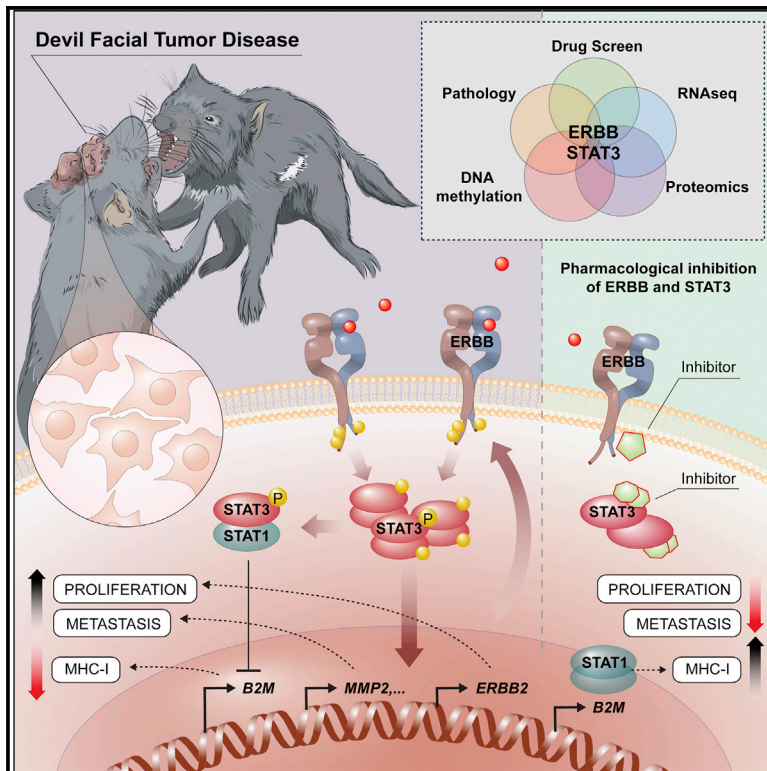


Cancer Cell

The ERBB-STAT3 Axis Drives Tasmanian Devil Facial Tumor Disease

Graphical Abstract



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In Brief

Kosack et al. identify the ERBB-STAT3 signaling axis as a central molecular driver of Tasmanian devil transmissible facial tumors. Inhibition of ERBB or STAT3 prevents tumor growth in xenograft models and restores MHC class I expression, suggesting a chemo-immunotherapeutic strategy to save Tasmanian devils.

Highlights

- Tasmanian devil proteome and DNA methylome reveal modulated tumor environment
- Identification of cancer-driving hyperactive ERBB-STAT3 axis as a druggable target
- ERBB-STAT3-driven suppression of MHC class I genes may facilitate immune evasion
- Successful pharmacological treatments of Tasmanian devil tumors in a xenograft model



The ERBB-STAT3 Axis Drives Tasmanian Devil Facial Tumor Disease

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SUMMARY

The marsupial Tasmanian devil (*Sarcophilus harrisi*) faces extinction due to transmissible devil facial tumor disease (DFTD). To unveil the molecular underpinnings of this transmissible cancer, we combined pharmacological screens with an integrated systems-biology characterization. Sensitivity to inhibitors of ERBB tyrosine kinases correlated with their overexpression. Proteomic and DNA methylation analyses revealed tumor-specific signatures linked to the evolutionary conserved oncogenic STAT3. ERBB inhibition blocked phosphorylation of STAT3 and arrested cancer cells. Pharmacological blockade of ERBB or STAT3 prevented tumor growth in xenograft models and restored MHC class I expression. This link between the hyperactive ERBB-STAT3 axis and major histocompatibility complex class I-mediated tumor immunosurveillance provides mechanistic insights into horizontal transmissibility and puts forward a dual chemo-immunotherapeutic strategy to save Tasmanian devils from DFTD.

INTRODUCTION

Cancer cells do not usually transmit between individuals. No human examples of transmissible cancers are known apart from rare iatrogenic cases during surgery and transplantation or maternal-fetal transmission (Isoda et al., 2009; Matser et al., 2018;

Metzger and Goff, 2016). Accordingly, horizontal transmission is not considered a hallmark of cancer (Hanahan and Weinberg, 2011). Yet, at least six species in the animal kingdom harbor clonal cancers which spread horizontally within populations (Metzger et al., 2016; Ostrander et al., 2016). These diseases include the fatal devil facial tumor disease (DFTD) in Tasmanian

Significance

Tasmanian devils are threatened with extinction by a horizontally transmitted cancer termed devil facial tumor disease (DFTD). Using an integrative systems-wide approach, this study identifies the ERBB-STAT3 signaling axis as a central molecular driver of DFTD. Pharmacological targeting of this axis in cell culture and in xenograft mouse models inhibited tumor growth and restored the expression of MHC class I genes. These results extend our understanding of the molecular signaling cascades driving DFTD and provide a potential explanation why horizontally transmitted DFTD cells are not rejected by the immune system. Pharmacological interference with ERBB-STAT3 may provide a much-needed addition to the ongoing efforts to save Tasmanian devils from transmissible cancer.



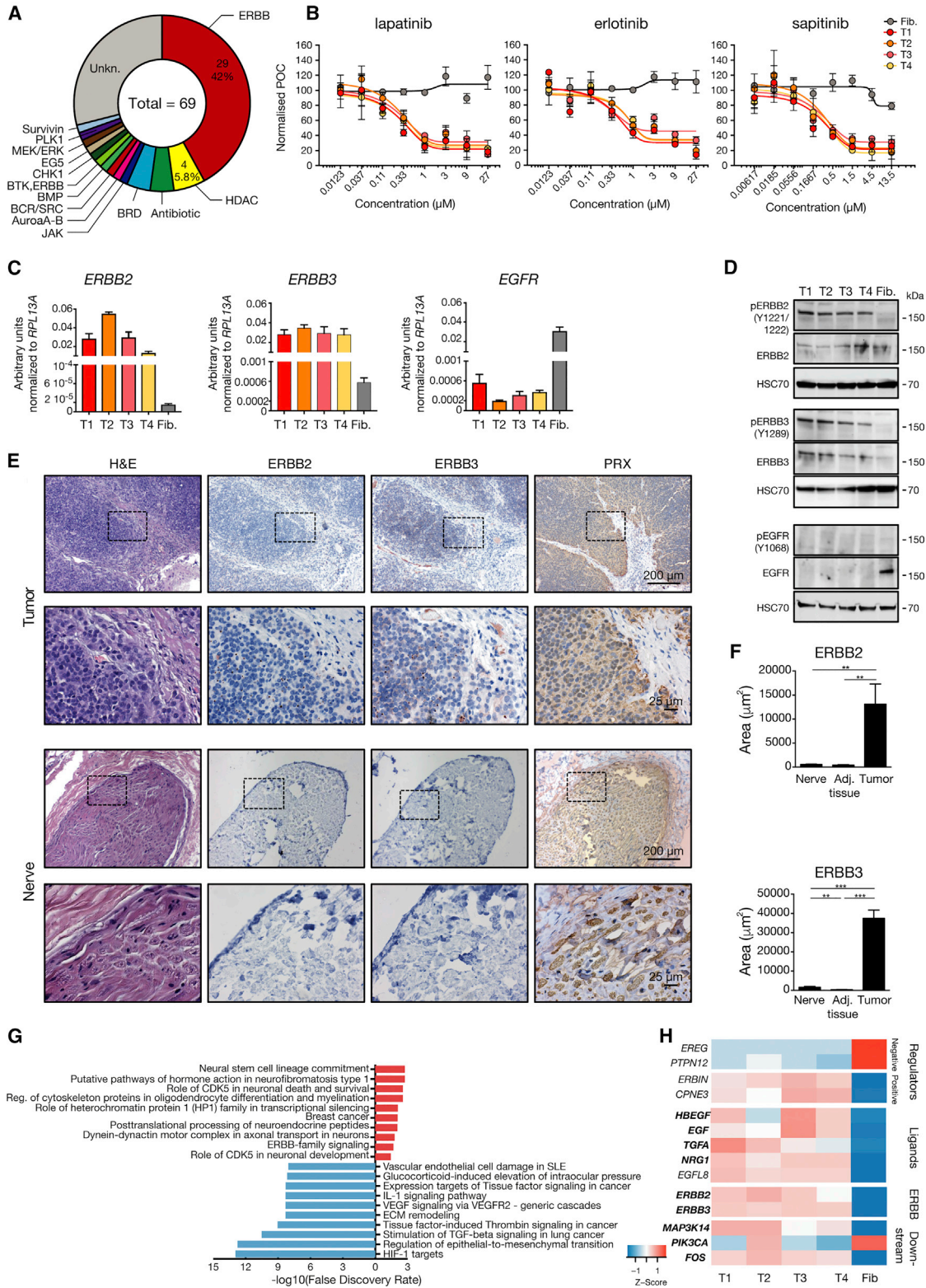


Figure 1. A Pharmacological Screen Identified ERBB-Specific Vulnerability of DFTD

(A) The targets of the 69 drug hits in the 4-point drug screen showing a reduction in cell viability in at least 1 of the 4 DFTD cell lines compared with healthy fibroblasts.

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devils (Murchison et al., 2010, 2012; Pearse and Swift, 2006; Stammnitz et al., 2018), a sexually transmitted sarcoma in dogs (Frampton et al., 2018; Murchison et al., 2014; Murgia et al., 2006), and leukemia-like cancers in mollusks (Metzger et al., 2015, 2016). Genetic studies provided invaluable insights into these unusual cancers but the molecular underpinnings of malignancy and transmissibility remain poorly understood.

DFTD is an allogeneic graft of Schwann cell origin, which is transmitted by direct transfer of living cancer cells from one individual to another as a result of biting behavior during feeding or mating interactions (Murchison et al., 2010; Pearse and Swift, 2006). Diseased devils succumb to the disease within months (Loh et al., 2006a), rendering DFTD a serious threat to the survival of the population of the largest living marsupial carnivore. Several mechanisms are thought to confer the tumor cells with the property of being successfully transmitted between individual Tasmanian devils, including the lack of rejection due to the low expression levels of major histocompatibility complex (MHC) class I genes and diminished genetic diversity (Siddle et al., 2007, 2013). Despite recent efforts, vaccines and treatments showed limited success against DFTD (Kreiss et al., 2015; Tovar et al., 2017). A second transmissible cancer was discovered in 2014 (Pye et al., 2016), which has been termed DFT2 to distinguish it from the first transmissible DFTD, now termed DFT1. In this study DFTD refers to DFT1, first identified in 1996 (Pearse and Swift, 2006).

In this study we investigate DFTD-specific aberrant signaling pathways in order to understand and exploit the underlying molecular wiring of this transmissible cancer for potential therapeutic avenues.

RESULTS

ERBB-Specific Vulnerability of DFTD Identified by Pharmacological Screening

To address the inherent limitations of working with non-model organisms such as the Tasmanian devil, we characterized cell lines and primary biopsies (Table S1) through an integrative and unbiased systems-biology approach consisting of pharmacological screens, transcriptomics, proteomics, and epigenomics. First, we performed a cell viability screen with over 2,500 selected compounds against 4 DFTD tumor cell lines (T1–T4) on an automated high-throughput screening platform to identify potential pharmacological vulnerabilities (Table S2). A fibroblast cell line of Tasmanian devil origin served as control (Murchison et al., 2012). Sixty-nine compounds killed at least one out of

four DFTD tumor cell lines, but did not affect the viability of fibroblasts, as measured by intracellular ATP levels (Figure 1A; Table S2). Interestingly, this unbiased approach yielded a substantial enrichment of tyrosine kinase inhibitors targeting selectively the ERBB receptors (29/69; 42%) including lapatinib, erlotinib, and sapitinib (Figure 1B, Table S2). In addition, we observed DFTD tumor cell-specific killing for inhibitors for histone deacetylases (HDAC), BET bromodomains and other potential therapeutic targets (Figure 1A, Table S2). The human ERBB family has four members (*EGFR*, *ERBB2*, *ERBB3*, and *ERBB4*) (Hynes and Lane, 2005), of which the devil genome has all orthologs annotated except *ERBB4*. Interestingly, DFTD cells expressed higher transcript levels of *ERBB2* and *ERBB3* compared with fibroblasts, while *EGFR* was barely detectable (Figure 1C). To validate this finding on the protein level, we tested antibodies with cross-species recognition (Table S1). Western blot analysis confirmed increased levels of total ERBB2 and ERBB3 in DFTD cell lines compared with fibroblasts (Figure 1D). The phosphorylated residues Y1221/1222 (ERBB2) and Y1289 (ERBB3) are conserved across species (Table S1), highlighting the evolutionary impact of tyrosine kinase signaling in cancer cells. Likewise, sequence alignments with >95% conserved amino acids in its protein kinase domain suggest that Tasmanian devil ERBB3 is a pseudokinase as known from other vertebrates. Phospho-site-specific antibodies provided evidence for persistent activation of ERBB2 and ERBB3 (Figure 1D). These results were corroborated by immunohistochemical detection of increased expression of ERBB2 and ERBB3 in primary tumor biopsies of diseased Tasmanian devils compared with adjacent non-tumor tissue (Figures 1E and 1F). Tumor cells were identified by the DFTD diagnostic marker Periaxin (PRX) (Murchison et al., 2010; Tovar et al., 2011). To control for the Schwann cell origin of DFTD, we also analyzed PRX-positive peripheral nerve tissue and found that DFTD tumors express elevated levels of ERBB2 and ERBB3 compared with nerve tissue (Figures 1E and F). Transcriptional profiling of DFTD tumor cell lines T1–T4 and fibroblasts revealed differentially expressed transcripts and predicted transcription factors driving this differential gene regulation (Figures S1A–S1D; Table S3). Among the predicted upregulated pathways was ERBB family signaling (Figure 1G). The genes driving this enrichment included *HBEGF*, *EGF*, and *NRG1* that encode ERBB ligands, *ERBIN* and *CPNE3* that encode positive regulators, and the proto-oncogene *FOS* (all increased expression), as well as *EREG* and *PTPN12* (decreased expression) that encode negative regulators (Figure 1G). We corroborated the differential expression of the ERBB ligands in

(B) Eight-point dose-response curves for lapatinib, erlotinib, and sapitinib in four tumor cell lines (T1–T4) and fibroblasts (Fib.) shown as normalized percentage of control (POC).

