

## **Response to Dr P. J. Kramer's article, 'Changing concepts regarding plant water relations', Volume 11, Number 7, pp. 565–568**

E.-D. SCHULZE, E. STEUDLE, T. GOLLAN & U. SCHURR Lehrstuhl Pflanzenökologie, Universität Bayreuth, BOX 10 12 51, D-8580 Bayreuth, West Germany

Looking back on a more than 50 year's experience in the field, Kramer is concerned that certain sound principles of plant water relations seem to be questioned which, in his opinion, has no adequate basis. Physical concepts tend to be modified or replaced by 'weaker' physiological or biochemical principles. To clarify his concern, Kramer questions the value of four topics which have been raised recently:

- (1) Cell volume or relative water content may be better parameters expressing metabolic effects on water status than water potential ( $\psi$ ). He suggests that water potential should be used because it is the wider concept based on thermodynamic laws.
- (2) Growth may occur despite turgor loss. Kramer reminds that this is not a new concept, since growth has always been considered to depend on both, water status and cell wall extensibility.
- (3) The importance of hydraulic vs biochemical effects of water stress, especially with respect to stomatal response.
- (4) The role of root vs shoot as the primary sensor for water stress. He thinks that observations of root signals result from very special experimental conditions which do not generally apply to situations in the field.

The considerations of Kramer are very useful in that they focus on recent issues in plant water relations from a long-term perspective. Nevertheless, his concern may also reflect a change in scientific thinking. While previously we were used to looking for the 'most important factor', we are trained today to consider multifactorial systems, non-linear processes and changes in the optimum response. This may contribute to the discrepancies discussed.

### **Water status of plants: cell volume vs water potential**

The discussion of the issue is not new. When the concept of water potential was introduced by Slatyer (1967) and others, some physiologists opposed (e.g. Walter & Kreeb, 1970) by pointing out that water potential may not be the best parameter for describing physiological processes. There is now a general agreement that water potential gradients are the driving force for the water transport in plants although modifications of the driving force may occur in the pres-

ence of interactions between water and solute flows which may be expressed by the formalism of irreversible thermodynamics (Dainty, 1963; Zimmermann & Steudle, 1978). Usually, these coupling effects are not very important for mature plant cells, but should be important for the uptake of nutrients into roots, for growth, and for phloem transport (Fiscus, 1975; Steudle, 1985; Steudle, Oren & Schulze, 1987). However, in spite of this, we agree with Kramer that the term water potential is a useful one. Of course, this does not mean that the availability of water as expressed by  $\psi$  will limit any metabolic reaction in the living plant. The terms relative water content and cell volume are also useful and should not be omitted.

In our opinion, it would be much more important, to work out the mechanisms by which plants respond to changes in water potential, relative water content, cell volume or other physiological variables rather than to erect a hierarchy of different parameters. For example, during osmoregulation, plants adjust to changes in water potential by varying their internal osmotic pressure in order to maintain (within certain limits) a constant turgor and/or volume. For algal cells both types of regulation (turgor and volume regulation) exist (Kauss, 1977; Zimmermann, 1978). It has been shown that physical parameters such as membrane stretching and compression transform changes of water potential, turgor, and volume into metabolic responses such as active membrane transport (Coster, Steudle & Zimmermann, 1977).

There are also examples of pressure dependent transport in higher plants (e.g. Steudle, Zimmermann & Zillikens, 1982; Steudle & Zimmermann, 1984). Nevertheless, there is no doubt that there is an active response of living cells or organs, such that they can maintain their metabolism in the presence of an adverse water potential in the surroundings.

