Supplementary Materials

Visualization of bacterial protein complexes labeled with fluorescent proteins and nanobody binders for STED microscopy

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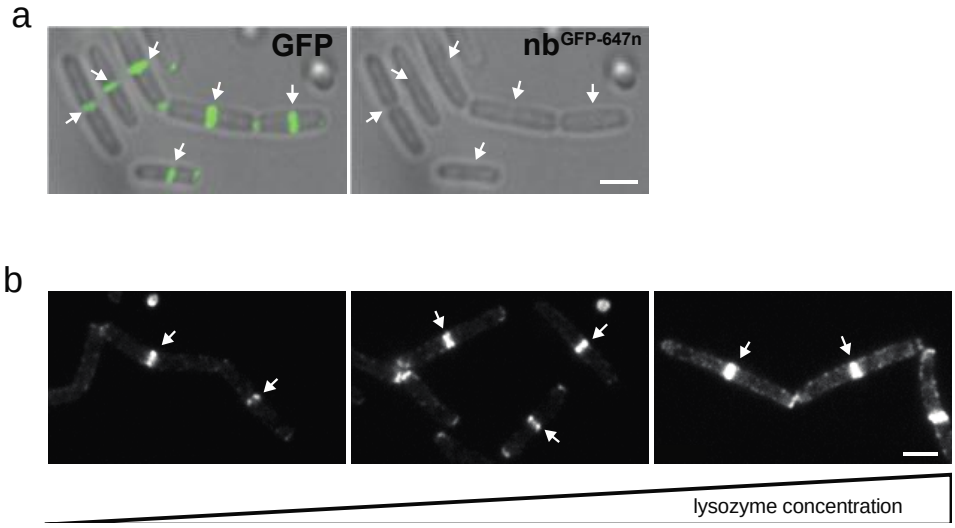
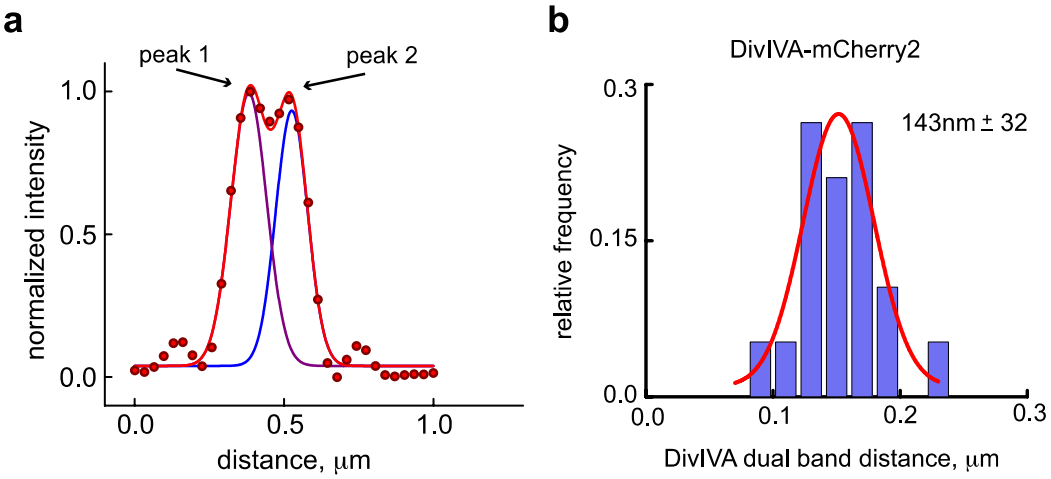
SM Figure 1.