(C) Expression of the ERBB family members quantified by real-time PCR (n = 3 replicates).

(D) Western blots of the three annotated ERBB proteins (total and phosphorylated).

(E) Histopathological analysis of tumor and peripheral nerve biopsies for H&E and immunohistochemistry (IHC) against Periaxin (PRX), ERBB2, and ERBB3 on serial consecutive sections. Dotted rectangles indicate magnified areas. Scale bars, 200 and 25 μ m.

(F) Quantification of ERBB2 and ERBB3 signal in tumor biopsies, adjacent tissue, and peripheral nerve tissue. Statistical significance was calculated by unpaired t test.

(G) MetaCore pathway maps enrichments separately for up- (red) and downregulated (blue) transcripts in tumor cell lines versus fibroblasts. Only the first ten pathways and with a false discovery rate ≤ 0.05 are reported.

(H) Heatmap of RNA-seq expression for genes driving the MetaCore pathway “ERBB-family signaling” (bold gene symbols) as well as known positive (*ERBIN* and *CPNE3*) and negative (*EREG* and *PTPN12*) regulators of the ERBB pathway and *EGFL8* in four DFTD cell lines (T1–T4) and fibroblasts (Fib.).

Graphs represent the mean \pm SEM. See also Figure S1 and Tables S1, S2, S3, and S4.

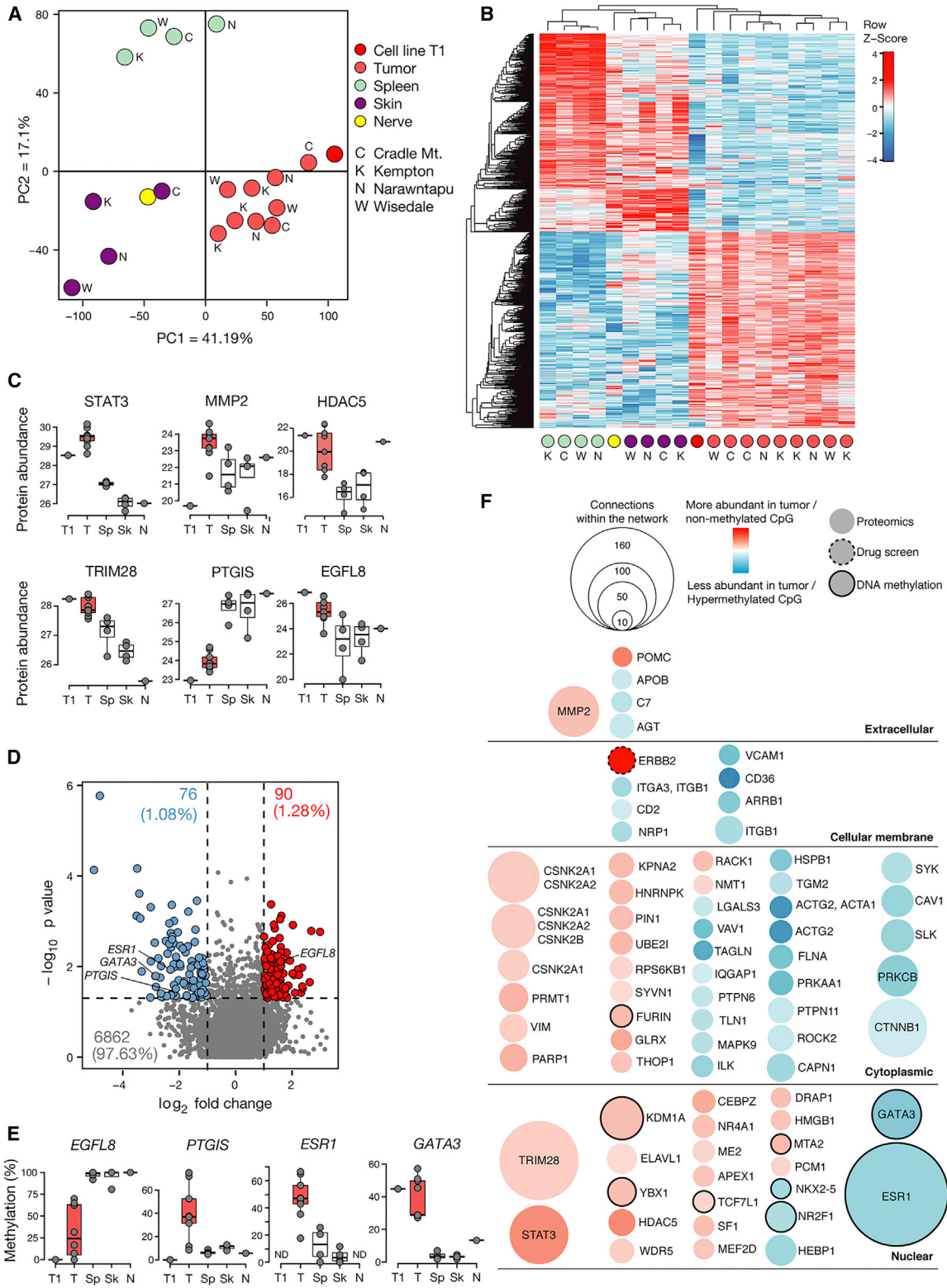


Figure 2. Integrative Systems-Level Analysis of DFTD

(A) Principal-component analysis of the 3,894 proteins quantified in all samples. Sampling locations are indicated in capital letters. Cell line denotes the DFTD cell line 06/2887 (T1) and “Nerve” stands for a healthy nerve biopsy.

(B) Hierarchical clustering of the 987 differentially modulated proteins between tumor and healthy biopsies.

(C) Boxplots of selected protein abundance across conditions in DFTD cell line (T1) and biopsies of tumor (T), spleen (Sp), skin (Sk), and nerve tissue (N).

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primary DFTD tumor tissue by real-time PCR (Figure S1E). Due to its marked upregulation, we also included *EGFL8*, which encodes the putative ERBB ligand epithelial growth factor-like domain multiple 8, in our analysis (Figures 1H, S1E, and S1F). Of note, variant analysis of our high-coverage RNA sequencing (RNA-seq) data did not provide any evidence for activating mutations in expressed devil orthologous genes of the involved pathways (Figures S1G–S1I; Table S4). Together, these data indicate that DFTD tumor cells, which express high levels of phosphorylated ERBB2 and ERBB3 and show activation of the ERBB signaling pathway, are exquisitely vulnerable to ERBB kinase inhibitors.

Characterization of DFTD by Integrated Proteomic and DNA Methylation Analysis

To unravel the involved signaling cascades in DFTD, we investigated global changes in protein abundance in primary biopsies of diseased devils by proteomic analysis. This approach included DFTD tumor tissue as well as healthy control tissue from skin and spleen from four Tasmanian devils from different geographical locations as well as peripheral nerve tissue and the DFTD tumor cell line T1 (Table S1). Overall, we identified 6,672 unique proteins across all samples searched against a Uniprot reference library of the devil (Figure S2A, Table S5). A total of 4,981 of these identified proteins were detected across more than 80% of the samples (Figures S2B and S2C). This unbiased expression proteomic approach was not specifically designed to enrich for hydrophobic transmembrane proteins such as ERBB2 and ERBB3. Principal-component analysis of the 3,894 proteins quantified in all samples distinguished the replicates according to the tissue of origin, with the first principal component accounting for 41.1% of the inter-sample variability differentiating tumor from healthy samples (Figure 2A). Upon differential analysis of tumor versus healthy tissues we defined a tumor-modulated signature of 987 proteins (Figure 2B; Table S5). Among the most prominent proteins overexpressed in tumor tissue we identified the oncogenic transcription factor STAT3 (Figures 2C and S2D). Downstream targets of STAT3, among which matrix metalloproteinase 2 (MMP2) (Figures 2C and S2E–S2G) (Xie et al., 2004) is also differentially modulated in the tumor biopsies. We detected MMP2, which is secreted as a proprotein, at high abundance in tumor biopsies but not in the DFTD cell line. This may be due to technical limitations or could indicate its extracellular secretion in the tumor microenvironment. Tumor tissue also expressed high levels of the histone deacetylase HDAC5 and the SUMO/ubiquitin E3 ligase TRIM28, which is linked to STAT3 signaling (Tsuruma et al., 2008), as well as low expression of the tumor suppressor PTGIS (Figure 2C). Further,

our proteomic analysis confirmed the high expression levels of the *EGFL8* in DFTD tumor cells (Figures 2C, 1H, and S1F). Pathway enrichment analysis highlighted downregulation of processes related to chemotaxis, cell adhesion, and cytoskeleton remodeling (Figures S2H and S2I; Table S5), some of which are negatively regulated by STAT3 (Kortylewski et al., 2005; Yu et al., 2007).

In complementation to the proteomic characterization we mapped DNA methylation by reduced representation bisulfite sequencing to depict the landscape of epigenetic regulation in the aforementioned primary biopsies from Tasmanian devils. Quantification of CpG methylation was performed by an unbiased *de novo* approach (Klughammer et al., 2015). For the sub-sequent analysis, due to the imperfect genome annotation of this non-model organism, we focused on the identification of methylated CpGs in promoter regions. DNA methylation marks readily distinguished tumor and healthy tissue and identified tumor-specific methylation signatures and their putative transcription factor binding sites (Figures S3A–S3J). Differential analysis of tumor versus healthy biopsies highlighted 166 candidate genes with different DNA methylation levels in their promoters (Figures 2D and 2E; Table S6), which included the tumor-specific hypomethylated *EGFL8* promoter as well as hypermethylated promoters of *ESR1*, *PTGIS*, and *GATA3* (Figure 2E). Comparative analysis by gene set enrichment revealed a high concordance of RNA-seq-derived transcript levels and proteomic data in the DFTD cell line (Figure S3K) and was in line with an inverse correlation pattern of methylated promoter regions and respective gene transcript level (Figures S3L and S3M). An integrative network analysis of the identified drug vulnerabilities, protein and methylation signatures revealed a high connectivity in the molecular wiring of DFTD (Figure 2F; Table S6). This suggested a critical involvement of central oncoprotein hubs consisting of STAT3, TRIM28, and others, which may be triggered by ERBB kinase action.

Molecular Dissection of ERBB-STAT3 Axis in DFTD

The proteomic tumor signatures revealed increased levels of STAT3, which can become activated by ERBB receptor tyrosine kinase signaling, as well as an enrichment of STAT3 target genes (Figures 2C, S2E, and S2F; Table S5). Due to the central roles of STAT3 in cancer and immunity (Villarino et al., 2017; Yu et al., 2014) and the remarkably conserved amino acid sequences of STAT3 between *H. sapiens* and *S. harrisi* (99.09%), we investigated the levels of expression and activation of STAT3 by western blot. STAT3 is activated by phosphorylation of residues Y705 and S727. Thus, we tested antibodies specific to these phosphorylated residues and found highly increased

(D) Volcano plot of genes with differentially methylated promoters between healthy and tumor biopsies (hypermethylated in tumor [blue], hypomethylated in tumor [red]).

(E) Boxplot of differentially methylated gene promoters for selected genes.

(F) Direct connection proteins network among the 987 tumor-modulated proteins, 166 tumor differentially methylated gene promoters, and *ERBB2* and *ERBB3* from the drug screen and RNA-seq. The direct network interactions were built with MetaCore based on protein-protein binding, transcriptional regulation, and phosphorylation interactions. Tumor signature proteins do not have a border, while methylation candidates are represented with a black border. The *ERBB2* candidate from the drug screen has a dashed black border. Of the 632 entities showing direct connectivity, only nodes with ten or more connections in MetaCore are displayed. The area of each entity is proportional to the number of connections within the network. Modulation on tumor versus healthy proteomics differential analysis, or healthy versus tumor for methylation, is colored from blue (down-modulated) to red (up-modulated).

Boxplot boundaries mark the first and third quartiles, whiskers extending to 1.5 interquartile range from the boundaries, with the median in the center. See also Figures S2 and S3 and Tables S5 and S6.