### **Growth: turgor vs wall extensibility and solute transport**

According to Lockhart (1965), growth is the result of a viscous flow of matter and a concomitant flow of water whereby the former needs a certain minimum wall stress (yield) to occur. Hence, both the hydraulic conductivity or the wall extensibility could, in principle, become rate limiting (Molz & Boyer, 1978; Steudle, 1985; Cleland, 1986). In the experiments of Boyer, Cavalieri & Schulze (1985), both parameters were

equally important. There are situations which do not allow growth despite sufficient turgor or which allow growth despite turgor loss. The recent experiments cited by Kramer only point to the fact, that 'extremes' may be more common than previously thought. In the basic experiments of Green, Erickson & Buggy (1971) on individual cells of *Nitella*, it became already clear that despite the limitation of growth by wall extensibility there was also a metabolic control of growth. This control could, in principle, work through a shift in the yield threshold or *via* solute transport, which, in turn, results in a coupling between water and solute flows (Stuedle, 1985). Recent unpublished experiments of Dr Hsiao carried out at Bayreuth on elongating maize roots support this view. Experiments with *Vigna* showed that growth was reduced when plants were grown at different air humidities despite higher leaf water contents in dry air (Nagarajah & Schulze, 1983). Again, growth appears to be not a purely physical process only driven by turgor and depending on wall mechanics. It seems to be regulated by physical *and* metabolic processes, whereby in a complex tissue or organ the decision which of the processes is rate limiting, is not easy and a more advanced model than that of Lockhart (1965) will be required in future (Shackel, Matthews & Morrison, 1987).

#### Water stress: hydraulic vs biochemical effects

Improved analytical methods show that many processes which were previously thought to be purely hydraulic in nature are in fact related to metabolism. In many cases, changes in plant water status are converted into ion fluxes. Stomatal response may be taken as an example. From the observation of Hsiao (1973) on potassium transport in guard cells, it is clear that most stomatal responses are associated with ion fluxes (Raschke, 1979). Even the stomatal responses to humidity, which were initially interpreted to be purely hydraulic (Lange *et al.*, 1971) are linked to potassium transport (Lösch, 1978). Only *transient* responses following perturbations appear to follow a 'pipe model' of plant water relations but they are generally regulated again by some metabolic action (Schulze, 1986).

Schulze & Hall (1982) pointed out that the same water potential may be reached by strongly transpiring well-watered plants at high photosynthesis or by water-stressed plants at low transpiration rates. The hydraulic effect in the well-watered plant was reversible and had no effect on plant performance, i.e. stomata were most widely open at the lowest potential. In contrast, the response to soil drought was non-reversible and indicated biochemical effects due to changes in root water status.

#### Primary signals of water stress: root vs shoot

The cuticle exhibiting a very low hydraulic conductivity (by a factor of  $10^4$ – $10^6$  less than a cell membrane;

Schönherr, 1982) uncouples the leaf from its environment. Thus, the response time of roots to changes in water potential of the environment should be much smaller than that of leaves. The initial observation on root effects was made in field experiments, where leaf conductance did not correlate with leaf water potential (Schulze *et al.*, 1980; Bates & Hall, 1981). Running (1980) correlated stomatal conductance with pre-dawn  $\psi$  as a measure of root water status. Turner, Schulze & Gollan (1985) and Gollan, Turner & Schulze (1985) found that stomatal conductance of a leaf in an environment of constant humidity did not respond uniformly to changes in bulk leaf water potential. If  $\psi$  was changed by transpiration of the whole plant, the stomatal response to  $\psi$  was less than the response to water potential at dry soil.

Even in plants where xylem water potential was close to zero and did not change while the soil was drying, leaf conductance decreased even though the leaves remained fully turgid (Gollan, Passioura & Munns, 1986). In these experiments, the root system was the only part that experienced a decrease in water potential. There was no difference in the initial phase of stomatal closure at drying soil between plants with or without controlled xylem water potential. Only at very dry soil there was an additional direct effect of leaf water potential on leaf conductance (Gollan, unpublished data).

The nature of the root to shoot communication of drying soil is still unclear but is not necessarily a hydraulic transmission. The ionic composition and the pH of the xylem sap changed with drying soil (Gollan, 1987). This could cause a release of ABA from the leaf internal storage into the apoplast (Cowan *et al.*, 1982). Also, at dry soil ABA is transported in the xylem sap at concentrations 100 times higher than at wet soil (Schurr, 1987). Associated with the increase in ABA concentration, stomatal conductance decreased but the sensitivity of the response was modulated by the ionic composition of the xylem sap (Gollan, 1987).

