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H₂S events in the Peruvian oxygen minimum zone facilitate enhanced dissolved Fe concentrations

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Dissolved iron (DFe) concentrations in oxygen minimum zones (OMZs) of Eastern Boundary Upwelling Systems are enhanced as a result of high supply rates from anoxic sediments. However, pronounced variations in DFe concentrations in anoxic coastal waters of the Peruvian OMZ indicate that there are factors in addition to dissolved oxygen concentrations (O₂) that control Fe cycling. Our study demonstrates that sediment-derived reduced Fe (Fe(II)) forms the main DFe fraction in the anoxic/euxinic water column off Peru, which is responsible for DFe accumulations of up to 200 nmol L⁻¹. Lowest DFe values were observed in anoxic shelf waters in the presence of nitrate and nitrite. This reflects oxidation of sediment-sourced Fe(II) associated with nitrate/nitrite reduction and subsequent removal as particulate Fe(III) oxyhydroxides. Unexpectedly, the highest DFe levels were observed in waters with elevated concentrations of hydrogen sulfide (up to 4 μmol L⁻¹) and correspondingly depleted nitrate/nitrite concentrations (<0.18 μmol L⁻¹). Under these conditions, Fe removal was reduced through stabilization of Fe(II) as aqueous iron sulfide (FeS_{aqu}) which comprises complexes (e.g., FeSH⁺) and clusters (e.g., Fe₂S₂[4H₂O]). Sulfidic events on the Peruvian shelf consequently enhance Fe availability, and may increase in frequency in future due to projected expansion and intensification of OMZs.

Iron (Fe) forms an important micronutrient, controlling marine phytoplankton growth and nitrogen fixation in vast regions of the global ocean^{1–3}. Shelf sediments are recognized as an important source of Fe to coastal waters and the open ocean^{4–6}. In particular, Fe supplied from shelf sediments in oxygen minimum zones (OMZs) results in highly elevated dissolved Fe (DFe) concentrations in coastal waters depleted in oxygen (O₂) to concentrations below 2 μmol L^{-1,7,8}, thereby providing a potential Fe source for offshore transport⁹. The strength of the Fe flux from shelf sediments is significantly enhanced in case sediments become euxinic (hydrogen sulfide (H₂S) present)^{10–12}.

Upwelling of nutrient-rich deep waters across the Peruvian shelf region results in extremely high primary productivity. The sediments and water column in this region are severely depleted in dissolved oxygen (O₂) as a result of bacterial respiration of sinking organic matter¹³, augmented by a sluggish ventilation accounting for a reduced lateral and vertical flux of oxygen into the Peruvian shelf region^{14,15}. The Peruvian sediment pore water concentrations of DFe, predominantly in the reduced form Fe(II), are in the 1–30 micromolar range and hence several orders of magnitude higher than in the overlying bottom waters¹⁶. The steep DFe concentration gradients and anoxic conditions of the bottom waters facilitate enhanced Fe fluxes out of the sediments reaching 10 to 866 μmol DFe m⁻² d^{-1,8}, which result in bottom water DFe concentrations of 20–300 nmol L^{-1,7,17}, with most of the DFe being present in the reduced form Fe(II)^{18–21}.

Iron(II) is formed in anoxic sediments by the dissimilatory reduction of Fe(III) oxides as part of a sequence of microbially mediated redox reactions^{22,23}. The layer of Fe(II) enrichment in the pore waters of the sediments is located above that of elevated H₂S levels. Below the Fe(II)-rich zone, H₂S is formed by the dissimilatory reduction of sulfate (SO₄²⁻) and reacts with dissolved Fe(II) to form a wide variety of aqueous Fe complexes ([FeSH]⁺, [Fe(SH)₂]⁰, [Fe(SH)₃]⁻, etc.) and clusters (e.g., Fe_nS_n⁰ | 4H₂O (n = 2 or 4)) (summarized as FeS_{aqu})²⁴ followed by amorphous particulate Fe(II) monosulfides (Fe(II)S) (e.g. mackinawite (FeS_m))^{25–27}. Aqueous Fe sulfide clusters are ligated directly to H₂O and the structure of the cluster is similar to the basic structure of mackinawite²⁴. Due

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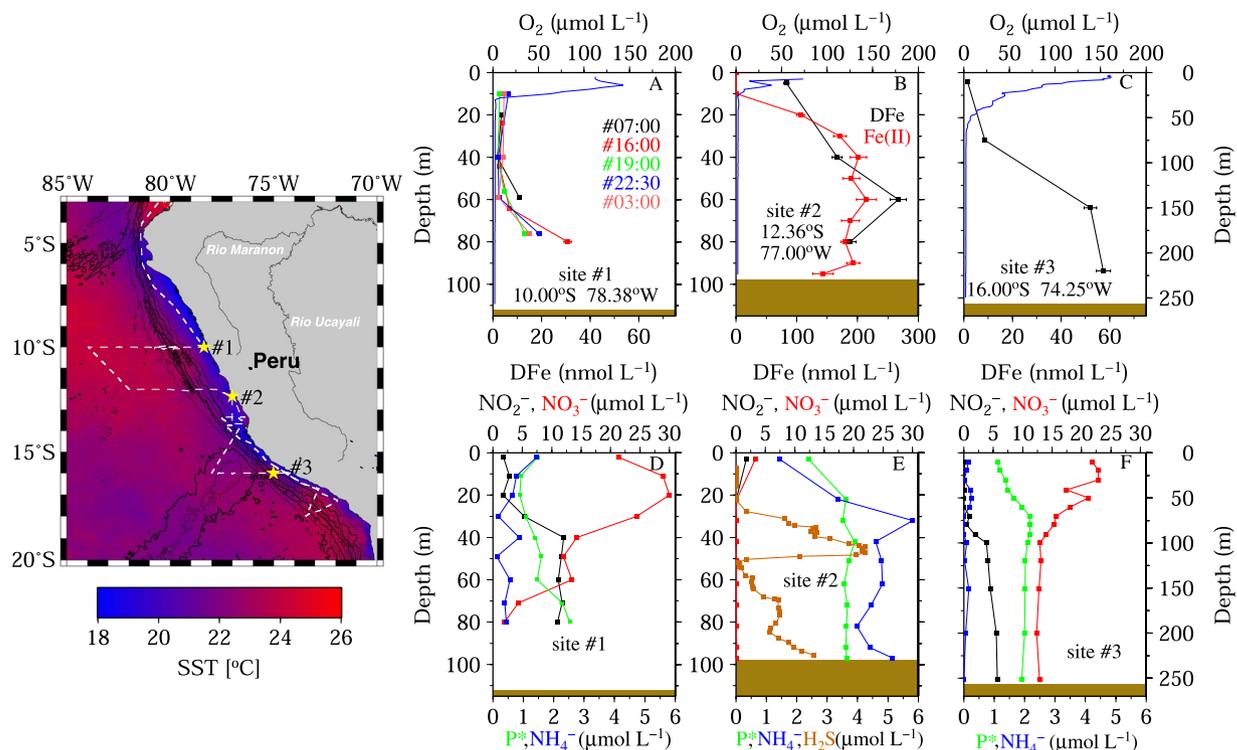


