

# Genetic Association and Brain Morphology Studies and the Chromosome 8p22 Pericentriolar Material 1 (*PCMI*) Gene in Susceptibility to Schizophrenia

Hugh M. D. Gurling, MBBS, MD, MPhil, FRCPsych; Hugo Critchley, MB, ChB, DPhil, MRCPsych; Susmita R. Datta, BSc; Andrew McQuillin, PhD; Ekaterina Blaveri, PhD; Srinivasa Thirumalai, MD, MRCPsych; Jonathan Pimm, MPhil, MBBS, MRCPsych; Robert Krasucki, BSc, MBBS, MRCPsych; Gursharan Kalsi, PhD; Digby Quested, MD, MRCPsych; Jacob Lawrence, MBBS, BSc, MRCPsych; Nicholas Bass, MB, ChB, BSc, MRCPsych; Khalid Choudhury, BSc; Vinay Puri, BSc; Owen O'Daly, BSc; David Curtis, MBBS, MD, PhD, MRCPsych; Douglas Blackwood, PhD, MRCPsych; Walter Muir, PhD, MRCPsych; Anil K. Malhotra, MD, PhD; Robert W. Buchanan, MD; Catriona D. Good, MB, ChB, FFRad, FRCR; Richard S. J. Frackowiak, MD, DSc, FRCP, FMedSci; Raymond J. Dolan, MD, FRCPsych, FMedSci

**Context:** There is evidence of linkage to a schizophrenia susceptibility locus on chromosome 8p21-22 found by several family linkage studies.

**Objectives:** To fine map and identify a susceptibility gene for schizophrenia on chromosome 8p22 and to investigate the effect of this genetic susceptibility on an endophenotype of abnormal brain structure using magnetic resonance imaging.

**Design:** Fine mapping and identification of a chromosome 8p22 susceptibility gene was carried out by finding linkage disequilibrium between genetic markers and schizophrenia in multiply affected families, a case-control sample, and a trio sample. Variation in brain morphology associated with pericentriolar material 1 (*PCMI*) alleles was examined using voxel-based morphometry and statistical parametric mapping with magnetic resonance imaging.

**Setting and Patients:** A family sample of 13 large families multiply affected with schizophrenia, 2 schizophrenia case-control samples from the United Kingdom and Scotland, and a sample of schizophrenic trios from the United States containing parents and 1 affected child with schizophrenia.

**Main Outcome Measures:** Tests of transmission disequilibrium between *PCMI* locus polymorphisms and schizophrenia using a family sample and tests of allelic association in case-control and trio samples. Voxel-based morphometry using statistical parametric mapping.

**Results:** The family and trio samples both showed significant transmission disequilibrium between marker D85261 in the *PCMI* gene locus and schizophrenia. The case-control sample from the United Kingdom also found significant allelic association between *PCMI* gene markers and schizophrenia. Voxel-based morphometry of cases who had inherited a *PCMI* genetic susceptibility showed a significant relative reduction in the volume of orbitofrontal cortex gray matter in comparison with patients with non-*PCMI*-associated schizophrenia, who, by contrast, showed gray matter volume reduction in the temporal pole, hippocampus, and inferior temporal cortex.

**Conclusions:** The *PCMI* gene is implicated in susceptibility to schizophrenia and is associated with orbitofrontal gray matter volumetric deficits.

*Arch Gen Psychiatry.* 2006;63:844-854

Author Affiliations are listed at the end of this article.

**T**HE EVIDENCE OF A MAJOR role for genetic susceptibility in the etiology of schizophrenia has been shown by family, twin, adoption, and genetic linkage studies. Rather than the family environment, specific, unique environmental factors, which include biological environment and stochastic effects that are not shared by siblings, have been implicated.<sup>1</sup> Two types of investigation have dominated research into schizophrenia. The first type, using structural brain imaging studies, has consistently

shown volumetric abnormalities in the brains of patients with schizophrenia. The other involves the systematic mapping of chromosomal regions in which genes that increase susceptibility to schizophrenia are present. A key finding has been the presence of locus heterogeneity with different genetic susceptibilities being involved in the etiology of schizophrenia in different multiply affected families.

Genetic linkage analyses to a chromosome 8p21-22 schizophrenia susceptibility locus (SCZD6, Online Mendelian Inheritance in Man [OMIM] 603013), under

the assumption of locus heterogeneity or admixture, confirm that this region of chromosome 8 harbors a susceptibility gene for schizophrenia. Three independent investigators<sup>2-4</sup> report lod scores above 3.00 and a fourth<sup>5</sup> confirms a lod score above 2.00. Once linkage has generally implicated a region of a chromosome, it is necessary to fine map exactly which gene is involved by tests of transmission disequilibrium and allelic association with schizophrenia to detect underlying linkage disequilibrium between marker alleles and disease alleles.

In an earlier attempt to fine map a specific susceptibility gene on 8p22, we carried out tests of allelic association in a smaller case-control sample with 15 markers that flanked the *PCMI* gene without finding any evidence of association.<sup>6</sup> We then carried out tests of transmission disequilibrium of markers genotyped in our family study and followed up these results in 2 case-control samples and then in a collection of trios consisting of both parents and 1 affected offspring. Once allelic association due to linkage disequilibrium between markers and schizophrenia is detected, it is possible to differentiate genetic subgroups of schizophrenia and to examine clinical or endophenotypic characteristics of genotypically defined groups. We chose the endophenotype of abnormal brain morphology to study in relation to genotype because it seemed a priori the most likely variable to be associated with underlying heterogeneous genetic effects.

The evidence that there are volumetric deficits in the brains of subjects with schizophrenia has been replicated many times both in first-onset illness and in chronically affected subjects.<sup>7-11</sup> Consistency is generally reported in the regional pattern of cortical and subcortical morphometric abnormalities affecting gray matter density and volume.<sup>9,10</sup> The diagnosis of schizophrenia encompasses symptom clusters putatively associated with dysfunction of distinct neuroanatomical substrates.<sup>12</sup> Methodological advances now enable unbiased automated analysis of structural magnetic resonance imaging (MRI) brain scans on a voxel-wide basis to quantify differences in brain morphology and tissue composition at a level of detail that cannot be achieved by macroscopic examination.<sup>13-19</sup> In this study, we describe an application of voxel-based morphometry (VBM) to test for differences in regional brain volume in patients with 8p22 *PCMI*-associated schizophrenia. A chromosome 8 subgroup was selected on the basis of having inherited specific marker alleles at the *PCMI* gene locus and compared with patients with schizophrenia without these alleles and with healthy matched control subjects.

## METHODS

### PATIENT AND CONTROL SAMPLES

The clinical sampling, diagnosis, and genotyping of the family sample for the testing of transmission disequilibrium between alleles at the *D8S261* locus and schizophrenia has been described previously.<sup>4</sup> Briefly, it consisted of 13 large, affected pedigrees containing cases diagnosed using Research Diagnostic Criteria (RDC). Pedigrees were selected on the basis of appearing to demonstrate a single source of schizophrenia or *DSM-III-R* schizotypal illness with unilineal transmission and no cases of

bipolar affective disorder. A pedigree was considered unilineal not just on the basis of the interview diagnoses with the Schedule for Affective Disorders and Schizophrenia–Lifetime Version (SADS-L) but also with all the information available from past records. Two affection classes were used for the extended transmission disequilibrium test (ETDT) and linkage analyses: core schizophrenia consists of schizophrenia, unspecified functional psychosis, and schizoaffective psychosis; schizophrenia spectrum consists additionally of schizoid and schizotypal personality disorder according to *DSM-III-R* criteria. Of the 182 interviewed individuals, 56 fell into the core schizophrenia category and an additional 12 fell into the spectrum category.

The sampling of the UK case-control sample has recently been described in a study showing a genetic effect on schizophrenia from the region of the *epsin 4* gene on chromosome 5.<sup>20</sup> We recruited a sample of unrelated volunteers with schizophrenia and unrelated “supernormal” controls from London and South England. This type of control, in which mental illness in the control subjects and their first-degree relatives is grounds for exclusion, will maximize the chance of finding a genetic difference between a schizophrenic group and a control group. All subjects in the case-control samples and the subsamples used for MRI scanning were included only if both parents were of white English, Irish, Welsh, or Scottish descent and if 3 out of 4 grandparents were of the same descent. One grandparent was allowed to be of Caucasian European origin but not of Jewish or non-European Union ancestry based on the European Union countries before the recent enlargement. Approval was obtained from the UK National Health Service multicenter and local research ethics boards, and all subjects signed an approved consent form after reading an information sheet.