The time lag for the transmission of a root signal is important. In herbaceous species the xylem water stream may reach velocities of  $100 \text{ mh}^{-1}$  (Ziegler, 1982). Due to the high conductance and rigidity of the xylem system changes in water potential will be transmitted rapidly. In contrast, the time lag for solutes to appear in the shoot of trees should last much longer. In conifers water moves at a rate of about  $1 \text{ mh}^{-1}$ . In this case, a hydraulic signal may be faster. Nobel (1983) showed that xylem and phloem water relations parameters are linked. If  $\psi$  decreases in the leaf, the turgor of sieve elements will also decrease. Plants may balance the change in  $\psi$  by increasing phloem loading. If a decrease of leaf water potential would lower the pressure in the phloem, a reversed phloem flow could occur, which was not observed using  $^{14}\text{C}$  (Goeschl *et al.*, 1988). However, it has been observed that the sap flow in the phloem stops in drying soil, probably, because of an influence on the unloading process in

the root. This would create an immediate signal to the leaves, even in conifers. Again, the originating site for this signal would be the root. The indication that such regulative mechanisms exist in the field was supported by the observation that the range between maximum and minimum water potential shift during the day even when the soil dries (Kappen *et al.*, 1975).

The term water stress is often used in a very general sense. Water stress situations in field and laboratory experiments differ in terms of duration, periodicity, severity of stress and in the stressed organ. In each case the relevance of chemical versus hydraulic signals has to be evaluated. We think that root signals are important in the field and in the forest, even if the water potential of the leaves is lower than that of the roots. Especially in situations where stress develops slowly, stress-induced metabolic signals could be integrated, which is hard to envisage for hydraulic signals. One example for such an integration is the fact that ABA concentration does not only increase in dry soil but also in plants with low nitrate nutrition irrespective of soil water status (Gollan, 1987). Therefore, root-to-shoot signals may be important in environments with periodically changing soil conditions as defining the thresholds for short-term responses (metabolic or hydraulic). Structurally these thresholds could be due to morphological changes (e.g. stomatal frequency) or to changes of the sensitivity of stomata (Gollan, 1987), especially if one proposes plant growth regulators to be involved in root to shoot communication (Schurr, 1987). In this context, stress intensity has to be considered. If roots are stressed so severely that hydraulic effects do occur in the shoot, other than metabolic effects in the root will also cause changes in the shoot. One has to think about 'hydraulic' and 'metabolically' communicated stresses, whereby the relative importance of the different signals depend on the physiological state of the plant.

## Conclusions

We see that numerous hydraulic effects are inter-related with ionic or biochemical events. The nature of the primary sensor could, in principle, be both hydraulic (turgor, hydraulic conductivity, elasticity etc) or metabolic (rate of biochemical reaction, active membrane transport, translocation etc). Changes in the nutrient supply may occur prior to or in parallel with changes in water relations. In spite of this, we cannot see that the concept of water transport based on thermodynamic principles is getting lost during this discussion. However, the complementation of these concepts seems to be necessary in order to relate plant hydraulics with solute relations and metabolic events. New experimental and theoretical approaches seem to be necessary in order to arrive at a more integrative view of plant water relations. If this is done properly, the result should not contradict the sound physical basis of water transport.