Figure 1. (Map) Sampling area of M77-3 (FS Meteor) in the Peruvian OMZ between December 2008 and January 2009. The cruise track is indicated by the dashed white line and the three Go-Flo sampling sites of this study are marked by the yellow stars and white station numbers. The colors show satellite sea surface temperature data (SST) in degree centigrade (MODIS). The GEBCO bathymetry is expressed by contour lines. The map was created by the open source software Generic Mapping Tool, GMT, Version 4 (<http://gmt.soest.hawaii.edu>). (A) Dissolved Fe concentration (DFe) in the water column at site 1 was sampled 5 times within 24 hours. O₂ concentration is given by the blue line. (D) Nitrate (NO₃⁻ in red), nitrite (NO₂⁻ in black), Ammonia (NH₄⁺ in blue), and P* (in green) determined at site 1. Results from site 1 (A and D) and site 3 (C and F) are similarly assembled, but in B additionally the Fe(II) concentrations are shown in red and in E the water content of H₂S is provided, a compound that was not detected at site 1 and 3.

to similar structural homologies found for aqueous zinc and copper sulfide clusters, it has been suggested that the structure of the cluster determines the form of the initial particulate phase^{28,29}. However, mackinawite formation is thermodynamically favored when the ion activity product of H₂S and Fe(II) exceeds the thermodynamic stability product of FeS_m ($\log K_{sp} = -3.6$)²⁶. This condition is typically met in euxinic sediments (e.g. Fe(II) > 30 μmol L⁻¹; H₂S > 10 μmol L⁻¹³⁰), and represents the initial step of Fe pyrite (FeS₂) formation^{25,27,31}.

The formation of FeS_m is also reported to occur in anoxic coastal waters and has been observed in permanently euxinic bottom waters of the Framvaren fjord (Norway), where Fe(II) and H₂S levels are in the millimolar range³². Because H₂S concentrations in these waters are extremely high, Fe sulfide precipitates in the water column as framboidal pyrite³³. It is, however, unclear whether favorable conditions for Fe sulfide precipitation also occur in the coastal OMZ regions off Peru. The Peruvian OMZ features extremely low O₂ concentrations (< 50 nmol kg⁻¹)³⁴⁻³⁷, with anoxic waters prevailing at depths between 10 to 500 m, which in near shore regions are in direct contact with shelf sediments²⁸. The positioning of the OMZ over euxinic shelf sediments facilitates benthic supply of H₂S and Fe(II), which accumulates periodically in the overlying waters^{7,38}. The transient presence of H₂S in Peruvian coastal waters have mainly been observed during the austral summer season and their occurrences have recently been associated with stagnant flow on the shelf³⁹. H₂S and DFe in the Peruvian OMZ waters have been reported to reach concentrations as high as 13 μmol L⁻¹ and 300 nmol L⁻¹, respectively, during the upwelling season in austral summer and the formation of Fe(II)S minerals resulting in DFe removal has been suggested⁴⁰.

Here we present new data from three locations with different environmental settings on the Peruvian shelf regarding DFe, Fe(II), H₂S, nitrate (NO₃⁻), nitrite (NO₂⁻) and ammonium (NH₄⁺) (with the sum of all N species = DIN), O₂, and dissolved inorganic phosphorous (DIP) expressed as P* (P* = DIP - DIN/16)⁴¹. We examine the mechanisms that control DFe in the Peruvian OMZ, under contrasting conditions of presence and absence of enhanced water column H₂S concentrations.

Results and Discussion

Water column anoxia occurred at all three study sites during the upwelling season in December/January 2008/09 (Fig. 1A–C). The surface mixed layer extended down to ca. 10 m depth at site 1, 20 m at site 2 and 60 m depth at site 3, and was assessed using a fixed density difference criterion ($\Delta\sigma = 0.125$)⁴². At all sites, O₂ concentrations decreased below the surface mixed layer to levels below the LOD of the CTD mounted O₂ sensors. At site 2, O₂

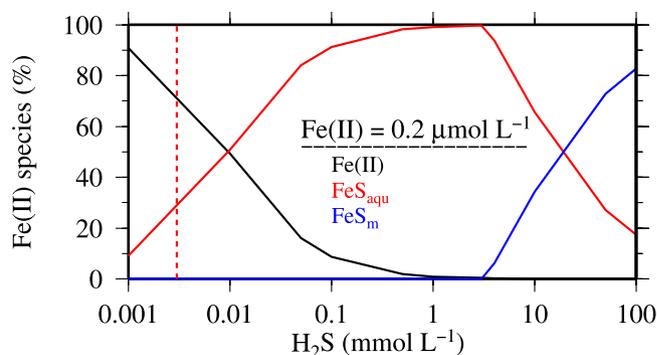


Figure 2. Distribution of Fe(II) species in percent (dissolved Fe(II) (black), soluble Fe (II) complexes/clusters (FeS_{aqu}) (red), and particulate FeS_m (mackinawite) (blue)) at a constant Fe(II) concentration of $0.2 \mu\text{mol L}^{-1}$ and under increasing hydrogen sulfide (H_2S) concentrations. The red dashed line represents the average H_2S concentration in subsurface waters <50 m at site 2 in the Peruvian OMZ.

was $75 \mu\text{mol L}^{-1}$ at 2 m depth, and decreased below 20 m to levels undetectable for the STOX sensor. The strong vertical oxygen gradient in the near-surface layers was a consequence of enhanced O_2 consumption by microbial organic matter remineralization. At all three sites the anoxic conditions extended down to the seafloor.

The DFe levels in our study region were considerably higher than those reported for other coastal regions where OMZs prevail, e.g. $1.2\text{--}6.3 \text{ nmol L}^{-1}$ on the north west African shelf⁴. The profiles of DFe concentrations at sites 1 and 3 differed pronouncedly from site 2 (Fig. 1A–C). Dissolved Fe concentrations at site 1 increased from 2 nmol L^{-1} in the surface layer to 30 nmol L^{-1} at 80 m depth. Over a period of 24 h, five separately sampled casts indicated that DFe profiles remained essentially constant (Fig. 1A). Similar to site 1, DFe concentrations at site 3 increased from 2 nmol L^{-1} in the surface layer to 60 nmol L^{-1} at 90 m depth. Bottom water DFe values at both sites agreed well with reported values for anoxic Peruvian shelf bottom waters of 30 to 60 nmol L^{-1} ^{17,19–21}. In contrast, DFe at site 2 increased from very high values of 80 nmol L^{-1} in the surface waters to ca. 200 nmol L^{-1} below 30 m depth, and remained relatively constant at ca. 200 nmol L^{-1} down to the seafloor (Fig. 1B). Below 30 m water depth, DFe occurred predominantly in the reduced Fe(II) form (Fig. 1B), an Fe species that has been analyzed just for site 2. Several studies with focus on Fe speciation have been already conducted in the Peruvian OMZ^{9,20,21}. In agreement with our results, these studies showed that the reduced Fe(II) form represented the main DFe species in the anoxic part of the water column. However, Fe(II) concentrations in the water column determined during those studies were lower than found during our present study ($\text{Fe(II)} = 15\text{--}73 \text{ nmol L}^{-1}$ ²⁰, $12\text{--}47 \text{ nmol L}^{-1}$ ¹⁹, $0.2\text{--}16 \text{ nmol L}^{-1}$ ²¹).