All 450 schizophrenic cases were first selected for having a clinical diagnosis of schizophrenia made by a psychiatrist recorded in National Health Service medical case notes according to the *International Classification of Diseases, 10th Revision (ICD-10)*, which is the same as the *DSM-IV* diagnosis of schizophrenia. Subjects with clear organic or symptomatic schizophreniform psychoses were excluded. All subjects were interviewed and data were collected with the SADS-L interview schedule.<sup>21</sup> Accuracy of data entries into the SADS-L was ensured by using medical case note information and all other available records. All cases were diagnosed as having schizophrenia according to the probable level of the RDC. The supernormal control subjects were also interviewed with the initial clinical screening questions of the SADS-L and selected on the basis of not having a family history of schizophrenia, alcoholism, or bipolar disorder and for having no past or present personal history of any RDC-defined mental disorder.

The US trio sample consisted of 100 unrelated patients meeting *DSM-III-R* criteria for schizophrenia or schizoaffective disorder as determined by structured diagnostic interview (Structured Clinical Interview for *DSM-III-R* [SCID]) conducted by trained raters with demonstrated interrater reliability. The research subjects were recruited from the Experimental Therapeutics branch of the National Institute of Mental Health and the Maryland Psychiatric Research Center of the University of Maryland School of Medicine, Baltimore. All subjects gave written informed consent to participate in the study under a protocol approved by the institutional review boards of the National Institute of Mental Health and the University of Maryland School of Medicine. After we obtained informed consent, the parents of each subject were also recruited for genetic testing.

The Scottish sample consisted of 200 unrelated cases of schizophrenia and 200 unrelated controls. Ethical approval was granted by the University of Edinburgh and the Royal Edinburgh Hospital, and all subjects gave informed written consent. These cases were diagnosed using the same clinical as-

essment (SADS-L) and diagnostic system (RDC) as those of the London sample. Accuracy of data entries into the SADS-L was ensured by using medical case note information and all other available records. The information on all cases was reviewed by a second psychiatrist who confirmed the RDC diagnoses. All cases were diagnosed as having schizophrenia according to the probable level of the RDC. The Scottish normal control subjects were also interviewed with the SADS-L and selected on the basis of not having a family history of schizophrenia, alcoholism, or bipolar disorder and for having no past or present personal history of any RDC-defined mental disorder. Ancestry was checked and all subjects were of white English, Irish, Scottish, or Welsh origin. The subgroups of research subjects with schizophrenia and controls employed for the MRI study were all selected only from London-based schizophrenic or control samples, and the imaging research was carried out with approval from the National Health Service multicenter research ethics committee, the National Hospital for Neurology and Neurosurgery and Institute of Neurology Joint Research Ethics Committee (University College London Hospitals National Health Service Trust), and the University College London Hospitals ethics committee.

## GENETIC METHODS

Novel CA dinucleotide repeat polymorphisms were identified by screening genomic DNA close to the marker D8S261. Primers to amplify these repeats were designed using the Primer3 software program.<sup>22</sup> The 3 (CA)<sub>n</sub> repeat markers were D8S261, D8S2615, and D8S2616. Their repeat fragment sizes, respective allele numbers, and heterozygosities are listed at <http://www.gdb.org> (the Genome Database). Genotyping methods for the single nucleotide polymorphism markers rs916550, rs412750, rs454755, rs208753, rs3780103, rs6991775, rs445422, rs412750, rs13276297, rs3214087, and rs370429 are at Ensembl, <http://www.ensembl.org>; the University of California, Santa Cruz, genome browser, <http://www.genome.ucsc.edu>; and the Single Nucleotide Polymorphism database at the National Center for Biotechnology Information, <http://www.ncbi.nlm.nih.gov/projects/SNP/>. Polymerase chain reaction conditions for amplification were as previously described.<sup>6</sup> The familial test of association was computed with the ETDT.<sup>23</sup> The CLUMP test of allelic association<sup>24</sup> was used for the case-control samples. All *P* values reported (including the ETDT test of the 100 US trios) are empirically derived using a Monte Carlo simulation approach. Subtests of the CLUMP program were the following: T1, Pearson  $\chi^2$  statistic of the raw contingency table; T2, the  $\chi^2$  statistic of a table with rare alleles grouped together to prevent small expected cell counts; T3, the largest of the  $\chi^2$  statistics of 2 × 2 tables each of which compares 1 allele against the rest grouped together; T4, the largest of the  $\chi^2$  statistics of all possible 2 × 2 tables comparing any combination of alleles against the rest. Tests of haplotype association were carried out with GENECOUNTING/LDPAIRS and confirmed using an empirical permutation test of significance.<sup>25,26</sup> The extent of linkage disequilibrium between markers was calculated with estimated haplotypes as incorporated into GENECOUNTING/LDPAIRS<sup>25,27</sup> (**Figure 1**).

## VOXEL-BASED MORPHOMETRY

There were 2 patient groups. One group with schizophrenia (SZ8) was selected on the basis that they had inherited marker alleles from any 2 markers that were associated with schizophrenia. This group comprised 14 subjects (mean ± SD age, 40.9 ± 8.4 years; 6 male and 8 female patients). A second patient group also had schizophrenia (SZ0) but had not inher-

ited any of the alleles associated with chromosome 8 schizophrenia (*n* = 13; mean ± SD age, 42.5 ± 8.4 years; 10 male and 2 female patients). All SZ8 and SZ0 schizophrenic research subjects had diagnoses of chronic or subchronic schizophrenia and at the time of scanning were all being treated with antipsychotic drugs. One of 14 SZ8 cases was recorded as being unresponsive to treatment. Two of 13 SZ0 cases were recorded as being unresponsive to treatment. All other SZ8 and SZ0 cases were partially responsive to treatment. None had good prognosis schizophrenia, and all were unemployed.

The mean ± SD age at onset for the SZ8 *PCMI*-associated group was 19.1 ± 5.7 years; the mean ± SD age at onset for the SZ0 group with non-*PCMI*-associated schizophrenia was 18.8 ± 3.1 years. Healthy control subjects, who were screened to exclude medication use and psychological, neurological, or systemic illness, were individually matched with patients with schizophrenia by age, sex, social class, and ethnicity. These research subjects were not genotyped and were not part of the normal control groups used in the case-control sample. These included 14 normal controls matched to patients with chromosome 8-associated schizophrenia (CON8) (mean ± SD age, 39.8 ± 10.0 years; 6 men and 8 women) and 13 normal controls matched to the patients with non-chromosome 8 schizophrenia (CON0) (mean ± SD age, 42.7 ± 11.7 years; 10 men and 3 women).

All subjects were scanned using an optimized 3D-MPRAGE sequence for enhanced T1 tissue contrast on a 2 Tesla Magnetom scanner (Siemens AG, Munich, Germany).<sup>16</sup> Data were analyzed using statistical parametric mapping and optimized VBM (Statistical Parametric Mapping program version 2a [SPM2a], <http://www.fil.ion.ucl.ac.uk/spm/software>) employing the framework of the general linear model.<sup>15-19</sup> Data preprocessing was carried out as follows. First, customized gray and white matter templates were created from the subject structural MRIs. This involved normalizing each structural MRI to the standard statistical parametric map T1 template, segmenting into gray and white matter and cerebrospinal fluid (CSF), and then smoothing each gray and white matter segment with an 8-mm, full width at half maximum, isotropic Gaussian kernel. Subsequently, the images were averaged to create gray and white matter templates in stereotactic space. A fully automated brain extraction procedure was then used to remove nonbrain tissue from the original structural MRIs,<sup>17,19</sup> which incorporates a segmentation step. The resulting extracted gray and white matter images were then normalized to the gray and white matter templates, respectively, thus removing any potential contribution of nonbrain voxels. Note that these templates were specific to the present study, created from the participants' own structural scans. The spatial normalization used the residual sum of squared differences as the matching criterion and included both affine transformations and a linear combination of smooth basis functions that modeled global nonlinear shape differences. We then applied the normalization parameters to the original structural images. The rationale for this iterative step was to facilitate optimal segmentation of fully normalized images.

