

Supplementary Materials for

***Australopithecus afarensis* endocasts suggest ape-like brain organization and prolonged brain growth**

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The PDF file includes:

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Other Supplementary Material for this manuscript includes the following:

(available at advances.sciencemag.org/cgi/content/full/6/14/eaaz4729/DC1)

Data file S1

Supplementary Materials

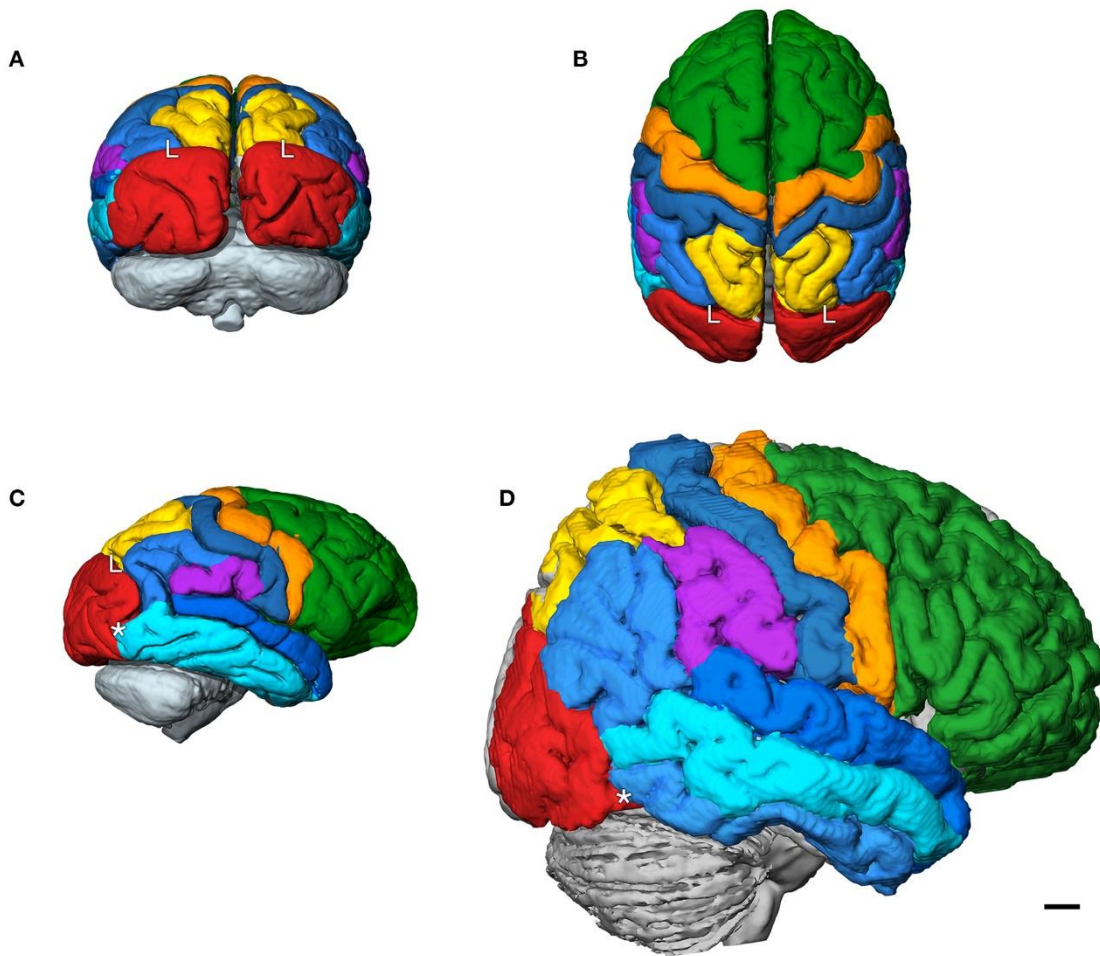


Fig. S1. Different brain organization in apes and humans. Whereas the lunate sulcus (L) is a prominent structure on ape brains, the morphology of the occipital lobe (red) is highly variable in *Homo sapiens*, who rarely display a “true” lunate sulcus. (A-C) Segmentation of a chimpanzee brain based on an MRI scan, in posterior, superior, and lateral views. (D) Modern human brain in lateral view. Some researchers have used the “occipital notch” in *H. sapiens* brains as the structure corresponding to the inferior and lateral end of the ape lunate sulcus (58, 59). The asterisk in (C) points to the inferior and lateral end of the lunate sulcus in *P. troglodytes*; the asterisk in (D) to the corresponding structure in *H. sapiens*. Scale bar is 1 cm.