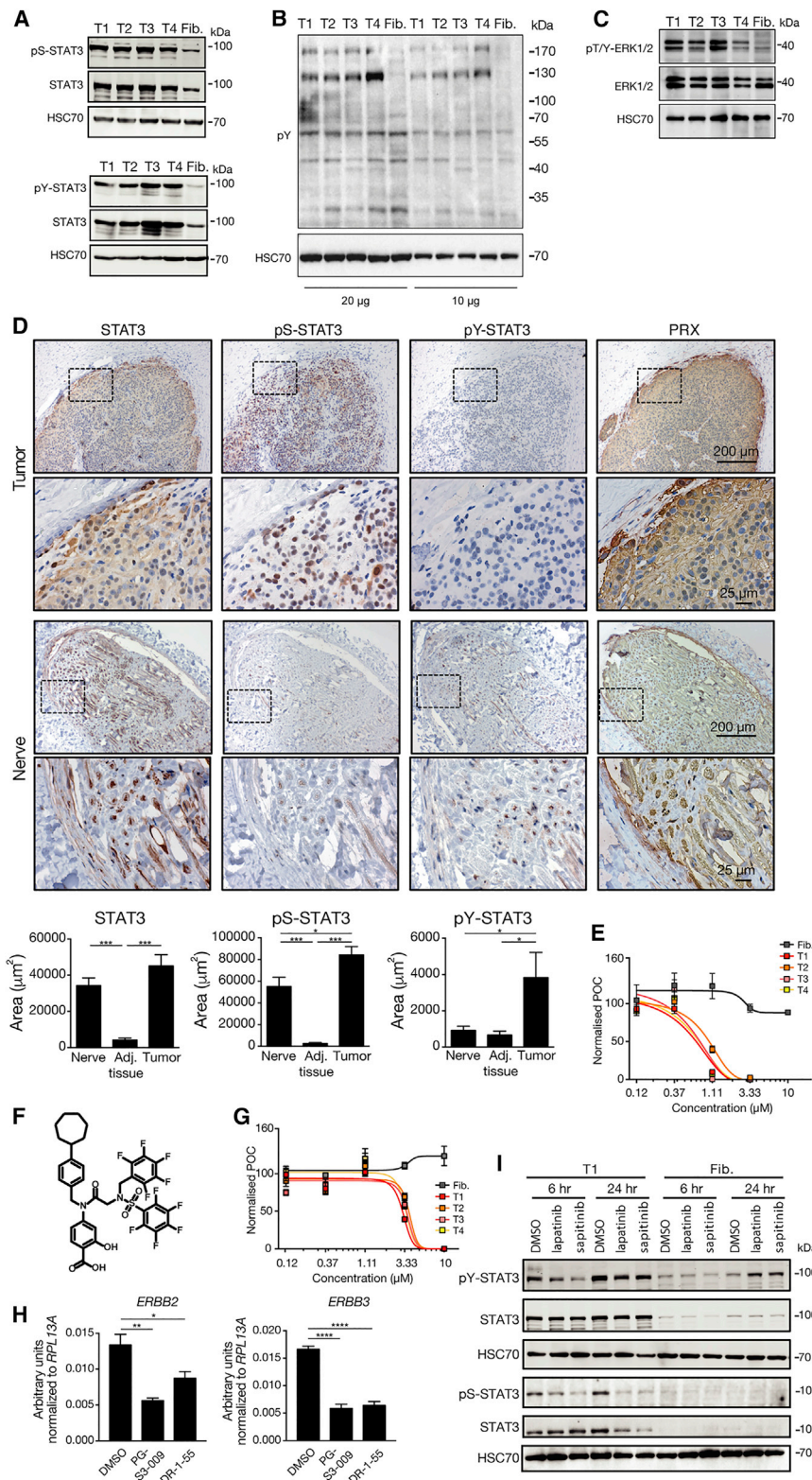


Figure 3. Molecular Dissection of ERBB-STAT3 Axis in DFTD

(A) Western blots of total STAT3, pS-STAT3, and pY-STAT3 of four DFTD cell lines (T1–T4) and fibroblasts (Fib.).

(B) Total protein phosphorylation immunoblots from lysates of four DFTD cell lines (T1–T4) and fibroblasts (Fib.) using a global anti-pY monoclonal antibody (4G10) in different input amounts.

(C) Western blots of total ERK1/2 and pT/Y-ERK1/2 of four DFTD cell lines (T1–T4) and fibroblasts (Fib.).

(D) Representative images of IHC for total STAT3, pS-STAT3, pY-STAT3, and PRX in primary tumor and peripheral nerve biopsy on serial consecutive sections and quantification of total STAT3, pS-STAT3, and pY-STAT3 in tumor biopsies, adjacent tissue, and peripheral nerve tissue. Dotted rectangles indicate magnified areas. Scale bars, 200 and 25 μm .

(E) Five-point dose-response curve of cell lines to the STAT3 inhibitor PG-S3-009 with DFTD and fibroblast cell lines.

(F) Structure of DR-1-55.

(G) Five-point dose-response curve of cell lines to the STAT3 inhibitor DR-1-55 with DFTD and fibroblast cell lines.

(H) DFTD cells treated with 2 μM PG-S3-009, 4 μM DR-1-55, or DMSO as control. Twenty-four hours after treatment, expression of *ERBB2* and *ERBB3* was measured by real-time PCR ($n = 3$ replicates).

(I) Western blots of total STAT3, pS-STAT3, and pY-STAT3 upon treatment with the ERBB inhibitors lapatinib (1 μM) and sapitinib (1 μM). Statistical significance was calculated by (D and H) unpaired t test. Graphs represent the mean \pm SEM. See also Figure S4.

phosphorylation compared with fibroblasts (Figure 3B), indicating increased tyrosine kinase signaling. ERBB family members activate RAS-RAF-MAPK/ERK, which in turn phosphorylates STAT3 at S727 (Chung et al., 1997). Indeed, we detected increased ERK1/2 phosphorylation in DFTD tumor cells (Figure 3C). STAT3 phosphorylation was corroborated by immunohistochemical stainings in primary tumor biopsies, showing higher levels of STAT3 phosphorylation at residues S727 and Y705 compared with adjacent non-tumor tissue or peripheral nerve tissue (Figure 3D).

Treatment with the covalent STAT3-selective inhibitor PG-S3-009 (Garg et al., 2017) resulted in DFTD tumor cell-specific killing (Figure 3E) and reduced STAT3 phosphorylation of residue Y705 (Figure S4C). In addition, treatment with DR-1-55, another selective STAT3 inhibitor

that covalently modifies a cysteine in the SH2 domain of STAT3 via a nucleophilic attack (Figures 3F, S4D, and S4E), reduced the expression of the STAT3 target gene *MMP2* (Figure S4F).

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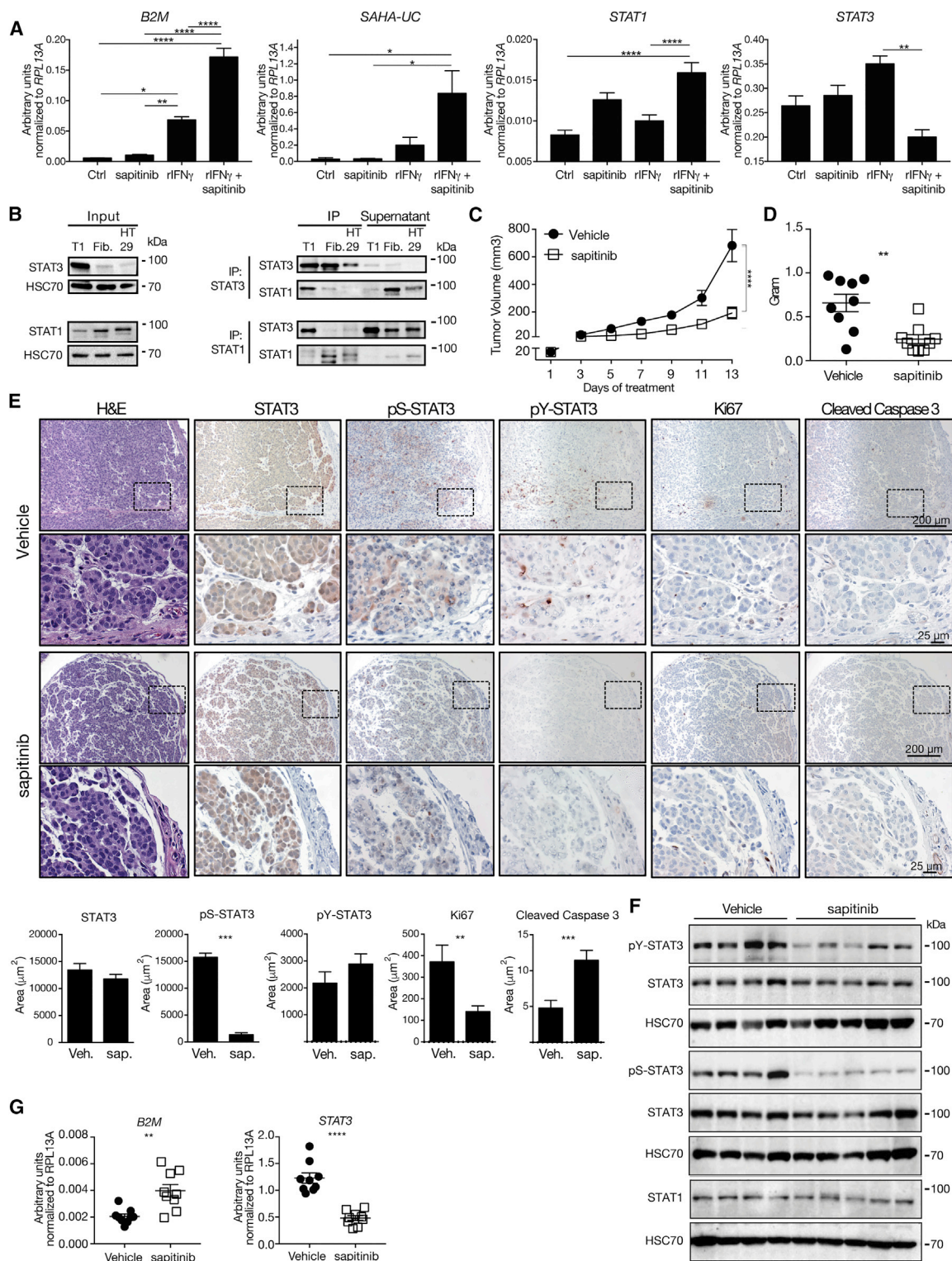


Figure 4. Blockade of ERBB Induces MHC Class I Gene Expression

(A) DFTD tumor cell line T1 was treated with recombinant interferon- γ (rIFN- γ) and/or 1 μ M sapitinib. Control cells were treated with solvents. Forty-eight hours after treatment, expression of *B2M*, *SAHA-UC*, *STAT1*, and *STAT3* were measured by real-time PCR (n = 3 replicates).

(B) Reciprocal co-immunoprecipitation of STAT3 and STAT1 followed by western blots for STAT3 and STAT1 in DFTD tumor cell line (T1), fibroblasts, and human HT29 colon cancer cells as control.

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We observed similar effects of DFTD tumor cell-specific killing with both STAT3 inhibitors (Figure 3G). Further, pharmacological inhibition of STAT3 by either PG-S3-009 or DR-1-55 led to reduced expression of *ERBB2* and *ERBB3* (Figure 3H). In line with this, DFTD tumor cells expressed higher levels of TRIM28 compared with fibroblasts (Figure S4G), and blockade of either ERBB signaling or STAT3 led to reduced transcription of *TRIM28* (Figure S4H). We also found abolished expression of the suppressor of cytokine signaling 1 (SOCS1) in DFTD tumor cells (Figures S4I and S4J), a known inhibitor of STAT3 activation (Song and Shuai, 1998). Intriguingly, treatment with ERBB inhibitors lapatinib and sapitinib inhibited serine and tyrosine phosphorylation of STAT3 (Figure 3I). In summary, our results suggest that DFTD tumor cells exhibit an ERBB-dependent constitutive activation of STAT3 in a positive feedforward loop.

Hyperactivated ERBB-STAT3 Reduces Expression of MHC Class I-Related Genes

Transmissibility of DFTD has been linked to reduced expression of MHC class I genes (Siddle and Kaufman, 2013), which prompted us to assess a potential link between hyperactive ERBB-STAT3 and MHC class I gene expression. To this end we stimulated DFTD tumor cells with recombinant autologous interferon γ (rIFN- γ). This induced the expression of *B2M* and *SAHA-UC*, one of Tasmanian devil's MHC class I genes, as shown previously (Figure 4A) (Siddle and Kaufman, 2013). Treatment with the ERBB inhibitor sapitinib alone was insufficient to induce MHC class I genes, which suggested an additional signaling requirement. Importantly, concomitant treatment of rIFN- γ with sapitinib amplified increased expression of *B2M* and *SAHA-UC* (Figures 4A and 4B). In addition, we observed increased expression of *STAT1* and a trend toward reduced *STAT3* expression upon treatment with rIFN- γ and sapitinib (Figures 4A and S5A). *B2M* and *SAHA-UC* are bona fide STAT1 target genes. Thus, we hypothesized that high levels of STAT3 may interfere with STAT1 transcriptional regulation. Indeed, reciprocal co-immunoprecipitation confirmed physical interaction between STAT3 and STAT1 (Figure 4B). This was corroborated by co-localization of STAT3 and STAT1 in DFTD tumor cells (Figures S5B and S5C).

