Centriole foci persist in starfish oocytes despite Plk1 inactivation or loss of microtubule nucleation activity

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Number of characters: 17943 characters without spaces, Material and Methods, References and Legends.

Key words: starfish, *Patiria miniata*, *Asterias forbesi*, centriole, inactivation, pericentriolar material (PCM), Plk1

ABSTRACT

Centrioles must be eliminated or inactivated from the oocyte to ensure that only the two functional centrioles contributed by the sperm are present in the zygote. Such removal can occur during oogenesis, as in *Drosophila* where departure of the Polo kinase from centrosomes leads to loss of microtubule nucleating activity and centriole removal. In other species, oocyte-derived centrioles are removed around the time of fertilization through incompletely understood mechanisms. Here, we use confocal imaging of live starfish oocytes and zygotes expressing markers of microtubule nucleating activity and centrioles to investigate this question. We first assay the role of Polo-like-kinase 1 (Plk1) in centriole elimination. We find that although Plk1 localizes around oocyte-derived centrioles, kinase impairment with BI-2536 does not protect centrioles from removal in the bat star *P. miniata*. Moreover, we uncover that all four oocyte-derived centrioles lose microtubule nucleating activity when retained experimentally in the zygote of the radiate star *A. forbesi*. Interestingly, two such centrioles nevertheless retain the centriolar markers mEGFP::PACT and pmPoc1::mEGFP. Together, these findings indicate that centrioles can persist when Plk1 activity is impaired, as well as when microtubule nucleating activity is lacking, uncovering further diversity in the mechanisms governing centriole removal.

INTRODUCTION

Centrioles are microtubule-based cylindrical organelles fundamental notably for recruiting the pericentriolar material (PCM), and thus for forming the centrosome, an important microtubule organizing center (MTOC) of animal cells (reviewed in Bornens, 2012; Gönczy and Hatzopoulos, 2019). Control of centriole inheritance at fertilization is critical to ensure bipolar spindle formation and faithful chromosome segregation during the first cell division of metazoan organisms. In most species, maternal centrioles are eliminated or inactivated from the oocyte, whereas two paternal centrioles are contributed to the zygote by the sperm (reviewed in Delattre and Gönczy, 2004; Manandhar *et al.*, 2005). The mechanisms by which maternal centrioles are removed are incompletely understood.

Most proliferating animal cells are born with two centrioles, each of which then seeds assembly of one procentriole that remains engaged in its vicinity until mitosis. At that time, the two centrosomes, each containing a centriole/procentriole pair, act as MTOCs and direct bipolar spindle formation. The centriole and procentriole within each centrosome then disengage from one another during mitosis, so that each resulting cell will be endowed with two centrioles. These two centrioles are structurally distinct, with the older centriole, also called the mother centriole, harboring appendages, which are absent from the younger, or daughter, centriole (reviewed in Firat-Karalar and Stearns, 2014; Loncarek and Bettencourt-Dias, 2018; Gönczy and Hatzopoulos, 2019) .

There are exceptions to the above canonical cycle of centriole inheritance. One such interesting case is encountered at fertilization (reviewed in Delattre and Gönczy, 2004; Manandhar *et al.*, 2005). If each gamete contributed a pair of centrioles, then the zygote would be endowed with four centrioles that would each duplicate during the first cell cycle, thus leading to the presence of four centrosomes and potentially tetrapolar spindle formation. In most metazoan species, including *Drosophila* and human beings, this problem is solved through the removal of centrioles during oogenesis (Mahowald and Strassheim, 1970; Szollosi *et al.*, 1972; Gard, 1994; Sathananthan *et al.*, 2001; Mikeladze-Dvali *et al.*, 2012). In other organisms, such as starfish, centrioles remain in the oocyte and

assemble the two meiotic spindles before being removed (Nakashima and Kato, 2001; Shirato *et al.*, 2006; Crowder *et al.*, 2015). In either scenario, the zygote inherits solely the two functional centrioles contributed by the sperm. Whilst being a fundamental feature of metazoan development, how centrioles are removed from the female gamete remains generally poorly understood.

One exception to such lack of understanding is in *Drosophila*. Here, regulation of the PCM by the kinase Polo is key for eliminating centrioles during oogenesis (Pimenta-Marques *et al.*, 2016). Polo localizes to centrosomes during early oogenesis but is no longer detectable at that location as oocytes mature; this is followed by PCM disassembly, loss of microtubule nucleating activity and then centriole elimination. Moreover, RNAi-mediated depletion of Polo results in precocious PCM disassembly and centriole elimination. Conversely, targeting excess Polo to centrioles enables PCM maintenance and prevents centriole elimination. Such persisting centrioles act as MTOCs and result in abnormal meiosis and abortive zygotic development (Pimenta-Marques *et al.*, 2016). The extent to which the contribution of Polo uncovered in flies will prove general is unclear, including in systems where maternal centrioles serve to assemble the two meiotic spindles and are eliminated or inactivated solely thereafter.

Starfish constitute an attractive model to analyze centriole removal from the oocyte, since in this system the process takes place in a stereotyped manner after oocyte maturation. In the bat star *Patiria miniata*, two of the four centrioles present in the mature oocyte are shed in the first polar body, whereas a further one is expelled with the second polar body, leaving a single centriole in the zygote (Fig. 1A, B). This remaining centriole is eliminated shortly after meiosis II completion. Monitoring of fluorescent fusion proteins specific for either mother or daughter centriole established that this last remaining unit is a daughter centriole (Borrego-Pinto *et al.*, 2016a). Moreover, mother and daughter centrioles show distinct behavior when experimentally retained in the *P. miniata* zygote through inhibition of cytokinesis with an actin-depolymerizing drug. The two oocyte-derived mother centrioles nucleate microtubules and persist, which, together with the two sperm-derived centrioles, leads to the formation of a tetrapolar spindle

during the first mitosis. By contrast, daughter centrioles do not act as MTOCs and are eliminated. Intriguingly, an analysis in the radiate star *Asterias forbesi* suggested an apparent difference with *P. miniata*, since all four centrioles experimentally retained in the *A. forbesi* zygote fail to sustain microtubules nucleation as observed by polarization microscopy (Sluder *et al.*, 1989, 1993). In the absence of molecular markers, however, it is not clear how this apparent difference relates to the fate of centrioles.