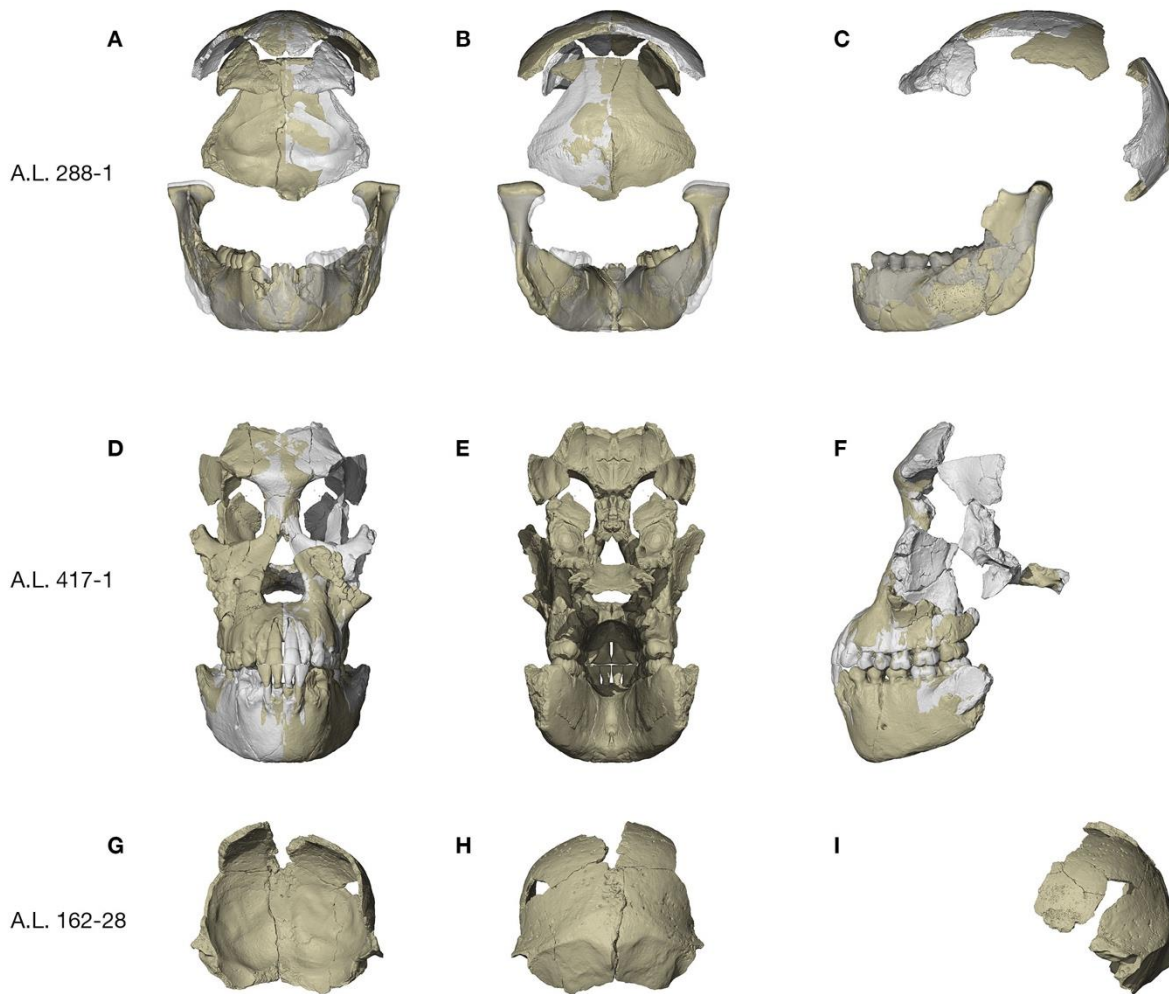


Fig. S2. Size comparison of small *A. afarensis* adults. (A-C) Reconstruction of A.L. 288-1 in anterior, posterior, and lateral view. Mirrored surfaces are shown in gray. (D-F) Reconstruction of A.L. 417-1 in anterior, posterior, and lateral view. (G-I) Original surface of A.L. 162-28 in anterior, posterior, and lateral view.

