

# Systems biology

# **TEPIC 2**—an extended framework for transcription factor binding prediction and integrative epigenomic analysis

Florian Schmidt () <sup>1,2,3,\*</sup>, Fabian Kern () <sup>1,2,4</sup>, Peter Ebert () <sup>2,3</sup>, Nina Baumgarten () <sup>1,2,5,6</sup> and Marcel H. Schulz () <sup>1,2,5,6,\*</sup>

<sup>1</sup>High throughput Genomics and Systems Biology, Cluster of Excellence on Multimodal Computing and Interaction, <sup>2</sup>Computational Biology and Applied Algorithmics, Max Planck Institute for Informatics, <sup>3</sup>Graduate School of Computer Science and <sup>4</sup>Chair for Clinical Bioinformatics, Saarland Informatics Campus, Saarbrücken 66123, Germany, <sup>5</sup>Institute for Cardiovascular Regeneration, Goethe University and <sup>6</sup>German Center for Cardiovascular Research, Partner site Rhein-Main, Frankfurt am Main 60590, Germany

\*To whom correspondence should be addressed.

Associate Editor: Bonnie Berger

Received on June 28, 2018; revised on September 4, 2018; editorial decision on September 27, 2018; accepted on October 8, 2018

#### Abstract

**Summary:** Prediction of transcription factor (TF) binding from epigenetics data and integrative analysis thereof are challenging. Here, we present TEPIC 2 a framework allowing for fast, accurate and versatile prediction, and analysis of TF binding from epigenetics data: it supports 30 species with binding motifs, computes TF gene and scores up to two orders of magnitude faster than before due to improved implementation, and offers easy-to-use machine learning pipelines for integrated analysis of TF binding predictions with gene expression data allowing the identification of important TFs. **Availability and implementation:** TEPIC is implemented in C++, R, and Python. It is freely available at https://github.com/SchulzLab/TEPIC and can be used on Linux based systems. **Contact:** fschmidt@mmci.uni-saarland.de or marcel.schulz@em.uni-frankfurt.de

Supplementary information: Supplementary data are available at Bioinformatics online.

# **1** Introduction

Transcription Factors (TFs) are key players of transcriptional regulation. Prediction of TF binding is essential to gain a deeper understanding of their function. While experimental identification of TF binding is possible through laborious and expensive ChIP-seq assays, several computational approaches have been proposed to identify TF binding sites (TFBSs) (Jayaram *et al.*, 2016). These predictions have been successfully augmented using epigenetics data (Cuellar-Partida *et al.*, 2012; Pique-Regi *et al.*, 2011; Sherwood *et al.*, 2014). As delineated in Supplementary Section 1, *TEPIC* 2 builds upon and extends the functionality of existing TFBS prediction tools. Among other features, *TEPIC* 2 allows the direct aggregation of TFBS predictions on the gene level and uses these scores to gain novel insights on cell type specific functions of TFs via several machine learning analysis. This is a unique feature not supported by competitive TFBS prediction approaches (Supplementary Tables S1 and S2). Compared to its predecessor, *TEPIC 2* has substantially lower runtime, contains an extended set of TF motifs, offers various means for downstream machine learning analyses as easy-to-use pipelines, and adds new functionalities to compute TF gene scores.

#### 2 Features

The core functionalities of *TEPIC 2* are to predict TFBS in user provided regions and to aggregate them to TF gene scores. The TF gene score computation has been modified to compute statistical features such as region length, region count, and the signal of an epigenetic assay within the considered regions. *TEPIC 2* can compute a binary binding assessment, i.e. a TF binds or does not bind, based on p-values obtained using a set of background regions of similar

1608

 $\ensuremath{\mathbb{C}}$  The Author(s) 2018. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

characteristics as the input set. This feature complements the continuous TF affinity values of TRAP, which are not suitable for all downstream applications (Supplementary Section 4).

Additionally, the aforementioned TF gene scores can be used in several integrative analysis workflows (Supplementary Section 7, 8 and 9). *INVOKE* refers to a sparse linear regression model to reveal key TFs potentially regulating transcription. It highlighted several known tissue-specific regulators in liver hepatocytes and CD4+ T cells (Schmidt *et al.*, 2017) and is also available as a web-server (Kehl *et al.*, 2017). Besides, *TEPIC 2* includes a sparse logistic regression classifier to infer TFs related to gene expression changes between samples (*DYNAMITE*). *DYNAMITE* has been successfully applied to discover regulators of CD4+ T cell differentiation (Durek *et al.*, 2012) to uncover master regulatory TFs from paired timeseries expression and epigenomics data (*EPIC-DREM*), which was used to analyze mesenchymal stem-cell differentiation of osteoblasts and adipocytes (Gerard *et al.*, 2018).

Furthermore, we considerably extended the set of TF motifs readily available in *TEPIC 2*. Now, this resource contains 30 species-specific and six taxonomy-specific sets from JASPAR (Mathelier *et al.*, 2016), as well as aggregated sets for humans, mice and vertebrates containing 561, 380 and 690 TF motifs (Supplementary Section 3). To streamline the training and interpretation of statistical models (Supplementary Fig. S1), we provide clustered versions of the merged TF motif files, representing families of binding motifs with high similarity (Pape *et al.*, 2008).

