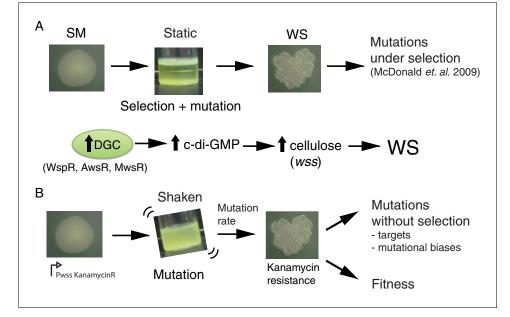




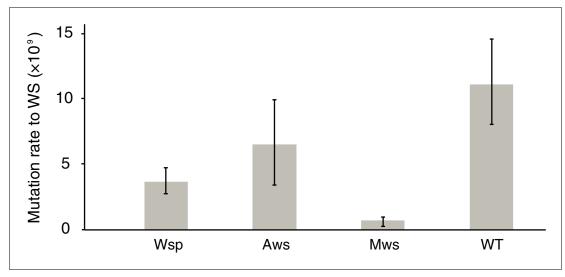
## Figures and figure supplements

Predicting mutational routes to new adaptive phenotypes

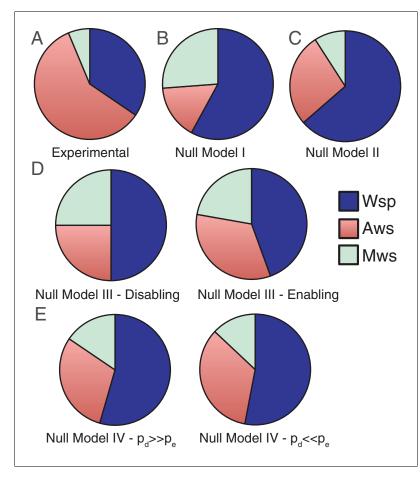
Peter A Lind et al



**Figure 1.** The *Pseudomonas fluorescens* SBW25 wrinkly spreader model. (A) Selection for access to oxygen allows wrinkly spreader (WS) mutants to invade the ancestral smooth (SM) population in static microcosms. WS mutants form a mat at the air-liquid interface through increased expression of the main structural component, cellulose, encoded by the wss operon. Expression of cellulose is controlled by the second messenger c-di-GMP, which is produced by diguanylate cyclases (DGCs). Mutations in the *wsp, aws* and *mws* operons, that activate their respective DGCs (WspR, AwsR, MwsR), are the primary mutational pathways to WS. (B) When a reporter construct connecting expression of the wss operon to resistance to kanamycin is used under shaken non-selective conditions, WS mutants can be isolated without the biasing influence of natural selection. This allows estimation of the mutation rate to WS and an unbiased spectrum of mutations defining the mutational target. Fitness can then be assayed in competition with a common reference strain. DOI: https://doi.org/10.7554/eLife.38822.003



**Figure 2.** Mutation rates to WS. Fluctuation tests were used to estimate the mutation rate to WS for the three common mutational pathways to WS. Error bars represent mean and 95% confidence intervals. All mutation rates are significantly different from all others (t-test p<0.05, see Materials and method section for details). Number of replicates n = 200 for Aws and Wsp, n = 400 for Mws, n = 100 for WT. DOI: https://doi.org/10.7554/eLife.38822.004



**Figure 3.** Comparisons of experimental data and null model predictions for the use of the Wsp, Aws and Mws pathways. (A) Experimental data from *Figure 2*. (B) Null Model I: number of base pairs as proxy for mutational target size. (C) Null Model II: number of genes as a proxy for mutational target size. (D) Null Model III: function and interactions between components determine mutational target size. To indicate the range of possible states we consider the situation with enabling mutations only (left hand panel) and disabling mutations only (right hand panel). (E) Null Model IV: as per null Model III, but interactions have both pleiotropic and continuous effects. To indicate the range of possible states we consider the probability of disabling mutational effects ( $p_{d}$ ) to be 10 times more common than enabling changes ( $p_{e}$ ),  $p_{d} = 0.001$ ,  $p_{e} = 0.0001$  (left hand panel) the probability of enabling mutational effects ( $p_{e}$ ) to be 10 times more common than disabling changes ( $p_{d}$ ),  $p_{d} = 0.001$ ,  $p_{e} = 0.0001$ ,  $p_{e$ 