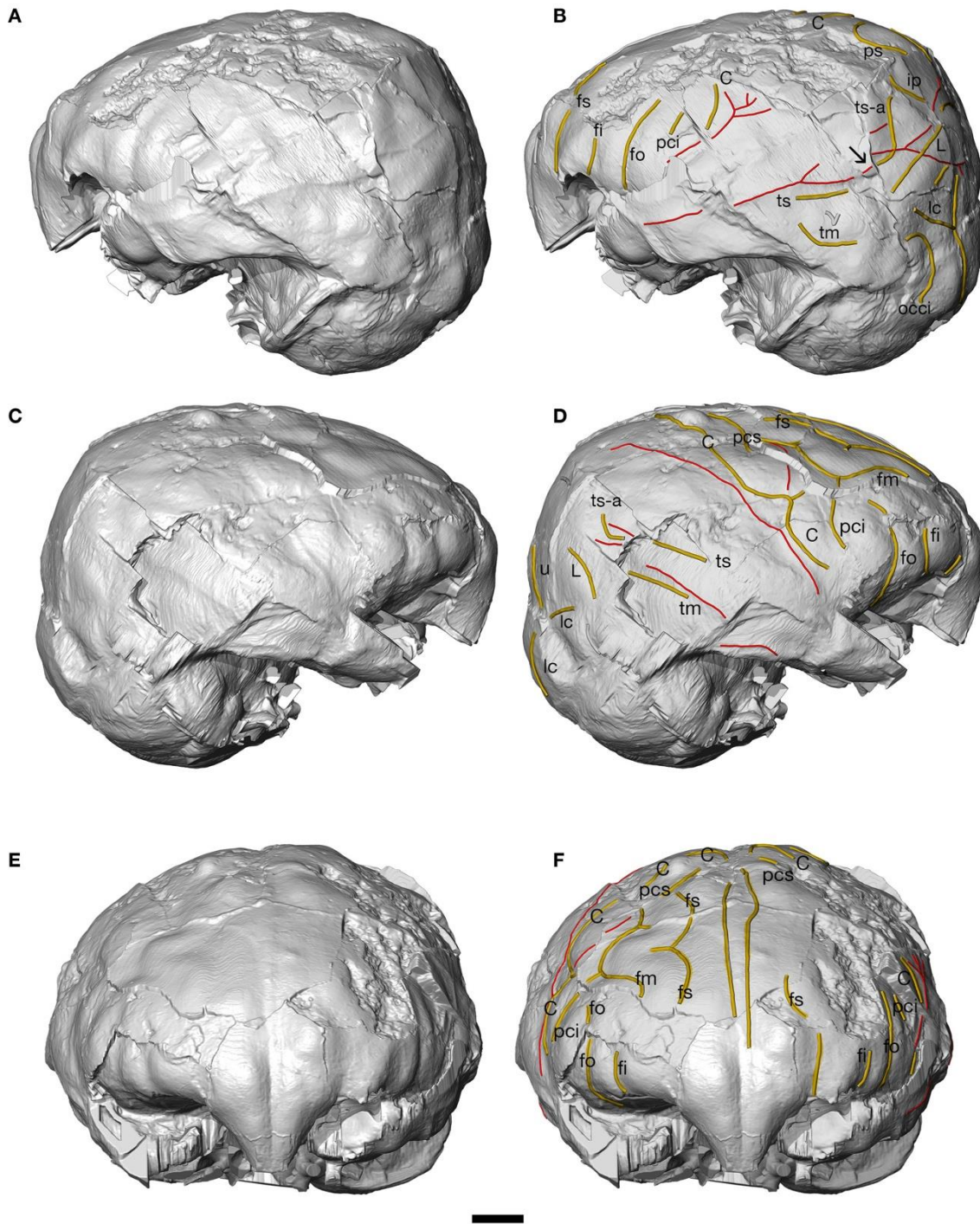


Fig. S3. Natural endocast of DIK-1-1. Lateral views of the unreconstructed DIK-1-1 endocast from the left (**A, B**) and the right (**C, D**) side, as well as in frontal view (**E, F**). Sulci are drawn as yellow lines, blood vessels as red lines. The arrow in (**B**) points to a blood vessel disrupted by the taphonomic displacement of the posterior part of the skull. *C* sulcus centralis, *fs* frontalis superior, *fm* frontalis medius, *fi* frontalis inferior, *fo* fronto-orbitalis, *h* horizontal ramus of *pci*, *ip* s. intraparietalis, *pci* praecentralis inferior, *pcs* praecentralis superior, *ps* parietalis superior, *pti* postcentralis inferior, *ptm* postcentralis medius, *pts* postcentralis superior, *L* s. lunatus, *ts* temporalis superior, *ts-a* ramus temporalis superior, *tm* temporalis medius, *occi* occipitalis inferior, *lc* s. calcarinus lateralis, *u* s. calcarinus ramus superior. Scale bar is 1 cm.