RESULTS AND DISCUSSION

Plk1 localizes at centrioles in starfish oocytes

We sought to investigate whether Polo-like-kinase 1 (Plk1) can protect centrioles from elimination in starfish, in a manner analogous to the role exerted by its homologue Polo in *Drosophila*. First, we determined whether Plk1 localizes to centrioles in *P. miniata* oocytes. To this end, we generated mRNAs coding for mEGFP tagged *P. miniata* and *H. sapiens* Plk1 proteins. Here and thereafter, oocytes were injected with *in vitro* transcribed mRNAs, in this case encoding mEGFP::pmPlk1, as well as the microtubule associated protein hsEB3::mCherry3 to monitor growing microtubules and thus MTOC activity. After overnight incubation to allow translation of the injected mRNAs, oocytes were matured with 1-methyladenine (1-MA) (Kanatani *et al.*, 1969), leading to resumption of cell cycle progression, nuclear envelope breakdown (NEBD) and then execution of the two meiotic divisions, which were filmed using time-lapse confocal microscopy (Figures 1A and 1B). The number of oocytes analyzed for each experiment and a summary of the results are provided in Table S1.

As shown in Figure 1C, we found that mEGFP::pmPlk1 localizes to the two poles of the meiosis I spindle in *P. miniata* oocytes (-25:15, insets 1 and 2). At the onset of meiosis II, the focused signal at the inner pole of the meiosis I spindle splits into two foci (00:00, insets 3 and 4) that then each localizes to the two poles of the meiosis II spindle (07:20, insets 5 and 6). We noted a difference in fluorescence intensity between these two mEGFP::pmPlk1 foci, with a brighter signal for the outer focus, closer to the plasma membrane (07:20, compare inset 5 and inset 6). In addition, we found that the inner focus of mEGFP::pmPlk1 is no longer detected shortly after extrusion of the second polar body (27:32, inset 7). Similar dynamics were observed for the human hsPlk1:mEGFP fusion protein (Fig. S1A). Such distributions mirror those reported for pan-centriolar components in *P. miniata* oocytes (Borrego-Pinto *et al.*, 2016a), indicating that Plk1 localizes initially to all centrioles or their immediate vicinity, albeit to different degrees. Moreover, we observed that both mEGFP::pmPlk1 and hsPlk1::mEGFP also label what appears to be kinetochores (Figure 1C, -25:15 and 07:20; Figure S1A, -19:34 and 16:00; arrows), in line with the distributions reported in other systems for this kinase family (reviewed in Archambault and Glover, 2009). We found analogous localizations at

spindle poles and kinetochores when examining hsPlk1::mEGFP in *A. forbesi* oocytes (Figure S1B).

Overall, we conclude that Plk1 is present initially at all four oocyte-derived centrioles in starfish oocytes, with more protein detected at mother centrioles, and is then lost from daughter centrioles before MTOC activity ceases.

Plk1 activity does not protect centrioles from elimination in *P. miniata* oocytes

We set out to test whether Plk1 protects centrioles from elimination in *P. miniata* oocytes. Plk1 is required for bipolar spindle formation in a wide range of systems, including in the starfish *Asterina pectinifera*, where function-blocking antibodies result in monopolar spindle assembly during meiosis I (Okano-Uchida *et al.*, 2003). However, the fate of centrioles was not addressed in that study. Here, analyzing oocytes expressing hsEB3::mCherry3 and the pan-centriolar marker pmCentrin2::mEGFP, we established that treatment with 10 µM of the Plk1 inhibitor BI-2536 (Lénárt *et al.*, 2007) at the time of 1-MA addition prevents bipolar spindle assembly during meiosis I (compare Figure 2A with Figure 2B). Instead of a bipolar spindle (Figure 2A), a diffuse crown of microtubules forms transiently around centrioles (Figure 2B). Thereafter, centrioles in drug-treated specimens typically move deeper into the oocyte proper and lose microtubule nucleation activity (Figure 2B 01:36:04, compare with Figure 2A, 01:40:55), although sometimes hsEB3::mCh3 can be present around one centriole at later times (Figure 2B, 02:02:52, inset 9).

If Plk1 activity were to protect centrioles from elimination, then BI-2536 treatment should provoke disappearance of the two mother centrioles and potentially precocious elimination of daughter centrioles. To best determine whether this is the case, we analyzed mildly centrifuged oocytes expressing hsEB3::mCherry3 and pmCentrin2::mEGFP. Centrifugation leads to nuclear detachment from the animal pole, where centrioles remain cortically anchored (Figure 2C; Figure S1C-F). Even though centrosomes nucleate microtubules in such centrifuged oocytes (Figure 2C, 01:29:23), the meiotic spindles do not capture the distant chromosomes and, consequently, polar bodies are not extruded. As a result, all four oocyte-derived centrioles remain typically close to the plasma membrane, thus facilitating their monitoring throughout meiosis (Matsuura and Chiba, 2004; Borrego-Pinto *et al.*, 2016a). In centrifuged control oocytes, two of the

four pmCentrin2::mEGFP foci persist and nucleate microtubules, whereas the other two lose MTOC activity and are no longer detectable by the end of meiosis II (Figure 2C, 02:11:13 and 03:00:48; Figure 2E; Figure S1G). Importantly, in centrifuged BI-2536-treated oocytes, all four centrioles have essential no MTOC activity as early as MI, but at least one and often two of the four retained pmCentrin2::mEGFP foci persist until the end of MII (Figure 2D, 01:31:03 and 03:02:28; Figure 2E; Figure S1H). Moreover, we observed that loss of the pmCentrin2::mEGFP foci corresponding to daughter centrioles occurred on average ~18 minutes earlier in BI-2536 treated oocytes than in the control condition (Figure 2E). This may not reflect a *bona fide* temporal shift, but instead reflect the loss of focused hsEB3::mCh3 signal and the typically dimmer pmCentrin2::mEGFP signal upon BI-2536 treatment, rendering daughter centriole tracking more challenging (Figure S1H).