To test the effects of ERBB inhibition *in vivo*, we transplanted DFTD tumor cells (T1) subcutaneously into the flanks of NOD/SCID gamma (NSG) mice and, 21 days later when tumors were palpable, started to administer 50 mg/kg sapitinib or vehicle once daily. DFTD xenografts in the control group proliferated rapidly after transplantation, while treatment with sapitinib effectively stalled tumor growth (Figures 4C, 4D, and S6A). No drug toxicity was observed as assessed by the serum concentration of the liver transaminases alanine aminotransferase and aspartate aminotransferase and the kidney parameter blood urea nitrogen (Figure S6B) and by histopathology (Figure S6C). The

observed anti-tumor effect of sapitinib was corroborated by histological analysis of tumor tissue for total STAT3, pS-STAT3, and pY-STAT3, as well as by staining for Ki67 and Cleaved Caspase 3 (Figure 4E). DFTD xenograft tumors from mice treated with sapitinib showed reduced STAT3 serine and tyrosine phosphorylation and increased expression of *B2M* (Figures 4F and 4G). Moreover, we also assessed the therapeutic effects of STAT3 inhibition by treating NSG mice 22 days after DFTD cell transplantation with 10 mg/kg DR-1-55 each second day. Similar to sapitinib, DR-1-55 resulted in reduced tumor growth (Figures 5A and 5B) without observed toxicity (Figures 5C and S6D). Treatment with DR-1-55 resulted in pronounced reduction of pY STAT3 (Figures 5D and 5E) and reduced MMP2 expression (Figure 5D). Further, DR-1-55 increased the expression of *B2M*, while *STAT3* expression was decreased (Figure 5F). In contrast to our results *in vitro* (Figure 4A), we noted that treatment of xenografted mice with sapitinib or DR-1-55 alone was sufficient to increase *B2M* (Figures 4G and 5F), possibly indicating additional host-derived signals *in vivo*. Taken together, our results derived from cell culture and xenograft models provide strong evidence for a central role of the ERBB-STAT3 axis in growth and immune evasion of DFTD.

DISCUSSION

ERBB signaling is influenced by ligand-induced activation and triggers downstream processes, such as context-dependent activation of transcriptional regulators, including members of the activator protein-1 family (AP-1, also bona fide STAT3 targets), ETS, and STAT3/5 transcription factors (Lemmon et al., 2014; Schneider and Yarden, 2016). ERBB family genes are frequently overexpressed, amplified or mutated in human solid cancers and ERBB family members are targets of clinical therapies (Appert-Collin et al., 2015; Bae and Schlessinger, 2010; Roskoski, 2014; Yarden and Pines, 2012). In Schwann cells, ERBB2/3 signaling regulates expansion and migration of progenitor cells as well as different functions in myelination and repair of axons (Corfas et al., 2004; Newbern and Birchmeier, 2010; Stassart et al., 2013). Moreover, we found increased expression of proteins associated with epithelial to mesenchymal transition (EMT) process in DFTD tumors that are also upregulated in Schwann cells upon nerve injury to support axon regeneration (Chen et al., 2007; Ferguson and Muir, 2000; Weiss et al., 2016). This included the EMT-inducing zinc-finger E-box-binding homeobox factor (ZEB2) (Comijn et al., 2001) and the STAT3 target MMP2, which enhances the degradation of extracellular matrix proteins and cancer cell invasion (Nistico et al., 2012). The regenerative properties of Schwann cells are highly linked to their plasticity whereby, upon nerve injury, they reversibly de-differentiate, acquire high motility,

(C and D) Tumor volume (C) and tumor weight (D) of DFTD tumor cell line T1 transplanted into NSG mice and treated with either vehicle or 50 mg/kg sapitinib once daily (bilateral tumors, n = 5 mice per group). One out of two representative experiments is shown.

(E) H&E and IHC analyses for total STAT3, pS-STAT3, pY-STAT3, Ki67, and Cleaved Caspase 3 of tumor tissues. Pictures shown are from contiguous sections. Dotted rectangles indicate magnified areas. Quantification of total STAT3, pS-STAT3, pY-STAT3, Ki67, and Cleaved Caspase 3. Scale bars, 200 and 25 μ m.

(F) Western blots for total STAT3, pY-STAT3, pS-STAT3, and STAT1 from representative xenograft tumors.

(G) Expression of *B2M* and *STAT3* by real-time PCR from xenograft tumor tissue.

Statistical significance was calculated by (A) one-way or (C) two-way ANOVA with Bonferroni correction or (D, E, and G) unpaired t test. Graphs represent the mean \pm SEM. See also Figures S5 and S6.

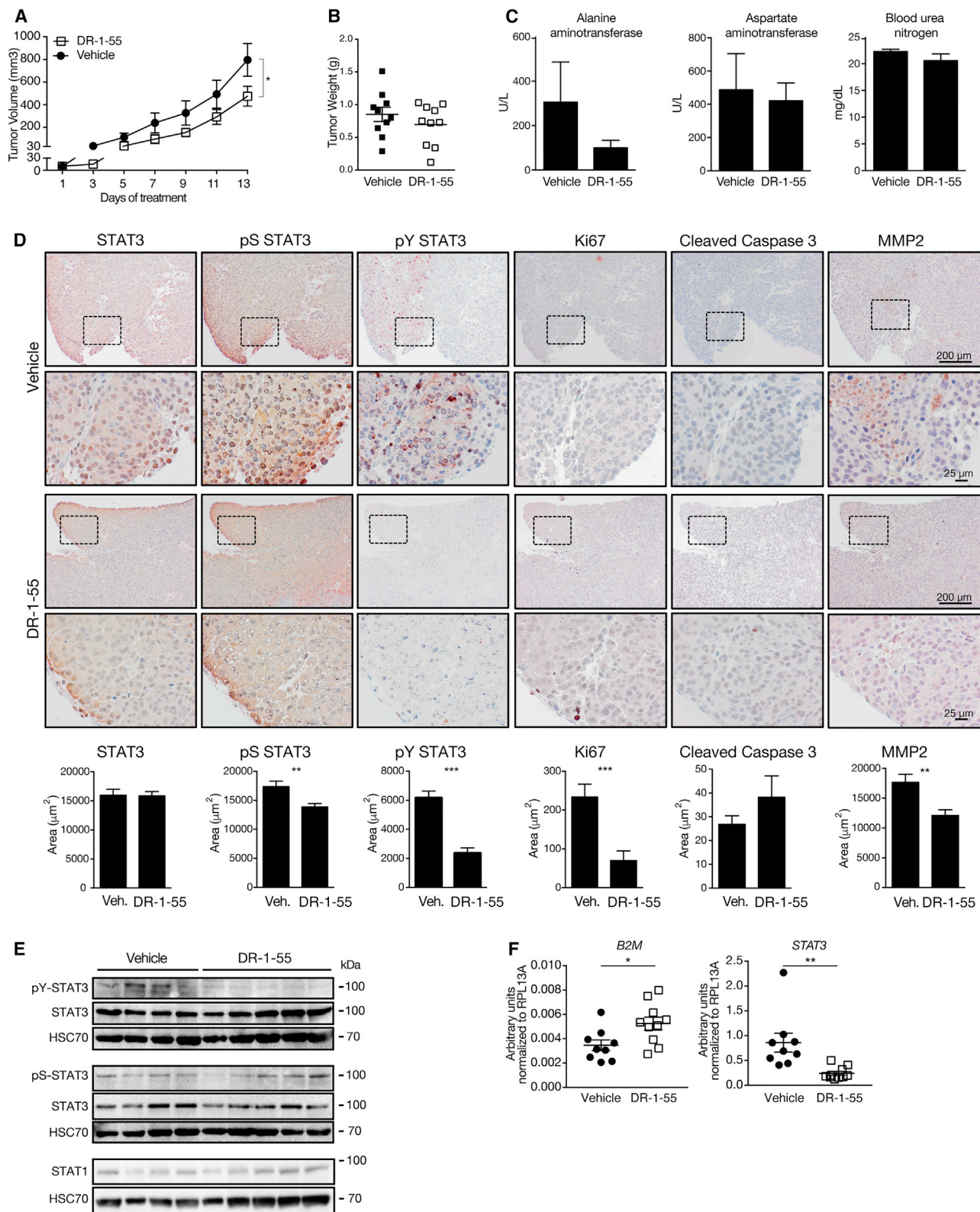


Figure 5. Xenograft Model with STAT3 Inhibitor DR-1-55

(A and B) Tumor volume (A) and tumor weight (B) of NSG mice transplanted with DFTD tumor cell line T1 and, 22 days after transplantation, treated with either vehicle or 10 mg/kg DR-1-55 each day (bilateral tumors, $n = 5$ mice per group).

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and guide the growth of the damaged axons (Kim et al., 2013). Thus, the identification of hyperactivated ERBB-STAT3 signaling in DFTD may suggest aberrant regulation of the Schwann cell-intrinsic repair program (Arthur-Farraj et al., 2017). Of note, DFT2 tumors are pathologically similar but stain-negative for the Schwann cell marker PRX (Pye et al., 2016) and display copy-number gains for PDGFR (Stammnitz et al., 2018). It will be interesting to investigate the involvement of similar driver tyrosine kinase-STAT3 pathways blocking MHC class I.

A major enigmatic question of DFTD concerns the molecular properties that are required for transmission between individuals to explain the lack of rejection. Changes in MHC class I expression and diversity have been described in transmissible tumors of devils and dogs (Belov, 2011). Of note, recent immunotherapy trials with MHC-induced DFTD cells showed immunogenicity *in vivo* (Tovar et al., 2017). Our data indicate that targeting of the hyperactivated ERBB-STAT3 axis re-establishes the expression of MHC class I, thus facilitating MHC-mediated tumor immunosurveillance in Tasmanian devils (Figure 6). Promoters of interferon-stimulated MHC class I-related genes are targets for STAT1, while STAT3 is known to interfere with transcription by sequestering STAT1 in the cytoplasm through heterodimerization (Friedrich et al., 2017; Nivarthi et al., 2016; Stancato et al., 1996). We hypothesize that the endogenous tonic interferon-STAT-MHC class I axis in DFTD is disrupted due to high STAT3 action promoting cancer cell proliferation, survival, and invasion. Interestingly, STAT3 overexpression was previously proposed as one of several candidate mechanisms of immune evasion in transplantation and transmissible tumors (Fassati and Mitchison, 2010). Next to the potential interference of STAT3 with expression of MHC-I, there may also be a role for TRIM28, which we found to be overexpressed in DFTD and which acts as a negative regulator of interferon signaling (Liang et al., 2011). Of note, reduced expression of MHC-I is expected to impair rejection by CD8⁺ T cells only but not by natural killer cells, which recognize cells in the absence of MHC-I. Natural killer cells of Tasmanian devils, however, lack the ability to directly recognize DFTD cells (Brown et al., 2011, 2016; Peel and Belov, 2018), which is likely to contribute to the lack of rejection of horizontally transmitted tumor cells. Together, our findings may bear relevance also for other transmissible cancers in higher organisms including dogs, whose transmissible tumor lacks B2M and shows low MHC class I surface expression (Cohen et al., 1984; Murgia et al., 2006).

We wish to acknowledge that molecular investigations of non-model organisms can be notoriously hampered by imperfect genome annotations, orthology inferences, and lack of reagents. Several layers of controls were included in this integrative and unbiased systems-biology approach. This was complemented by comparative pathology for which we exploited the high degree of conservation of key oncoproteins driving human cancer,

cross-validating a similar driver oncogene scenario in Tasmanian devils.

Our data indicate a positive feedback loop between ERBB receptor tyrosine kinases and STAT3. The underlying mechanism of maintained STAT3 activation, the role of specific STAT3-activating kinases and the apparent lack of negative regulation requires further investigation. Notably, STAT3 S727 phosphorylation is often constitutive in cancer cells associated with hyperactive RAS-RAF signaling, and modulates ATP production in mitochondria through the respiratory chain (Gough et al., 2009; Wegrzyn et al., 2009). Serine phosphorylation of STAT3 can be catalyzed by many kinases including p38 MAPK, ERK, JNK, CDKs, mTOR, or PKC in response to growth factors such as EGF, PDGF, or Insulin (Chung et al., 1997). Sunitinib efficiently blocked S727 phosphorylation of STAT3 in DFTD cells, while STAT3 inhibition tended to show stronger effects on phosphorylation at residue Y705. Thus, an inhibition of growth factor signaling through sunitinib combined with STAT3 inhibition could be beneficial to synergistically block survival, proliferation, and metabolic functions of STAT3. Not mutually exclusively, the involved processes may include recently detected copy gains of ERBB3 (Hayes et al., 2017; Taylor et al., 2017) leading to enhanced ERBB2-ERBB3 heterodimer activity, the secretion of ligands of the ERBB family, and/or blunted negative regulation by phosphatases or the SOCS pathway.