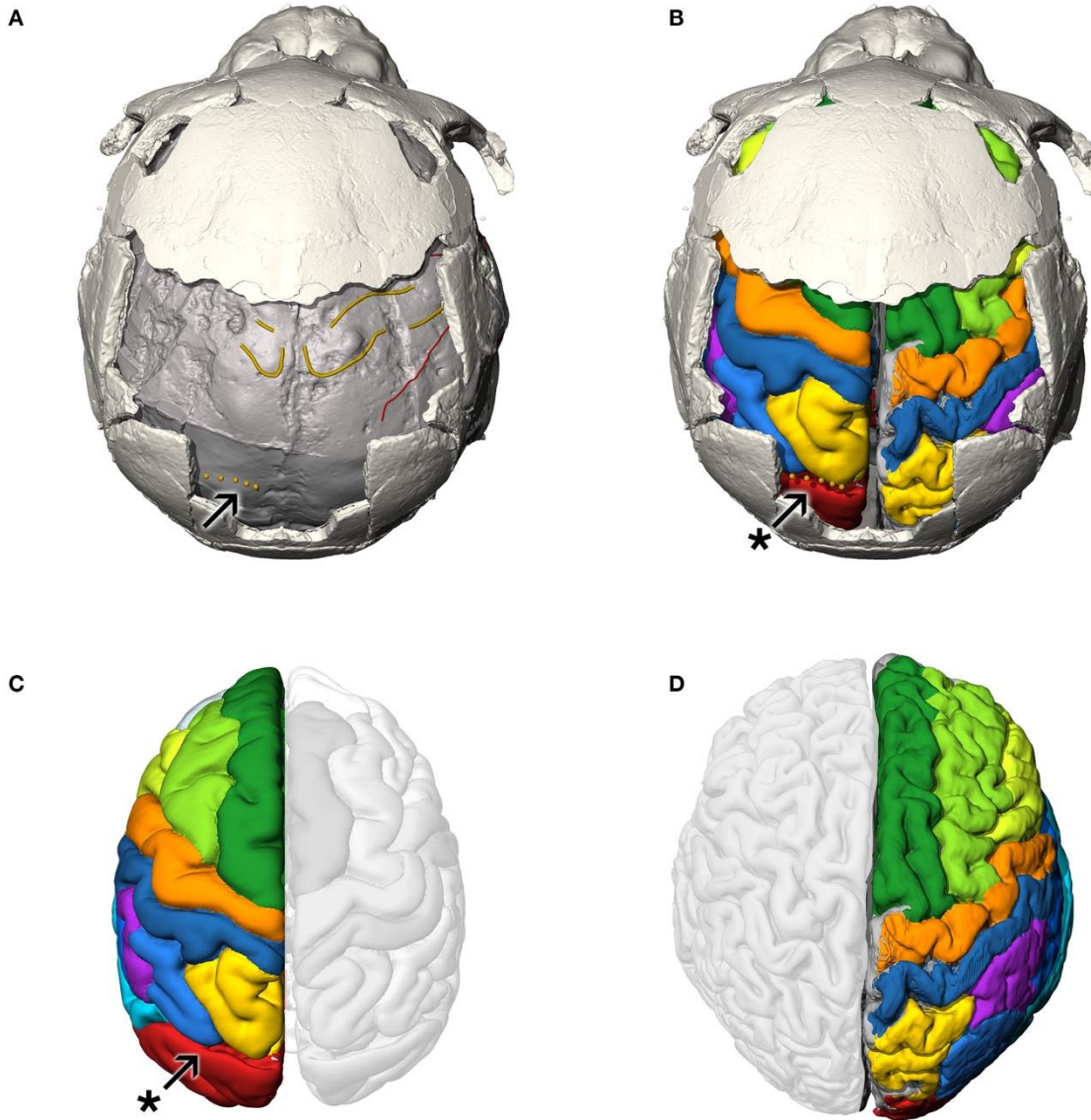


Fig. S4. Ape-like lunate sulcus in DIK-1-1. (A) The reconstructed DIK-1-1 endocranium shows a clear impression of a lunate sulcus (L; shown as an orange dotted line). (B) Thin-plate spline warping the left hemisphere of a chimpanzee brain (C) and the right hemisphere of a human brain (D) to match the neurocranial shape of the DIK-1-1 reconstruction shows that its lunate sulcus matches the chimpanzee prediction perfectly (arrow with asterisk). Morphology and position of the lunate sulcus in DIK-1-1 are therefore ape-like.