The original segmentation, implicit in the brain extraction step, was performed on affine coregistered images (not in stereotactic space) whereas the Bayesian priors for segmentation were fully normalized (using affine and nonlinear spatial transformations) and were in stereotactic space. Segmentation of fully normalized images (in stereotactic space) is thus optimal. The spatially normalized structural images were resliced to a final voxel size of approximately 1.5 × 1.5 × 1.5 mm<sup>3</sup> and optimally segmented into gray and white matter and CSF and non-CSF partitions. Segmentation with SPM2a employs a mixture model cluster analysis (after correcting for nonuniformity in image intensity) to identify voxel intensities matching particular tissue types combined with an a priori knowledge of the spatial distribution of these tissues in normal subjects, derived from probability.

We incorporated an additional step to correct for volume changes introduced during nonlinear spatial transformations. This involved modulating the image partitions (eg, the gray and white matter segments) by the Jacobian determinants derived from the spatial normalization step. In effect, an analysis of modulated data tests for regional differences in the absolute amount (volume) of gray/white matter whereas an analysis of unmodulated data tests for regional differences in concentration of gray/white matter in the voxel.<sup>18</sup> Finally, all the normalized, segmented, modulated, and unmodulated images were smoothed with a 12-mm, full width at half maximum, isotropic Gaussian kernel. This conditions the residuals to conform more closely to the Gaussian random field model underlying the statistical process used for adjusting *P* values.<sup>18,19</sup>

Global measures of gray matter, white matter, and CSF volumes were calculated from modulated segmented images to test for group differences in overall tissue compartment volumes. Second, separate analytic design matrices were constructed to test for regional differences in gray and white matter tissue compartments between patients with schizophrenia and control subjects. In these analyses, subject data were entered as 4 groups (SZ8, SZ0, CON8, CON0). Age, sex, and total intracranial tissue volume (derived from the sum of the global measures of compartmental volumes) were entered into these analyses as confounding covariates. Thus, we tested for between-group differences that were independent of age, sex, and overall head size.

Statistical parametric maps were constructed to test for morphological differences between subjects with schizophrenia and controls. Data are reported at a significance of  $P < .05$ , corrected for multiple comparisons using the false discovery rate correction.<sup>14</sup> Note that all these regional effects survived a voxel-wise threshold significance of  $P < .001$ , uncorrected for multiple comparisons. We tested first for global differences in tissue compartment volumes. We then tested for regional differences in gray and white matter volumes between all patients with schizophrenia and controls to study the main effect of schizophrenia. Subsequently, we tested for group differences between SZ8 and SZ0 patients and controls in gray matter tissue volume, constraining the analysis to voxels identified as showing a significant main effect of schizophrenia. Where plotted (Figures 2, 3, and 4), the effect size parameter estimates are given in arbitrary units proportional to the percentage difference in volume-adjusted signal. It is the relative differences rather than the scaling of these values that is the key metric.

## RESULTS

### GENETIC ASSOCIATION

We reanalyzed the data from our previously published whole genome scan in schizophrenia families using markers on chromosome 8 that were approximately 10 centimorgans apart. The test of association (ETDT) in this family sample between alleles of the marker D8S261 and schizophrenia, but not the other chromosome 8 markers, was found to be significant after taking multiple alleles into account ( $P = .006$ , data not shown). We followed this up in the case-control sample and found that D8S261 was also significantly associated with schizophrenia (Table 1, empirical  $P = .009$ ). The linkage disequilibrium relationships between these markers computed with GENECOUNTING/LDPAIRS as in Figure 1 showed that all markers associated with schizophrenia were in significant linkage disequilibrium ( $P < .001$ ) with

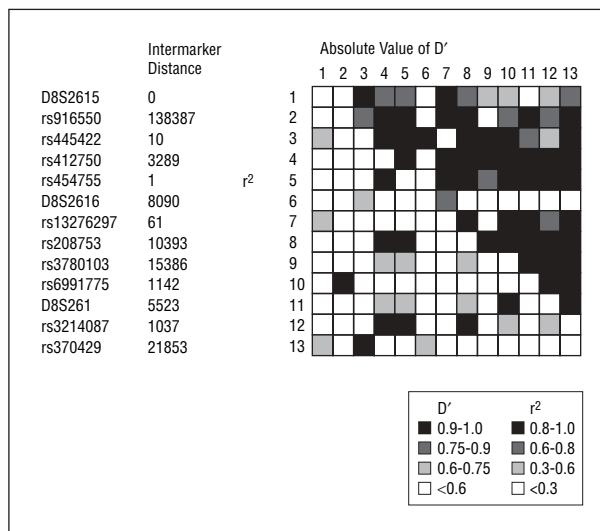
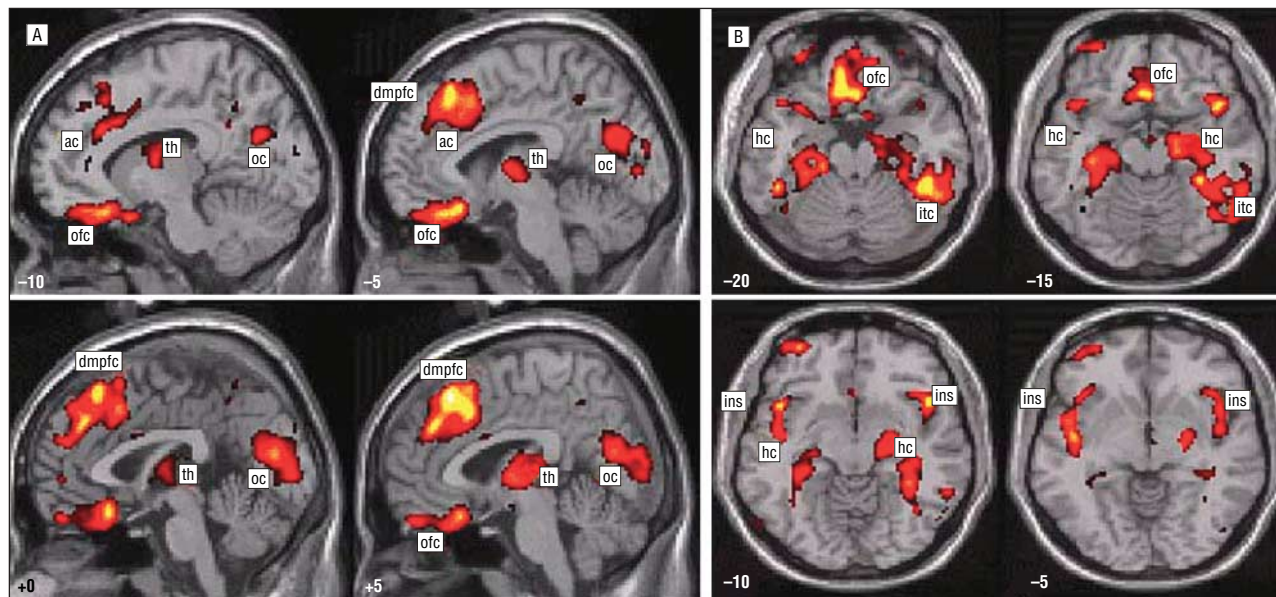


Figure 1. Tests of pairwise linkage disequilibrium between markers within the PCMI gene.

1 or more of the other associated markers (Figure 1). We then genotyped 12 other closely linked markers within the PCMI gene (Figure 5). Three of these markers also demonstrated significant allelic association. The associated markers were rs445422, rs13276297, and rs370429. The markers D8S2616 and rs3214087 both showed a trend toward association with *P* values of .07. Markers D8S2615, rs916550, rs412750, rs454755, rs208753, rs3780103, and rs6991775 did not show evidence for association with schizophrenia. The markers showing positive tests of single marker allelic association with schizophrenia are in Table 1. These markers were combined into haplotypes and these showed significant association with schizophrenia as in Table 2. All the associated markers were within or very close to the PCMI gene and no other genes in the region could be implicated (Figure 5). Empirically derived significance values for these tests of allelic and haplotypic association are in Table 1 and Table 2.