We cannot exclude that a potential Plk1-dependent mechanism modulating centriole elimination requires only minute kinase activity. Moreover, whereas the *Drosophila* genome encodes a single Polo kinase, there is a second Polo-like kinase in *P. miniata* that is ~ 40% identical to Plk1 (sequences IDs PMI_003306 and PMI_004640 from EchinoBase aligned with ClustalW), which could potentially act redundantly with Plk1. However, 10 µM BI-2536 is expected to readily inhibit both Plk1 and Plk2, considering that the corresponding IC50s for their human counterparts are of 0.83 nM and 3.5 nM, respectively (Steegmaier *et al.*, 2007).

Overall, within the time frame of this experiment, these findings lead us to conclude that inhibiting Plk1 activity in *P. miniata* oocytes is sufficient to provoke loss of MTOC activity but not to trigger the elimination of mother centrioles or a drastic precocious disappearance of daughter centrioles.

Mother centrioles are extruded into polar bodies of *A. forbesi* oocytes

Early observations raised the possibility that centriole fate may differ in the radiate star *A. forbesi* compared to the bat star *P. miniata* (Sluder *et al.*, 1989, 1993; see Introduction). We set out to investigate this potential difference using molecular markers. As a first step, we tested whether the two mother centrioles are systematically directed to the first and second polar bodies in *A. forbesi*, as they are in *P. miniata* (Borrego-Pinto *et al.*, 2016a). We injected oocytes with mRNA encoding a fusion protein between mEGFP and the mother centriole-specific

component Odf2. We found that, like in *P. miniata,* the strong focused signals of pmOdf2::mEGFP were systematically present on both poles of the meiosis I spindle and on the pole closest to the plasma membrane of the meiosis II spindle (Figure 3). Therefore, mother centrioles are systematically extruded in polar bodies in *A. forbesi* as they are in *P. miniata*.

Retained oocyte-derived centrioles lose MTOC activity upon meiosis II exit in *A. forbesi*

We next set out to investigate whether oocyte-derived centrioles experimentally retained in *A. forbesi* zygotes exhibit MTOC activity. To this end, we injected oocytes with mRNA encoding hsEB3::mCherry3 to label growing microtubules. Following 1-MA-induced maturation and NEBD, we added sperm to activate development and then Latrunculin B to block extrusion of the two polar bodies. MTOC activity of the centrioles thus retained in the zygote was assessed from meiosis I until the first mitotic division by time-lapse confocal microscopy. We found that the meiosis I spindle forms normally, with a pair of centrioles at each pole, and that centrioles disengage normally thereafter (not shown); this is followed by the formation of two small bipolar spindles with one centriole at each pole during meiosis II (Figure 4A, 10:00). Importantly, we found that all oocyte-derived centrioles lose their MTOC activity upon exit from meiosis II (Figure 4A, 30:47 and 4B), leading to the formation of a bipolar spindle driven exclusively by sperm-derived centrioles during the first mitotic division in the zygote (Figure 4A, 77:35). This is in stark contrast to *P. miniata*, where a tetrapolar spindle assembles following retention of oocyte-derived centrioles in the fertilized zygote (Borrego-Pinto *et al.*, 2016a). We conclude that mother centrioles retained in the *A. forbesi* zygote do not function as MTOCs, in contrast to the situation in *P. miniata*.

Centriolar markers persist at retained oocyte-derived centrioles in *A. forbesi* zygotes

At least three scenarios could explain the lack of MTOC activity of centrioles retained in *A. forbesi* zygotes. First, all four centrioles could be eliminated. Second, all four centrioles could be retained, but without exhibiting MTOC activity. Third, daughter centrioles could be removed but mother centrioles could persist without nucleating microtubules. To distinguish between these possibilities, we set out to

monitor centriolar markers from meiosis II until the first mitosis in *A. forbesi* zygotes treated with Latrunculin B (Figures 5 and S2; Movie S1). To follow centrioles, we injected mRNAs coding for a fusion protein between mEGFP and PACT, a small protein fragment derived from human Pericentrin and AKAP450 that marks centrioles across a range of organisms (Gillingham and Munro, 2000). As shown in Figure 5, we found that mEGFP::PACT labels initially all four centrioles localizing at the poles of the two bipolar spindles formed during meiosis II upon Latrunculin B treatment (Figure 5A, 3:30; Figure 5B, centrioles 1-4 and Figure 5C). After meiosis II exit, the MTOC activity of all four centrioles is lost, as monitored with hsEB3::mCherry3, whereas all four mEGFP::PACT foci persist initially (Figure 5A, 29:09; Figure 5B, centrioles 1-4; Figure 5C). Thereafter, two of the four mEGFP::PACT foci, presumably corresponding to the daughter centrioles, are no longer detectable (Figure 5A, 59:14; Figure 5B, centrioles 1 and 2; Figure 5C). Importantly, we found in addition that the two other foci, which we surmise correspond to the two mother centrioles, remain present at least until the first mitosis and were still observed as far as the second mitosis in some oocytes that were analyzed until then (data not shown). Upon onset of the first mitotic division, these two oocyte-derived units approach the sperm-derived bipolar spindle, probably through the action of minus-end directed motors, without regaining MTOC activity (Figure 5A, 59:14; Figure 5B, centrioles 3 and 4; Figure 5C).

We sought to extend the above observation to another centriolar marker. Therefore, we investigated oocytes injected with mRNA coding for pmPoc1::mEGFP, a centriolar component and microtubule binding protein conserved from green algae to human beings, including starfish (Keller *et al.*, 2005; Borrego-Pinto *et al.*, 2016a). As reported in Figure S2, we found that pmPoc1::mEGFP labels all four centrioles and microtubules also in *A. forbesi* zygotes treated with Latrunculin B. Although the weaker pmPoc1::mEGFP centriolar signal compared to mEGFP::PACT sometimes prevented us from determining with certainty the number of oocyte-derived centrioles that persist until mitosis, there was usually at the least one left (Figure S2C, Table S1). Analysis of the minute foci harboring mEGFP::PACT and pmPoc1::mEGFP using serial section electron microscopy would be needed to ascertain whether they retain the full native architecture of centrioles. Regardless, the findings to date establish that foci of

oocyte-derived centriolar components can remain present until mitosis in *A. forbesi*.