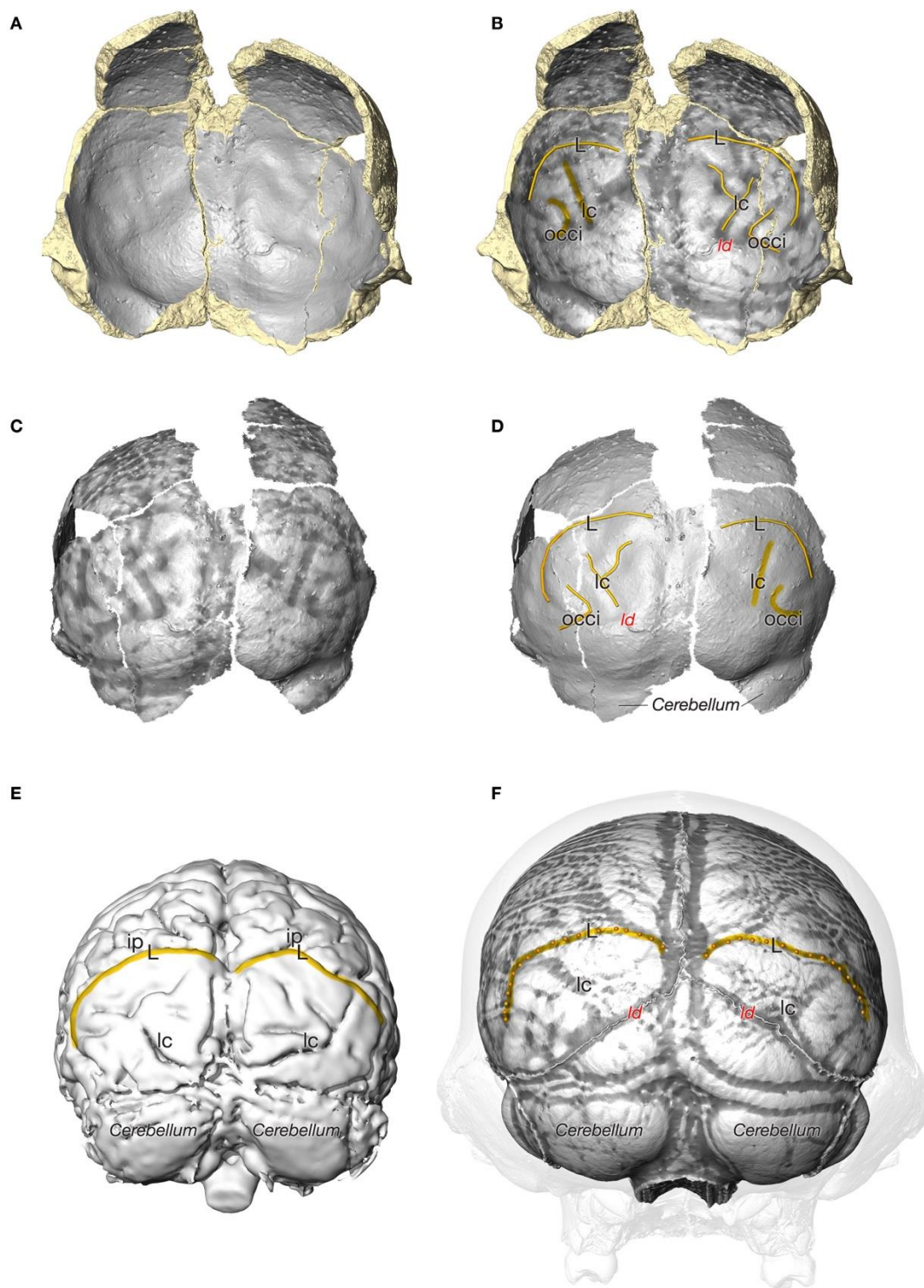


Fig. S5. Endocranial morphology of A.L. 162-28. The micro-CT data of the specimen reveal a clear impression of an ape-like lunate sulcus (L) on the left and right side. Sulcal labels as in Fig. 3; *ld*: lambdoidal suture. (A, B) Anterior view. The endocranial surface is shown in gray. (C, D) Posterior view of the endocranial surface. (E) Posterior view of a chimpanzee brain based on an in-vivo MRI scan (“Amanda” from the Yerkes National Primate Research Center). (F) Chimpanzee endocast based on a post-mortem CT scan (*Pan troglodytes verus* from the Tai forest). In (B, C, F) we superimposed a grayscale gradient based on the local curvature to visually enhance the sulcal impressions.