## References

- Bates, L.M. & Hall, A.E. (1981) Stomatal closure with soil water depletion not associated with changes in bulk leaf water status. *Oecologia*, **41**, 62–65.
- Boyer, J.S., Cavalieri, A.J. & Schulze, E.-D. (1985) Control of the rate of cell enlargement: excision, wall relaxation, and growth-induced water potentials. *Planta*, **163**, 527–543.
- Cleland, R.E. (1986) The role of hormones in wall loosening and plant growth. *Australian Journal of Plant Physiology*, **13**, 93–104.
- Coster, H.G.L., Steudle, E. & Zimmermann, U. (1977) Turgor pressure sensing in plant cell membranes. *Plant Physiology*, **58**, 636–643.
- Cowan, I.R., Raven, J.A., Hartung, W. & Farquhar, G.D. (1982) A possible role for abscisic acid in coupling stomatal conductance and photosynthetic carbon metabolism in leaves. *Australian Journal of Plant Physiology*, **9**, 489–498.
- Dainty, J. (1963) Water relations of plant cells. *Advances in Botanical Research*, **1**, 279–326.
- Fiscus, E.L. (1975) The interaction between osmotic and pressure induced water flow in plant roots. *Plant Physiology*, **55**, 917–922.
- Goeschl, J.D., Fares, Y., Magnuson, C.E., Scheld, H.W., Strain, B.R., Jaeger, C.H. & Nelson, C.E. (1988) Short-lived isotope kinetics: a window to the inside. In *Research Instrumentation for the 21st Century* (ed. G.R. Beecher), pp. 21–52. Beltsville Symposium in Agricultural Research.
- Gollan, T. (1987) *Wechselbeziehungen zwischen Abscisinsäure, Nährstoffhaushalt und pH im Xylemsaft und ihre Bedeutung für die stomatische Regulation bei Bodenaustrocknung*. Dissertation, Universität Bayreuth.
- Gollan, T., Passioura, J.B. & Munns, R. (1986) Soil water status affects the stomatal conductance of fully turgid wheat and sunflower leaves. *Australian Journal of Plant Physiology*, **13**, 459–464.
- Gollan, T., Turner, N.C. & Schulze, E.-D. (1985) The response of stomata and leaf gas exchange to vapor pressure deficits and soil water content. III: In the sclerophyllous woody species *Nerium oleander*. *Oecologia*, **65**, 356–362.
- Green, P.B., Erickson, R.O. & Buggy, J. (1971) Metabolic and physical control of cell elongation rate-*in vivo* studies in *Nitella*. *Plant Physiology*, **47**, 423–430.
- Hsiao, T.C. (1973) Effects of water deficit on guard cell potassium and stomatal movement. *Plant Physiology Supplement*, **51**, 9.
- Kappen, L., Oertli, J.J., Lange, O.L., Schulze, E.-D., Evenari, M. & Buschbom, U. (1975) Seasonal and diurnal course of water relations of the arido-active plant *Hammada scoparia* in the Negev desert. *Oecologia*, **21**, 175–192.
- Kauss, H. (1977) Biochemistry of osmotic regulation. In *International Review of Biochemistry and Plant Biochemistry II*, Volume XIII (ed. D.H. Northcote), pp. 119–140. University Park Press, Baltimore.
- Lange, O.L., Lösch, R., Schulze, E.-D. & Kappen, L. (1971) Responses of stomata to changes in humidity. *Planta*, **100**, 76–86.
- Lockhart, J.A. (1965) An analysis of irreversible plant cell elongation. *Journal of Theoretical Biology*, **8**, 264–276.
- Lösch, R. (1978) Veränderungen im stomatären Kaliumgehalt bei Änderungen von Luftfeuchte und Umgebungstemperatur. *Berichte der Deutschen Botanischen Gesellschaft*, **91**, 645–656.
- Molz, F.J. & Boyer, J.S. (1978) Growth-induced water potentials in plant cells and tissues. *Plant Physiology*, **62**, 423–429.
- Nagarajah, S. & Schulze, E.-D. (1983) Responses of *Vigna unguiculata* (L.) Walp. to atmospheric and soil drought. *Australian Journal of Plant Physiology*, **10**, 385–394.
- Nobel, P.S. (1983) *Biophysical Plant Physiology and Ecology*. W.H. Freeman and Company, San Francisco.
- Raschke, R. (1979) Movements of stomata. In *Encyclopedia of Plant Physiology Vol. 7, Physiology of Movements* (eds W. Haupt & M.E. Feinlieb), pp. 383–441. Springer-Verlag, Berlin.
- Running, S.W. (1980) Environmental control of leaf conductance in conifers. *Canadian Journal of Forest Research*, **6**, 385–394.
- Schönherr, J. (1982) Resistance of plant surfaces to water loss: transport properties of cutin, suberin and associated lipids. In *Encyclopedia of Plant Physiology Vol. 12B, Physiological Plant*