Concluding remarks

We set out to investigate whether the persistence of MTOC activity from the PCM surrounding oocyte-derived centrioles retained in *P. miniata* zygote (Borrego-Pinto *et al.*, 2016a) may reflect a mechanism analogous to that operating during *Drosophila* oogenesis (Pimenta-Marques *et al.*, 2016). However, we found that centrioles can persist in *P. miniata* zygotes with impaired Plk1 activity, as well as in *A. forbesi* without them nucleating microtubules, uncovering diversity in the mechanisms governing centriole removal across metazoan organisms. We note that whereas centrioles are eliminated during oogenesis in *Drosophila*, removal occurs after the meiotic divisions in starfish. Such differential timing may explain why Plks play a role in flies and seemingly not in starfish for ensuring proper centriole number at fertilization.

What mechanisms ensure that oocyte- and sperm-derived centrioles are endowed with different fates in the newly fertilized zygote? In *P. miniata*, MAP kinase activity suppresses the formation of sperm asters during meiosis and thus prevents them from interfering with active oocyte-derived centrioles driving meiotic spindle formation (Stephano and Gould, 2000). A related phenomenon operates in the clam *Spisula soldissima*, where γ-tubulin and MTOC activity of sperm centrioles is lost during meiosis I and regained only during meiosis II (Wu and Palazzo, 1999). Moreover, during physiological polyspermy in the newt *Cynops pyrrhogaster*, multiple sperm cells initially enter the oocyte, but only two centrioles develop a large aster that then drive mitotic spindle formation, with the remaining centrioles degenerating (Iwao *et al.*, 2002)**.** It will be interesting to unravel how specific centrioles can be earmarked for retention or removal in the same cell in these diverse settings. In starfish, it will also be interesting to explore whether variations in PCM and centriolar components, as well as in activities present in the cytoplasm, could explain the differential ability of persisting mother centrioles to nucleate microtubules during mitosis in the two species (Figure 5D), and thus further unravel the diversity of mechanisms governing centriole removal.

MATERIALS AND METHODS

mRNA generation

mRNAs were synthesized and purified as described (Borrego-Pinto *et al.*, 2016b). Briefly, the ORF of the protein of interest in frame with either an N- or C-terminal fluorescent tag was subcloned into pGEMHE for *in vitro* transcription reactions (Borrego-Pinto *et al.*, 2016b). Capped mRNAs were synthesized from linearized templates using the AmpliCap-Max T7 High Yield Message Maker kit (CellScript), and a poly-A tail added using the A-Plus Poly(A) Polymerase Tailing Kit (CellScript). Purified mRNAs (typically 2-6 µg/µl) were diluted in 11 µl RNAse-free water (Borrego-Pinto *et al.*, 2016b).

Starfish and gamete collection

Patiria miniata (previously known as *Asterina miniata)* was purchased from Monterey Abalone Company (Monterey, USA), and *Asterias forbesi* collected from the waters of Cape Cod by the Marine Resource Center (MRC) of the Marine Biological Laboratory (MBL) (Woods Hole, USA). Animals were maintained in sea water tanks at 16-20°C at the MRC or at EMBL's Marine Facility (Heidelberg, Germany). Ovaries were dissected from female animals, washed for 20 minutes in calcium-free sea-water supplemented with phenylalanine (437 mM NaCl, 9 mM KCl, 22.9 mM $MgCl_2$, 25.5 mM $MgSO_4$, 2.1 mM $NaHCO_3$, 50 mM phenylalanine; pH 8), after which oocytes were collected upon treatment of ovary pieces with ~100 µM acetylcholine as described (Terasaki *et al.*, 1994 and http://mterasaki.us/panda/injection/). Healthy-looking oocytes were selected and kept in filtered seawater at ~14°C to be used within two days of extraction. Sperm was obtained by extracting testis fragments from male animals; such fragments were kept dry for several days at 4°C and tested daily before use.

Microinjection, maturation, drug treatments and fertilization

Oocytes were mounted in Kiehart-Ellis chambers and injected using mercury back-filled needles (Terasaki, 1994; Borrego-Pinto *et al.*, 2016b). The amount of injected mRNAs was calibrated using an ocular micrometer and the optimum determined empirically for each mRNA. Injected oocytes were incubated overnight at ~14°C in a

humidified Petri dish to allow protein expression. Meiosis resumption was then induced by 10 µM 1-methyladenine (1-MA, Acros Organics). To prevent polar body extrusion, oocytes were treated with 250 nM Latrunculin B (EMD Biosciences) in seawater 5-10 min after sperm addition, since fertilization depends on actin polymerization. BI-2536 (https://www.axonmedchem.com/product/1129) was diluted 1:1000 in sea water from a 10 mM stock in 100% DMSO, and added to oocytes simultaneously with 1-MA. Control oocytes were exposed to 1:1000 DMSO in seawater. We noted that treatment with 10 µM BI-2536 did not result in a noticeable difference in the timing of NEBD compared to DMSO control in *P. miniata* oocytes observed under a dissecting microscope. For *in vitro* fertilization, sperm was diluted 1:1000 - 1:8000 in seawater depending on the motility on the day of the experiment, and added 30-45 min after 1-MA addition.

Centrifugation and drug treatments

Oocyte centrifugation was performed in a clinical centrifuge at 2400 rpm for 1h at 4°C (Multifuge 3 L-R, Heraeus) after placing the oocyte chamber into a plastic holder in a 50ml Falcon tube filled with sea water (see Figure S1B) (Matsuura and Chiba, 2004; Borrego-Pinto *et al.*, 2016a). After centrifugation, the oocyte chamber was placed into a 35mm µ-dish (Vitaris) for 1-MA addition and BI-2536 treatment.

Microscopy and image processing

After overnight incubation and experiment-specific handling following maturation, injected oocytes were transferred to a confocal microscope for multi-position dual color time-lapse imaging. Depending on the experiment, data were acquired with a 40x 1.25NA water immersion lens on a Nikon A1+ confocal, a 40x HCX PL APO 1.10 NA water immersion lens on a Leica SP5II or a 40x C-Apochromat NA 1.2 water immersion lens on a Zeiss LSM780 confocal microscope. Z-stacks of the relevant portion of the oocyte were captured typically every micron at ~3-5 minutes intervals. Brightness/contrast adjustments were performed using Fiji in a uniform manner for all time points in a given oocyte, and panels assembled using Adobe Illustrator.