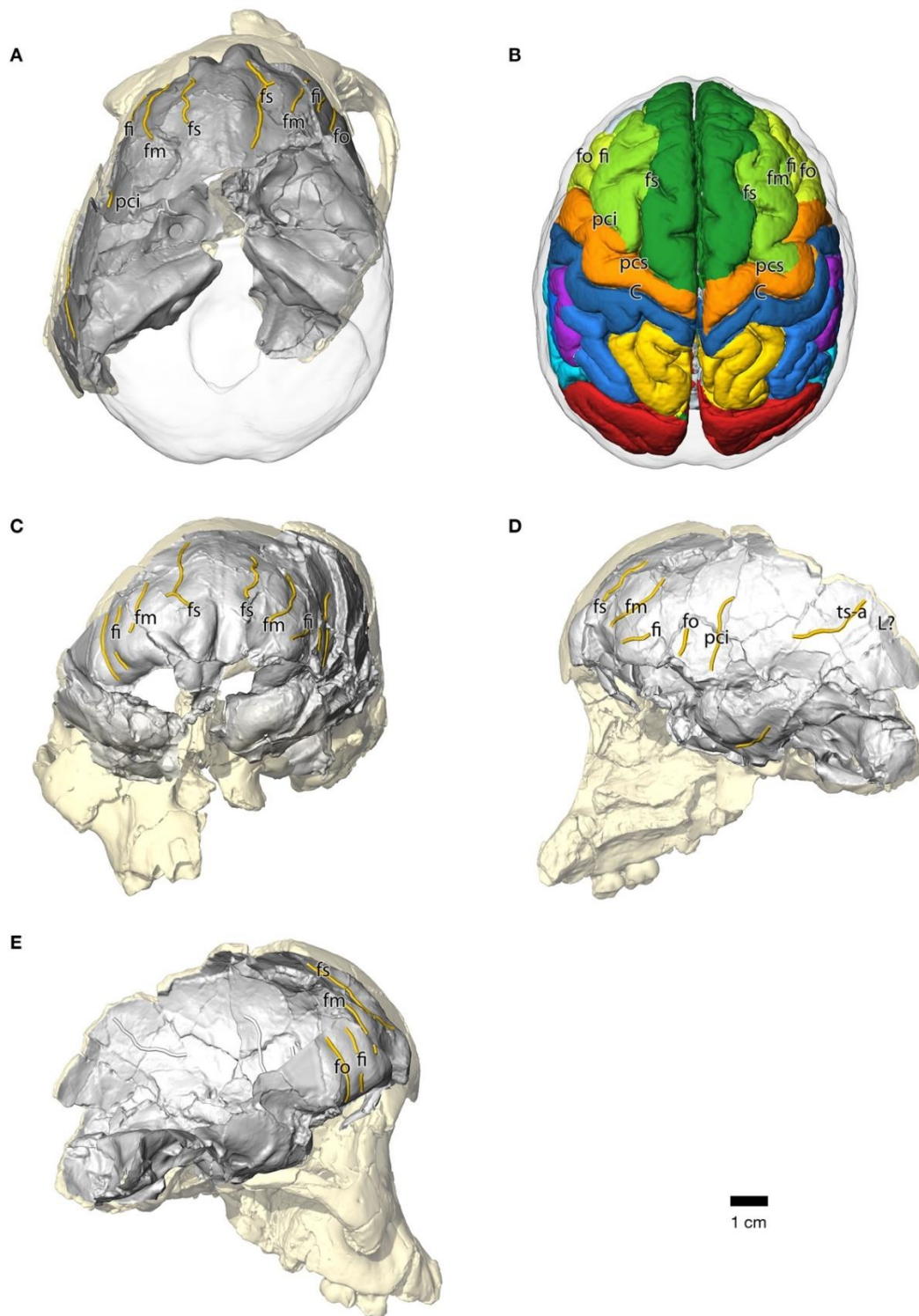


Fig S6. Endocast of A.L. 333-105. (A) Superior view of the unreconstructed endocast of A.L. 333-105. Sulci are drawn as yellow lines (for labels see Methods section). Semitransparent gray surface shows outline of reconstructed endocasts. The original bone is also shown as a beige semitransparent surface. (B) superior view of an infant *Pan troglodytes* brain from MRI data and an endocasts (semitransparent surface) from CT data of the same individual. Colors approximate the boundaries between major convolutions of the brain. (C-E) as in (A) showing frontal, left lateral and right lateral views, respectively. Gray lines in (E) show *ts-a*, *pci*, and part of *fo* on the opposite (left) side. Scale bar is 1 cm.

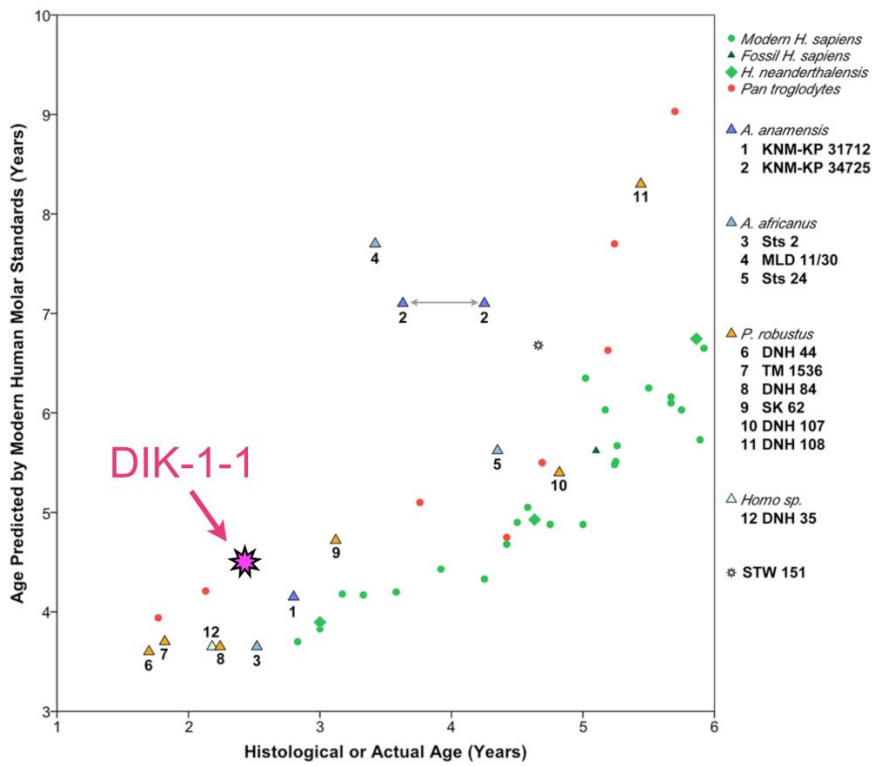


Fig. S7. Ages predicted from modern human molar calcification standards compared to known- or histologically-derived ages. DIK-1-1 is plotted as a pink star. The method and comparative sample are described in (23).