Histopathologically, DFTD presents as undifferentiated pleomorphic tumor cells with fibrous stroma and occasional infiltrating immune cells (Howson et al., 2014; Loh et al., 2006a, 2006b). It is, thus, conceivable that the complex underlying interactions within the tumor microenvironment may contribute to the activation of the ERBB-STAT3 axis. The integrated and unbiased systems-level analysis presented here is expected to provide a critical foundation for further investigations and raises general questions about the tumor biology and transmissibility of such cancers in other species. The implicated canonical cancer signatures apparently do not suffice to give rise to transmissible cancers in humans. Thus, it should be emphasized that the occurrence of transmissible cancers in mammals is likely to depend on a complex combination of molecular as well as non-molecular context-dependent features, such as aggressive behavior, tissue wounding, and population dynamics (Epstein et al., 2016).

Blocking the ERBB-STAT3 axis may present a promising drug target whose interference arrests cancer cells and at the same time leads to increased tumor surveillance through re-expression of MHC class I (Garrido et al., 2016). While pharmacological treatments come with inherent logistic limitations for wildlife diseases, this rationalized therapeutic strategy—possibly in combination with a vaccine against DFTD and/or immunotherapeutic interventions—offers a much-needed expansion of the so far limited measures to preserve the Tasmanian devil from extinction.

(C) Serum concentration of alanine aminotransferase, aspartate aminotransferase, and blood urea nitrogen.

(D) Tumor tissue immunohistochemically stained and quantified for total STAT3, pS-STAT3, pY-STAT3, Ki67, Cleaved Caspase 3, and MMP2. Pictures shown are from contiguous sections. Dotted rectangles indicate magnified areas. Scale bars, 200 and 25 μ m.

(E) Western blots for total STAT3, pY-STAT3, pS-STAT3, and STAT1 from representative xenograft tumors.

(F) Expression of *B2M* and *STAT3* by real-time PCR from xenograft tumor tissue.

Statistical significance was calculated by (A) two-way ANOVA with Bonferroni correction or (B–D and F) unpaired t test. Graphs represent the mean \pm SEM. See also Figure S6.

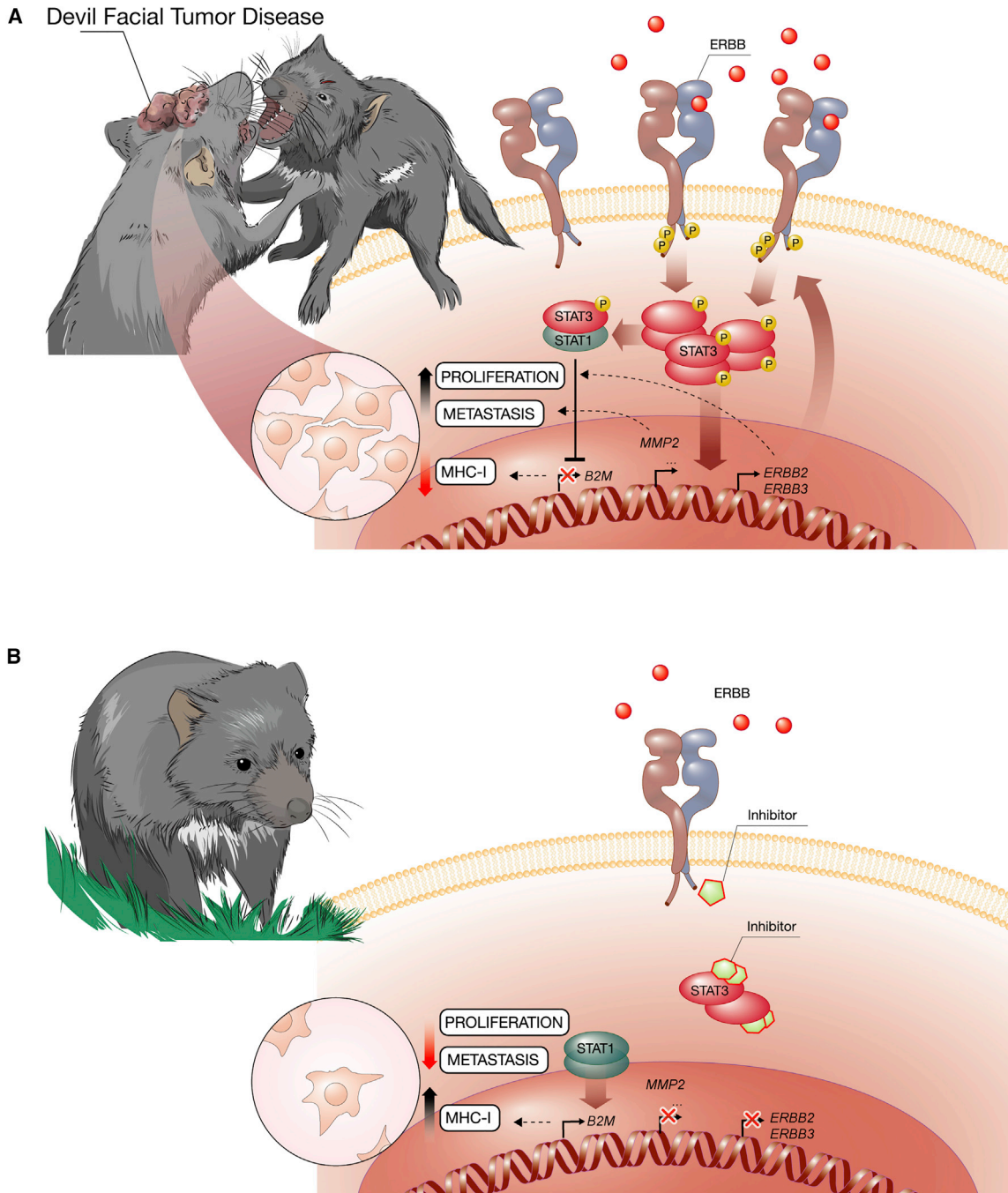


Figure 6. Working Model for the Impact of the ERBB-STAT3 Axis in DFTD

(A) Aggressive social interactions in the highly inbred population of Tasmanian devils enabled the rapid spread of DFTD with fatal consequences. The hyperactive ERBB-STAT3 axis induces the expression of downstream metastasis-related genes (i.e., *MMP2*) while suppressing the expression of MHC class I genes (i.e., *B2M*). We hypothesize that highly abundant phosphorylated STAT3 protein traps unphosphorylated STAT1 proteins in heterodimers, thereby preventing the transcriptional regulation of STAT1 downstream target genes such as *B2M*. This may contribute to immune evasion and the known lack of tumor rejection upon horizontal transmission.

(B) Interference with the ERBB-STAT3 axis by using either ERBB inhibitors or STAT3 inhibitors results in killing of DFTD tumor cells.

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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SUPPLEMENTAL INFORMATION

Supplemental Information includes six figures, six tables, and one video and can be found with this article online at <https://doi.org/10.1016/j.ccell.2018.11.018>.

A video abstract is available at <https://doi.org/10.1016/j.ccell.2018.11.018#mmc8>.

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AUTHOR CONTRIBUTIONS

L.K., B.W., A.O., B.A., B.V., K.P., A.L., A.R., J.K., and E.D.d.A. conducted the experiments and analyzed the data. A.P., P.M., K.K., H.B., C.B., S.K., K.L.B., R.M., and A.B. analyzed the data. D.A.R., J.P., G.T., S.A., P.T.G., H.V.S., G.M.W., and E.P.M. provided the material. L.K., A.P., B.V., R.M., and A.B. wrote the manuscript. A.B. conceived and supervised the study.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
anti-HSC70 monoclonal mouse	Santa Cruz	Cat#sc-7298
anti-EGFR monoclonal mouse	Santa Cruz	Cat#sc-373746
anti-EGFR monoclonal mouse	BD Biosciences	Cat#610016
anti-phospho-EGFR (Y1068) monoclonal rabbit	Cell Signaling Technology	Cat#3777
anti-HER/ErbB2 monoclonal mouse	Santa Cruz	Cat#sc-7301
anti-HER2/ErbB2 monoclonal rabbit	Cell Signaling Technology	Cat#4290
anti-phospho-HER2/ErbB2 (Y1221/1222) monoclonal rabbit	Cell Signaling Technology	Cat#2243
anti-HER3/ErbB3 monoclonal rabbit	Cell Signaling Technology	Cat#12708
anti-phospho-HER3/ErbB3 (Y1289) monoclonal rabbit	Cell Signaling Technology	Cat#2842
anti-Periaxin/PRX	Sigma Aldrich	Cat#HPA001868
anti-STAT3 monoclonal mouse	BD Biosciences	Cat#610189
anti-STAT3 monoclonal mouse	Cell Signaling Technology	Cat#9139
anti-phospho-STAT3 (Y705) polyclonal rabbit	Cell Signaling Technology	Cat#9131
anti-phospho-STAT3 (Y705) polyclonal rabbit	Cell Signaling Technology	Cat#9134
anti-pY (4G10)	Merck Millipore	Cat#05-321
anti-ERK1/2 monoclonal rabbit	Cell Signaling Technology	Cat#4695
anti-phospho-ERK1/2 (T202/Y204) monoclonal rabbit	Cell Signaling Technology	Cat#4370
anti-STAT1	Cell Signaling Technology	Cat#9172
anti-Kap-1/TRIM28 rabbit polyclonal	Merck Millipore	Cat#ABE1859
anti-EGFL8 rabbit polyclonal	Abcam	Cat#ab58650
anti-SOCS1	Cell Signaling Technology	Cat#3950
anti-B2M	Siddle et al., 2013	N/A
anti-phospho-STAT1 (Y701) (58D6) monoclonal rabbit	Cell Signaling Technology	Cat#9167
Ki67 (NCL-Ki67p)	Leica Biosystems	Cat#Ki67-MM1-L-CE
Cleaved Caspase 3 (Asp175)	Cell Signaling Technology	Cat#9661S
ECL anti rabbit IgG (NA934V)	GE Healthcare	Cat#NA934
anti-mouse (NA931) HRP	GE Healthcare	Cat#NA931
anti-MMP2 polyclonal rabbit	Abcam	Cat#ab37150
Biological Samples		
Devil facial tumor disease 1 (DFT1) biopsy: 08/0038, Mid Tumour T1	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/0195, Mid Tumour T2	This paper	N/A
Tasmanian devil spleen biopsy: 08/0195, Spleen	This paper	N/A
Tasmanian devil skin biopsy: 08/0195, Skin	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/0289, Early Tumour T2	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/0289, Mid Tumour T1	This paper	N/A
Tasmanian devil spleen biopsy: 08/0289, Spleen	This paper	N/A
Tasmanian devil skin biopsy: 08/0289, Skin	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/0590, Mid Tumour T2	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/0590, Late Tumour T1	This paper	N/A

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REAGENT or RESOURCE	SOURCE	IDENTIFIER
Tasmanian devil spleen biopsy: 08/0590, Spleen	This paper	N/A
Tasmanian devil skin biopsy: 08/0590, Skin	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/1818, Early Tumour T3	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/1818, Mid Tumour T2	This paper	N/A
Devil facial tumor disease 1 (DFT1) biopsy: 08/1818, Late Tumour T1	This paper	N/A
Tasmanian devil spleen biopsy: 08/1818, Spleen	This paper	N/A
Tasmanian devil skin biopsy: 08/1818, Skin	This paper	N/A
Tasmanian devil nerve biopsy: Christine	This paper	N/A
Tasmanian devil nerve biopsy: Curley Shirley	This paper	N/A
Chemicals, Peptides, and Recombinant Proteins		
Recombinant devil interferon gamma (rIFN γ)	Siddle et al., 2013	N/A
Kinase Inhibitor Drug Library	TargetMol and Cayman Chemical	Cat# L1600 and Cat# 10505 respectively
2K Drug Library	Sdelci et al., 2016	N/A
Sapitinib	Adooq Bioscience	Cat#A10116
Lapatinib	Adooq Bioscience	Cat#A10514
PG-S3-009	Garg et al., 2017	N/A
DR-1-55	This paper	N/A
Deposited Data		
Proteomic Data	This paper	PRoteomics IDEntification database accession number 1-20180126-165173
DNA Methylation Data	This paper	Gene Expression Omnibus database accession number GSE108160
RNA Seq Data	This paper	Gene Expression Omnibus database accession number GSE108107
Experimental Models: Cell Lines		
Devil facial tumor disease 1 (DFT1) cell line: 06/2887, Strain 1	Deakin et al., 2012	N/A
Devil facial tumor disease 1 (DFT1) cell line: 05/2569, Strain 2	Deakin et al., 2012	N/A
Devil facial tumor disease 1 (DFT1) cell line: 06/0368, Strain 3	This paper	N/A
Devil facial tumor disease 1 (DFT1) cell line: 07/0192, Strain 4	Ingles and Deakin, 2015	N/A
Tasmanian devil fibroblast cell line	Murchison et al., 2012	N/A
Experimental Models: Organisms/Strains		
NOD scid gamma (NSG) mice	The Jackson Laboratory	Cat#005557
Oligonucleotides		
<i>RPL13A</i> forward: 5'-CCCCACAAGACCAAGCGAGGC-3'	Siddle et al., 2013	N/A
<i>RPL13A</i> reverse: 5'-ACAGCCTGGTATTTCAGCCAACC-3'	Siddle et al., 2013	N/A
<i>EGFR</i> forward: 5'-GCAGATAGCCAAGGGTATGAGTTACC-3'	This paper	N/A
<i>EGFR</i> reverse: 5'-TTTTGCCAGCCCAAATCTGT-3'	This paper	N/A
<i>ERBB2</i> forward: 5'-GGAACCCAAGTGTGCACAGG-3'	This paper	N/A
<i>ERBB2</i> reverse: 5'-TGGCATCAGCAGGCAGGTA-3'	This paper	N/A
<i>ERBB3</i> forward: 5'-TACATGGTCATGGTTAAGTGCTGG-3'	This paper	N/A
<i>ERBB3</i> reverse: 5'-GGTGGATCTCGGGCCATT-3'	This paper	N/A
<i>MHC-1 (SAHA-UC; KY194695)</i> forward: 5'-CCGTGGGC TACGTGGACGATCAGC-3'	Siddle et al., 2013	N/A
<i>MHC-1 (SAHA-UC; KY194695)</i> reverse: 5'-GTCGTAGG CGAACTGAAG-3'	Siddle et al., 2013	N/A
<i>B2M</i> forward: 5'-TGTGCATCCTCCCTACCTGGAGG-3'	Siddle et al., 2013	N/A