ACKNOWLEDGEMENTS

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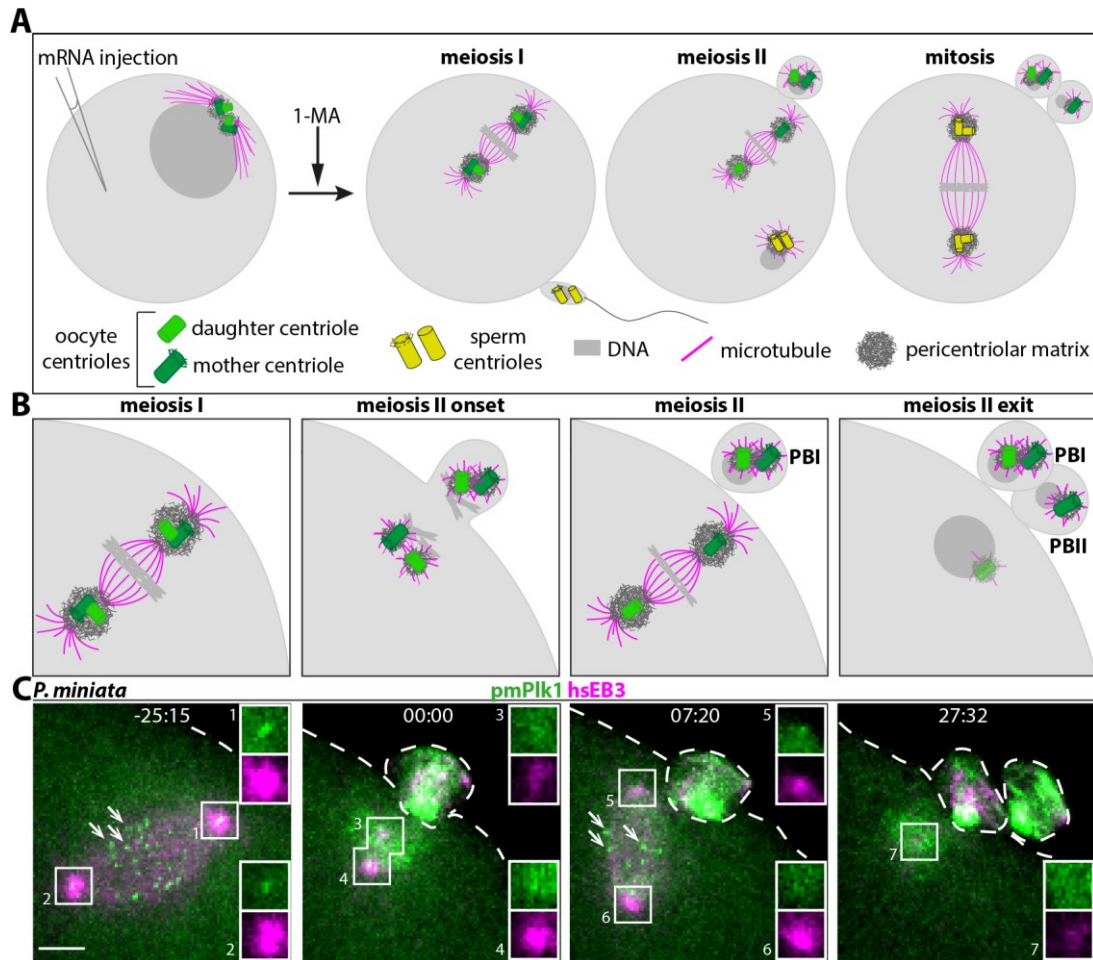
Figure legends

Figure 1. Centriole behavior and Plk1 distribution in *P. miniata* oocytes.

(A and B) Schematic of centriole fate during meiotic divisions of *P. miniata* starfish oocyte, together with overview of experimental procedure utilized in this work, not to scale; B shows higher magnification views of the region with meiotic spindles. Arrested oocytes are injected with mRNA(s) coding for the protein(s) of interest; meiotic resumption is induced by 1-Methyladenine (1-MA), followed by fertilization, depending on the experiment, and then by confocal time-lapse microscopy. During the first meiotic division, pairs of centrioles, each containing a mother centriole (dark green, bearing appendages) and a daughter centriole (light green) are present at the poles of the spindle (B, meiosis I, metaphase represented). First polar body (PBI) extrusion results in the removal of 2n DNA and of a pair of centrioles from the oocyte (B, meiosis II onset). The two remaining centrioles then disengage from one another and drive the formation of the meiosis II spindle (B, meiosis II, metaphase represented). The mother centriole is invariably positioned towards the plasma