- Ecology II. Water relations and carbon assimilation* (eds O.L. Lange *et al.*), pp. 153–179. Springer Verlag, Berlin.
- Schulze, E.-D. (1986) Carbon dioxide and water vapor exchange in response to drought in the atmosphere and in the soil. *Annual Review of Plant Physiology*, **37**, 247–274.
- Schulze, E.-D. & Hall, A.E. (1982) Stomatal responses, water loss and CO<sub>2</sub> assimilation rates of plants in contrasting environments. In *Encyclopedia of Plant Physiology* Vol. 12B, *Physiological Plant Ecology II. Water relations and carbon assimilation* (eds O.L. Lange *et al.*), pp. 181–230. Springer Verlag, Berlin.
- Schulze, E.-D., Hall, A.E., Lange, O.L., Evenari, M., Kappen, L. & Buschbom, U. (1980) Long-term effects of drought on wild and cultivated plants in the Negev desert. I: Maximal rates of net photosynthesis. *Oecologia*, **45**, 11–18.
- Schurr, U. (1987) *Beziehungen zwischen Abscisinsäure aus der Wurzel und der stomatären Leitfähigkeit im Tagesgang und bei Bodenaustrocknung*. Diplomarbeit, Universität Bayreuth.
- Shackel, K.A., Matthews, M.A. & Morrison, J.C. (1987) Dynamic relation between expansion and cellular turgor in growing grape (*Vitis vinifera* L.) leaves. *Plant Physiology*, **84**, 1166–1171.
- Slatyer, R.O. (1967) *Plant-water relationships*. Academic Press, London.
- Stedle, E. (1985) Water transport as a limiting factor in extension growth. In *Control of leaf growth* (eds Baker, N. R., Davies, W.J. & Ong, C.K.), SEB Seminar Series Vol. 27, pp. 35–55. Cambridge University Press, Cambridge.
- Stedle, E. & Zimmermann, U. (1984) Water relations of plant cells: further development of the pressure probe and of techniques for measuring pressure-dependent transport. In *Membrane transport in plants* (eds Cram, W.J. *et al.*), pp. 73–82. Academia, Praha.
- Stedle, E., Oren, R. & Schulze, E.-D. (1987) Water transport in maize roots. *Plant Physiology*, **84**, 1220–1232.
- Stedle, E., Zimmermann, U. & Zillikens, J. (1982) Effect of cell turgor on hydraulic conductivity and elastic modulus of *Eloдея* leaf cells. *Planta*, **154**, 371–380.
- Turner, N.C., Schulze, E.-D. & Gollan, T. (1985) The response of stomata and leaf gas exchange to vapor pressure deficits and soil water content. II: In the mesophytic herbaceous species *Helianthus annuus*. *Oecologia*, **65**, 348–355.
- Walter, H. & Kreeb, K. (1970) Die Hydratation und Hydratur des Protoplasmas der Pflanzen und ihre öko-physiologische Bedeutung. *Protoplasmatologia* Vol. II.C.6. Springer Verlag, Wien, New York.
- Ziegler, H. (1982) Flüssigkeitsströme in Pflanzen. In *Biophysik* (eds W. Hoppe *et al.*), pp. 652–663. Springer Verlag, Berlin Heidelberg New York.
- Zimmermann, U. (1978) Physics of turgor-and osmoregulation. *Annual Review of Plant Physiology*, **29**, 121–148.
- Zimmermann, U. & Stedle, E. (1978) Physical aspects of water relations of plant cells. *Advances in Botanical Research*, **6**, 45–117.

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.

This document is a scanned copy of a printed document. No warranty is given about the accuracy of the copy. Users should refer to the original published version of the material.