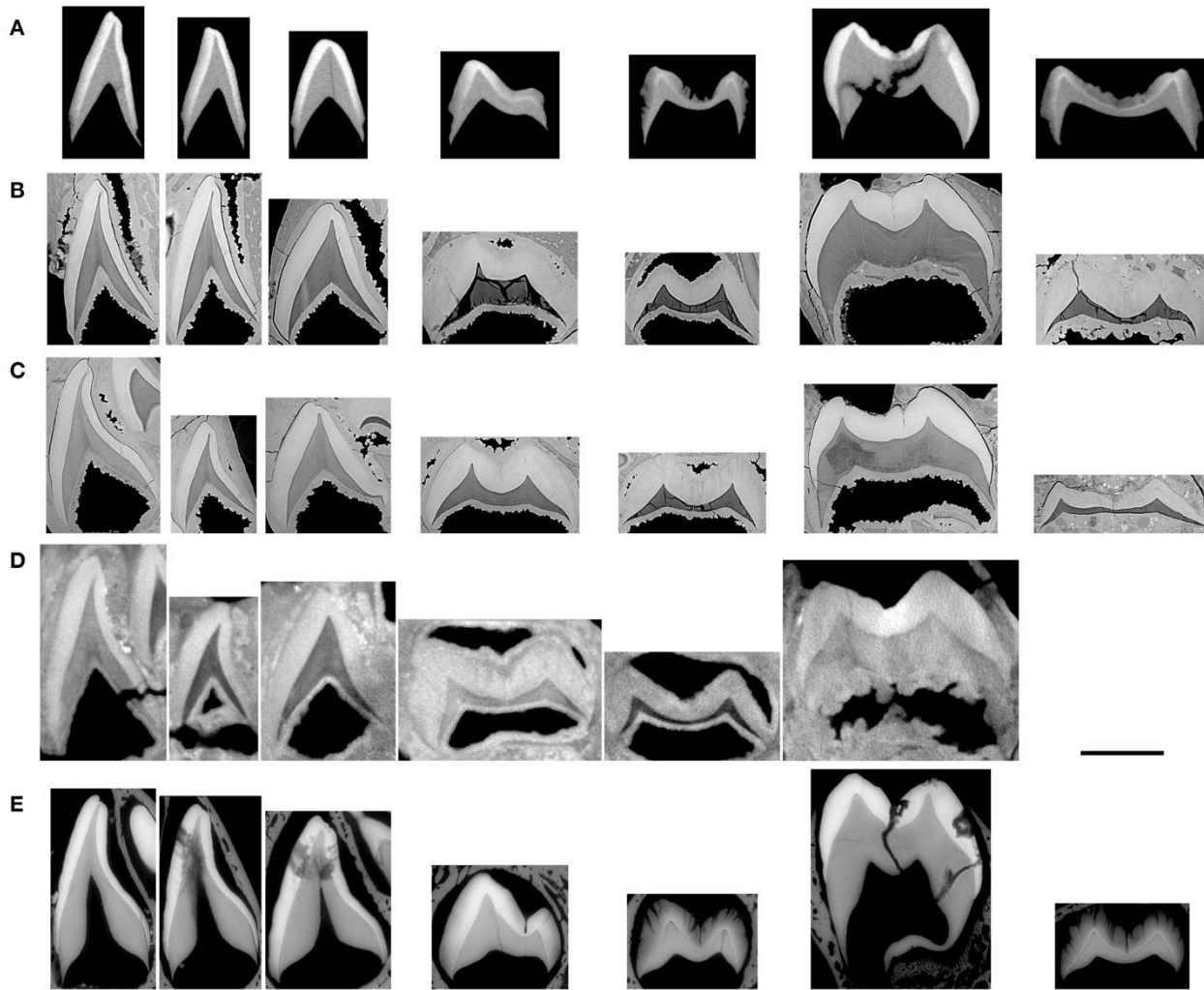


Fig S8. Chimpanzee-equivalent age at death estimation. Teeth from left to right: I1, I2, C, P3, P4, M1, M2. (A) Lower permanent teeth of a *P. troglodytes verus* specimen from the Tai Forest, Ivory Coast (MPI-EVA 06/16 [11777], Bambou), aged 2.13 years at death. The specimen was imaged using a Skyscan 1173 microtomograph. (B, C) Lower and upper permanent teeth of DIK-1-1 (*A. afarensis*), respectively, imaged at the ESRF, on the ID19 beamline. (D) Upper permanent teeth of A.L. 333-105 (*A. afarensis*), imaged with a Skyscan 1173 microtomograph. The M2 is not preserved. Both *A. afarensis* specimens are similar to the chimpanzee in overall developmental stage and pattern. However, in enamel thickness and overall dental shape they more closely resemble modern humans, justifying the use of the thick enamel model for score conversion from micro CT slices to radiographs. (E) Lower permanent teeth of a modern human (CCEC-30001165, musée des Confluences, Lyon), imaged at the ESRF beamline BM05. Comparison of modern and fossil specimens shows that fossilization has substantially modified the original degree of mineralization of the dental germs: incomplete crowns appear to be fully mineralized (e.g., P4s and M2s), and normal density gradients are not visible at the developing cervix of all other teeth. Scale is 5 mm.