Two nonoverlapping sets of 3 markers were combined into separate haplotypes (PCM1A1/2 and PCM1B1/2) and all 4 haplotypes showed significant association with schizophrenia (Table 2). Replication of evidence for association with the 3 positive markers was attempted in 2 independent samples. The US replication sample of 100 parent offspring trios including 100 cases of schizophrenia showed that the same dinucleotide allele (142 base pairs [bp]) at the D8S261 locus was also associated with schizophrenia (empirical,  $P = .03$ ) as in Table 3. A Scottish sample of 200 cases and 200 controls failed to show any significant association between schizophrenia and 3 microsatellite markers (D8S2615, D8S261, D8S2616) near or within the PCMI locus. Other markers flanking the PCMI locus, including D8S2615, rs916550, rs412750, rs454755, rs208753, rs3780103, and rs6991775, showed no individual allelic association with schizophrenia in the London sample. However, markers rs454755, rs3780103, rs6991775, and rs6991775 all showed association with schizophrenia when combined into the haplotypes PCM1A1/2 and PCM1B1/2.



**Figure 2.** Gray matter differences associated with main effect of schizophrenia. The figure depicts the anatomical locations in red-yellow of significantly ( $P < .05$ , corrected) less gray matter volume in groups of patients with schizophrenia (SZ8 and SZ0) relative to matched controls (CON8 and CON0). Data representing group differences are plotted on sagittal (A) and axial (B) sections of a normalized template brain. Distance in millimeters lateral (x coordinate) to the anterior commissure and vertical (z coordinate) to the anterior commissure/posterior commissure line are given at the bottom left corners of these sections. Marked regional foci of gray matter differences are marked as follows: ac indicates anterior cingulate; dmpfc, dorsal medial prefrontal cortex; ofc, orbitofrontal cortex; th, thalamus; oc, occipital cortex; hc, hippocampus; ins, insula; and itc, inferior temporal lobe.

The research subjects with schizophrenia were selected for the MRI study on the basis that they had inherited alleles associated with schizophrenia at any 2 of the marker loci previously shown to be associated with schizophrenia. These were allele length 142 bp at marker D8S261, allele length 205 bp at marker D8S2616, allele T at rs445422, allele T at rs13276297, and allele A at rs370429. In analyses of structural MRI scans, patients with schizophrenia had significantly reduced total and relative gray matter volumes compared with controls, and this was apparent in both *PCMI*-associated cases (SZ8) and non-*PCMI*-associated (SZ0) patients (mean  $\pm$  SD volumes; controls, 715.8  $\pm$  69 mL; SZ8, 644  $\pm$  63 mL; SZ0, 677  $\pm$  75 mL;  $F_2 = 5.2$ ,  $P < .05$ ). Mean  $\pm$  SD percentage gray of gray + white matter volume were controls, 63.9%  $\pm$  0.02%; SZ8, 62.8%  $\pm$  0.01%; and SZ0, 63.3  $\pm$  0.02%;  $F_2 = 5.2$ ,  $P < .05$ . Patients with schizophrenia and controls did not differ in total white matter volume ( $F_2 = 1.1$ ,  $P = .25$ ) or CSF volume ( $F_2 = 3.0$ ,  $P = .097$ ). Within the schizophrenic group, we observed no significant difference between SZ8 and SZ0 patient groups in these measures of global tissue volumes.

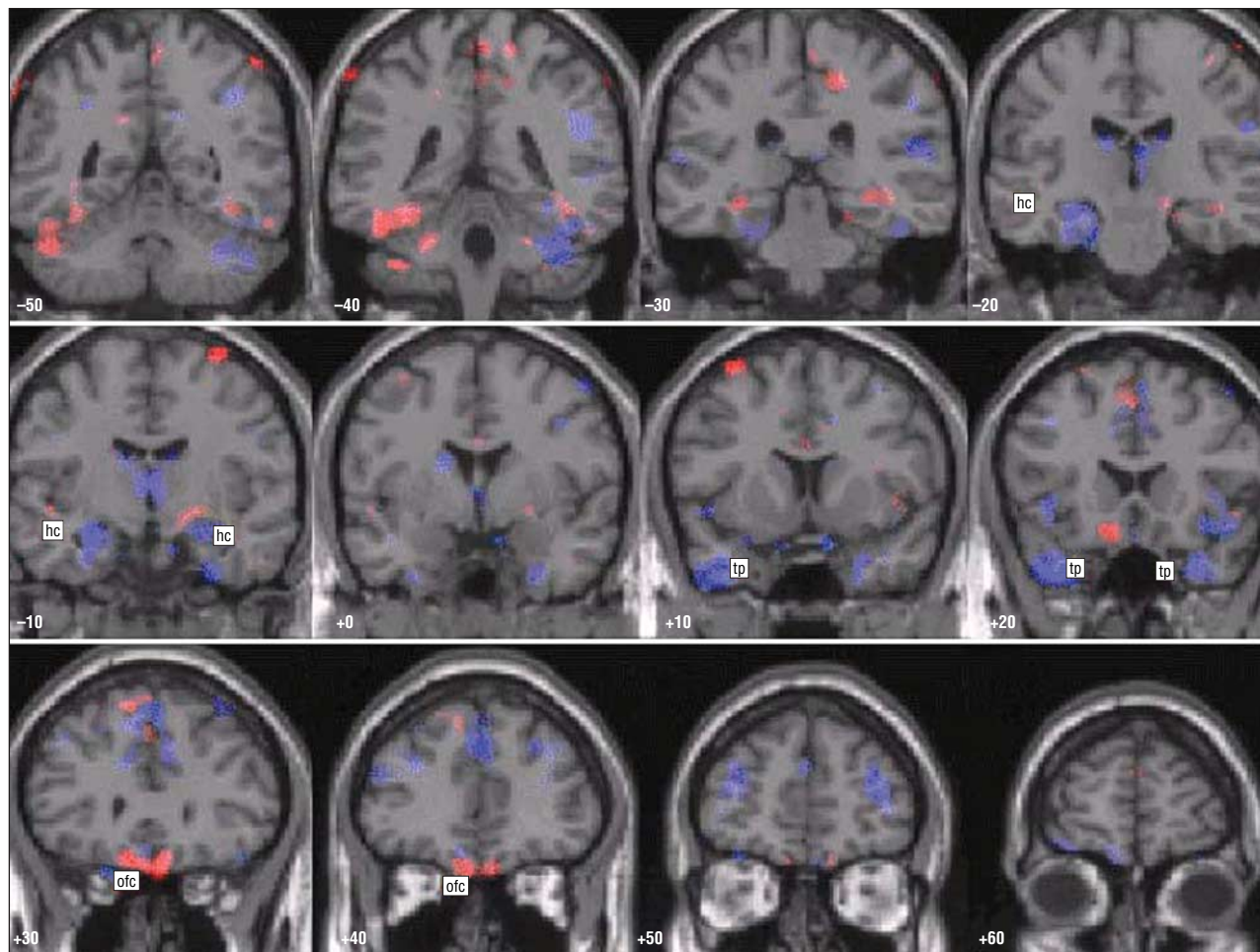
### BRAIN MORPHOLOGY

We examined for group differences in regional brain morphology in separate analyses of gray matter controlling for total intracranial volume, age, and sex. Compared with controls, patients with schizophrenia (combined SZ8 and SZ0 groups) showed a distributed pattern of gray matter volume deficit, encompassing right dorsolateral prefrontal cortex, bilateral dorsomedial and orbitomedial prefrontal cortices, insulae, temporal poles, medial thalamus, posterior hippocampi, inferior temporal (fusiform) cortices, occipital (striate) cortex, and lateral cerebellar hemi-

spheres (Figure 2). Descriptive analysis of trends identified only 1 region in patients with schizophrenia showing a trend in regional gray matter volume augmentation relative to controls. This cluster was located within the primary visual cortex ( $-3$ ,  $-59$ ,  $3$ ;  $P < .001$ , uncorrected,  $Z = 3.69$ ).

We identified differences between SZ8 and SZ0 patients in the relative distribution of gray matter within those brain areas showing a main effect of diagnosis. Patients in the SZ8 group compared with matched controls had relatively lower gray matter volume, shown in red in Figure 3, located maximally in the bilateral orbitofrontal cortices ( $P < .05$ , corrected). In contrast, SZ0 patients had significantly less gray matter, shown in blue in Figure 4, compared with matched controls in temporal poles, particularly on the left, medial temporal lobe (anterior hippocampus), right inferior temporal lobe, medial thalamus, right cerebellum, and dorsomedial prefrontal cortex ( $P < .05$ , corrected for main effect of schizophrenia) (Figure 3).