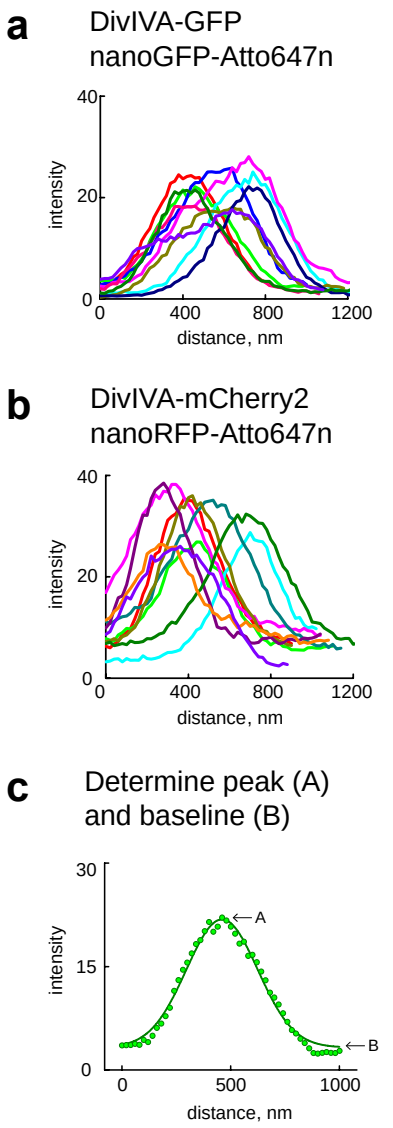
SM Figure 1. Effect of lysozyme treatment on nanobody binder entry into *B. subtilis*. (a) Overlay of bright field (gray) and fluorescence signal from GFP (left) in green, and NB^{GFP}-Atto647n (right) in *B. subtilis* cells expressing DivIVA-GFP (1803) treated with 0 mg/ml of lysozyme. Arrows indicate division septa where DivIVA-GFP is localized Scale bar 2 μm. **(b)** Fluorescence signal from NB^{RFP}-Atto647n in *B. subtilis* cells expressing DivIVA-PAmCherry *(*JB37) treated with 0.04, 0.2, and 0.4 mg/ml of lysozyme, left to right. Arrows indicate NB^{RFP}-atto647n signal at division septa. Scale bar 3 μm. Images were taken with a diffraction-limited microscope. In (a), GFP imaging was performed with an exposure of 0.2s, ND filter = 32%, and NB^{GFP}-Atto647n imaging was performed with an exposure time of 0.5s, ND filter = 32%. In (b), exposure was 0.2s, ND filter = 50% for NB^{GFP}-atto647n imaging.

SM Figure 2.



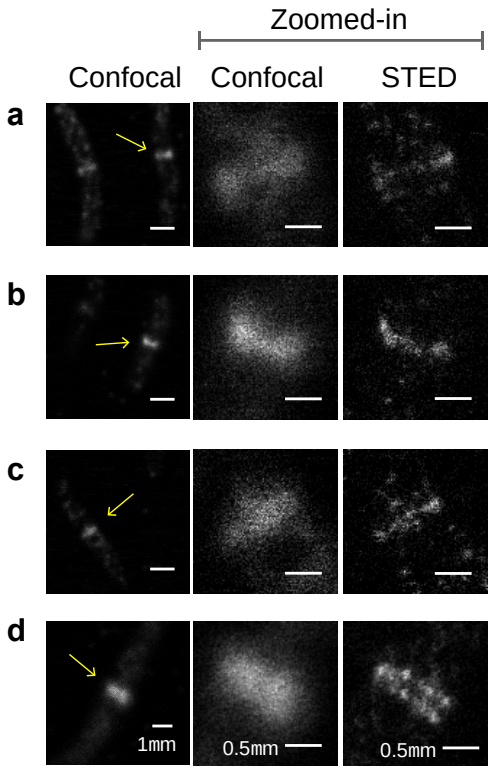
SM Figure 2. Histogram of distance between the DivIVA dual band in SIM images. Intensity profiles of the mCherry2 fluorescent signal along the longitudinal axis of cells from *B. subtilis* cells expressing DivIVA-mCherry2 were obtained using Fiji. **(a)** To extract the center position of each band along the longitudinal axis, the sum of two Gaussian functions were fitted (solid line) to this intensity profile (circles), using the software OriginPro 9.1G. Then, the values for the two center parameters (peak 1 and peak 2) of the Gaussians were subtracted to calculate their distance. **(b)** Mean and STD of the distance distribution are indicated in the histogram.

SM Figure 3



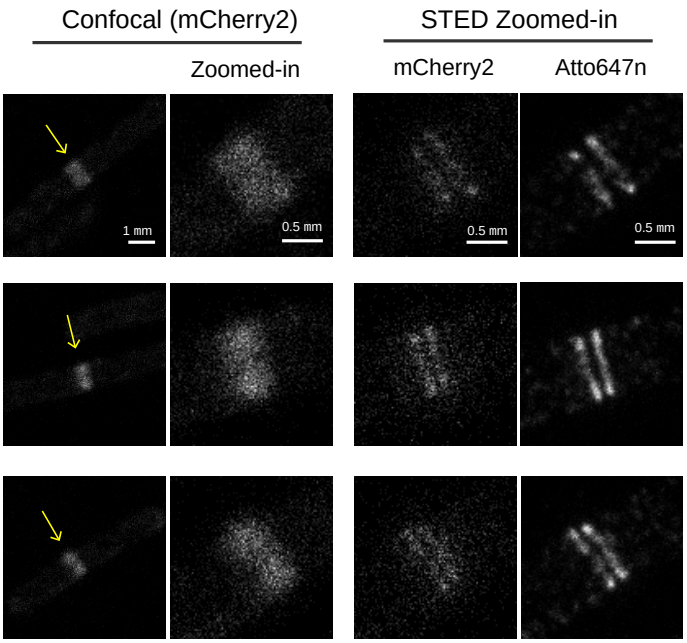
SM Figure 3. Determination of signal-to-background ratio. (a, b) Intensity profiles of the fluorescent signal from the fluorophore conjugated nanobody-along the longitudinal axis of cells from *B. subtilis* cells expressing either DivIVA-GFP or DivIVA-mCherry2. Each line represents the signal distribution from one cell, which was obtained by drawing a line along the longitudinal axis of cells using ImageJ. **(c)** Each intensity profile (circles) was fitted with a single peak gaussian function (continuous line), peak height at center point (A) and baseline (B) were obtained using the software OriginPro 9.1G.