The water column at site 2 featured enhanced H_2S concentrations, coinciding with high DFe levels (Fig. 1B,E). However, an increase in H_2S concentrations at site 2 was observed with depth including a mid-depth maximum at 50 m ($\sim 4 \mu\text{mol L}^{-1}$; Fig. 1E) and $\sim 3 \mu\text{mol L}^{-1}$ near the seafloor. Elevated DFe concentrations of up to 300 nmol L^{-1} during a H_2S event reaching $\sim 10 \mu\text{mol L}^{-1}$ on the Peruvian shelf in 2012 have also been reported by Scholz, *et al.*⁷, with DFe and H_2S being released by anoxic sediments.

Removal of DFe through the formation of Fe(II) sulfide minerals has been reported for euxinic sediments²⁷, deep-sea hydrothermal vent systems⁴³, euxinic fjord waters³², and anoxic waters in the bottom boundary layer of the Peruvian OMZ⁴⁰. Using Visual MINTEQ. 3.1⁴⁴, we calculated the species distribution of dissolved Fe(II), aqueous sulfide (FeS_{aqu} ; $\log K = 5.62$ ²⁴), and Fe(II)S in the crystal structure of mackinawite ($\log K = -3.6$ ²⁴), an ubiquitous mineral in low temperature aqueous environments, to determine if Fe(II)S formation in the water column at site 2 was feasible (Fig. 2) (concentrations of model parameters used are listed in supplementary material S1). At 13°C and pH 7.65, which is typical for subsurface waters on the Peruvian shelf^{45,46}, and concentrations of Fe(II) of 200 nmol L^{-1} and H_2S levels of $3 \mu\text{mol L}^{-1}$, which reflect the bottom water composition at site 2, the ion activity product of the educts ($\log IAP = -5.466$) is below the typical apparent solubility product of mackinawite of $\log K'_{sp} = -3.6$ ⁴⁴ (undersaturation with respect to mackinawite: $\log IAP - \log K'_{sp} < 0$). To validate the robustness of the model a sensitivity test was executed by increasing pH and temperature which would increase $\log IAP$ and enhance the likelihood of mackinawite formation. At 40°C and pH 8.5, obviously very unrealistic conditions for the Peruvian OMZ, the $\log IAP$ increased slightly to -4.61 . This suggests that under the transient conditions typically encountered, the formation of mackinawite is thermodynamically unfavorable in the anoxic bottom waters off Peru.

The mackinawite formation will only commence at Fe(II) and H_2S concentrations in the higher micromolar range. Our modelling indicates that at pH 7.65, 13°C and $200 \text{ nmol Fe(II) L}^{-1}$ more than 2 mmol L^{-1} H_2S are required to facilitate the formation of mackinawite (Fig. 2). Further, noticeably less H_2S is required ($\sim 10 \mu\text{mol L}^{-1}$) at higher Fe(II) levels of $>3 \mu\text{mol L}^{-1}$ to attain mackinawite saturation. These required enhanced H_2S and Fe(II) concentrations for the formation of Fe(II) sulfide minerals in seawater are in agreement with other modelling studies ($\text{Fe} \geq 3 \mu\text{mol L}^{-1}$; $\text{H}_2\text{S} \geq 50 \mu\text{mol L}^{-1}$ ⁴⁷) and earlier observations in anoxic regions such as the Framvaren fjord ($\text{Fe} \geq 2 \mu\text{mol L}^{-1}$; $\text{H}_2\text{S} \geq 1 \text{ mmol L}^{-1}$ ^{32,48}), Black Sea ($\text{Fe} \geq 0.3 \mu\text{mol L}^{-1}$; $\text{H}_2\text{S} \geq 30 \mu\text{mol L}^{-1}$ ^{49,50}), and Baltic Sea ($\text{Fe} \geq 2.34 \mu\text{mol L}^{-1}$; $\text{H}_2\text{S} \geq 52 \mu\text{mol L}^{-1}$ ⁵¹).

In H_2S free seawater more than 75% of Fe(II) occurs as a truly dissolved free cation, while the remainder will form complexes with hydroxide (OH^-), carbonate (CO_3^{2-}), and chloride (Cl^-) ions⁵². In the presence of an excess

of H_2S over Fe(II) and above pH 7.5, FeS_{aqu} becomes the dominant Fe(II) species²⁴ (Fig. 2). Formation of thiols in sediments⁵³ may result in Fe(II)-thiol complexes, with thiols also facilitating reduction of Fe(III)⁵⁴. Organic ligands complexing with Fe(III) serve a similar purpose in oxic waters to stabilize Fe⁵⁵. In the sulfide containing Peruvian waters at site 2, the Fe distribution in the water column was therefore controlled by the total Fe flux from the sediments rather than the equilibrium concentrations of their solid phases. Prolonged sulfidic periods will therefore result in an increase in DFe concentrations in the anoxic water column, as formation of Fe(II)S precipitates like mackinawite is unfavorable at the Fe(II) and H_2S concentrations so far observed in this region.