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REAGENT or RESOURCE	SOURCE	IDENTIFIER
<i>B2M</i> reverse: 5'-CATTGTTGAAAGACAGATCGGACCGC-3'	Siddle et al., 2013	N/A
<i>STAT1</i> forward: 5'-GGAAAAGCAAGACTGGGACTATGC-3'	This paper	N/A
<i>STAT1</i> reverse: 5'-GCGGCTATAGTGCTCATCCAA-3'	This paper	N/A
<i>STAT3</i> forward: 5'-GGAAGCTGACCCAGGTAGTGC-3'	This paper	N/A
<i>STAT3</i> reverse: 5'-CGGCAGGTCAATGGTATTGC-3'	This paper	N/A
<i>TRIM28</i> forward: 5'-AAGTGGCGCACCTCCATCC-3'	This paper	N/A
<i>TRIM28</i> reverse: 5'-CCCCTTGTGAGCTCCTT-3'	This paper	N/A
<i>EGF</i> forward: 5'-TATGACTGGTACCGGCCCTG-3'	This paper	N/A
<i>EGF</i> reverse: 5'-TGCCAGCATTAGCTACCACTTGT-3'	This paper	N/A
<i>NRG1</i> forward: 5'-CAGATACTCGTGCAAGTGCCC-3'	This paper	N/A
<i>NRG1</i> reverse: 5'-TGCAGATGCCAGTGATGGTC-3'	This paper	N/A
<i>EGFL8</i> forward: 5'-TCCATACAGCAAGGGAGTTGC-3'	This paper	N/A
<i>EGFL8</i> reverse: 5'-ATCCGCTGACCTGCACACA-3'	This paper	N/A
<i>HBEGF</i> forward: 5'-GGCTGCTCATGTTCAGGTACC-3'	This paper	N/A
<i>HBEGF</i> reverse: 5'-TTTCCATCAGTGGGCAATAGG-3'	This paper	N/A
<i>MMP2</i> forward: 5'-CAGACAAAGAGTTGGCTGTACAATACC-3'	This paper	N/A
<i>MMP2</i> reverse: 5'-CCTTCAGCACAAACAGGTTGC-3'	This paper	N/A

CONTACT FOR REAGENT AND RESOURCE SHARING

Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Andreas Bergthaler (abergthaler@cemm.oew.ac.at).

EXPERIMENTAL MODEL AND SUBJECT DETAILS**Cell Culture and Tissue Biopsies**

DFTD cells were grown from primary cell cultures derived from fine needle aspirates that have been collected from the wild (see Table S1) (Deakin et al., 2012; Ingles and Deakin, 2015; Murchison et al., 2012). Devil Facial Tumor cell lines T1-4 were grown in RPMI (Gibco 21875-034) supplemented with 10% Fetal bovine serum (PAA A15-101), 1% Pen Strep Glutamine (Gibco 10378-016) and 50 μ M 2-Mercaptoethanol (Sigma M-3148). Fibroblasts were grown in Advanced DMEM (Gibco 12491-015) supplemented with 10% Fetal bovine serum (PAA A15-101), 1% Pen Strep Glutamine (Gibco 10378-016). Cells were grown at 35°C in 5% CO₂ and lifted with either PBS containing 1 mM EDTA or 0.05% Trypsin-EDTA (Gibco 25300-054). CHO supernatants containing recombinant devil interferon gamma (rIFN γ) was used 1:3 diluted (Siddle et al., 2013). Control cells received supernatants of wild type CHO cells. The ERBB inhibitors sapitinib (Adooq Bioscience, Cat# A10116) and lapatinib (Adooq Bioscience, Cat# A10514) were dissolved in DMSO and used at the indicated concentrations. The STAT3 inhibitors PG-S3-009 and DR-1-55 were dissolved in DMSO and used at the indicated concentrations.

Primary biopsies from Tasmanian devils were obtained from the Department of Primary Industries, Parks, Water and Environment (DPIPWE) (Tasmanian Government, Australia). All animal procedures were performed under a Standard Operating Procedure approved by the Secretary, Wildlife Management Branch, Tasmanian Government Department of Primary Industries, Parks, Water and the Environment (DPIPWE), in agreement with the DPIPWE Animal Ethics Committee. Biopsies were received frozen on dry ice and stored in liquid nitrogen until processed or embedded in paraffin blocks (Table S1).

Mouse Xenograft Studies

NOD scid gamma (NSG) mice (JAX # 005557) were maintained under pathogen-free conditions at the University of Veterinary Medicine, Vienna. Mice were at the age of 8-14 weeks at the time of cell implantation. Mice were kept at 12 hr/12 hr light cycle and received standard food and water *ad libidum*. All animal experiments were approved by the institutional ethics and animal welfare committee of the University of Veterinary Medicine, Vienna and the national authority according to §§ 26ff. of Animal Experiments Act, TVG 2012 BMWFW-68.205/0130-WF/V/3b/2016. The experimental design and number of mice assigned to each treatment arm were based on prior experience with similar models and provided sufficient statistical power to discern significant differences.

For sapitinib treatment mice were matched according to initial tumor size and randomized to treatment with sapitinib or vehicle (ddH₂O + 1% Tween80). No mice were excluded from the analysis. Mice were implanted subcutaneously in both flanks with 1x10⁶ Devil Facial Tumor cell line 1 (Table S1) in 100 μ l PBS, using a 27G needle. Tumor growth was measured every second day using Vernier calipers for the duration of the experiment and tumor volumes were calculated with the following formula: tumor

volume = (length × width²)/2. Treatment was initiated when the average tumor volume reached approximately 3 mm³ and experiments were terminated once tumor volume in control group reached 1 cm³. 1 out of 10 transplanted tumors did not grow in each group of mice (n=5). Tumors were resected and used for analysis of tumor weight, immunohistochemistry, real-time PCR and immunoblotting as described.

In the case of DR-1-55 treatment, the mice were randomly divided into two groups and, once the tumor volume reached approximately 3 mm³, the mice were treated each second day with DR-1-55 (10 mg/kg) or vehicle (1% DMSO, 5% Tween80, 50% PEG400 in PBS) until tumors in control group reached approximately 1 cm³.

METHODS DETAILS

Drug Viability Screen

We used a combined library of selected 1847 drugs (Sdelci et al., 2016) and 684 kinase inhibitors (Targetmol catalog no. L1600 and Cayman Chemical item no 10505), which were transferred onto 384-well plates using an acoustic liquid handler (Echo, Labcyte). 5000 cells per well of DFTD cell lines T1-4, and 2500 cells per well of fibroblasts were added on top of the drugs (50 nl in DMSO) with a dispenser (Thermo Fisher Scientific) to a total of 50 μl/well and incubated at 37°C. Cell viability was measured after 72 hr using the CellTiter-Glo® Luminescent Cell Viability Assay (Promega G7573) in a multilabel plate reader (EnVision, PerkinElmer). Initially, all drugs were tested on DFTD cell line #1 at a single dose (typically 10 μM). 434 drug hits with effects on cell viability were subsequently tested in a 4-dose response in triplicate wells of DFTD cell lines 1 to 4 as well as fibroblasts. Drug candidates were selected based on the difference between the Area Under the Curve (AUC) of each tumor cell line to the control fibroblast higher than 50. The addition of 2 standard deviations of the mean AUC of each strain should also result in a lower value than the subtraction of two standard deviations from the fibroblast AUC (Equation 1). This yielded 69 candidates that killed at least one DFTD cell line. Out of those, 41 drug candidates killed at least three out of four tumor cell lines but not fibroblasts. These 41 drugs were re-tested as 8-point dose-response in triplicates on the aforementioned five cell lines and cell viability was assessed by CellTiter-Glo as described previously. The STAT3 inhibitors PG-S3-009 (Garg et al., 2017) and DR-1-55 were tested separately as 5-point dose-response curves in triplicates. The percentage of control (POC) was calculated by using linear regression for each plate individually, setting the mean signal of the negative control wells (DMSO) to 100% and the mean signal of the positive control wells to 0%.

$$\overline{AUC}_{\text{fibroblast}} - 2Sd(\overline{AUC}_{\text{fibroblast}}) > \overline{AUC}_{\text{tumor strains}} - 2Sd(\overline{AUC}_{\text{tumor strains}}) \quad (\text{Equation 1})$$

Equation 1: Equation identifying significant drugs killing the DFTD tumor strains compared to the fibroblast. AUC stands for area under the curve. Sd stands for standard deviation.

DNA and RNA Isolation

For DNA methylation analysis, approximately 20 mg of primary Tasmanian devil tissue was isolated and homogenised using the Tissue Lyser II (Qiagen, Hilden, Germany, 12 x 30 sec, 30 Hz). DNA and RNA were isolated using the AllPrep DNA/RNA Mini Kit (Qiagen, 80204), according to the manufacturer's instructions. For expression analysis from cell lines, total RNA was isolated from approximately 1x10⁶ cells using QIAzol lysis reagent according to the manufacturer's instructions (Qiagen).

Real-Time PCR

Isolated RNA was reverse transcribed into cDNA using the First Strand cDNA Synthesis Kit (Fermentas) according to the manufacturer's instructions. Subsequent gene expression was then analysed using SYBR Select Master Mix (Applied Biosystems; 4472908). We designed and used the following gene-specific primers: 5'- CCCCACAAGACCAAGCGAGGC -3' and 5'- ACAGCCTGGTATTC CAGCCAACC -3' for *RPL13A* (Siddle et al., 2013), 5'- GCAGATAGCCAAGGGTATGAGTTACC-3' and 5'- TTTTGCCAGCCCA AAATCTGT-3' for *EGFR*, 5'- GGAACCCAAGTGTGCACAGG-3' and 5'- TGGCATCAGCAGGCAGGTA-3' for *ERBB2*, and 5'- TACATG GTCATGGTTAAGTGCTGG-3' and 5'- GGTGGATCTCGGGCCATT-3' for *ERBB3*, 5'- CCGTGGGCTACGTGGACGATCAGC -3' and 5'- GTCGTAGGCGAACTGAAG -3' for *MHC-1* (SAHA-UC; KY194695) (Siddle et al., 2013), 5'-TGTGCATCCTCCCTACCTGGAGG -3' and 5'-CATTGTTGAAAGACAGATCGGACCGC -3' for *B2M* (Siddle et al., 2013), 5'- GGAAAAGCAAGACTGGGACTATGC -3' and 5'- GCGGCTATAGTGCTCATCAA -3' for *STAT1*, 5'- GGAAGCTGACCCAGGTAGTGC -3' and 5'- CGGCAGGTCAATGGTATTGC -3' for *STAT3*, 5'-AAGTGCGCACCTCCATCC-3' and 5'-CCCCTGTTGAGCTCCTT-3' for *TRIM28*, 5'-TATGACTGGTACCGGCC CTG-3' and 5'-TGCCAGCATTAGCTACCACTTGT-3' for *EGF*, 5'-CAGATACTCGTGCAAGTGCCC-3' and 5'-TGCAGATGCCAGT GATGGTC-3' for *NRG1*, 5'-TCCATACAGCAAGGGAGTTTGC-3' and 5'-ATCCGCTGACCTGCACACA-3' for *EGFL8*, 5'-GGCTGCTC ATGTTCCAGGTACC-3' and 5'-TTCCATCAGTGGGCAATAGG-3' for *HBEGF*, 5'-CAGACAAAGAGTTGGCTGTACAATACC-3' and 5'-CCTTCAGCACAAACAGTTGC-3' for *MMP2*. Designed forward primers span exon-exon junctions where possible.

