

Index

List of figures	VII
List of abbreviations	VII
1 Introduction.....	- 1 -
1.1 Fertilization in mammals	- 1 -
1.2 Zona pellucida glycoproteins.....	- 4 -
1.3 Ion channels and transporters required for sperm fertility	- 8 -
1.3.1 CatSper, the principal sperm Ca²⁺ channel	- 8 -
1.3.2 Slo3 potassium channel	- 10 -
1.3.3 The proton channel Hv1	- 10 -
1.3.4 Sodium-proton exchanger family	- 11 -
1.4 ZP signaling	- 12 -
1.5 Aim of this PhD thesis.....	- 14 -
2 Materials and methods	- 15 -
2.1 Materials.....	- 15 -
2.1.1 Chemicals	- 15 -
2.1.2 Antibodies	- 15 -
2.2 Escherichia coli culture	- 17 -
2.2.1 Bacterial strains and vectors	- 17 -
2.2.2 Composition and preparation of E. coli culture media	- 17 -
2.2.3 Amplification of E. coli cultures for plasmid preparation	- 17 -
2.2.4 Generation of competent E. coli cells	- 18 -
2.3 Preparation of nucleic acids	- 18 -
2.3.1 Mini-preparation of plasmid DNA via alkaline lysis	- 18 -
2.3.2 Midi- and maxi-preparation of plasmid DNA	- 18 -
2.3.3 Preparation of genomic DNA from mouse tissue	- 19 -
2.4 Separation, purification and quantification of plasmid DNA and DNA fragments	- 19 -
2.4.1 Agarose-gel electrophoresis	- 19 -
2.4.2 Elution of DNA from agarose gels	- 20 -
2.4.3 Ethanol precipitation	- 20 -

2.4.4	Purification with SureClean.....	- 20 -
2.4.5	Photometric quantification of nucleic acid concentration	- 21 -
2.4.6	Quantification of nucleic acid concentration by agarose-gel electrophoresis.	- 21 -
2.5	Modification of nucleic acids.....	- 21 -
2.5.1	Restriction digest of plasmid DNA	- 21 -
2.5.2	Ligation of DNA fragments	- 21 -
2.5.3	Transformation	- 21 -
2.6	Polymerase chain-reaction (PCR)	- 22 -
2.6.1	Primer synthesis	- 22 -
2.6.2	PCR conditions	- 22 -
2.7	Mammalian cell culture	- 23 -
2.7.1	Sterile work.....	- 23 -
2.7.2	Cell lines	- 23 -
2.7.3	Continuous culture of HEK293, HEK293T cells, and hybridoma cells	- 24 -
2.7.4	Cryopreservation of mammalian cell lines	- 24 -
2.7.5	Transient transfection	- 24 -
2.7.6	Stem cell culture	- 25 -
2.8	Immunofluorescence.....	- 25 -
2.8.1	Immunocytochemistry	- 25 -
2.8.2	Sectioning of frozen tissue	- 25 -
2.8.3	β-galactosidase staining of testis sections	- 26 -
2.9	Protein preparation	- 26 -
2.9.1	Protein preparation from mammalian cells	- 26 -
2.9.2	Protein preparation from mouse tissue	- 26 -
2.9.3	Protein quantification with bicinchoninic acid.....	- 27 -
2.10	Purification of proteins from cell supernatant	- 27 -
2.10.1	Batch purification via Ni-NTA agarose.....	- 27 -
2.10.2	Large-scale protein purification using the ÄKTA system	- 28 -
2.10.3	Buffer exchange	- 28 -
2.10.4	Protein quantification	- 28 -
2.10.5	PNGase digestion.....	- 28 -

2.11	Separation and detection of specific proteins.....	- 29 -
2.11.1	Reducing SDS-polyacrylamide gel electrophoresis	- 29 -
2.11.2	Mass spectrometry	- 30 -
2.11.3	Transfer and immobilization of proteins by Western blotting	- 30 -
2.11.4	Immunostaining of immobilized proteins	- 31 -
2.12	Laboratory animals	- 32 -
2.12.1	Captive care and breeding.....	- 32 -
2.12.2	Isolation of native mouse zona pellucida	- 33 -
2.13	Mouse and human sperm experiments.....	- 33 -
2.13.1	Mouse sperm preparation	- 33 -
2.13.2	In-vitro fertilization	- 34 -
2.13.3	Human sperm preparation	- 35 -
2.13.4	Sperm membrane protein preparation	- 36 -
2.13.5	Acrosome reaction assay.....	- 36 -
2.13.6	Antigen retrieval for ICC on sperm	- 37 -
2.13.7	STORM analysis of sperm flagellar proteins.....	- 37 -
2.13.8	Flagellar beat analysis	- 38 -
2.13.9	Electrophysiological recordings from human sperm	- 38 -
2.14	Fluorometric measurements in sperm	- 39 -
2.14.1	Fluorescent Ca^{2+} indicator CAL520.....	- 39 -
2.14.2	Fluorescent pH_i indicator BCECF	- 41 -
2.14.3	Ca^{2+} and pH_i fluorimetry in multi-well plates	- 43 -
2.14.4	Stopped-flow device.....	- 43 -
2.14.5	Ca^{2+} and pH_i fluorimetry in the stopped-flow device.....	- 44 -
2.14.6	Calculation of EC_{50} values from dose-response curves	- 46 -
3	Results	- 47 -
3.1	The action of zona pellucida glycoproteins in mouse sperm	- 47 -
3.1.1	Isolation and functional characterization of native mouse zona pellucida glycoproteins	- 47 -
3.1.2	ZP-evoked $[\text{Ca}^{2+}]_i$ responses	- 49 -
3.1.3	ZP-evoked pH_i responses.....	- 51 -

3.1.4	Molecular mechanism underlying the ZP-evoked pH_i response	- 56 -
3.1.5	The ZP-evoked pH_i response in mouse involves the NHA1 Na⁺/H⁺ exchanger	- 60 -
3.1.6	The ZP-induced acrosome reaction in mouse sperm requires a polarized membrane potential and alkalinization	- 65 -
3.1.7	Summary	- 66 -
3.1.8	Heterologous expression and characterization of mouse zona pellucida glycoproteins	- 67 -
3.2	The action of human zona pellucida glycoproteins in human sperm	- 72 -
3.2.1	Heterologous expression and characterization of human zona pellucida glycoproteins	- 72 -
3.2.2	ZP-evoked [Ca²⁺]_i responses	- 73 -
3.2.3	Species-specificity of heterologous mouse and human ZP glycoproteins	- 77 -
3.2.4	ZP-evoked pH_i responses	- 78 -
3.2.5	Molecular mechanism underlying the ZP glycoprotein-evoked pH_i response.	- 80 -
3.2.6	ZP glycoproteins directly activate human CatSper	- 85 -
4	Discussion	- 88 -
4.1	ZP signaling in mouse and human sperm	- 89 -
4.2	Regulation of ZP-evoked acrosome reaction	- 91 -
4.3	Which roles play different ZP glycoproteins in ZP signaling?	- 92 -
4.4	Outlook	- 93 -
5	References	- 96 -
6	Appendix	- 113 -
7	Acknowledgement	- 115 -