The formation of FeS_{aqu} stabilizes DFe via an increased soluble pool (as opposed to oxygenated waters in which DFe is dominated by Fe(III)-ligands and colloids⁵⁶), and hence facilitates vertical diffusive DFe fluxes in the Peruvian OMZ. Vertical diffusive DFe fluxes were determined for site 2 by combining eddy diffusivities determined from microstructure measurements sampled in the same region during January 2012 (FS Meteor cruise M92³⁹) and DFe concentration from this study. Altogether, 102 microstructure profiles were collected at shelf stations with bottom depth between 80 m and 100 m. The microstructure shear and temperature measurements were performed using a MSS90-D profiler (S/N 32, Sea & Sun Technology). Standard processing procedures were used to determine the rate of kinetic energy dissipation of turbulence in the water column (please see Schafstall *et al.*⁵⁷ for a detailed description). Subsequently, eddy diffusivities were determined from $K_p = \Gamma \varepsilon N^{-2}$ where N is stratification and Γ is mixing efficiency for which a value of 0.2 was used (Fig. S1)⁵⁸. Between mid-depth waters (40 m) and the surface (10 m), an average eddy diffusivity of $K_p = 3.1 \times 10^{-4} \text{ m}^2 \text{ s}^{-1}$ was obtained (Fig. S1). We employed the method of de Jong *et al.*⁵⁹ and obtained a vertical diffusive DFe flux $F_{\text{DFe}} = K_p \frac{\partial \text{DFe}}{\partial z}$ (with z being depth) from mid-depth waters at 40–60 m into the surface mixed layer of $101 \mu\text{mol m}^{-2} \text{ d}^{-1}$. Upper and lower 95% confidence limits were determined to be $164 \mu\text{mol m}^{-2} \text{ d}^{-1}$ and $69 \mu\text{mol m}^{-2} \text{ d}^{-1}$ based on a Gaussian error propagation⁵⁷. This vertical diffusive flux is in a range similar to reported benthic DFe fluxes ($10\text{--}866 \mu\text{mol m}^{-2} \text{ d}^{-1}$) from shallow Peruvian shelf sediments⁸. It indicates an important role of FeS_{aqu} in the anoxic waters of the Peruvian OMZ, which through stabilization facilitates Fe supply to surface ocean phytoplankton communities on the shelf and possibly further afield by filaments and mesoscale eddies that move off shore, away from the coast zone⁶⁰.

In the absence of H_2S in the anoxic water column at sites 1 and 3, DFe concentrations were 72 to 94% lower in bottom waters than at site 2. Vertical diffusive water column fluxes calculated using a K_p of $3.1 \times 10^{-4} \text{ m}^2 \text{ s}^{-1}$ at sites 1 and 3 were concurrently reduced by 88 to 97% ($7.60 \pm 4.13 \mu\text{mol m}^{-2} \text{ d}^{-1}$), which indicates that supply and removal processes in anoxic water columns may differ and may depend on controls other than O_2 .

Organic matter remineralization processes in anoxic environments are mainly controlled by anaerobic microbial processes involving nitrogen, sulfur and Fe for electron-transfer reactions⁶¹. Of relevance for the Peruvian OMZ is the reduction of nitrate, coupled with the oxidation of H_2S by filamentous bacteria of the family *Desulfobulbaceae*^{62,63}. A similar coupling of anaerobic microbial denitrification with Fe(II) oxidation has been documented for chemolithotrophic organisms in freshwater sediments^{64,65}, suboxic aquifers⁶⁶, marine coastal sediments⁶⁷, and for the anoxic water column off the coast of Peru, where Fe(II) oxidizers such as *Marinobacter aquaeolei* are active⁷.

Transient accumulations of H_2S have been reported for anoxic Peruvian, Namibian and Indian shelf waters that were depleted in nitrate/nitrite^{68–72}. These euxinic waters form the extreme end point of ocean conditions, with fully oxygenated waters being the opposite end point⁷³. We suggest that the coupled process of anaerobic microbial denitrification and H_2S oxidation depletes nitrate and allows DFe to accumulate in anoxic bottom waters in the Peruvian OMZ. The extent of anaerobic microbial denitrification in the Peruvian anoxic environments is mainly controlled by H_2S levels and less by Fe(II) given that H_2S concentrations are an order of magnitude higher than those of Fe(II).

Depending on H_2S and nitrate concentrations, two distinct and one advanced scenarios can be envisaged (Fig. 3). Scenario 1: Under conditions of an anoxic water column (no measurable H_2S) above the sediments and in the presence of nitrate and nitrite (i.e. denitrification and anammox has not fully removed nitrate and nitrite) bacteria at the sediment-water interface will reduce these nitrogen species and thereby utilize both H_2S and Fe(II). During that process all the sulfide and a significant fraction of the Fe(II) will be oxidized. Insoluble Fe(III)oxyhydroxide particles are consequently formed near the sediment-bottom water interface, leading to an accumulation of reactive solid Fe phases near the sediment surface. Any Fe(II) that diffuses across the sediment-bottom water interface into the water column is then oxidized by $\text{O}_2/\text{H}_2\text{O}_2$ or, in the absence of O_2 , via nitrate-dependent Fe(II) oxidizing microbes with the Fe remaining either as organically complexed Fe(III) (FeL), or is lost by scavenging processes and insoluble particle formation, as was documented by Heller *et al.*¹⁸. This situation was observed at sites 1 and 3 (Fig. 1A–C and F) and is illustrated in Fig. 3 (left schematic).

Scenario 2: Reductive removal of the nitrate and nitrite pool in the sediment, in association with H_2S oxidation, allows enhanced sediment-water H_2S and Fe(II) fluxes resulting in an accumulation of these species in the water column. The oxycline in the upper water serves as a removal filter, with e.g. oxidation of Fe(II) to Fe(III) in the presence of oxygen and hydrogen peroxide⁷⁴. This scenario is proposed for site 2 (Fig. 1B and E) and resulted in the sulfidic event with H_2S concentrations of up to $4 \mu\text{M}$ and Fe(II) concentrations of up to 200 nM (Fig. 3, middle schematic). We assume that such a scenario is temporarily confined. Advanced scenario 2: Long lasting release of sedimentary H_2S and Fe(II) raises concentrations of both compounds in the water column, with highest concentrations expected at the sediment-bottom water interface. We hypothesize that under such conditions, $\log IAP > \log K'_{sp}$, resulting in the formation of mackinawite at the sediment-bottom water interface (Fig. 3, right schematic). Due to a lack of observational data, we do not know if such an advanced scenario can occur in the more turbulent Peruvian OMZ, or if it just arises in water bodies as stagnant as the Black Sea⁴⁹ and the Framvaren fjord³².