We formally tested for morphometric differences between SZ8 and SZ0 in the pattern of regional gray matter decreases in an interaction analysis in which differences between SZ8 and matched controls were compared with differences between SZ0 and matched controls. The search volume for this analysis, as shown in Figure 4, was constrained using a mask derived from the main effect of diagnosis. Grey matter volume was significantly lower in SZ8 compared with SZ0 patients within medial orbitofrontal cortex bilaterally (x, y, z coordinates [millimeters] from anterior commissure;  $-7.5$ ,  $34.5$ ,  $-22.5$  and  $6$ ,  $34$ ,  $-28$ ;  $P < .05$ , corrected). Patients in the SZ0 group showed a significant paucity of gray matter compared with SZ8 patients in the left hippocampus ( $-27$ ,  $-12$ ,  $-1$ ), the bilateral temporal poles ( $-37$ ,  $21$ ,  $-42$  and  $39$ ,  $19.5$ ,  $-34.5$ ),



**Figure 3.** Group differences in regional gray matter. Regional gray matter differences between the group of patients with alleles associated with chromosome 8 schizophrenia (SZ8) and their matched controls (CON8) and between the group of patients with schizophrenia but none of the alleles associated with chromosome 8 schizophrenia (SZ0) and their matched controls (CON0). Separate T contrasts were used to test for differences between SZ8 and CON8 and between SZ0 and CON0. These analyses were constrained using masking to regions showing a significant main effect of schizophrenia. Group data are plotted on axial sections of a normalized template brain scan. Regions showing significant differences ( $P < .05$ , corrected) between SZ8 and CON8 subjects are depicted in red, and those between SZ0 and CON0 subjects are shown in blue. The axial sections highlight the different patterns of regional gray matter volume deficit associated with these 2 schizophrenic groups; the orbitofrontal deficits in SZ8 patients and the temporal polar and hippocampal deficit in the SZ0 patients are labeled. The distance anterior in millimeters (y coordinate) from the anterior commissure is given in the bottom left corner of each section. ofc indicates orbitofrontal cortex; tp, temporal pole; and hc, hippocampus.

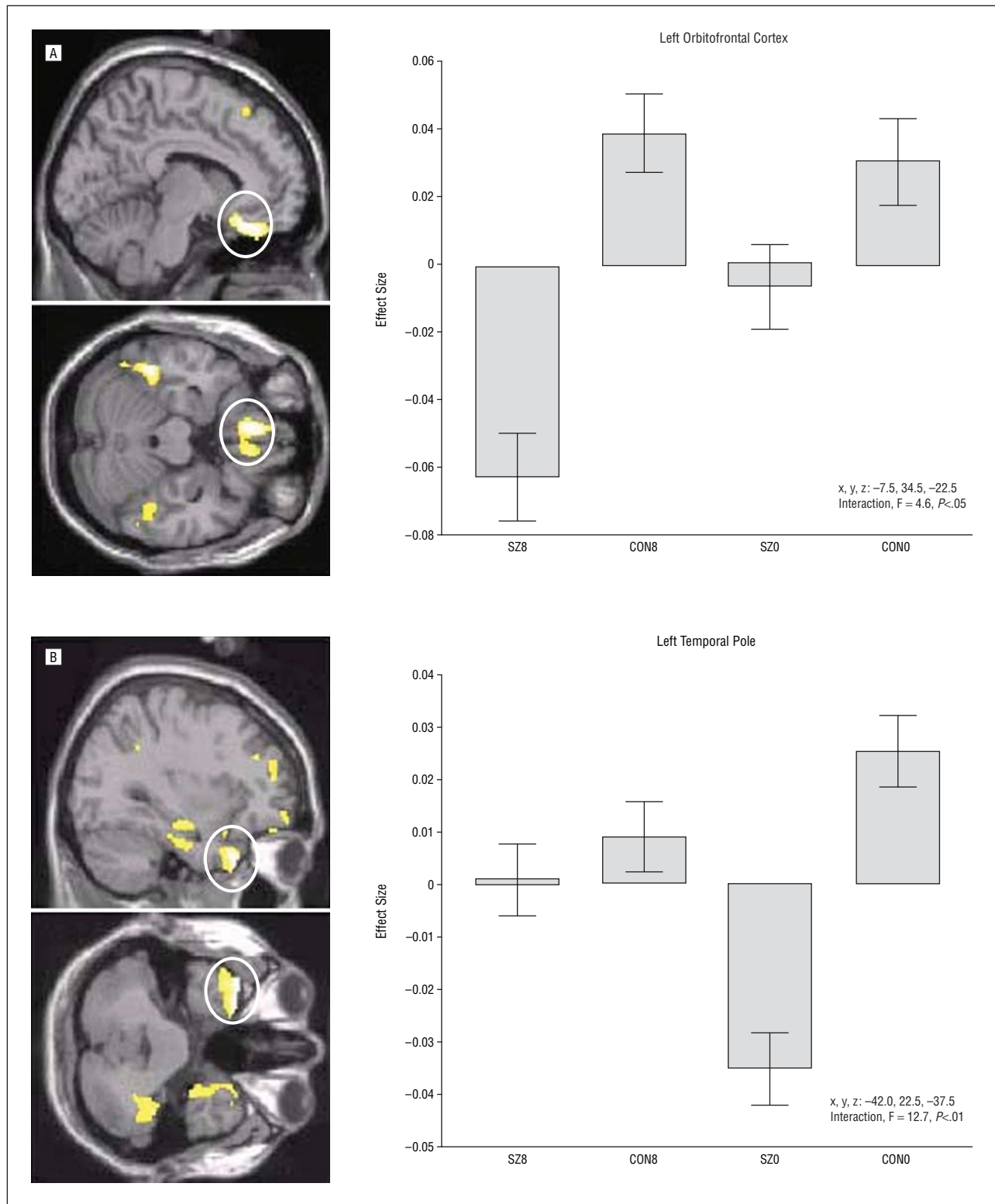
and the right dorsolateral prefrontal cortex (30, 46.5, 19.5) ( $P < .05$ , corrected) (Figure 1).

#### COMMENT

The interpretation of the level of statistical significance in the allelic association studies should consider both multiple alleles and multiple markers. In the study of brain morphology, the number of statistical tests carried out should be considered. In the single marker tests of allelic association, we used empirical tests of significance, which do not need further correction for multiple alleles. For the global tests of haplotypic association with schizophrenia, further correction for both multiple alleles and multiple markers are not required because the permutation test significance takes both into account. The finding of extended transmission disequilibrium between D8S261 with schizophrenia in the family linkage sample was the reason to select this and other markers within the *PCMI* gene for genotyping in the case-control sample. In addition,

the marker D8S261 had been independently studied and implicated in schizophrenia in the National Institute of Mental Health trio sample and not as a result of the positive ETDT in the UK family sample or positive association in the UK case-control sample. The other markers showing association in the UK case-control sample were selected for being in linkage disequilibrium with each other. Taking into account that D8S261 had already been implicated in schizophrenia in prior studies and the fact that the additional markers genotyped were not independent tests of association, it can be argued that further corrections of significance values for testing association with multiple markers in the UK case-control sample are not needed.

Our results demonstrate that a chromosomal locus showing replicated evidence of linkage with schizophrenia in several family studies can be identified by observing linkage disequilibrium between markers and schizophrenia in a case-control association sample in which the cases have not been selected for a positive family history of schizophre-



**Figure 4.** Between-group gray matter differences in orbitofrontal cortex and temporal pole. A, Orbitofrontal gray matter volume deficit in the group of patients with *PCM1* alleles associated with chromosome 8 schizophrenia (SZ8), showing involvement of the olfactory gyrus. Adjacent and to the right are plotted the effect sizes of the different parameter estimates. This is given in arbitrary units proportional to the percentage difference in volume-adjusted signal. It is the relative differences rather than the scaling of these values that is the key metric. A, There was relative reduction in orbitofrontal gray matter (at location  $x, y, z: -7.5, 34.5, -22.5$ ) in both the SZ8 group and the group of patients with schizophrenia with none of the chromosome 8 *PCM1* alleles (SZ0). However, the difference was marked for the SZ8 patients, reaching significance for an interaction, (CON8–SZ8)–(CON0–SZ0). B, Temporal pole gray matter volume deficit in SZ0 patients. Locations of gray matter differences between SZ0 patients and other subjects are illustrated on sagittal and axial sections of a template brain. Adjacent is a plot of parameter estimates for the peak of these differences at the left temporal pole. The greater difference in temporal lobe gray matter volume existed between SZ0 and CON0 subjects. The interaction, (CON0–SZ8)–(CON8–SZ8), was significant. CON8 indicates research subjects who are normal controls matched to the SZ8 cases; CON0, research subjects who are normal controls matched to the SZ0 cases.

