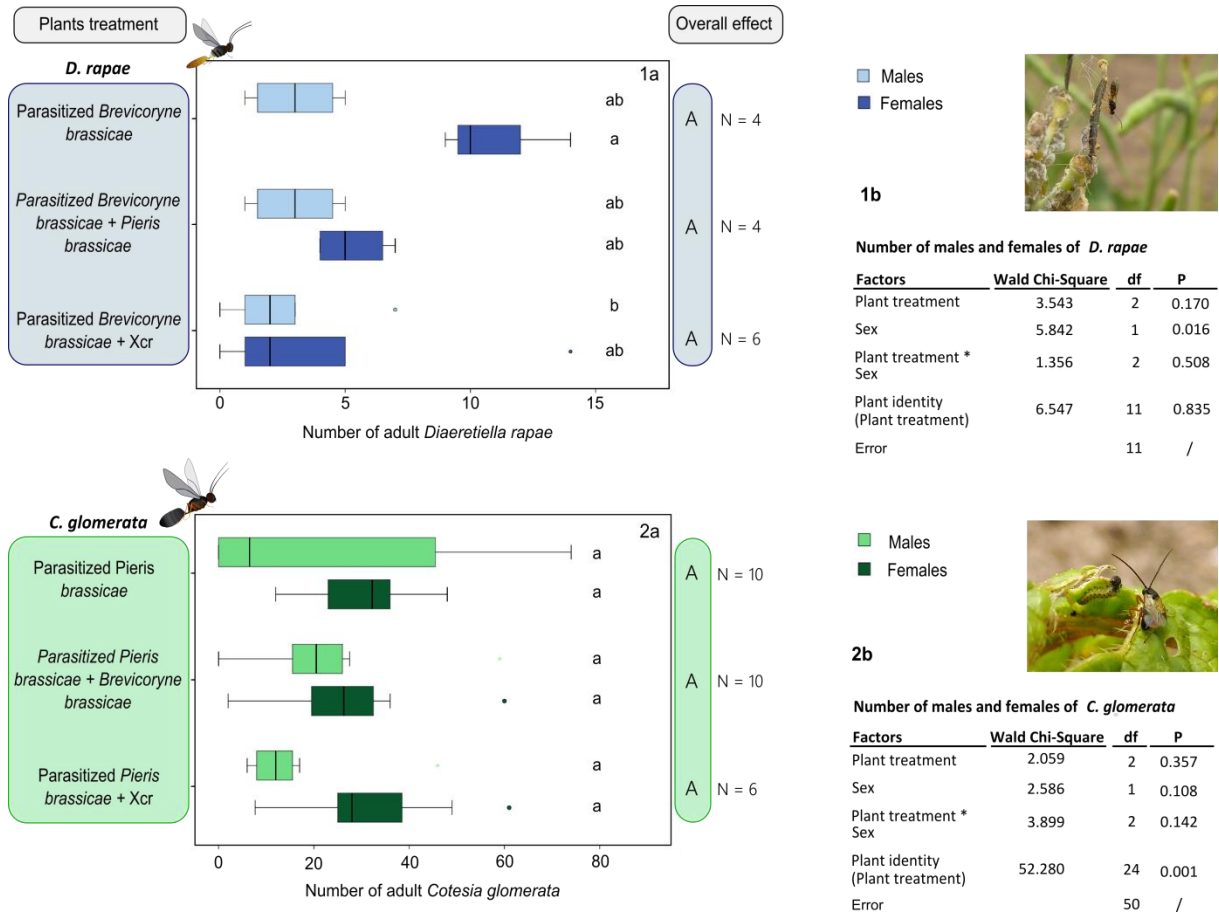


Fig. S4 Number of adults *Diaeretiella rapae* and of adults *Cotesia glomerata* that emerged from *Brevicoryne brassicae* aphids and *Pieris brassicae* caterpillars, respectively, reared on flowering *Brassica nigra* plants exposed to single or dual attack.

(a) Number of males and females *D. rapae* (median, 1st and 3rd quartiles, \pm SD) and (a) of males and females *C. glomerata* (median, 1st and 3rd quartiles, \pm SD) that emerged from their respective herbivorous hosts. Hosts of the parasitic wasps were reared on plants exposed to single or simultaneous dual attack by *B. brassicae*, *P. brassicae*, and/or *Xanthomonas campestris* pv. *raphani* (Xcr). (b, d) Statistics; overall effects of the treatment were tested with a general linear model with a poisson distribution using likelihood function and chi-square test. The Bonferroni *post-hoc* test was used for pairwise comparisons at the 0.05 significance level. Uppercase letters indicate overall significant differences between treatments, lowercase letters indicate significant differences between each treatment for males and females at the 0.05 level. N, Number of plant replicates. Outliers are represented by circles (out) and stars (far out).

References

Kramell R, Schmidt J, Schneider G, Sembdner G, Schreiber K. 1988. Synthesis of n-(jasmonoyl)amino acid conjugates. *Tetrahedron* 44(18): 5791-5807.