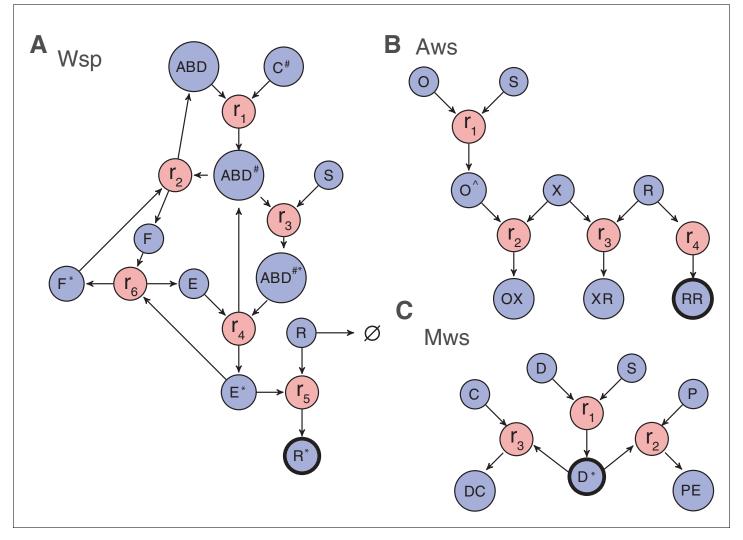


Figure 4. Reaction diagrams for null models III and IV. Reaction diagrams show the interactions governing production of a wrinkly spreader in (A) Wsp, (B) Aws and (C) Mws pathways. The blue circles indicate molecular components, the red circles indicate reactions, and arrows indicate which molecular components are reactants and which are products. Full descriptions of the models including all reactions and the resulting systems of differential equations are found in *Figure 4—figure supplement 1* for Wsp, *Figure 4—figure supplement 2* for Aws and *Figure 4—figure supplement 3* for Mws. The black edged circles are the reporter proteins, which is the activated forms of the diguanylate cyclases that directly determine whether a pathway generates a wrinkly spreader. Increased production of any reporter leads to increased c-di-GMP concentration causing increased cellulose production and thereby a wrinkly spreader.

4	B , O	C
1. Methylated WspC activates the WspABD complex via methylation	1. C# + ABD $\xrightarrow{r_1}$ ABD#	$\frac{\partial [ABD]}{\partial t} = r_2 [F^*] [ABD^{\#}] - r_1 [ABD] [C^{\#}]$
2. Phosphorylated WspF inactivates a methylated WspABD complex by demethylation	2. F* + ABD# $\xrightarrow{r_2}$ F + ABD	$\frac{\partial [ABD\#]}{\partial t} = r_1 [ABD][C\#] - r_2 [F^*][ABD\#] - r_3 S[ABD\#] + r_4 [E][ABD\#]$
3. A signal S can activate a methylated WspABD	3. ABD + S $\xrightarrow{r_3}$ ABD#*	$\frac{\partial [ABD\#^*]}{\partial t} = r_3 S[ABD\#] - r_4 [E][ABD\#^*]$
complex via an autophosphorylation reaction	4 ABD#* + E $\xrightarrow{r_4}$ ABD# +E*	$\frac{\partial [E^*]}{\partial t} = r_4[E][ABD\#^*] - r_5[E^*][R] - r_6[E^*][F]$
4. A methylated and phosphorylated WspABD complex can activate WspE by phosphorylation		$\frac{\partial [R^*]}{\partial t} = r_5 [R] [E^*]$
5. Phosphorylated WspE can phosphorylate WspR to an active form	5. $E^* + R \xrightarrow{r_5} R^* + E$	$\frac{\partial[E]}{dt} = r_6[E^*][F] - r_4[E][ABD\#^*] + r_5[R][E^*]$
6. Phosphorylated WspE can phosphorylate WspF	6. $E^* + F \xrightarrow{r_6} F^* + E$	$\frac{\partial [F]}{\partial t} = r_2[F^*][ABD^{\#}] - r_6[E^*][F]$
7. WspR is degraded or bound by a competitor at constant concentration (mathematically equivalent).	7. R $\xrightarrow{.01} \emptyset$	$\frac{\partial [F^*]}{\partial \partial t} = r_6[E^*][F] - r_2[F^*][ABD\#]$
The rate (0.01) is chosen to provide a steady, slow decay. Whithout this all WspR is converted to R*		$\frac{\partial[R]}{\partial t} = -r_5[R][E^*]01[R]$