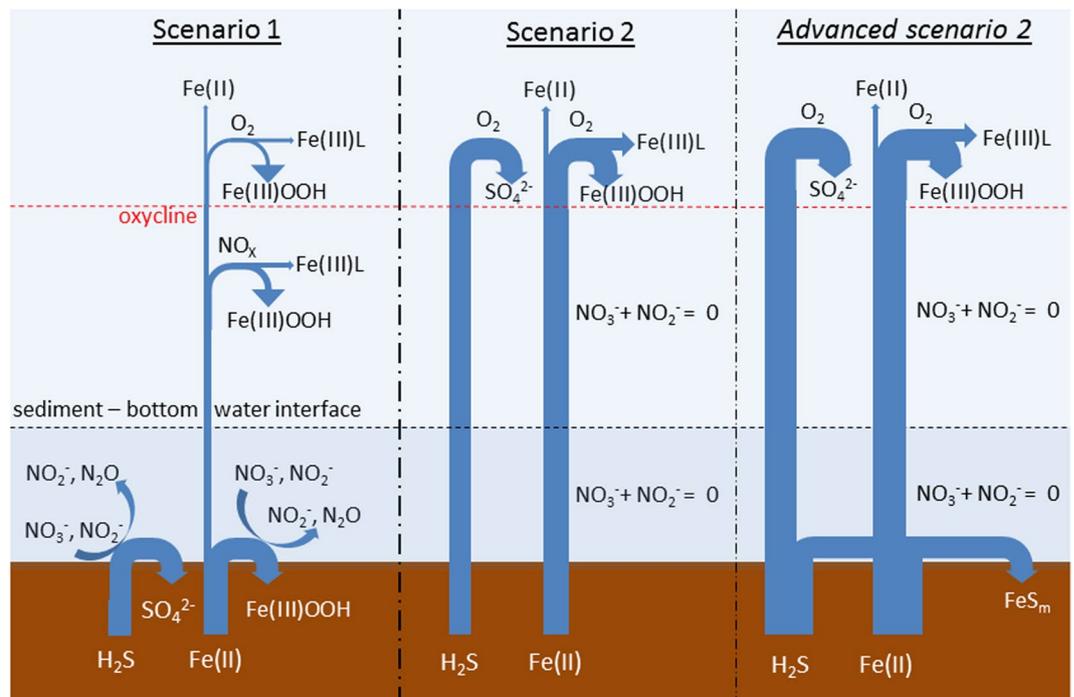


Figure 3. The left part of the sketch illustrates Scenario 1: H_2S and the majority of Fe(II) are removed at the sediment-bottom water interface by oxidizing both compounds with NO_3^- and NO_2^- to sulfate (SO_4^{2-}) and iron oxyhydroxides (Fe(III)OOH). In the water column, Fe(II) is oxidized by NO_3^- and O_2 with subsequent formation of particulate Fe(III)OOH and organically complexed Fe (Fe(III)L). A small fraction of Fe(II) is kept in solution by photochemical reduction processes in the surface. The middle part of the sketch illustrates Scenario 2: After H_2S reduced most of the $\text{NO}_3^-/\text{NO}_2^-$ present in the sediments, H_2S and Fe(II) can diffuse without restriction across the sediment-water interface and upwards in the water column until they are oxidized at the oxycline, with subsequent formation of particulate Fe(III)OOH , organically complexed Fe (Fe(III)L) and sulfate (SO_4^{2-}). Similar to Scenario 1, a small fraction of Fe(II) is kept in solution by photochemical reduction processes in surface waters. The right part of the sketch illustrates Advanced scenario 2: Similar to scenario 2, but due to long lasting sedimentary release, H_2S and Fe(II) concentrations increase in the water column. We hypothesize that elevated concentrations of H_2S and Fe(II) in the millimolar range facilitate the precipitation of mackinawite (FeS_m) at the sediment-bottom water interface.

The coupling of Fe(II) and nitrite in OMZs is under debate in the scientific community. Hong and Kester¹⁹ found a linear relationship between nitrite and Fe(II) for samples from the OMZ along the Peruvian shelf and suggested this represented a common sediment source. More recently several trace metal studies conducted in open ocean OMZs observed a deep Fe(II) maximum that coincided with elevated nitrite and DFe ($\text{Fe(II)} + \text{Fe(III)}$) levels at the same depths. In the Peruvian OMZ a filament with elevated Fe(II) and nitrite concentrations in the center of the otherwise DFe rich OMZ has been located in the anoxic core zone between 300 m and 400 m water depth^{9,20,75}. The filament extended from the coast to ca. 1,000 km off-shore and was related to off-shore Fe transport. A similar pattern has also been described for the OMZ in the Arabian Sea^{76,77} and both were interpreted as a coupling between nitrate reduction and Fe(II) accumulation, with Fe(III) being microbially reduced⁷⁸. In contrast, Rickard and Luther III²⁴, Scholz, *et al.*⁷, and Heller, *et al.*¹⁸ suggested a role for nitrite in the oxidation of Fe(II) and an accumulation of particulate Fe oxyhydroxides in the anoxic water column off Peru.

Periodic sulfidic events occur in the Peruvian OMZ. Our observations indicate that the present conditions are not favorable for the formation of mackinawite in the anoxic Peruvian waters given that Fe(II) and H_2S concentrations remain consistently too low to exceed the solubility product of Fe(II) sulfide minerals. At micromolar H_2S concentrations, aqueous Fe sulfide complexes and clusters become the dominant Fe(II) species and buffer DFe through prevention of scavenging, thereby enhancing Fe solubility in the euxinic water column.

Oxygen minimum zones are projected to expand and intensify as a consequence of reduced oxygen solubility related to ocean warming, increased stratification of the water column^{15,79}, changes in oxygen consumption via biotic respiration, and changes in the large-scale overturning circulation⁸⁰. This will likely result in more frequent H_2S events and associated enhanced Fe(II) concentrations in coastal OMZs⁷². For the eastern tropical South Pacific off Peru it has been shown that eddies frequently forming at the coast can transport coastal waters far offshore within days and weeks⁸¹. This scenario may enhance the supply of DFe to local surface waters and potentially to the Fe depleted South Pacific gyre system^{3,82,83}, with positive feedbacks for primary productivity and nitrogen fixation.

Materials and Methods

Sampling locations and sampling. Seawater samples for trace metal analysis were collected at three sites on the Peruvian shelf during the upwelling season in December and January 2008/09 (RV *Meteor* cruise M77-3). Sites were located at 10.00°S, 78.38°W (1), 12.33°S, 77.00°W (2) and 16.00°S, 74.24°W (3) (Fig. 1). The water depth was 112 m at site 1, 98 m at site 2 and 255 m at site 3. Samples for trace metal analysis were collected using trace metal clean Go-Flo bottles (General Oceanics) attached to a Kevlar wire. Go-Flo bottles were deployed once at sites 2 and 3, while at site 1 the Go-Flo bottles were deployed five times within 24 h. After recovery, bottles were transferred into a clean lab container. The seawater samples were filtered using cartridge filters (0.2 µm, Sartobran 300, Sartorius) and dispensed into acid cleaned 1 L low density polyethylene (LDPE, Nalgene) bottles. The samples were then acidified with quartz distilled hydrochloric acid to pH 1.8 (17.8 µmol H⁺ L⁻¹). Unfiltered seawater samples for on-board Fe(II) analysis were dispensed into opaque acid cleaned 60 mL LDPE bottles under normal filtered air. The Fe(II) analysis was carried out immediately after sample collection. All sample handling was performed in a laminar flow hood.

Sample analysis. Dissolved Fe concentrations were determined half a year later using graphite furnace atomic absorption spectrometry (Perkin Elmer, 4100 ZL) following Grasshoff *et al.*⁸⁴. The blank and limit of detection (LOD) (three times the standard deviation of the blank measurement) for Fe concentrations were 0.104 nmol L⁻¹ and 0.079 nmol L⁻¹, respectively. The accuracy of the analytical procedure was evaluated by the analysis of certified seawater standard NASS-5 (National Research Council of Canada) and SAFe. Our Fe values agree well with the certified values for NASS 5 and the SAFe data (NASS 5: 26.3 ± 1.1 nmol kg⁻¹ (certified: 25.7 ± 2.0 nmol kg⁻¹); SAFe S: 0.112 ± 0.013 nmol kg⁻¹ (census: 0.093 ± 0.008 nmol kg⁻¹); SAFe D2: 0.83 ± 0.13 nM Fe (census: 0.933 ± 0.023 nmol kg⁻¹)). The precision of the method is 3–5%.