**Table 1. Single Marker Allelic Associations With Schizophrenia in the London-Based Case-Control Sample**

Marker	Distance to Next Marker, bp	Variant Bases or Marker Fragment Sizes With Allele Frequencies Below													$\chi^2$	CLUMP Empirical P Value		
		C	T															
rs445422	0	C	T														5.76	.02
Controls		868	12															
Cases		758	24															
D8S2616	11 380	213	211	209	207	205	203	201	199	197	195	193	191	189			10.09	.07*
Controls		1	5	28	57	95	539	42	101	4	5	0	1	0				
Cases		0	12	26	46	120	457	49	101	5	5	0	0	1				
rs13276297	61	C	T														5.50	.02
Controls		840	16															
Cases		710	28															
D8S261	32 534	128	130	132	134	136	138	140	142	144	146	148					10.47	.009†
Controls		39	266	8	51	308	108	78	10	4	1	13						
Cases		38	206	9	54	297	107	55	28	3	1	12						
rs3214087	1037	Ins	Del														3.24	.07
Controls		202	656															
Cases		180	720															
rs370429	21 853	A	G														6.38	.01
Controls		12	856															
Cases		25	753															

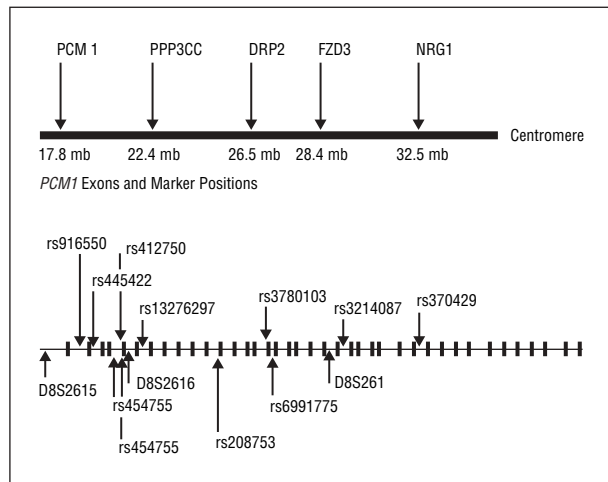
Abbreviation: bp, base pair.

\*Denotes most likely alleles to be associated with schizophrenia in each haplotype.

†Permutation test empirical P value.

nia. The fact that we were able to detect linkage disequilibrium in a sample of 450 cases of schizophrenia and find evidence in support of this in an independent sample of 100 US trio cases suggests that the chromosome 8p22 schizophrenia susceptibility alleles may be present in most European populations. As with other reported genetic associations with schizophrenia, it is clear that only a small fraction of research subjects with schizophrenia have a genetic susceptibility from the *PCMI* gene locus. This confirms the general findings of linkage studies, which are explicable only by assuming the presence of extensive locus heterogeneity for genes involved in schizophrenia. We attribute the failure to find association in the Scottish sample as being due to a different admixture of genetic subtypes in Scotland or possibly because only a limited set of 3 markers could be genotyped in this sample. The UK case-control sample employed for this study has previously been shown to exhibit allelic association to the *epsin 4* gene on chromosome 5, which encodes enthoprotin, a clathrin-associated protein involved in vesicle endocytosis in the brain and elsewhere.<sup>20</sup>

The gene *PCMI* is involved in the maintenance of centrosome integrity and the regulation of the microtubule cytoskeleton. Its protein structure bears similarities to the structural myosin proteins, which are microtubule-associated proteins involved in axon guidance, synaptogenesis, functioning of the synapse, and intracellular transport along axons and dendrites.<sup>28</sup> It is of note that a different gene disrupted in schizophrenia (*DISC1*), known to straddle a translocation breakpoint on chromosome 1 that cosegregates with schizophrenia and other types of psychiatric disorder in a single large Scottish pedigree,<sup>29</sup> also has similarities to structural proteins such as myosins. It seems likely that *PCMI* has a role in the development of the nervous system and neuronal activity. For example, it is known that *PCMI* interacts with the brain-specific protein huntingtin-



**Figure 5.** Map of putative chromosome 8p21-22 schizophrenia susceptibility loci on 8p and diagram of the genomic structure of *PCMI* showing the relative positions of exons, introns, and genetic markers. Distances are shown in megabases of DNA. *PCMI* indicates pericentriolar material 1; *PPP3CC*, calcineurin; *DRP2*, dihydropyrimidinase-related protein 2; *FZD3*, frizzled; and *NRG1*, neuregulin.

associated protein 1 (*HAPI*) that binds variably to huntingtin in relation to the number of glutamine repeat lengths present. *PCMI* also interacts with cytoskeletal, vesicular, and motor proteins to mediate interactions among these different molecules.<sup>28</sup>

Our neuroimaging findings further validate the genetic data in so far as we demonstrate distinct differences in regional gray matter abnormalities in patients with chromosome 8p22 *PCMI*-associated schizophrenia (SZ8) compared with non-chromosome 8 *PCMI*-associated cases (SZ0). The significance levels reported were corrected for multiple tests and are therefore con-



**Table 2. Haplotypic Associations With Schizophrenia**

Haplotype Label	No. of Marker Loci	Alleles Increased in Haplotypes Showing Association With Schizophrenia	Frequency in Cases, %	Frequency in Controls, %	Global Log Likelihood Ratio $\chi^2$	Permutation Significance
PCM1A1	3 Marker	rs445422 allele C, 87366_66 allele T, rs370429 allele G*	29	17	14.66	.003
PCM1A2		rs445422 allele T, 87366_66 allele C, rs370429 allele A*	23	12		
PCM1B1	3 Marker	rs454755 allele A, rs3780103 allele G, rs6991775 allele A*	40	21	18.28	.002
PCM1B2		rs454755 allele G, rs3780103 allele A, rs6991775 allele A*	7	0		
PCM1C1	4 Marker	rs454755 allele A, 87366_66 allele C, rs3780103 allele G, rs6991775 allele A*	1	0	28.72	.002
PCM1C2		rs454755 allele G, 87366_66 allele C, rs3780103 allele A, rs6991775 allele A*	5	2		

\*Denotes most likely alleles to be associated with schizophrenia in each haplotype.

**Table 3. Transmission of Individual Alleles at Marker D8S261 and TDT Analysis in the US Trio Sample\***

Transmission	Dinucleotide Repeat Lengths and Allele Counts									
Allele size, bp	130	132	134	136	138	140	142	144	146	148
Allele counts transmitted	6	3	27	0	8	33	25	12	6	1
Allele counts not transmitted	4	11	38	3	3	34	10	11	6	1

Abbreviations: bp, base pairs; TDT, transmission disequilibrium test.

\* $\chi^2_3$  for allele-wise TDT = 18.88,  $P = .03$ .  $\chi^2_{25}$  for genotype-wise TDT = 43.21,  $P = .01$ .

servative. A larger sample size, which would reduce the standard error and increase confidence in the results, would require another 450 cases of schizophrenia to be sampled and genotyped at the *PCMI* locus to test for association. Therefore, replication in an independent sample is desirable.