Western Blotting

Approximately 5x10⁶ cells from DFTD strains 1-4 and 2.5x10⁶ Tasmanian devil fibroblasts were pelleted (260 g, 5 min, 4°C; Table S1), washed three times in cold PBS, snap frozen in liquid nitrogen and frozen at -80°C until processed. Sample preparation and Western blotting was performed using standard techniques. Nitrocellulose membranes (0.45 μm Amersham Protran 10600002, GE Healthcare, Buckinghamshire, UK) were incubated with the following antibodies in the dilution as indicated (see also Table S1): specific

anti-phospho-STAT3 (Y705) polyclonal rabbit (1:1000; 9131; Cell Signaling Technology, Cambridge, UK), anti-STAT3 monoclonal mouse (1:1000; 610189; BD Biosciences, Franklin Lakes, NJ, USA) or (9139; Cell Signaling; 1:1000), anti-phospho-STAT3 (S727) polyclonal rabbit (1:1000; 9134; Cell Signaling Technology, Cambridge, UK), anti-STAT1 (rabbit; 9172; Cell Signaling; 1:1000), anti-phospho-STAT1 (Y701) monoclonal rabbit (1:1000; 9167; Cell Signaling Technology), anti-EGFL8 polyclonal rabbit (1:1000; ab58650; Abcam, Cambridge, UK), anti-phospho-EGFR (Y1068) monoclonal rabbit (1:1000; 3777; Cell Signaling Technology), anti-EGFR monoclonal rabbit (1:1000; sc-373746; Santa Cruz, Dallas, TX, USA), anti-phospho-HER2/ERBB2 (Y1221/1222) monoclonal rabbit (1:1000; 2243; Cell Signaling Technology), anti-ERBB2 monoclonal mouse (1:1000, sc-7301; Santa Cruz Biotechnology), anti-phospho-HER3/ERBB3 (Y1289) monoclonal rabbit (1:1000; 2842; Cell Signaling Technology), anti-HER3/ERBB3 monoclonal rabbit (1:1000; 12708; Cell Signaling Technology), anti-HSC70 monoclonal mouse (1:1000; sc-7298; Santa Cruz), anti-TRIM28 polyclonal rabbit (1:1000; ABE1859; Millipore); anti-ERK1/2 monoclonal rabbit (1:1000; 4695; Cell Signaling Technology, Cambridge, UK), anti-phospho-ERK1/2 (T202/Y204) monoclonal rabbit (1:1000; 4370; Cell Signaling Technology, Cambridge, UK), anti-pY (4G10; Merck Millipore; 1:1000), anti-SOCS (1:1000; 3950; Cell Signaling Technology, Cambridge, UK), anti-B2M (1:1000; (Siddle et al., 2013)), ECL anti-rabbit IgG (NA934V) or anti-mouse (NA931) HRP (1:10000; GE Healthcare, Buckinghamshire, UK).

Immunoprecipitation

Cells were lysed in HE buffer (10 mM HEPES (pH 7.35), 1 mM EDTA) supplemented with protease inhibitors using a dounce tissue grinder. Human HT-29 cells (ATCC HTB-38) served as control. For immunoprecipitation, 1 mg protein lysate was incubated with 2 μ g of anti-STAT3 (9139; Cell Signaling) or anti-STAT1 (9172; Cell Signaling) at 4°C overnight and immunoprecipitated with 25 μ l Dynabeads Protein G (10004D; Thermo Fisher Scientific, Waltham, MA, USA) for 2 hr at 4°C. Beads were washed 3x with HE buffer and samples were eluted in 40 μ l Laemmli buffer at 95°C for 10 min.

Histology

DFTD tissues of diseased animals (328T1 and 463T1) were fixed in 10% neutral buffered formalin and paraffin-embedded. 2 μ m FFPE consecutive tumor sections were stained with Hematoxylin (Merck, Darmstadt, Germany) and Eosin G (Carl Roth). For immunohistochemical stainings, heat-mediated antigen retrieval was performed in citrate buffer at pH 6.0 (S1699; Dako, Agilent, Santa Clara, CA, USA), EDTA at pH 8.0 or TE at pH 9.0. Sections were stained with antibodies specific to STAT3 (1:200; pH 6; 9139; Cell Signaling Technology), phospho-STAT3 (S727) monoclonal rabbit (1:80; pH 9; 9134; Cell Signaling Technology); EGFR monoclonal mouse (1:300; 610016; pH 9; BD Biosciences), HER2/ERBB2 monoclonal rabbit (1:200, 4290; Cell Signaling Technology), HER3/ERBB3 monoclonal rabbit (1:200, 12708; Cell Signaling Technology), Periaxin/PRX (1:200, HPA001868, Sigma Aldrich), MMP2 (1:200; ab37150, Abcam), Ki67 (1:1000; NCL-Ki67p; Novocastra, Leica Biosystems;) or Cleaved Caspase 3 (Asp175) (1:200, 9661S, Cell Signaling Technology) using standard protocols (see also [Table S1](#)). Images were photographed using an Olympus BX 53 microscope, and were quantified using HistoQuest™ software (TissueGnostics GesmbH, Vienna, Austria).

Immunofluorescence

Cells were grown on sterile glass coverslips, rinsed with PBS, fixed in 4% paraformaldehyde for 10 min and permeabilized using 0.5% Triton X-100 in PBS for 8 min. Cells were blocked in 3% BSA+0.1% Triton in PBS for 1 hr, incubated with primary antibodies (anti-STAT1 1:500, 9172 Cell Signaling Technology; anti-STAT3 1:1500, 9139 Cell Signaling Technology) for 1 hr or at 4°C overnight, washed and probed with the secondary antibodies conjugated to Alexa Fluor 488 and Alexa Fluor 568 (Molecular Probes) for 1 hr. Cells were stained with DAPI and mounted in VECTASHIELD Antifade Mounting Medium with DAPI (Vector Laboratories). The images were acquired with a Zeiss LSM 880 Confocal Laser Scanning Microscope.

Mass-Spectrometry Based Proteomics

Sample Preparation for MS

Approximately 1×10^7 cells from DFT1 tumor cell line T1 were pelleted (260g, 5 min, 4°C), washed three times in PBS and frozen at -80°C until processed. Primary biopsies were thawed and placed in a petri dish. Using a scalpel blade, 5×5 to 5×8 mm pieces of tissue were excised from the solid mass and placed in a 2 ml Eppendorf tube. Depending on the size of the piece of tissue, 500–1000 μ l of lysis buffer (50 mM HEPES, pH 8.0, 2% SDS, 1 mM PMSF, and protease inhibitor cocktail (Sigma-Aldrich)) was added to the tumour and skin samples. Samples were homogenised using the Tissue Lyser II (Qiagen, Hilden, Germany) for 4×2 min, 30 Hz. For some samples, it was necessary to repeat the homogenization procedure. Spleen samples were pre-cleared of blood before tissue lysis. Tissue pieces were placed in a 2 ml Eppendorf tube containing 1 ml red blood cell (RBC)-lysis buffer (eBioscience, San Diego, USA). Spleen tissue was homogenised using the Tissue Lyser II (Qiagen, Hilden, Germany) for 3×30s, centrifuged at 20000 g for 10 min and supernatant containing the lysed red blood cells removed. Lysis was performed at room temperature (RT) for 20 min. Lysed samples were heated at 99°C for 5 min and then cooled to RT. The cell lysate was sonicated using a Covaris S2 high performance ultrasonicator (Covaris Inc., Brighton, UK). The lysate was centrifuged at 20000 g for 15 min at 20°C, and the protein extract was collected from the supernatant. Total protein content of the whole tissue lysates was determined using the BCA protein assay kit (Pierce Biotechnology, Rockford, IL) following the recommendations of the manufacturer. The assay was performed in a 96-well plate using 10 μ l of each lysate and standard protein. The samples were measured in triplicates. Bovine serum albumin (BSA) (Pierce Biotechnology, Rockford, IL) was used as the standard protein.

Filter-Aided Sample Preparation (FASP)

100 µg total protein per tissue was used for FASP digestion. Dithiothreitol (DTT; SIGMA-Aldrich Chemie, Vienna, Austria) was added to sample to a final concentration of approx. 83 mM. After incubation of the samples at 99°C for 5 min, FASP digestion was performed using a 30 kDa molecular weight cut-off filter (Microcon-30, Ultracel YM-30, Merck-Millipore Co., Cork, IRL) (Wisniewski et al., 2009). Briefly, 200 µL 8 M urea in 100 mM Tris-HCl (pH 8.5) (UA) was added to the samples. If the volume exceeded 50 µL, then 400 µL UA was added. After equilibration of the filter units with 200 µL UA and centrifugation at 14000×g for 15 min, the lysed samples were applied in steps of 250 µL to the filter unit and centrifuged at 14000 g for 15 min at 20°C to remove SDS. Any remaining SDS was exchanged by urea in a second washing step with 200 µL UA. The proteins were alkylated with 100 µL 50 mM iodoacetamide (Sigma-Aldrich Chemie, Vienna, Austria) for 30 min at RT. Afterwards, three washing steps with 100 µL UA solution were performed, followed by three washing steps with 100 µL 50 mM TEAB buffer (Sigma-Aldrich, Vienna, Austria). Proteins were digested with Trypsin overnight at 37°C. Peptides were recovered using 40 µL 50 mM TEAB buffer followed by 50 µL of 0.5 M NaCl (Sigma-Aldrich, Vienna, Austria).

Two-dimensional liquid chromatography was performed by reverse-phase chromatography at high and low pH. FASP digests were purified by solid-phase extraction (SPE) (MacroSpin Columns, 30–300 µg capacity, Nest Group Inc. Southboro, MA, USA) and reconstituted in 23 µL 5% acetonitrile, 10 mM ammonium formate. Peptides were separated on a Gemini-NX C18 (150 × 2 mm, 3 µm, 110 Å, Phenomenex, Torrance, US) using a 30 min gradient from 5 to 90% acetonitrile containing 10 mM ammonium formate buffer, pH 10, at a flow rate of 100 µL/min, using an Agilent 1200 HPLC system (Agilent Biotechnologies, Palo Alto, CA). Details of the methodology are as described previously (Bennett et al., 2011). Ten time-based fractions were collected. Samples were acidified by the addition of 5 µL 5% formic acid. Solvent was removed in a vacuum concentrator, and samples were reconstituted in 5% formic acid. Liquid chromatography mass spectrometry was performed on a hybrid linear trap quadrupole (LTQ) Orbitrap Velos mass spectrometer (ThermoFisher Scientific, Waltham, MA) using the *Xcalibur version 2.1.0* coupled to an Agilent 1200 HPLC nanoflow system (dual pump system with one trap-column and one analytical column) via a nanoelectrospray ion source using liquid junction (Proxeon, Odense, Denmark). Solvents for HPLC separation of peptides were as follows: solvent A consisted of 0.4% formic acid (FA) in water, and solvent B consisted of 0.4% FA in 70% methanol and 20% 2-propanol. From a thermostatted microautosampler, 8 µL of the tryptic peptide mixture were automatically loaded onto a trap column (Zorbax 300SB-C18 5 µm, 5×0.3 mm, Agilent Biotechnologies) with a binary pump at a flow rate of 45 µL/min. 0.1% trifluoroacetic acid (TFA) was used for loading and washing the precolumn. After washing, the peptides were eluted by back-flushing onto a 16 cm fused silica analytical column with an inner diameter of 50 µm packed with C18 reversed phase material (ReproSil-Pur120 C18-AQ, 3 µm, Dr. Maisch GmbH, Ammerbuch-Entringen, Germany). The peptides were eluted from the analytical column with a 27 min gradient ranging from 3% to 30% solvent B, followed by a 25 min gradient from 30% to 70% solvent B, and finally a 7 min gradient from 70% to 100% solvent B at a constant flow rate of 100 nL/min (Bennett et al., 2011). The analyses were performed in a data-dependent acquisition mode, and dynamic exclusion for selected ions was 60s. A top 15 collision-induced dissociation (CID) method was used, and a single lock mass at *m/z* 445.120024 (Si(CH₃)₂O)₆) was employed (Olsen et al., 2005). Maximal ion accumulation time allowed in CID mode was 50 ms for MSⁿ in the LTQ and 500 ms in the C-trap. Automatic gain control (AGC) was used to prevent overfilling of the ion traps and was set to 5000 in MS² mode for the LTQ and 10⁶ ions for a full MS¹ FTMS scan. Intact peptides were detected in the Orbitrap Velos at a resolution of 60000 resolution (at *m/z* 400). The threshold for switching from MS¹ to MS² was 2000 counts.

DNA Methylation Analysis

DNA methylation profiling by RRBS was performed as described previously using 100 ng of genomic DNA isolated from RNA-later preserved tissue samples through the Allprep DNA/RNA Mini kit (QIAGEN) (Klughammer et al., 2015). Methylated and unmethylated spike-in controls were added in a concentration of 0.1% to assess bisulfite conversion efficiency independent of CpG context. DNA was digested using the restriction enzymes MspI and TaqI in combination (as opposed to only MspI in the original protocol) in order to increase genome-wide coverage. Restriction enzyme digestion was followed by fragment end repair, A-tailing, and adapter ligation. The amount of effective library was determined by qPCR, and samples were multiplexed in pools of 13 with similar qPCR C_t values. The pools were then subjected to bisulfite conversion followed by library enrichment by PCR. Enrichment cycles were determined using qPCR and ranged from 9 to 13 (median: 11). After confirming adequate fragment size distributions on Bioanalyzer High Sensitivity DNA chips (Agilent), libraries were sequenced on Illumina HiSeq 3000/4000 machines in a 50 bp single-read setup.