Figure 4—figure supplement 1. Wsp model. (A) Description of functional interactions (B) Description of molecular reaction (C) Differential equations describing the dynamics of the Wsp pathway. The activity of WspA is modulated by methylation, where it is activated by the CheR-like methyltransferase WspC (PFLU1221). The CheB-like methylesterase WspF (PFLU1224) functions as a negative regulator. Modulation of WspR activity through changes in oligomerization state and clustering is not explicitly included in the model (*De et al., 2008; Huangyutitham et al., 2013*), but can be interpreted as changes in the rate of WspR activation.

## eLIFE Research article

Α	1. A signal S binds to AwsO and activates it	<b>B</b> $_{1.S+O} \xrightarrow{r_1} O^{\wedge}$	$\mathbf{C}_{\frac{\partial[X]}{\partial t} = -r_2[X][O^{\wedge}] - r_3[X][R]}$
	2. Activated AwsO binds to AwsX to form a complex that sequesters AwsX	2. $X + O^{\wedge} \xrightarrow{r_2} OX$	$\frac{\partial [XR]}{\partial t} = r_3[X][R]$
	3. AwsX binds AwsR to form a complex that sequesters AwsX	3. X + R $\xrightarrow{r_3}$ XR	$\frac{\partial[O]}{\partial t} = -r_1 S[O]$ $\frac{\partial[O^A]}{\partial t} = r_1 S[O] - r_1 [X][OA]$
	4. AwsR dimerizes (active form)	4. R + R $\xrightarrow{r_4}$ RR	$\frac{\partial [O^{\Lambda}]}{\partial t} = r_1 S[O] - r_2 [X][O^{\Lambda}]$ $\frac{\partial [OX]}{\partial t} = r_2 [X][O^{\Lambda}]$
			$\frac{\partial[R]}{\partial t} = -r_3[X][R] - r_4[R][R]$
			$\frac{\partial [RR]}{\partial t} = r_4[R][R]$

Figure 4—figure supplement 2. Aws model. (A) Description of functional interactions (B) Description of molecular reaction (C) Differential equations describing the dynamics of the Aws pathway. Release of AwsX mediated repression results in a conformational shift that in the model is represented as formation of an active dimer.

<b>A</b> 1. A signal S binds to DGC domain and activates it	$\mathbf{B}_{1.S+D} \xrightarrow{r_{1}} D^{*}$	$\mathbf{C}  \frac{\partial [D]}{\partial t} = -r_1[D]S$
2. Activated DGC interacts with PDE to inactivate it	2. $D^* + P \xrightarrow{r_2} DP$	$\frac{\partial [D^*]}{\partial t} = r_1[D]S - r_2[D^*][P] - r_3[D^*][C]$
3. Activated DGC interacts with C and to form alternative complex DC	3. D* +C $\xrightarrow{r_3}$ DC	$\frac{\partial [P]}{\partial t} = -r_2 [D^*][P]$ $\frac{\partial [DP]}{\partial t} = r_2 [D^*][P]$ $\frac{\partial [C]}{\partial t} = -r_3 [D^*][C]$ $\frac{\partial [DC]}{\partial t} = r_3 [D^*][C]$

Figure 4—figure supplement 3. Mws model. (A) Description of functional interactions (B) Description of molecular reactions (C) Differential equations describing the dynamics of the Aws pathway.