Our evidence suggests that brain regions showing the greatest differences between the schizophrenic groups are subsumed within a wider matrix of regional brain abnormalities that span other genetic susceptibilities to schizophrenia. Nevertheless, the peak locations of reduced gray matter volume in SZ8 and SZ0 groups predict important patterns of neuropsychological deficits or vulnerability that can perhaps inform etiopathological mechanisms. Selective gray matter deficit in orbitofrontal cortex, observed in the SZ8 group, is likely to preferentially compromise neural mechanisms supporting reward-related processing and motivational behaviors. In such cases, symptomatology of the patient may be biased toward affective and behavioral features. In contrast, patients with non-chromosome 8 *PCMI*-associated schizophrenia may exhibit more mnemonic and perceptual features.<sup>12</sup> Relevant to this is the recent research showing that families with evidence for linkage to 8p21-22 had significantly more affective deterioration, poorer outcome, more thought disorder, and fewer depressive symptoms than affected individuals from non-8p21-22-linked families.<sup>30</sup>

There is evidence of a second schizophrenia susceptibility locus called neuregulin 1 (*NRG1*) on chromosome 8p21-22<sup>31</sup> and 2 replications were successful.<sup>32-34</sup> This locus is about 20 million bases toward the centromere from the *PCMI* 8p22 locus (Figure 5). Gerber et

al<sup>35</sup> also found evidence of allelic association with schizophrenia on chromosome 8p21 at or near the calcineurin (*PPP3CC*) gene locus. Two Japanese research groups now report<sup>36,37</sup> association with schizophrenia at the frizzled (*FZD3*) gene locus suggesting that there may be 4 loci increasing susceptibility on the short arm of chromosome 8. This is compatible with the multiple lod score peaks along the short arm of chromosome 8 in our previous genome linkage scan of schizophrenia.<sup>4</sup> In the case-control sample in which we have implicated the *PCMI* gene, we have also found weak but significant evidence for association between schizophrenia and the *NRG1* gene (Susmita Datta, PhD, unpublished data, January 2006) but no evidence for association between markers at the *PPP3CC* or *FZD3* genes and schizophrenia (Susmita Datta, PhD, unpublished data, January 2006). In addition to the *epsin 4* gene association, 2 further allelic associations with markers at the *FXVD6* (phosphohippolin) gene on chromosome 11q23.3 (Khalid Choudhury, BSc, unpublished data, January 2006) and the *KIST* serine threonine kinase (*UHMK1*) gene on chromosome 1q23.3<sup>38</sup> have been found in the same sample. All of these associations have been found by following up localizations originally made in our family linkage analyses.

Further collections of genotyped patients are now needed to confirm the *PCMI* 8p22 allelic association with schizophrenia and to replicate the finding of brain morphology changes associated with *PCMI*-associated marker alleles. Modest but statistically significant levels for *PCMI* marker association with schizophrenia have been found in this study. This is compatible with the fact that linkage studies have already proven heterogene-

ity of linkage in schizophrenia. The linkage studies can only be explicable if one assumes that no single schizophrenia susceptibility gene has an effect in more than 5% to 10% of families. By inference, this would also apply to case-control samples as well. High levels of significance really would not be expected from a case-control sample if only 5% to 10% of cases shared a common genetic susceptibility.

Because some of the single nucleotide polymorphism marker alleles associated with schizophrenia are at both low and high frequencies, sufficient numbers of cases and controls are needed to obtain a good chance of replication. Assuming complete linkage disequilibrium between markers and disease alleles for a significance of  $P < .05$  and a power of 80% of replicating the result (ie, 1 in 4) with a low-frequency allele present in 5% of controls and 10% of cases, then 600 cases and 600 controls should be enough. For a high-frequency disease-associated allele present in 50% of cases and in 45% of controls, then 1300 cases and 1300 controls are needed. We can now sequence DNA from cases who have inherited *PCMI* gene marker alleles associated with schizophrenia to define the precise etiological base pair changes or mutations increasing susceptibility to schizophrenia. If these can be found, then exactly how *PCMI* shapes regional brain structure and function can be understood. The alternative splicing of *PCMI* messenger RNA, interaction of *PCMI* with other proteins, posttranslational modification, phosphorylation, and glycosylation may all play a role in mediating genetic effects on the disease.

**Submitted for Publication:** May 26, 2005; final revision received October 14, 2005; accepted December 1, 2005. **Author Affiliations:** Molecular Psychiatry Laboratory, Department of Mental Health Sciences, University College London Medical School, Windeyer Institute of Medical Sciences, London, United Kingdom (Drs Gurling, Datta, McQuillin, Blaveri, Pimm, Krasucki, Kalsi, Lawrence, Bass, Choudhury, Puri, and O'Daly); Functional Imaging Laboratory, Wellcome Department of Cognitive Neurology, Institute of Neurology, University College London (Drs Buchanan, Critchley, Good, Frackowiak, and Dolan); Berkshire Healthcare National Health Service Trust, Reading, United Kingdom (Dr Thirumalai); Camden and Islington Mental Health and Social Care Trust, St Pancras Hospital, London; West London Mental Health Trust, Hammersmith and Fulham Mental Health Unit, St Bernard's Hospital, London (Dr Quested); Queen Mary College, University of London and East London and City Mental Health Trust, Royal London Hospital, Whitechapel, London (Dr Curtis); Department of Psychiatry, University of Edinburgh, Royal Edinburgh Hospital, Morningside Park, Scotland, United Kingdom (Drs Blackwood and Muir); Experimental Therapeutics Branch of the National Institute of Mental Health, Bethesda, Md (Dr Malhotra); Maryland Psychiatric Research Center of the University of Maryland School of Medicine, Baltimore (Dr Buchanan); Departement des Etudes Cognitives, Ecole Normale Supérieure, Paris, France (Dr Frackowiak). Dr Quested is currently with the Department of Psychiatry, University of Oxford, Warneford Hospital, Headington, Oxford, United Kingdom. Dr Malhotra is cur-

rently with Psychiatry Research, Hillside Hospital of Long Island Jewish Medical Center, Glen Oaks, New York.

**Correspondence:** Hugh M. D. Gurling, MBBS, MD, MPhil, FRCPsych, Molecular Psychiatry Laboratory, Department of Mental Health Sciences, University College London, Windeyer Institute of Medical Sciences, 46 Cleveland St, London, W1T 4JF, United Kingdom (h.gurling@ucl.ac.uk).

**Financial Disclosure:** A patent application that was filed by University College London was later transferred to H. M. D. Gurling.

**Funding/Support:** The research was funded by the European Science Foundation, the Wellcome Trust, the Neuroscience Research Charitable Trust, and a Priory Hospital research lectureship to J.L. The work of R.J.D. and R.S.J.F. is funded by Wellcome Trust Programme grants. The work of C.D.G. was funded by a Wellcome Career development award and that of H.C. by a Wellcome Clinician Scientist Fellowship.

## REFERENCES

- Gurling HMD. The genetics of the schizophrenias. Genetics of Mental Disorders, Part II: Clinical Issues. In: Papadimitriou GN, Mendlewicz J, eds. *Balliere's Clinical Psychiatry, International Practice and Research*. 1996;2:15-46.
- Blouin JL, Dombroski BA, Nath SK, Lasseter VK, Wolyniec PS, Nestadt G, Thornquist M, Ullrich G, McGrath J, Kasch L, Lamacz M, Thomas MG, Gehrig C, Radhakrishna U, Snyder SE, Balk KG, Neufeld K, Swartz KL, DeMarchi N, Papadimitriou GN, Dikeos DG, Stefanis CN, Chakravarti A, Childs B, Housman DE, Kazazian HH, Antonarakis S, Pulver AE. Schizophrenia susceptibility loci on chromosomes 13q32 and 8p21. *Nat Genet*. 1998;20:70-73.
- Brzustowicz LM, Honer WG, Chow EWC, Little D, Hogan J, Hodgkinson K, Bassett AS. Linkage of familial schizophrenia to chromosome 13q32. *Am J Hum Genet*. 1999;65:1096-1103.
- Gurling HM, Kalsi G, Brynjolfson J, Sigmundsson T, Sherrington R, Mankoo BS, Read T, Murphy P, Blaveri E, McQuillin A, Petursson H, Curtis D. Genomewide genetic linkage analysis confirms the presence of susceptibility loci for schizophrenia, on chromosomes 1q32.2, 5q33.2, and 8p21-22 and provides support for linkage to schizophrenia, on chromosomes 11q23.3-24 and 20q12.1-11.23. *Am J Hum Genet*. 2001;68:661-673.
- Kendler KS, MacLean CJ, O'Neill FA, Burke J, Murphy B, Duke F, Shinkwin R, Easter SM, Webb BT, Zhang J, Walsh D, Straub RE. Evidence for a schizophrenia vulnerability locus on chromosome 8p in the Irish Study of High-Density Schizophrenia Families. *Am J Psychiatry*. 1996;153:1534-1540.
- Blaveri E, Kalsi G, Lawrence J, Quested D, Moorey H, Lamb G, Kohen D, Shiwach R, Chowdhury U, Curtis D, McQuillin A, Gramoustianou ES, Gurling HM. Genetic association studies of schizophrenia using the 8p21-22 genes: preproenkephalin (PNOC), neuronal nicotinic cholinergic receptor alpha polypeptide 2 (CHRNA2) and arylamine N-acetyltransferase 1 (NAT1). *Eur J Hum Genet*. 2001;9:469-472.
- Weinberger DR, Torrey EF, Neophytides AN, Wyatt RJ. Structural abnormalities in the cerebral cortex of chronic schizophrenic patients. *Arch Gen Psychiatry*. 1979;36:935-939.
- Weinberger DR. Computed tomography (CT) findings in schizophrenia: speculation on the meaning of it all. *J Psychiatr Res*. 1984;18:477-490.
- Harvey I, Ron MA, Du Boulay G, Wicks D, Lewis SW, Murray RM. Reduction of cortical volume in schizophrenia on magnetic resonance imaging. *Psychol Med*. 1993;23:591-604.
- Lim KO, Tew W, Kushner M, Chow K, Matsumoto B, DeLisi LE. Cortical gray matter volume deficit in patients with first-episode schizophrenia. *Am J Psychiatry*. 1996;153:1548-1553.
- Wright IC, Rabe-Hesketh S, Woodruff PW, David AS, Murray RM, Bullmore ET. Meta-analysis of regional brain volumes in schizophrenia. *Am J Psychiatry*. 2000;157:16-25.
- Liddle PF. Schizophrenic syndromes, cognitive performance and neurological dysfunction. *Psychol Med*. 1987;17:49-57.
- Worsley KJ, Marrett S, Neelin P, Vandal AC, Friston K, Evans AC. A unified statistical approach for determining significant signals in images of cerebral activation. *Human Brain Mapping*. 1996;4:58-73.
- Genovese CR, Lazar NA, Nichols T, Tew W. Thresholding of statistical maps in