membrane and thus extruded in the second polar body (PBII), together with 1n DNA (B, meiosis II exit). The remaining daughter centriole then loses MTOC activity and is eliminated (depicted as fading away in meiosis II exit panel). Fertilization results in the sperm contributing 1n DNA and a pair of centrioles (yellow) to the zygote. Sperm-derived centrioles then duplicate, leading to two centriole pairs that recruit PCM (dark grey) and govern bipolar spindle formation during the first mitosis (A, mitosis). (C) Still images from dual color time-lapse confocal microscopy of *P. miniata* oocyte expressing mRNAs encoding the microtubule marker hsEB3::mCherry3 (in magenta throughout the paper) and mEGFP::pmPlk1 (green), which localizes at centrioles (insets) and kinetochores (arrows point to three of them at metaphase of meiosis I and II). Here and in other figures, images are maximum intensity projections of selected z-planes spanning the region of interest, and insets 1.4x magnified single z-plane of the boxed regions; the oocyte plasma membrane is indicated with a dashed line. Moreover, unless stated otherwise, time is indicated in minutes:seconds starting from centriole disengagement at meiosis II onset and scale bars are 5 µm.
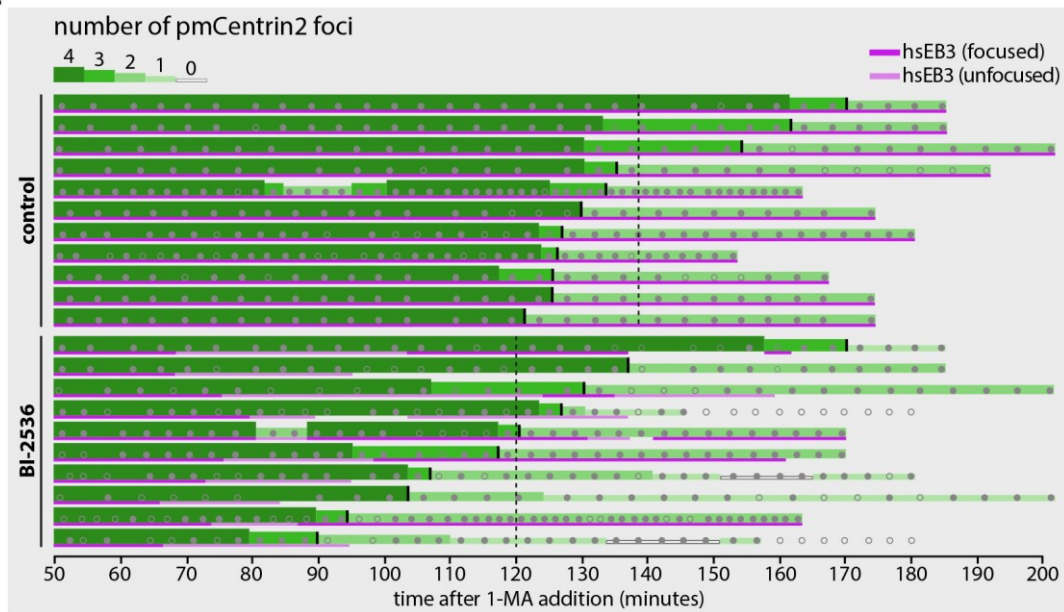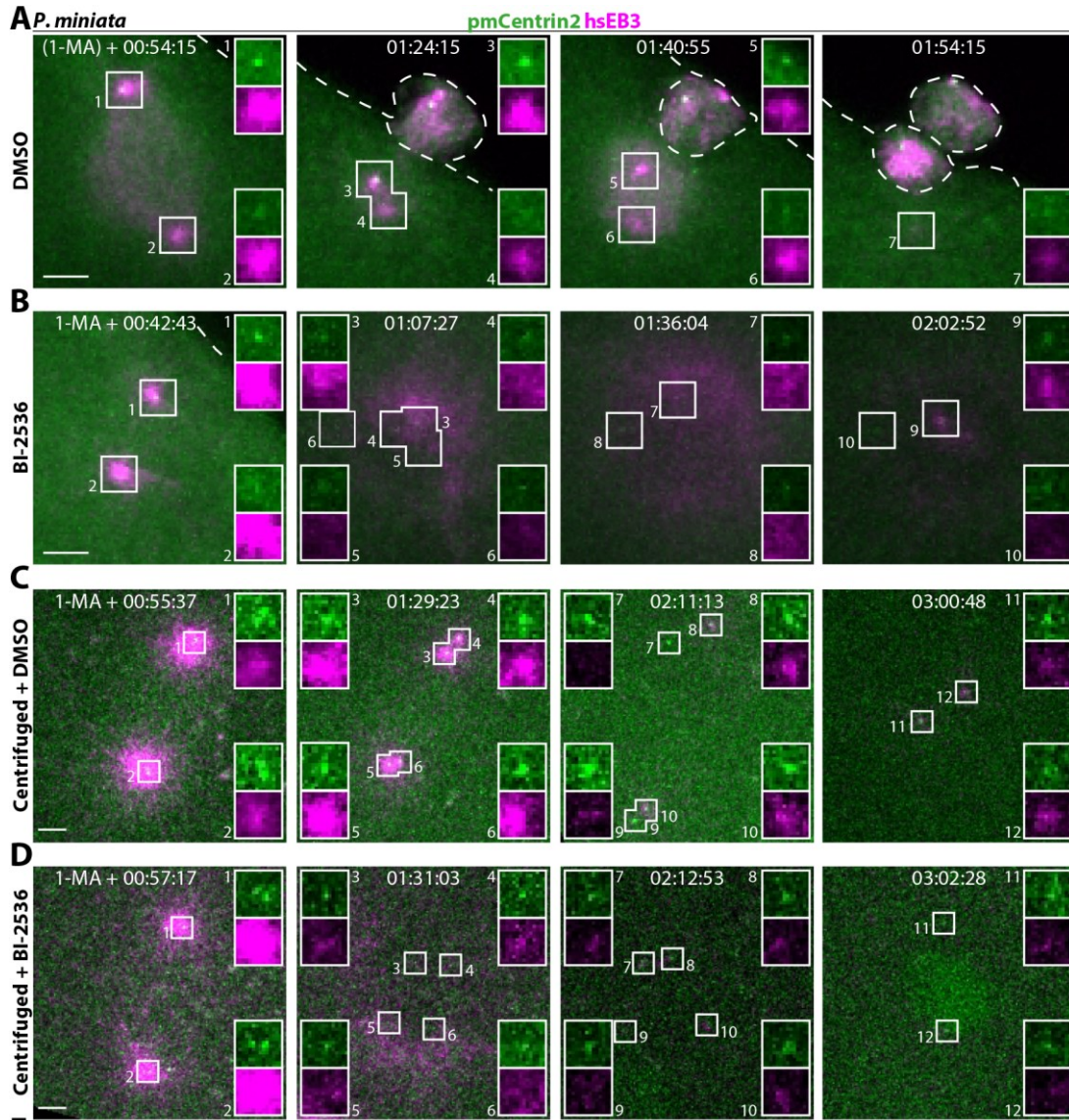
Figure 2. Plk1 inhibition does not provoke centriole elimination in *P. miniata.*