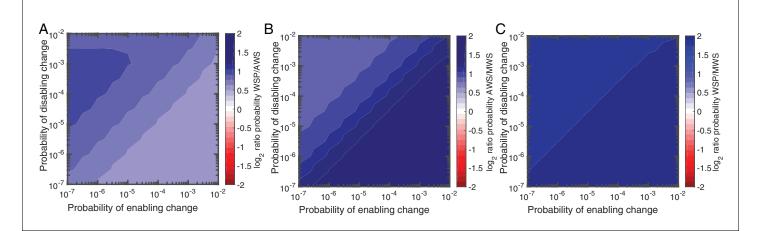
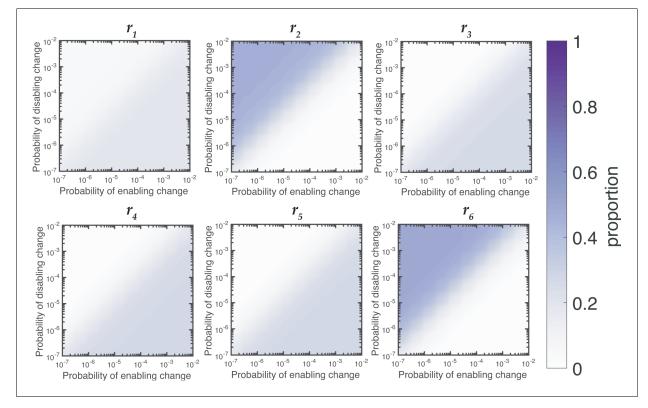
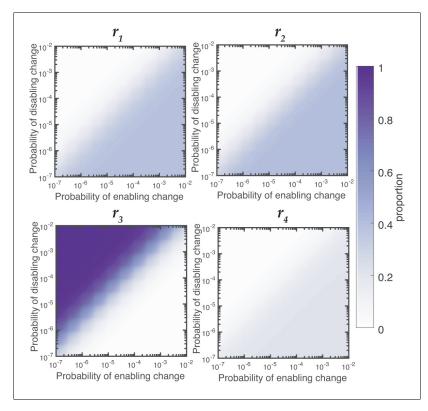
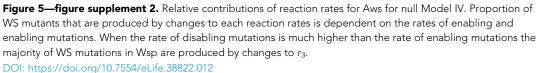


Figure 5. Null Model 4 predictions of the probability of using the Wsp, Aws and Mws pathways. (A) Probability of Wsp relative to Aws. (B) Probability of Mws relative to Aws. (C) Probability of Mws relative to Wsp. The relative contributions of individual reactions rates are available in *Figure 5—figure supplement 1* for Wsp and *Figure 5—figure supplement 2* for Aws. Sensitivity analysis is shown in *Figure 5—figure supplement 3*. DOI: https://doi.org/10.7554/eLife.38822.010



**Figure 5—figure supplement 1.** Relative contributions of reaction rates for Wsp for null Model IV. Proportion of WS mutants that are produced by changes to each reaction rates is dependent on the rates of enabling and enabling mutations. When the rate of disabling mutations is much higher than the rate of enabling mutations the majority of WS mutations in Wsp are produced by changes to  $r_2$  and  $r_6$ . DOI: https://doi.org/10.7554/eLife.38822.011