- functional neuroimaging using the false discovery rate. *Neuroimage*. 2002; 15:870-878.
15. Deichmann R, Good CD, Turner RF. Inhomogeneity compensation in structural brain imaging. *Magn Reson Med*. 2002;47:398-402.
  16. Deichmann R, Good CD, Josephs O, Ashburner J, Turner R. Optimization of 3-D MP-RAGE sequences for structural brain imaging. *Neuroimage*. 2000;12:112-127.
  17. Ashburner J, Friston KJ. Why voxel-based morphometry should be used. *Neuroimage*. 2001;14:1238-1243.
  18. Good CD, Johnsrude IS, Ashburner J, Henson RN, Friston KJ, Frackowiak RS. A voxel-based morphometric study of ageing in 465 normal adult human brains. *Neuroimage*. 2001;14:21-36.
  19. Ashburner J, Friston KJ. Voxel-based morphometry: the methods. *Neuroimage*. 2000;11:805-821.
  20. Pimm J, McQuillin A, Thirumalai S, Lawrence J, Quedsted D, Bass N, Lamb G, Moorey H, Datta SR, Kalsi G, Badacsonyi A, Kelly K, Morgan J, Punukollu B, Curtis D, Gurling H. The epsilon 4 gene on chromosome 5q, which encodes the clathrin-associated protein entrophin, is involved in the genetic susceptibility to schizophrenia. *Am J Hum Genet*. 2005;76:902-907.
  21. Endicott J, Spitzer RL. A diagnostic interview: the schedule for affective disorders and schizophrenia. *Arch Gen Psychiatry*. 1978;35:837-844.
  22. Rozen S, Skaletsky H. Primer3 on the WWW for general users and for biologist programmers. *Methods Mol Biol*. 2000;132:365-386.
  23. Sham PC, Curtis D. An extended transmission/disequilibrium test (TDT) for multi-allele marker loci. *Ann Hum Genet*. 1995;59:323-336.
  24. Sham PC, Curtis D. Monte Carlo tests for associations between disease and alleles at highly polymorphic loci. *Ann Hum Genet*. 1995;59:97-105.
  25. Zhao JH, Curtis D, Sham PC. Model-free analysis and permutation tests for allelic associations. *Hum Hered*. 2000;50:133-139.
  26. Zhao JH, Lissarrague S, Essioux L, Sham PC. GENECOUNTING: haplotype analysis with missing genotypes. *Bioinformatics*. 2002;18:1694-1695.
  27. Terwilliger JD, Ott J. *Handbook for Human Genetic Linkage*. Baltimore, Md: Johns Hopkins University Press; 1994.
  28. Engelender S, Sharp AH, Colomer V, Tokito MK, Lanahan A, Worley P, Holzbaur EL, Ross CA. Huntingtin-associated protein 1 (HAP1) interacts with the p150Glued subunit of dynactin. *Hum Mol Genet*. 1997;6:2205-2212.
  29. Millar JK, Christie S, Anderson S, Lawson D, Hsiao-Wei Loh D, Devon RS, Arveiler B, Muir WJ, Blackwood DH, Porteous DJ. Genomic structure and localisation within a linkage hotspot of Disrupted In Schizophrenia 1, a gene disrupted by a translocation segregating with schizophrenia. *Mol Psychiatry*. 2001;6:173-178.
  30. Kendler KS, Myers JM, O'Neill FA, Martin R, Murphy B, MacLean CJ, Walsh D, Straub RE. Clinical features of schizophrenia and linkage to chromosomes 5q, 6p, 8p, and 10p in the Irish Study of High-Density Schizophrenia Families. *Am J Psychiatry*. 2000;157:402-408.
  31. Stefansson H, Sigurdsson E, Steinthorsdottir V, Bjornsdottir S, Sigmundsson T, Ghosh S, Brynjolfsson J, Gunnarsdottir S, Ivarsson O, Chou TT, Hjaltason O, Birgisdottir B, Jonsson H, Gudnadottir VG, Gudmundsdottir E, Bjornsson A, Ingvarsson B, Ingason A, Sigfusson S, Hardardottir H, Harvey RP, Lai D, Zhou M, Brunner D, Mutel V, Gonzalo A, Lemke G, Sainz J, Johannesson G, Andresson T, Gudbjartsson D, Manolescu A, Frigge ML, Gurney ME, Kong A, Gulcher JR, Petursson H, Stefansson K. Neuregulin 1 and susceptibility to schizophrenia. *Am J Hum Genet*. 2002;71:877-892.
  32. Stefansson H, Sarginson J, Kong A, Yates P, Steinthorsdottir V, Gudfinnsson E, Gunnarsdottir S, Walker N, Petursson H, Crombie C, Ingason A, Gulcher JR, Stefansson K, St Clair D. Association of neuregulin 1 with schizophrenia confirmed in a Scottish population. *Am J Hum Genet*. 2003;72:83-87.
  33. Yang JZ, Si TM, Ruan Y, Ling YS, Han YH, Wang XL, Zhou M, Zhang HY, Kong QM, Liu C, Zhang DR, Yu YQ, Liu SZ, Ju GZ, Shu L, Ma DL, Zhang D. Association study of neuregulin 1 gene with schizophrenia. *Mol Psychiatry*. 2003;8:706-709.
  34. Iwata N, Suzuki T, Ikeda M, Kitajima T, Yamanouchi Y, Inada T, Ozaki N. No association with the neuregulin 1 haplotype to Japanese schizophrenia. *Mol Psychiatry*. 2004;9:126-127.
  35. Gerber DJ, Hall D, Miyakawa T, Demars S, Gogos JA, Karayiorgou M, Tonegawa S. Evidence for association of schizophrenia with genetic variation in the 8p21.3 gene, PPP3CC, encoding the calcineurin gamma subunit. *Proc Natl Acad Sci U S A*. 2003;100:8993-8998.
  36. Katsu T, Ujike H, Nakano T, Tanaka Y, Nomura A, Nakata K, Takaki M, Sakai A, Uchida N, Imamura T, Kuroda S. The human frizzled-3 (FZD3) gene on chromosome 8p21, a receptor gene for Wnt ligands, is associated with the susceptibility to schizophrenia. *Neurosci Lett*. 2003;353:53-56.
  37. Yang J, Si T, Ling Y, Ruan Y, Han Y, Wang X, Zhang H, Kong Q, Li X, Liu C, Zhang D, Zhou M, Yu Y, Liu S, Shu L, Ma D, Wei J, Zhang D. Association study of the human FZD3 locus with schizophrenia. *Biol Psychiatry*. 2003;54:1298-1301.
  38. Puri V, McQuillin A, Choudhury K, Datta S, Pimm J, Thirumalai S, Krasucki R, Lawrence J, Quedsted D, Bass N, Moorey H, Morgan J, Punukollu B, Kandasami G, Curtis D, Gurling H. Fine mapping by genetic association implicates the chromosome 1q23.3 gene UHMK1, encoding a serine/threonine protein kinase, as a novel schizophrenia susceptibility gene. *Biol Psychiatry*. 2006 In press.