Fe(II) concentrations were measured on samples collected at site 2 immediately upon collection by a chemiluminescence flow injection analysis following the method of Croot and Laan⁸⁵, which has a LOD of 0.1 nmol L⁻¹ Fe(II).

Nutrient and O₂ samples were obtained using Niskin bottles (General Oceanics) on a stainless steel CTD rosette deployed at the same locations. Nutrient samples were analyzed for NO₃⁻ and DIP using an autoanalyzer (TRAACS800, Bran&Lubbe) following Grasshoff *et al.*⁸⁶. NO₂⁻ was determined spectrophotometrically⁸⁶ and NH₄ was analyzed fluorometrically⁸⁷ on board. Oxygen concentrations in the water column were measured by a Seabird O₂ sensor that was calibrated with oxygen concentrations determined from CTD water samples (Winkler method⁸⁴). The latter caused a LOD of 2 µmol kg⁻¹. Additionally, at site 2 a switchable trace O₂ sensor (STOX) with a LOD of 50 nmol kg⁻¹ was used^{36,37}. For H₂S measurements seawater samples were collected with a pump-CTD³⁸ and were analyzed spectrophotometrically⁸⁸. At the pH of seawater HS⁻ represents the dominant hydrogen sulfide species, which is why the term H₂S for our study refers to the sum of H₂S, HS⁻, and S²⁺.

Data availability. The dataset generated during this study is available in the GEOMAR-OSIS repository (<https://portal.geomar.de/de/osis>).

References

- Schlösser, C. *et al.* Seasonal ITCZ migration dynamically controls the location of the (sub)tropical Atlantic biogeochemical divide. *PNAS* **111**, 1231–1232 (2014).
- Boyd, P. W. *et al.* Mesoscale iron enrichment experiments 1993–2005: Synthesis and future directions. *Science* **315**, 612–617 (2007).
- Browning, T. J. *et al.* Nutrient co-limitation at the boundary of an oceanic gyre. *Nature* **551**, 242–246 (2017).
- Milne, A. *et al.* Particulate phases are key in controlling dissolved iron concentrations in the (sub)tropical North Atlantic. *Geophys. Res. Lett.* **44**, 2377–2387 (2017).
- Elrod, V. A., Berelson, W. M., Coale, K. H. & Johnson, K. S. The flux of iron from continental shelf sediments: A missing source for global budgets. *Geophys. Res. Lett.* **31**, 1–4 (2004).
- Croot, P. L. & Hunter, K. A. Trace metal distributions across the continental shelf near Otago Peninsula, New Zealand. *Mar. Chem.* **62**, 185–201 (1998).
- Scholz, F. *et al.* Nitrate-dependent iron oxidation limits iron transport in anoxic ocean regions. *Earth Planet. Sc. Lett.* **454**, 272–281 (2016).
- Noffke, A. *et al.* Benthic iron and phosphorus fluxes across the Peruvian oxygen minimum zone. *Limnol. Oceanogr.* **57**, 851–867 (2012).
- Kondo, Y. & Moffett, J. W. Iron redox cycling and subsurface offshore transport in the eastern tropical South Pacific oxygen minimum zone. *Mar. Chem.* **168**, 95–103 (2015).
- Berner, R. A. Stability Fields of Iron Minerals in Anaerobic Marine Sediments. *J. Geol.* **72**, 826–834 (1964).
- Raiswell, R. & Canfield, D. E. Rates of reaction between silicate iron and dissolved sulfide in Peru Margin sediments. *Geochim. Cosmochim. Acta.* **60**, 2777–2787 (1996).
- Poulton, S. W., Krom, M. D. & Raiswell, R. A revised scheme for the reactivity of iron (oxyhydr)oxide minerals towards dissolved sulfide. *Geochim. Cosmochim. Acta.* **68**, 3703–3715 (2004).
- Kalvelage, T. *et al.* Aerobic Microbial Respiration In Oceanic Oxygen Minimum Zones. *PLOS ONE* **10**, e0133526 (2015).
- Brandt, P. *et al.* On the role of circulation and mixing in the ventilation of oxygen minimum zones with a focus on the eastern tropical North Atlantic. *Biogeosciences* **12**, 489–512 (2015).
- Stramma, L., Johnson, G. C., Firing, E. & Schmidtko, S. Eastern Pacific oxygen minimum zones: Supply paths and multidecadal changes. *J. Geophys. Res.* **115**, C09011 (2010).
- Scholz, F. *et al.* Early diagenesis of redox-sensitive trace metals in the Peru upwelling area - response to ENSO-related oxygen fluctuations in the water column. *Geochim. Cosmochim. Acta.* **75**, 7257–7276 (2012).
- Bruland, K. W., Rue, E. L., Smith, G. J. & DiTullio, G. R. Iron, macronutrients and diatom blooms in the Peru upwelling regime: brown and blue waters of Peru. *Mar. Chem.* **93**, 81–103 (2005).
- Heller, M. I. *et al.* Accumulation of Fe oxyhydroxides in the Peruvian oxygen deficient zone implies non-oxygen dependent Fe oxydation. *Geochim. Cosmochim. Acta.* **211**, 174–193 (2017).
- Hong, H. & Kester, D. R. Redox state of iron in the offshore waters of Peru. *Limnol. Oceanogr.* **31**, 512–524 (1986).
- Vedamati, J., Goepfert, T. J. & Moffett, J. W. Iron speciation in the eastern tropical South Pacific oxygen minimum zone off Peru. *Limnol. Oceanogr.* **59**, 1945–1957 (2014).