Transcriptome Expression Analysis and Variant Calling

The amount of total RNA was quantified using Qubit 2.0 Fluorometric Quantitation system (Life Technologies) and the RNA integrity number (RIN) was determined using Experion Automated Electrophoresis System (Bio-Rad). RNA-seq libraries were prepared with TruSeq Stranded mRNA LT sample preparation kit (Illumina) using Sciclone and Zephyr liquid handling robotics (PerkinElmer). Library amount was quantified using Qubit 2.0 Fluorometric Quantitation system (Life Technologies) and the size distribution was assessed using Experion Automated Electrophoresis System (Bio-Rad). For sequencing 6 libraries were pooled, diluted and sequenced on Illumina HiSeq 3000/4000 using 75 bp paired-end chemistry. Base calls provided by the Illumina Realtime Analysis software were converted into BAM format using Illumina2bam and demultiplexed using BamIndexDecoder (<https://github.com/wtsi-npg/illumina2bam>).

Measurement of Biochemistry Parameters

Serum was prepared by centrifugation of the whole blood for 20 min at 7000 rpm. Serum concentration of alanine aminotransferase, aspartate aminotransferase and blood urea nitrogen was measured using a chemistry analyzer (IDEXX VetTest 8008, IDEXX GmbH, Ludwigsburg, Germany).

QUANTIFICATION AND STATISTICAL ANALYSIS

Immunohistochemistry and Immunofluorescence Quantification

Immunohistochemical images were taken with Olympus BX 53 microscope and the quantification was performed using HistoQuest TM software (TissueGnostics GesmbH, Vienna Austria). Quantification of immunofluorescence was carried out using a co-localization pipeline of CellProfiler (version 3.1.5). Briefly, illumination was corrected using the "Regular" function and "Fit polynomial" as smoothing method. Subsequently, images were aligned with the "Mutual information" option and objects were identified with the following settings: size (pixels) 3-15, Threshold strategy: global, Threshold method: Otsu, with three classes thresholding, pixels in the middle intensity class were assigned as background and objects touching the image boundary were discarded. Objects were classified as co-localized if they touched, overlapped or enclosed each other and as not co-localized when none of the above applied. Percentages were calculated on a per image basis and for each condition 10 images were used. Data shown is representative of three similar experiments. Displayed is the percentage of STAT1 objects that were colocalized with a STAT3 object.

Mass-Spectrometry Based Proteomics - Quantification

The data were analyzed using Proteome Discoverer version 2.2 against the Uniprot Tasmanian Devil fasta database version 2016.11 including isoforms obtained by VARSPLIC (Kersey et al., 2000) and appended with known contaminants (91064 sequences total). The precursor masses were first recalibrated using the recalibration node (parameters: precursor mass tolerance: 20 ppm, fragment mass tolerance 0.5 Da, Carbamidomethylation of cysteine as static modification). The recalibrated data were then searched using Sequest HT (Eng et al., 1994) and Mascot (v2.3.02, MatrixScience, London, U.K.) (Perkins et al., 1999) search engines with precursor mass tolerance being 4 ppm, fragment ion tolerance was 0.3 Da, methionine oxidation used as dynamic modification, and carbamidomethylation of cysteine as static modification in both search engines. Moreover protein N-terminal acetylation was considered as dynamic modification in Sequest HT. Minimum length of peptides was set to 6 and maximum number of missed cleavages was set to two in Sequest HT and to one in Mascot. Percolator (Kall et al., 2007) was used to filter peptide-spectra matches (PSMs) at 1% false discovery rate (FDR) and QUALITY (Kall et al., 2009) to filter identified peptides at 1% FDR. Identified proteins were also filtered at 1% FDR by considering sum of negative logarithm of posterior error probabilities of connected PSMs. Matches against reversed fasta database were used to estimate FDR at PSM, peptide, and protein level. Abundance of proteins was quantified using the Minora feature detection node and integrating area under the MS1 chromatogram.

Principal component analysis was performed on 3894 out of 6672 proteins quantified in all 19 samples. We further focused on 4981/6672 proteins quantified in at least 80% of the samples. The protein abundance data was normalized by variance stabilizing transformation (Huber et al., 2002). Missing values are imputed on normalized abundance values with the k Nearest Neighbors (kNN) algorithm implemented in the R Bioconductor package impute (Hastie et al., 2017).

For each protein with a missing abundance in any of the samples, the kNN algorithm identifies the set of 10 most similar proteins based on non-missing abundance values. Missing abundance is then imputed as an average abundance in that set. An average of the signal is then performed on the k closest neighbours. Differential analysis, tumor versus the healthy, was performed between biopsies (excluding the tumor cell line) using the limma Bioconductor package (Ritchie et al., 2015). Proteins were considered as differentially modulated if their adjusted p value was ≤ 0.05 and their absolute \log_2 Fold Change was ≥ 1 between tumor and healthy biopsies. Hierarchical clustering of the 987 differentially modulated proteins was performed with Pearson's distance measure and the average clustering method.

DNA Methylation Analysis and Quantification

The DNA methylation (RRBS) data were analyzed using the RefFreeDMA pipeline as described previously to avoid potential biases in read mapping and methylation calling related to the scaffold assembly status of the published Tasmanian Devil reference genome sarHar1 (Klughammer et al., 2015). In brief, a custom *ad-hoc* reference genome was deduced directly from the RRBS sequencing reads and used for read mapping and methylation calling. Based on the thus produced DNA methylation profiles, differential DNA methylation analysis was performed as part of the RefFreeDMA pipeline and as originally described in (Assenov et al., 2014). Gene annotations were transferred by mapping the deduced genome fragments to the published scaffold-level Tasmanian Devil genome sarHar1, downloaded from the UCSC genome browser. A gene annotation file was produced by joining the UCSC transcript annotation file with the Ensembl v86 transcript annotation file of sarHar1 based on the common Ensembl transcript identifiers. Gene promoters were defined as the region between 5000 bases downstream and 2500 bases upstream of the transcription start site. 487540 (11.37%) of the individual CpGs are situated in promoters of annotated genes. For promoter methylation analysis we focused on a total of 69754 CpGs that are also covered in at least 80% of the samples. The significance of differential methylation throughout promoters or deduced genome fragments was assessed by combining the p values for single CpGs within the respective promoter regions or deduced genome fragments using an extension of the Fisher's method as described previously (Assenov et al., 2014; Klughammer et al., 2015).

Western Blot Quantification

Immunoblots were quantified using ImageJ (2.0.0) software. Three independent immunoblots were used for each quantification. Phospho-STAT3 levels were normalized to STAT3 and to loading control HSC70.

Transcriptome Expression Analysis and Variant Calling

Paired-end reads were trimmed for adaptor sequences and filtered with the trimmomatic tool (Bolger et al., 2014). Trimmed reads were aligned on the version 7.0 of the Tasmanian Devil with the STAR aligner (Dobin et al., 2013). Counting of reads on annotated transcripts (*Sarcophilus_harrisii*.DEVIL7.0.90.gtf from Ensembl) was performed with featureCounts (Liao et al., 2014). The DESeq2 Bioconductor library has been used for counts normalization and differential analysis between the transcriptomes of the four DFTD and fibroblast cell lines (Love et al., 2014). Differentially expressed genes were identified based on the following cutoffs: an average minimum expression value between conditions of 50 reads, an absolute log fold change of 1, and an adjusted p value of maximum 0.05. The pipeline for variant calling is based on the GATK version 3.7, following the best-practices (Van der Auwera et al., 2013). Called variants were annotated with SNPEff v4.2 (Cingolani et al., 2012). Variants are filtered on strand bias (<30), quality (QD > 2), coverage (DP >= 10) and allele frequency (AF = 0.5).

Network Analysis

We performed Transcription Factor and Pathway Maps enrichment on the differentially modulated entities (proteins, genes) with the MetaCore™ (Thomson Reuters, version 6.32 build 69020; cut-off for p value of enrichment 0.05). The results are reported based on the z-score of enrichment for the transcription factors and as -log₁₀ of the False Discovery Rate for the Pathway Maps. The 987 differentially modulated proteins were integrated together with the 166 genes with differentially modulated promoters and the candidates ERBB2 and ERBB3 from the drug-screen at the level of MetaCore interactions (Build Network functionality). Only high-confidence direct interactions between pairs of genes are considered: protein binding, transcription factor regulation, other functional interaction. 632 of the previous candidates form a direct network connection.

Motif Enrichment Analysis

We filtered individual CpG regions with less than 8 or more than 200 reads. On the remaining CpGs, we only kept those with less than 20% missing values across the 19 samples. Modulated individual CpGs between healthy and tumor samples were selected based on an adjusted p value smaller than 0.05, and a log fold-change greater than 1 for the UP dataset (4299 CpGs), and smaller than -1 for the DOWN dataset (1727 CpGs). Fasta sequences of these fragments were extracted. We performed motif enrichment analysis with the AME tool from the MEME package (McLeay and Bailey, 2010). We used the motif JASPAR CORE 2014 Vertebrate database (Mathelier et al., 2014). The motif enrichment for each fasta file accounts for background sequence composition by using the other sequence file as control.

Integrating DFTD Quantifications across Techniques

The DFTD tumor cell line T1 was the only sample quantified with all three techniques: RNA-seq, proteomics and methylation. We kept only the annotated (no SNO) genes from the RNA-seq data and computed the expression as the number of counts divided by the length of the gene as returned by featureCounts (Liao et al., 2014). The genes were ranked according to expression from the most to the least expressed. The top 15% most abundant proteins (817) were selected. We performed a Gene Set Enrichment Analysis (GSEA) (Mootha et al., 2003; Subramanian et al., 2005). Pre-ranked between the top DFTD 15% most abundant proteins and the ranked gene expression data using default parameters. For the annotated gene promoters with CpG fragments, we computed the average methylation level in the promoter regions defined as -5 kb to 2.5 kb around the TSS. Similarly, we ranked the genes according to their promoter methylation level. We selected the top 15% least methylated gene promoters (936) and performed GSEA Pre-ranked on the ranked RNA-seq expression. Alternatively, we ranked the annotated genes detected in the DFTD cell line by both RNA-seq and methylation data with a random method for ties. We compared the ranks of methylation and transcriptomics using the hexbin plot in R (hexbin R package (version 1.27.2). A Spearman correlation is performed between the ranks of the two datasets.

To study potential effects on chemotaxis and cytoskeleton remodeling, we performed MetaCore Pathway Enrichments on all proteins that were modulated, be it UP or DOWN ($\text{abs}(\log\text{FC}) \geq 1$, $\text{adj.p.val} \leq 0.05$; Table S5). Subsequently, we extracted proteins that were differentially modulated and led to the enrichments in the following pathways linked to chemotaxis and cytoskeleton remodeling: Cytoskeleton remodeling_Integrin outside-in signaling (4), Chemotaxis_SDF-1/CXCR4-induced chemotaxis of immune cells (5), Cytoskeleton remodeling_Regulation of actin cytoskeleton organization by the kinase effectors of Rho GTPases (9) and Cell adhesion_Role of tetraspanins in the integrin-mediated cell adhesion (13). The respective proteins are reported in Table S5 the chemotaxisGenes.xlsx file together with their respective modulation in the Proteomics and RNAseq datasets. Next, we generated a heatmap based on the protein abundance values of these proteins across the biopsies, cell line, nerve, skin and spleen (clustering distance : pearson, clustering method: average). On the right genes that were found as being bound by STAT3 across any of the ENCODE Chip-seq datasets (Rouillard et al., 2016) were color-coded. In addition, we color-coded whether the genes were found and/or modulated in the RNA-seq data (DFTD tumor cell lines T1-T4 vs. fibroblasts).

Characterization of the STAT3 Inhibitor DR-1-55

To characterize drug-target binding, we performed a ^{19}F NMR protein-ligand experiment as done previously (Ali et al., 2016; Garg et al., 2017; Wingelhofer et al., 2018). Incubation of purified recombinant STAT3 with DR-1-55 results in line broadening of the ^{19}F peaks from the compound, indicating a longer correlation time for the drug, which is a consequence of protein-drug binding. Concomitantly, a release of fluoride ions is observed, the by-product of a covalent reaction of the protein and a pentafluorobenzene containing molecule. In addition, we performed a thermal shift assay and observed a dose-dependent decrease in the denaturation temperature of recombinant STAT3, indicative of covalent protein-ligand binding interaction (de Araujo et al., 2017).

DATA AND SOFTWARE AVAILABILITY

The data reported in this paper are tabulated in the [Supplemental Information](#) and archived at the following databases: proteomic data in the PRoteomics IDEentification (PRIDE) database with accession number (1-20180126-165173), DNA methylation data in the Gene Expression Omnibus (GEO) database with accession number (GSE108160) and RNA-seq data in the GEO database with accession number (GSE108107).