(A-D) Still images from dual color time-lapse confocal microscopy of *P. miniata* oocytes expressing hsEB3::mCherry3 to mark microtubules and mEGFP::pmCentrin2 to mark centrioles, treated with either 0.1% DMSO as a control or 10 µM BI-2536 in 0.1% DMSO, both added simultaneously with 1-MA. A, B: non-centrifuged oocytes; note that grey levels were adjusted differently in A and B, which stem from independent experimental series. C, D: centrifuged oocytes. (E) Oocyte-derived centriole number (green) and MTOC activity (magenta) over time as monitored by mEGFP::pmCentrin2 foci and hsEB3::mCherry3 (magenta), respectively, in DMSO (C) or BI-2536-treated (D) centrifuged oocytes. Each line corresponds to one oocyte. Centriole number is indicated with different shades of green and different line thicknesses. Dark or light magenta lines indicate whether microtubules are focused around centrioles or instead diffuse, respectively. In BI-2536 oocytes, hsEB3::mCh3 is usually diffuse before disappearing completely (n= 5/10) or sometimes refocusing in a very limited area around centrioles (n= 5/10). Small grey discs: actual data points; grey circles: ambiguous data points, either because foci are out-of-focus or because foci disappear and reappear within 3 frames. Small vertical black marks indicate for each oocyte when the number of detected foci drops below 3, and vertical dashed lines the resulting average time of dropping below 3 for each condition (138 minutes for DMSO control, 120 minutes for BI-2536-treated). Time is indicated in hours:minutes:seconds after 1-MA addition, as meiosis II onset is difficult to discern in BI-2536 treated oocytes. In A, the parentheses around 1-MA denote the fact that timing in this particular oocyte is merely estimated from the actual timing of NEBD and the observation that NEBD occurs ~30 min after 1-MA addition.
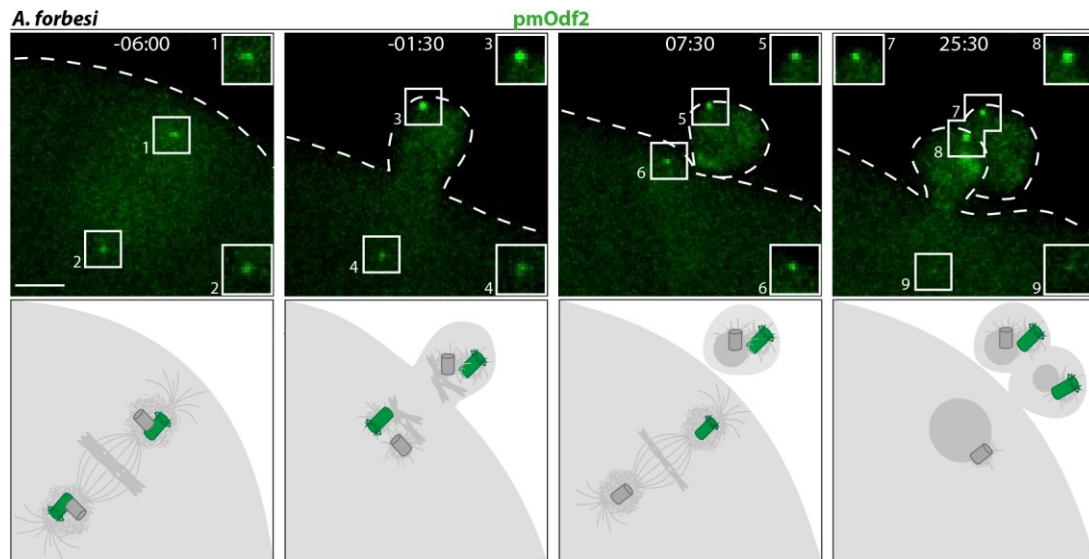
Figure 3. Mother centrioles are extruded into polar bodies in *A. forbesi* oocytes.
Top: still images from time-lapse confocal microscopy of *A. forbesi* oocyte expressing the mother centriole marker pmOdf2::mEGFP. A bright focus is observed in each polar body. In some oocytes, as illustrated here, a weak focus can also be detected at the inner-most centriole at the end of meiosis II. Bottom: corresponding schematic representation.
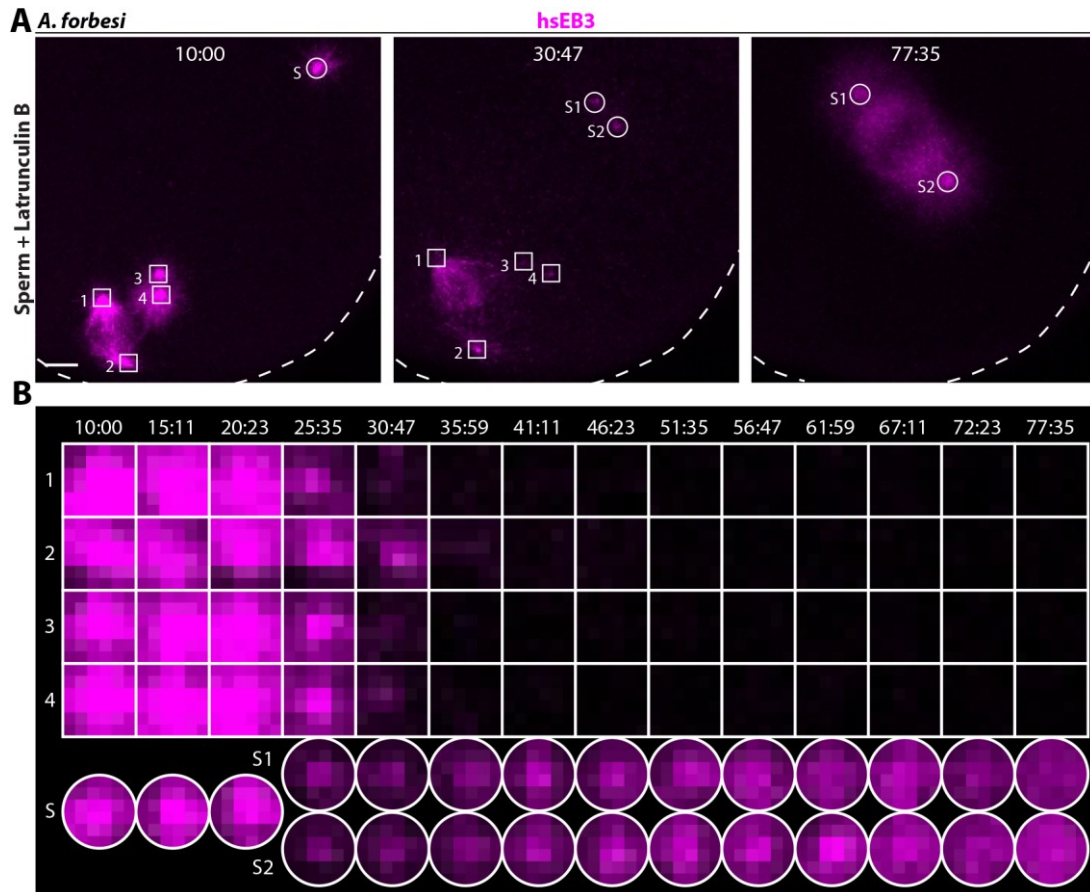
Figure 4. Oocyte-derived centrioles lose MTOC activity when retained in *A. forbesi* zygote.