21. Chever, F. *et al.* Total dissolvable and dissolved iron isotopes in the water column of the Peru upwelling regime. *Geochim. Cosmochim. Acta* **162**, 66–82 (2015).
22. Bonneville, S., Behrends, T. & Van Cappellen, P. Solubility and dissimilatory reduction kinetics of iron(III) oxyhydroxides: A linear free energy relationship. *Geochim. Cosmochim. Acta* **73**, 5273–5282 (2009).
23. Raiswell, R. & Canfield, D. E. The Iron Biogeochemical Cycle Past and Present. *Geochem. Perspect.* **1**, 1–2 (2012).
24. Rickard, D. & Luther, G. W. III. Chemistry of Iron sulfids. *Chem. Rev.* **107**, 514–562 (2007).
25. Rickard, D., Schoonen, M. A. A. & Luther, G. W. In *Geochemical Transformations of Sedimentary Sulfur* Vol. 612 ACS Symposium Series Ch. 9, 168–193 (American Chemical Society, 1995).
26. Rickard, D. The solubility of FeS. *Geochim. Cosmochim. Acta* **70**, 5779–5789 (2006).
27. Butler, I. B., Archer, C., Vance, D., Oldroyd, A. & Rickard, D. Fe isotope fractionation on FeS formation in ambient aqueous solution. *Earth Planet. Sci. Lett.* **236**, 430–442 (2005).
28. Luther, G. W., Theberge, S. M. & Rickard, D. T. Evidence for aqueous clusters as intermediates during zinc sulfide formation. *Geochim. Cosmochim. Acta* **63**, 3159–3169 (1999).
29. Luther, G. W., Rickard, D. T., Theberge, S. & Oldroyd, A. Determination of Metal (Bi)Sulfide Stability Constants of Mn²⁺, Fe²⁺, Co²⁺, Ni²⁺, Cu²⁺, and Zn²⁺ by Voltammetric Methods. *Environ. Sci. Technol.* **30**, 671–679 (1996).
30. Sell, K. S. & Morse, J. W. Dissolved Fe²⁺ and ΣH₂S Behavior in Sediments Seasonally Overlain by Hypoxic-to-anoxic Waters as Determined by CSV Microelectrodes. *Aquat. Geochem.* **12**, 179–198 (2006).
31. Kraal, P., Burton, E. D. & Bush, R. T. Iron monosulfide accumulation and pyrite formation in eutrophic estuarine sediments. *Geochim. Cosmochim. Acta* **122**, 75–88 (2013).
32. Skei, J. M. Formation of framboidal iron sulfide in the water of a permanently anoxic fjord - Framvaren, South Norway. *Mar. Chem.* **23**, 345–352 (1988).
33. Butler, I. B. & Rickard, D. Framboidal pyrite formation via the oxidation of iron (II) monosulfide by hydrogen sulphide. *Geochim. Cosmochim. Acta* **64**, 2665–2672 (2000).
34. Stramma, L. *et al.* Observed El Niño conditions in the eastern tropical Pacific in October 2015. *Ocean Science* **12**, 861–873 (2016).
35. Thomsen, S. *et al.* Do submesoscale frontal processes ventilate the oxygen minimum zone off Peru? *Geophys. Res. Lett.* **43**, 8133–8142 (2016).
36. Revsbech, N. P. *et al.* Determination of ultra-low oxygen concentrations in oxygen minimum zones by the STOX sensor. *Limnol. Oceanogr.: Methods* **7**, 371–381 (2009).
37. Kalvelage, T. *et al.* Nitrogen cycling driven by organic matter export in the South Pacific oxygen minimum zone. *Nature Geosci* **6**, 228–234 (2013).
38. Schunck, H. *et al.* Giant Hydrogen Sulfide Plume in the Oxygen Minimum Zone off Peru Supports Chemolithoautotrophy. *PLoS ONE* **8**, 1–18 (2013).
39. Sommer, S. *et al.* Depletion of oxygen, nitrate and nitrite in the Peruvian oxygen minimum zone cause an imbalance of benthic nitrogen fluxes. *Deep-Sea Res. I* **112**, 113–122 (2016).
40. Scholz, F., McManus, J., Mix, A. C., Hensen, C. & Schneider, R. R. The impact of ocean deoxygenation on iron release from continental margin sediments. *Nature Geosci* **7**, 433–437 (2014).
41. Deutsch, C., Sarmiento, J. L., Sigman, D. M., Gruber, N. & Dunne, J. P. Spatial coupling of nitrogen inputs and losses in the ocean. *Nature* **445**, 163–167 (2007).
42. Monterey, G. I. & deWitt, L. M. Seasonal variability of global mixed layer depth from WOD98 temperature and salinity profiles. *NOAA Technical Memorandum NMFS*, 1–61 (2000).
43. Hsu-Kim, H., Mullaugh, K. M., Tsang, J. J., Yucel, M. & Luther, G. W. Formation of Zn- and Fe-sulfides near hydrothermal vents at the Eastern Lau Spreading Center: implications for sulfide bioavailability to chemoautotrophs. *Geochemical Transactions* **9**, 1–14 (2008).
44. Gustafsson, J. P. *Visual MINTEQ ver. 3.1*, <https://vminteq.lwr.kth.se> (2014).
45. Feely, R. A., Sabine, C. L., Hernandez-Ayon, J. M., Janson, D. & Hales, B. Evidence for upwelling of corrosive “acidified” water onto the continental shelf. *Science* **320**, 1490–1492 (2008).
46. León, V. *et al.* pH como un Trazador de la Variabilidad Biogeoquímica en el Sistema de Humboldt. *Boletín del IMARPE* 26/1–2 (2011).
47. Dryssen, D. Metal complex formation in sulfidic seawater. *Mar. Chem.* **15**, 285–293 (1985).
48. Landing, W. M. & Westerlund, S. The solution chemistry of iron(II) in Framvaren fjord. *Mar. Chem.* **23**, 329–343 (1988).
49. Jørgensen, B. B., Fossing, H., Wirsén, C. O. & Jannasch, H. W. Sulfide oxidation in the anoxic Black Sea chemocline. *Deep-Sea Res.* **38**, S1083–S1103 (1991).
50. Kononov, S., Samodurov, A., Oguz, T. & Ivanov, L. Parameterization of iron and manganese cycling in the Black Sea suboxic and anoxic environment. *Deep-Sea Res. I* **51**, 2027–2045 (2004).
51. Kremling, K. The behavior of Zn, Cd, Cu, Ni, Co, Fe, and Mn in anoxic baltic waters. *Mar. Chem.* **13**, 87–108 (1983).
52. Millero, F. J., Yao, W. & Aicher, J. The speciation of Fe(II) and Fe(III) in natural waters. *Mar. Chem.* **50**, 21–39 (1995).
53. Mopper, K. & Taylor, B. F. In *Organic Marine Geochemistry* ACS Symposium Series 324–339 (ACS, 1986).
54. Hamed, M. Y. & Silver, J. Studies on the reactions of ferric iron with glutathione and some related thiols. Part II. Complex formation in the pH range three to seven. *Inorg. Chim. Acta* **80**, 115–122 (1983).
55. Gledhill, M. & Buck, K. N. The Organic Complexation of Iron in the Marine Environment: A Review. *Frontiers Microbiol.* **3**(69), 1–17 (2012).
56. Bergquist, B. A., Wu, J. & Boyle, E. A. Variability in oceanic dissolved iron is dominated by the colloidal fraction. *Geochim. Cosmochim. Acta* **71**, 2960–2974 (2007).
57. Schafstall, J., Dengler, M., Brandt, P. & Bange, H. Tidal-induced mixing and diapycnal nutrient fluxes in the Mauritanian upwelling region. *J. Geophys. Res.: Oceans* **115**, 1–19 (2010).
58. Osborn, T. R. Estimates of the local-rate of vertical diffusion from dissipation measurements. *J. Phys. Oceanogr.* **10**, 83–89 (1980).
59. de Jong, J. *et al.* Natural iron fertilization of the Atlantic sector of the Southern Ocean by continental shelf sources of the Antarctic Peninsula. *J. Geophys. Res.* **117**, 1–25 (2012).
60. Schütte, F. *et al.* Characterization of “dead-zone” eddies in the eastern tropical North Atlantic. *Biogeosciences* **13**, 5865–5881 (2016).
61. Stams, A. J. M. *et al.* Exocellular electron transfer in anaerobic microbial communities. *Environ. Microbiol.* **8**, 371–382 (2006).
62. Marzocchi, U. *et al.* Electric coupling between distant nitrate reduction and sulfide oxidation in marine sediment. *ISME J* **8**, 1682–1690 (2014).
63. Canfield, D. E. *et al.* A Cryptic Sulfur Cycle in Oxygen-Minimum-Zone Waters off the Chilean Coast. *Science* **330**, 1375–1378 (2010).
64. Kappler, A., Schink, B. & Newman, D. K. Fe(III) mineral formation and cell encrustation by the nitrate-dependent Fe(II)-oxidizer strain BoFeN1. *Geobiology* **3**, 235–245 (2005).
65. Pantke, C. *et al.* Green Rust Formation during Fe(II) Oxidation by the Nitrate-Reducing Acidovorax sp. Strain BoFeN1. *Environ. Sci. Technol.* **46**, 1439–1446 (2012).
66. Jewell, T. N. M., Karaoz, U., Brodie, E. L., Williams, K. H. & Beller, H. R. Metatranscriptomic evidence of pervasive and diverse chemolithoautotrophy relevant to C, S, N and Fe cycling in a shallow alluvial aquifer. *ISME J* **10**, 2106–2117 (2016).