Table S1. Comparative data for lower first molar metaconid crown formation times.

| | Time in Days (Max– Min) | Source |
|-----------------------------------|-------------------------|------------|
| <i>A. afarensis</i> (DIK-1-1) | 603 | This study |
| <i>A. afarensis</i> (A.L. 333-52) | 737 | (60) |
| <i>P. troglodytes</i> | 633 (601–682) | (61) |
| <i>H. neanderthalensis</i> | 846 (788–885) | (37) |
| Early SA <i>Homo</i> sp. | 876 | (23) |
| <i>G. gorilla</i> | 888 (714–1004) | (23) |
| <i>H. sapiens</i> (recent) | 930 (842-990) | (37) |

Table S2. Technical parameters. Synchrotron scanning of DIK-1-1 at the ESRF.

| | | | | | |
|--------------------------------|--|--|---|--|--|
| Voxel size (μm) | 45.7 | 30.3 | 4.94 | 0.678 | 0.678 |
| Sample | complete skull | complete skull / dental developmental level | permanent teeth dental germ | LLI1 enamel long period lines periodicity | LLI1 enamel long period lines periodicity |
| Beamline | ID17 | ID19 | ID19 | ID19 | ID19 |
| Optic | 45 μm taper optic ID17 with cerium doped taper protection | 30 μm Hasselblad optic from ID19 | 5 μm Rodenstock optic ID19 | Optic Peter revolver with 10x 0.3 N.A. and 2x eyepiece | Optic Peter revolver with 10x 0.3 N.A. and 2x eyepiece |
| Date | March 2010 | March 2010 | March 2010 | March 2010 | March 2010 |
| Average energy (keV) | 70 | 100 | ~ 71 | 60 | 60 |
| Filters (mm) | vitreous carbon 2.56 Al 4.2 Cu 2 | vitreous carbon 2.56 Al 4.2 Cu 2 | Diamond 1mm Al 2mm Cu 1mm | Diamond 1mm Al 2 mm Cu 0.14 mm | Diamond 1mm Al 2 mm Cu 0.14 mm |
| Propagation distance (mm) | 4500 | 5000 | 5000 | 150 | 150 |
| Monochromator | Si 111 curved double Laue | Si 111 curved double Laue | polychromatic beam | single bounce multilayer W/B4C 2.5 nm | single bounce multilayer W/B4C 2.5 nm |
| Sensor | FReLoN 2K14 (62) | FReLoN 2K14 (62) | FReLoN 2K14 | FReLoN 2K14 | FReLoN 2K14 |
| Camera mode | Frame transfer mode, no shutter | Frame transfer mode, no shutter | Frame transfer mode, no shutter | Full Frame, mechanical X-ray shutter | Full Frame, mechanical X-ray shutter |
| Scintillator | Gadox 60 | LuAG:Ce 750 μm | Gadox 5 μm | GGG:Eu 20 μm | GGG:Eu 20 μm |
| Insertion device (ID) | W150+W125 | W150 | W150 | U32 | U32 |
| ID Gap (mm) | 25 / 40 | 27 | 65 | 11.5 | 11.5 |
| Machine filling mode | 16 bunches (max 90 mA) | 16 bunches (max 90 mA) | 16 bunches (max 90 mA) | 16 bunches (max 90 mA) | 16 bunches (max 90 mA) |
| Number of Projections | 3999 | 5000 | 6000 | 2499 | 5000 |
| References every N projections | 3499 | 5000 | 6000 | 100 | 250 |
| Scan geometry | 360 degrees, step by step scan, vertical series, absorption protocol with glass balls (36) | 360 degrees, half acquisition, step by step scan, vertical series, double scan | 360 degrees, half-acquisition, continuous scan, vertical series | 360 degrees, vertical series, continuous scan | 360 degrees, half-acquisition, continuous scan |
| Exposure time (s) | 0.15 | 0.1 | 0.3 | 0.7 | 1 |
| Time per scan (min) | 22.28 | 23.00 | 40.43 | 41.86 | 100.61 |
| Number of scans | 40 | 62 | 92 | 5 | 2 |
| Reconstruction | modified Paganin delta/beta 1000 + ring correction (63, 64) | modified Paganin delta/beta 1000 + ring correction (63, 64) | edge detection | Paganin delta/beta 50 (for denoising only) | Paganin delta/beta 50 (for denoising only) |

Data file S1. SI_Gunz_et_al_Dental_Scoring. Dental score conversion from microtomographic to radiographic data.

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