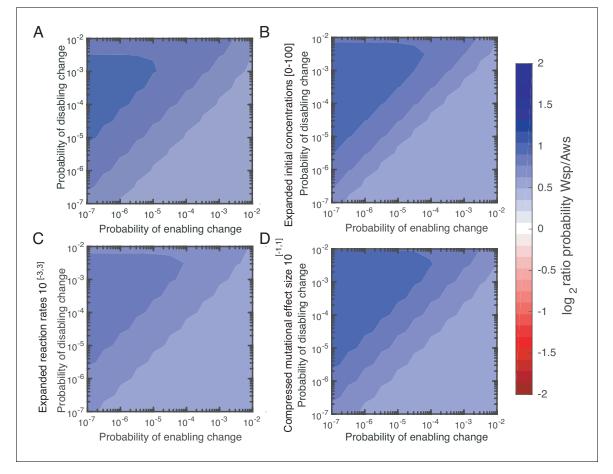
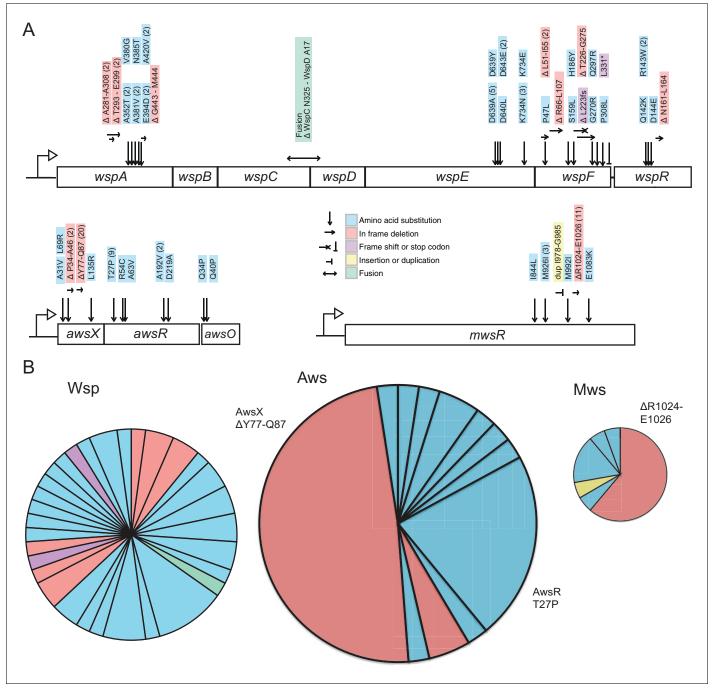


Figure 5—figure supplement 3. Parameter sensitivity analysis. To assess the effect of the chosen parameter (A) ranges on our results, we redid our sampling procedure for WSP for three different parameter regimes: (B) an expanded range for initial concentrations [0–100], (C) an expanded range for reaction rates 10 [-3,3], (D) a compressed range for mutational effect size 10 [-1,1]. We found that our qualitative results are robust to these changes. DOI: https://doi.org/10.7554/eLife.38822.013



**Figure 6.** Mutational targets. (A) 105 independent mutations in the *wsp* (n = 46), *aws* (n = 41) and *mws* (n = 18) operons were identified. Numbers of independent mutants are shown in brackets. Full details on the mutations are available in Figure 5—source data 1. (B) Diversity of mutations with area proportional to mutation rate (*Figure 2*). Two mutations (AwsX  $\Delta$ Y77-Q87 and AwsR T27P) contribute 41% of all mutations to WS suggesting that these are mutational hot spots.

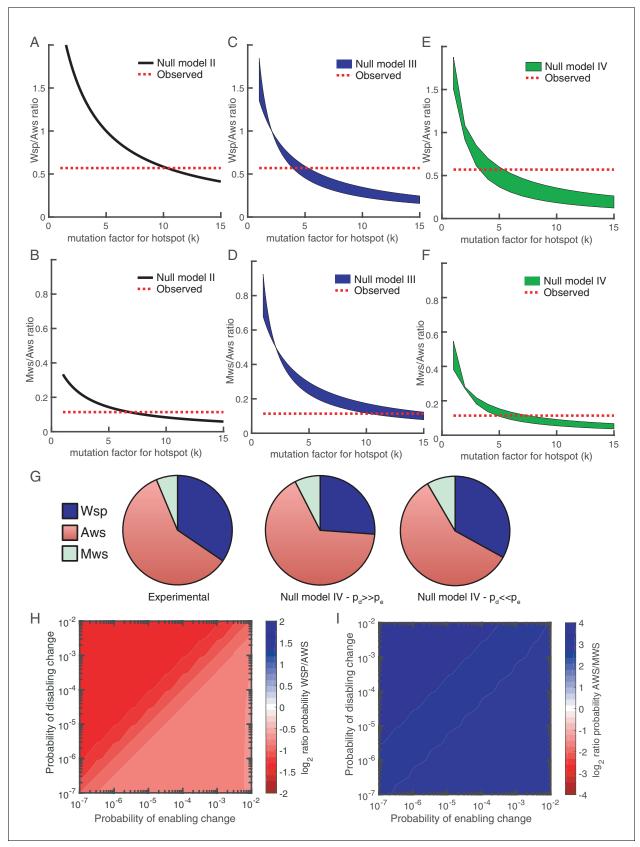
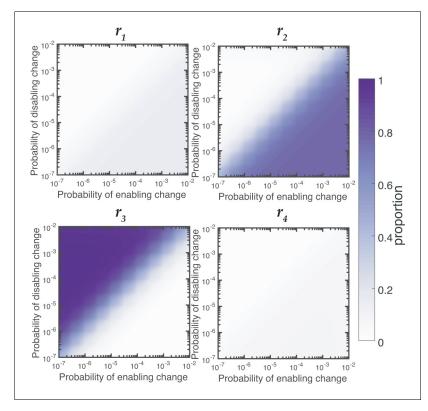