SM Figure 4



SM Figure 4. Visualization of FtsZ in *Escherichia coli with STED microscopy.* Strain EC484 containing GFP tagged FtsZ was visualized using NB^{GFP}-Atto647n via confocal and STED microscopy. Four examples are shown in **(a-d).** Confocal images in the Left column show FtsZ localization in single cells. Yellow arrows indicate FtsZ localization at division septa for **(a-d)**, and their corresponding zoomed-in confocal and STED images are shown in the Middle and Right columns, respectively.

SM Figure 5



SM Figure 5. Comparison of mCherry2 and Atto647n for STED imaging. *B. subtilis* expressing DivIVA-mCherry2 treated with the immunolabeling protocol using the binder NB^{RFP}-Atto647n (as described in Material and Methods) were imaged with STED microscopy. Yellow arrows indicate DivIVA localization at division septa, and their corresponding zoomed-in confocal and STED images are shown in the Middle and Right columns.

SM Table I. Cellular background for *B. subtilis* expressing mCherry2 and GFP fusion protein

FP	Fused to	Signal to background ratio (A/B)	Cellular Background (B/A*100)
GFP	DivIVA	2.6	40 %
mCherry2	DivIVA	2.9	35 %

Signal-to-background ratio and Cellular background were calculated as described in the main text and SM Figure 3. n = 31.

SM Table II: STED sample preparation reagents and conditions. Lists of nanobodies binders used in this study, the targets the binders imaged, the dilutions employed, Figures in the Main Text where imaging using each binder appears, and their Cat. No.

Binder	Target imaged	Dilution	Corresponding Figure	Cat. No. *
NB^{GFP}-atto594	DivIVA-GFP	1:250	3, 4	gba594
NB^{GFP}-atto647n	DivIVA-GFP	1:250	2, 3, 4	gba647n
NB^{GFP}-star635p	DivIVA-GFP	1:250	3, 4	gbas635p
NB^{RFP}-atto594	DivIVA-mCherry2	1:250	3, 4	rba594
NB^{RFP}-atto647n	DivIVA-mCherry2	1:250	2, 3, 4	rba647n
NB^{RFP}-star600	DivIVA-PAmCherry	1:250	4	n/a$

*Cat. No. for Chromotek GmbH products.
$ Gifted

SM Table III: Properties of STED dyes used in this study. Lists of dye molecules and their net charge, hydrophobicity, molecular weight, $\lambda_{excitation}$ and $\lambda_{emission}$.

Dye	Net charge (after coupling)	Hydrophobicity	Molecular weight (g/mol)	λ_{abs}, nm	λ_{fl}, nm	*REF
Atto595	-1	Very hydrophilic	1,389	603	626	Atto-tec
Atto647n	+1	Moderately hydrophilic	843	646	664	Atto-tec
Star600	0 (zwitterionic)	hydrophilic	880.9	604	623	Abberior
Star635p	-3	hydrophilic	1,030.8	638	651	Abberior
mCherry2	n.a.	n.a.	26,700	585	610	[43]

*obtained from dye information pages provided by Atto-tec of Abberior companies

SM Materials and Methods

SM Table III: Lists bacterial strains used in this study.

Name	Genotype	Species	Citation
1803	*divIVA::(PdivIVA*-gfp *divIVA* $^{+}$ cat*)*	*B. subtilis*	[44]
JS99	*divIVA*::divIVA-mCherry2 spec	*B. subtilis*	This study
JB37	*divIVA*::divIVA-PAmCherry spec	*B. subtilis*	[45]
EC484	P_{208}-*ftsZ-gfp leu*::Tn*10*	*E. coli*	[46]

SM References (also in Main Text):

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44. Hamoen, L. W.; Meile, J.-C.; de Jong, W.; Noirot, P.; Errington, J. SepF, a novel FtsZ-interacting protein required for a late step in cell division. *Mol Microbiol* **2006**, *59*, 989–999.
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