## **3 Implementation**

TEPIC 2 uses a parallelized C++ implementation of TRAP (Roider *et al.*, 2007) that is considerably faster than the previous *R* implementation. Runtime was further reduced by using more efficient search algorithms and by enabling pre-filtered analyses of samples in minutes (Fig. 1a, Supplementary Table S5, Supplementary Fig. S2 and Supplementary Section 5). We evaluated the accuracy of TFBS predictions from *TEPIC 2* using TF footprints called with HINT-BC (Gusmao *et al.*, 2016) on ENCODE data (The ENCODE Project Consortium, 2012). In comparison to established tools for TFBS prediction using epigenomics data (Cuellar-Partida *et al.*, 2012; Sherwood *et al.*, 2014), *TEPIC* performs favorably in terms of area under the precision recall curve (AUPR) (Fig. 1b, Supplementary Fig. S3 and Supplementary Section 2. The machine learning pipelines



**Fig. 1.** (a) Runtime comparison of *TEPIC* to *TEPIC 2* using a subset of 458 human TFs. While the original implementation ran up to 1300 minutes to compute TFBS, *TEPIC 2* is able to compute TF affinities for peaks in the vicinity of genes in at most 15 minutes. We used four cell line samples and three primary human hepatocyte samples (LiHe1–3) to conduct the runtime experiments. (b) We compared *TEPIC* TF affinities computed in footprints called with HINT-BC (Gusmao *et al.*, 2016) in four different cell-lines in terms of AUPR against PIQ (Sherwood *et al.*, 2014) and an extension of the widely used method Fimo, called Fimo-Prior (Cuellar-Partida *et al.*, 2012). Notably, TF affinities computed with TEPIC outperform both PIQ and Fimo-Prior

included in *TEPIC 2* are implemented in *R*. Both workflows deliver results that are easy to interpret, also for non-expert users, due to automated figure generation and extensive documentation. As input, the pipelines require standard file formats, e.g. *bed* files for candidate TFBS and tab delimited *txt* files containing gene expression data. *TEPICs* full functionality is brought to the user via *start-to-finish* pipelines, which are automatically installed with *TEPIC 2*.

#### 4 Conclusion

TEPIC 2 is a fast and easy-to-use tool for TFBS prediction combined with integrative analysis capabilities for gene expression and epigenomic data. TFBS prediction and downstream machine learning pipelines for various analysis settings allow a deep, seamless exploration of epigenomic datasets supporting data driven hypothesis generation about the role of individual TFs in complex regulatory landscapes.

#### Acknowledgements

We thank Helge Roider who wrote the original C++ implementation of TRAP, as well as the DEEP and ENCODE consortia for processing and providing the data used in this project.

#### Funding

This work has been supported by the German Federal Ministry of Education and Research in Germany (BMBF) [01DP17005, 01KU1216A] and the Cluster of Excellence on Multimodal Computing and Interaction (DFG) [EXC248].

Conflict of Interest: none declared.

## References

- Cuellar-Partida, G. *et al.* (2012) Epigenetic priors for identifying active transcription factor binding sites. *Bioinformatics*, 28, 56–62.
- Durek, P. et al. (2016) Epigenomic profiling of human CD4+ T cells supports a linear differentiation model and highlights molecular regulators of memory development. *Immunity*, 45, 1148–1161.
- Gerard, D. et al. (2018) Temporal epigenomic profiling identifies AHR and GLIS1 as super-enhancer controlled regulators of mesenchymal multipotency. bioRxiv.
- Gusmao, E. et al. (2016) Analysis of computational footprinting methods for DNase sequencing experiments. Nat. Methods, 13, 303–309.
- Jayaram, N. et al. (2016) Evaluating tools for transcription factor binding site prediction. BMC Bioinformatics. doi: 10.1186/s12859-016-1298-9.
- Kehl, T. et al. (2017) RegulatorTrail: a web service for the identification of key transcriptional regulators. Nucleic Acids Res., 45, W146–W153.
- Mathelier, A. et al. (2016) JASPAR 2016: a major expansion and update of the open-access database of transcription factor binding profiles. Nucleic Acids Res., 44, D110–D115.
- Pape,U.J. et al. (2008) Natural similarity measures between position frequency matrices with an application to clustering. *Bioinformatics*, 24, 350–357.
- Pique-Regi, R. et al. (2011) Accurate inference of transcription factor binding from DNA sequence and chromatin accessibility data. Genome Res., 21, 447–455.
- Roider, H.G. *et al.* (2007) Predicting trancription factor affinities to DNA from a biophysical model. *Bioinformatics*, 23, 134–141.
- Schmidt, F. et al. (2017) Combining transcription factor binding affinities with open-chromatin data for accurate gene expression prediction. Nucleic Acids Res., 45, 54–66.
- Schulz, M.H. et al. (2012) DREM 2.0: improved reconstruction of dynamic regulatory networks from time-series expression data. BMC Syst. Biol., 6, 104.
- Sherwood,R.I. *et al.* (2014) Discovery of directional and nondirectional pioneer transcription factors by modeling DNase profile magnitude and shape. *Nat. Biotechnol.*, **32**, 171–178.
- The ENCODE Project Consortium (2012) An integrated encyclopedia of DNA elements in the human genome. *Nature*, **489**, 741457–741474.