(A) Still images from time-lapse confocal microscopy of *A. forbesi* zygote treated with Latrunculin B to retain all centrioles, monitoring MTOC activity using hsEB3::Cherry3 from meiosis II onset until mitosis. Here, as well as in Figure 5, oocyte-derived centrioles are bounded by squares and sperm-derived centrioles by circles. Note that hsEB3::mCherry3 is lost from the surroundings of all four oocyte-derived centrioles, whereas the two sperm-derived centrioles (S and then S1, S2) exhibit MTOC activity. Scale bar: 10 µm. (B) Single confocal z-planes corresponding to the zygote shown in A and showing differential MTOC activity of oocyte-derived versus sperm-derived centrioles from meiosis II to mitosis. Each image is 3.73x3.73 µm.
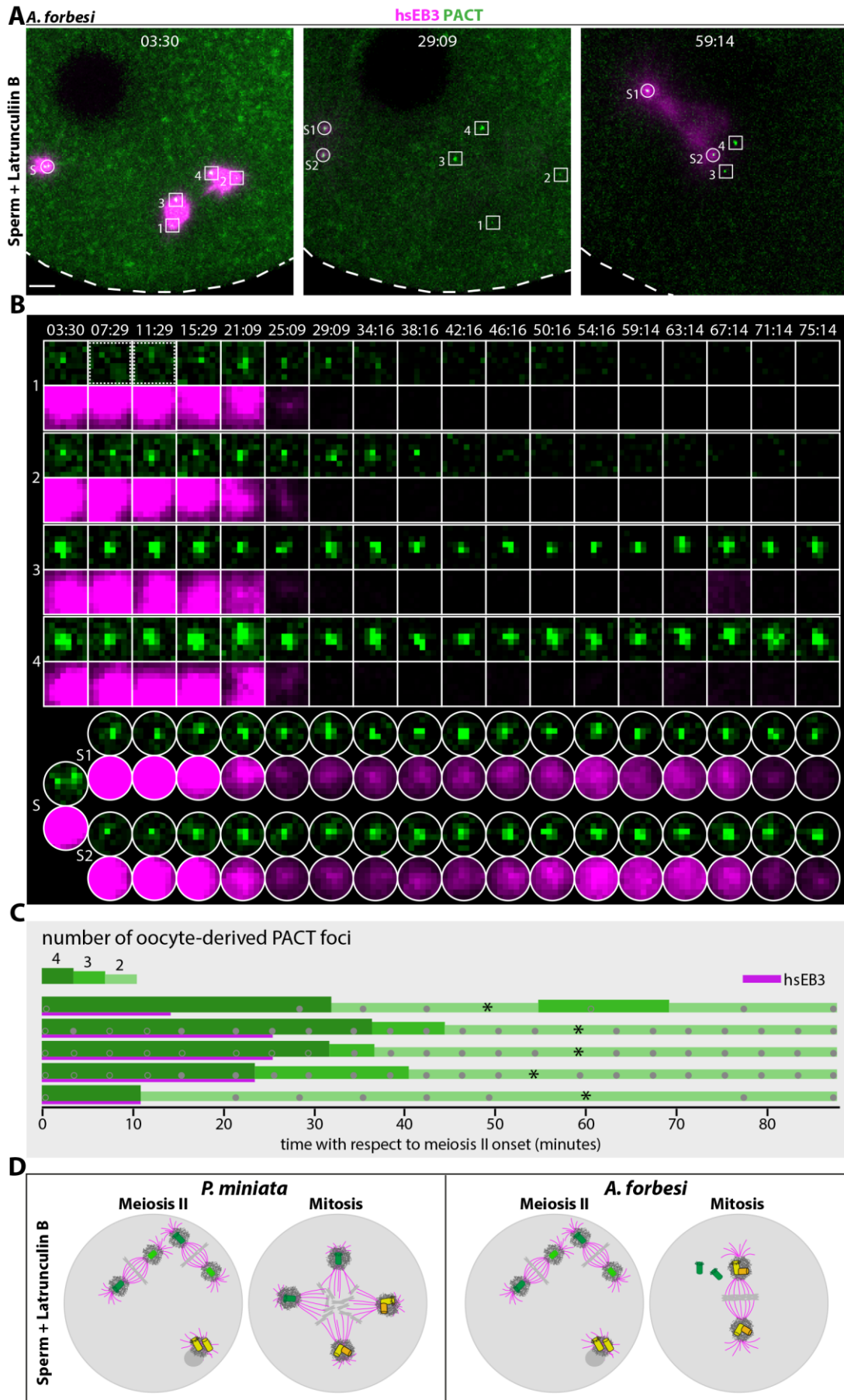
Figure 5. Two retained foci of mEGFP::PACT persist until mitosis in *A. forbesi*.

(A) Still images from dual color time-lapse confocal microscopy of *A. forbesi* zygote treated with Latrunculin B to retain all centrioles, monitoring microtubules labeled with hsEB3::mCherry3 and centrioles marked with mEGFP::PACT from meiosis II onset until mitosis. Scale bar: 10µm; see also Movie S1. (B) Single confocal z-planes showing MTOC activity and presence of mEGFP::PACT foci in oocyte-derived and sperm-derived centrioles from meiosis II until mitosis. Images boxed by a dashed line for centriole 1 indicate time frames when this centriole was slightly out of focus. Note that the little signal for hsEB3::mCherry3 of centriole 3 at time point 67:14 stems from sperm-derived MTOC activity. Each image is 3.98x3.98 µm. (C) Oocyte-derived centriole number over time as monitored by mEGFP::PACT foci (green) and MTOC activity as monitored by hsEB3::mCherry3 (magenta). Each line corresponds to one oocyte, with asterisks indicating mitosis onset. Centriole number is indicated with different shades of green and line thicknesses. Grey filled discs: data points; grey circles: ambiguous data points due to mEGFP::PACT not being yet detected at all centrioles, to foci being out-of-focus or to the presence of multiple, probably spurious, foci. (D) Schematic representation of centriole fate in *P. miniata* based on Borrego-Pinto *et al.*, 2016a and *A. forbesi* zygotes treated with Latrunculin B. The same color code is used as in Figure 1. See text for details.