Figure 7. Revised models including mutational hot spot. Null Models II (A, B), III (C, D) and IV (E, F) can be revised to take into account the mutational hot spot in AwsX by including a mutation factor (k). Each plot (A-F) shows the predicted ratios of Wsp/Aws or Aws/Mws compared to the experimental *Figure 7 continued on next page* 

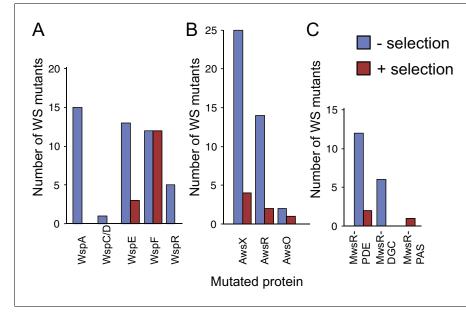


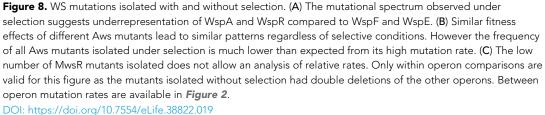
## Figure 7 continued

results (dashed red line). For null models III and IV a range is shown that depends on the probabilities of enabling ( $p_e$ ) and disabling ( $p_d$ ) changes. (G) Comparison of experimental data (left) and revised predictions from null model IV (middle, right), incorporating a five-fold increase in  $p_e$  and  $p_d$  for  $r_2$ and  $r_3$  in the Aws differential equation system. Middle pie chart use  $p_d = 0.001$ ,  $p_e = 0.0001$  and right pie chart  $p_d = 0.0001$ ,  $p_e = 0.0001$ ,

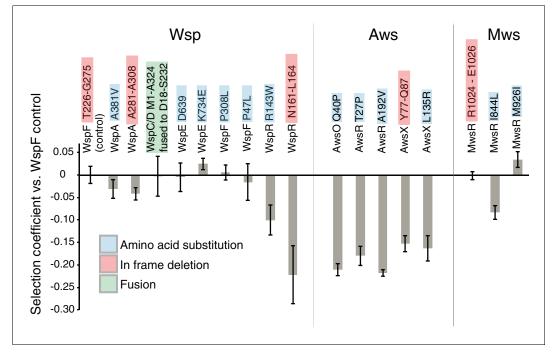


**Figure 7—figure supplement 1.** Relative contributions of reaction rates for Aws for null Model IV with a five-fold increase in mutation factor for  $r_2$  and  $r_3$ . Proportion of WS mutants that are produced by changes to each reaction rates is dependent on the rates of enabling and enabling mutations. When the rate of disabling mutations is much higher than the rate of enabling mutations the majority of WS mutations in Wsp are produced by changes to  $r_3$ . DOI: https://doi.org/10.7554/eLife.38822.017





Lind et al. eLife 2019;8:e38822. DOI: https://doi.org/10.7554/eLife.38822



**Figure 9.** Fitness of different WS mutants. Competitive fitness against a WspF  $\Delta$ T226-G275 reference strain was measured for representative mutations in the Wsp, Aws, Mws pathways. Pairwise competitions were performed in quadruplicates and error bars represent ±one standard deviation. Full data, including statistical tests are available in *Figure 9—source data 1*.