67. Laufer, K., Røy, H., Jørgensen, B. B. & Kappler, A. Evidence for the Existence of Autotrophic Nitrate-Reducing Fe(II)-Oxidizing Bacteria in Marine Coastal Sediment. *Appl. Environ. Microbiol.* **82**, 6120–6131 (2016).
68. Dugdale, R. C., Goering, J. J., Barber, R. T., Smith, R. L. & Packard, T. T. Denitrification and hydrogen sulfide in the Peru upwelling region during 1976. *Deep-Sea Res.* **24**, 601–608 (1977).
69. Naqvi, S. W. A. *et al.* Increased marine production of N₂O due to intensifying anoxia on the Indian continental shelf. *Nature* **408**, 346–349 (2000).
70. Borchers, S. L., Schnetger, B., Böning, P. & Brumsack, H. J. Geochemical signatures of the Namibian diatom belt: Perennial upwelling and intermittent anoxia. *Geochem. Geophys. Geosyst.* **6**, Q06006 (2005).
71. Brüchert, V., Currie, B. & Peard, K. R. Hydrogen sulphide and methane emissions on the central Namibian shelf. *Prog. Oceanogr.* **83**, 169–179 (2009).
72. Lavik, G. *et al.* Detoxification of sulphidic African shelf waters by blooming chemolithotrophs. *Nature* **457**, 581–584 (2009).
73. Ulloa, O., Canfield, D. E., DeLong, E. F., Letelier, R. M. & Stewart, F. J. Microbial oceanography of anoxic oxygen minimum zones. *PNAS* **109**, 15996–16003 (2012).
74. Rose, A. L. & Waite, T. D. Kinetic Model for Fe(II) Oxidation in Seawater in the Absence and Presence of Natural Organic Matter. *Environ. Sci. Technol.* **36**, 433–444 (2002).
75. Thamdrup, B., Dalsgaard, T. & Peter Revsbech, N. Widespread functional anoxia in the oxygen minimum zone of the eastern South Pacific. *Deep-Sea Res.* **165**, 36–45 (2012).
76. Kondo, Y. & Moffett, J. W. Dissolved Fe(II) in the Arabian Sea oxygen minimum zone and western tropical Indian Ocean during the inter-monsoon period. *Deep-Sea Res.* **173**, 73–83 (2013).
77. Moffett, J. W., Goepfert, T. J. & Naqvi, S. W. A. Reduced iron associated with secondary nitrite maxima in the Arabian Sea. *Deep-Sea Res.* **154**, 1341–1349 (2007).
78. Moffett, J. W. & Goepfert, T. J. & Wajih A. Naqvi, S. Reduced iron associated with secondary nitrite maxima in the Arabian Sea. *Deep-Sea Res.* **154**, 1341–1349 (2007).
79. Stramma, L. *et al.* Expansion of oxygen minimum zones may reduce available habitat for tropical pelagic fishes. *Nature Climate Change* **2**, 33–37 (2012).
80. Shepherd, J. G., Brewer, P. G., Oschlies, A. & Watson, A. J. Ocean ventilation and deoxygenation in a warming world: introduction and overview. *Philos Trans A Math Phys Eng Sci* **375**, 1–9 (2017).
81. Thomsen, S. *et al.* The formation of a subsurface anticyclonic eddy in the Peru-Chile Undercurrent and its impact on the near-coastal salinity, oxygen, and nutrient distributions. *J. Geophys. Res.: Oceans* **121**, 476–501 (2016).
82. Moore, C. M. *et al.* Processes and patterns of oceanic nutrient limitation. *Nat Geosci* **6**, 701–710 (2013).
83. Letscher, R. T., Primeau, F. & Moore, J. K. Nutrient budgets in the subtropical ocean gyres dominated by lateral transport. *Nat. Geosci.* **9**(815), 815–821 (2016).
84. Grasshoff, K., Erhardt, M. & Kremling, K. Methods of seawater analysis. 3 edn, Vol. 3 Edition (Wiley-VCH, 1999).
85. Croot, P. L. & Laan, P. Continuous shipboard determination of Fe(II) in Polar waters using flow injection analysis with chemiluminescence detection. *Anal. Chim. Acta* **466**, 261–273 (2002).
86. Hansen, H. P. & Koroleff, F. In Methods of seawater analysis (eds Grasshoff, K., Kremling, K. & Ehrhardt, M.) 159–228 (Wiley VCH, Weinheim, 1999).
87. Holmes, R. M., Aminot, A., Kerouel, R., Hooker, B. A. & Peterson, B. J. A simple and precise method for measuring ammonium in marine and freshwater ecosystems. *Can. J. Fish. Aquat. Sci.* **56**, 1801–1808 (1999).
88. Cline, J. D. Spectrophotometric determination of hydrogen sulfide in natural waters. *Limnol. Oceanogr.* **14**, 454–458 (1969).

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Author Contributions

C.S. and P.L.C. designed the research; C.S., P.L.C. and M.D. collected the samples/data; C.S., G.L. and P.S. analyzed the samples; C.S., P.L.C., G.L., M.F. and E.A. wrote the manuscript.

Additional Information

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