Supplementary Information for:

Biocatalytic routes to stereo-divergent iridoids

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	NsNEPS2*
Wavelength	1.00
Resolution range	49.54 - 1.85 (1.916 - 1.85)
Space group	P 21 21 21
Unit cell	68.753 106.68 142.876 90 90 90
Total reflections	1077784 (57590)
Unique reflections	88386 (7393)
Multiplicity	12.2 (7.8)
Completeness (%)	97.81 (82.82)
Mean I/sigma(I)	23.33 (2.05)
Wilson B-factor	36.43
R-merge	0.05726 (0.7775)
R-meas	0.05975 (0.8335)
R-pim	0.01681 (0.2918)
CC1/2	0.999 (0.798)
CC*	1 (0.942)
Reflections used in refinement	88361 (7392)
Reflections used for R-free	4417 (369)
R-work	0.1718 (0.3073)
R-free	0.1974 (0.3428)
CC(work)	0.969 (0.870)
CC(free)	0.962 (0.803)
Number of non-hydrogen atoms	8195
macromolecules	7467
ligands	280
solvent	552
Protein residues	1031
RMS(bonds)	0.008
RMS(angles)	0.89
Ramachandran favored (%)	97.85
Ramachandran allowed (%)	2.15
Ramachandran outliers (%)	0.00
Rotamer outliers (%)	0.00
Clashscore	5.77
Average B-factor	39.2
macromolecules	38.89
ligands	36.35
solvent	44.31

Supplementary Table 1. NsNEPS2 crystal structure data collection and refinement statistics.

Statistics for the highest-resolution shell are shown in parentheses.

Supplementary Table 2. Primers used for cloning *N. sibirica* genes from cDNA.

NEPS	Primer FW	Primer RV
NsNEPSL	AAGTTCTGTTTCAGGGCCCGGCGAACAATTTCCTCATGC	ATGGTCTAGAAAGCTTTATTTTGGAGGGGTGACG
NsNEPS1A	AAGTTCTGTTTCAGGGCCCGGCAAGCATTGTAAATCCGG	ATGGTCTAGAAAGCTTTATGTTGTTGAAGGTGCAACG
NsNEPS1B	AAGTTCTGTTTCAGGGCCCGGCAAGCATTGTAAATCCGG	ATGGTCTAGAAAGCTTTAGGATGAAGGAGCAAAGAATG
NsNEPS2	AAGTTCTGTTTCAGGGCCCGGGCCACAAGAAGAAGCTC	ATGGTCTAGAAAGCTTTATGAATGGGCGGCGAAT
NsNEPS4A	AAGTTCTGTTTCAGGGCCCGGCAAGCATTGTAAATCCGG	ATGGTCTAGAAAGCTTTATGTTGTTGAAGGTGCAACG
NsNEPS4B	AAGTTCTGTTTCAGGGCCCGGCAAGCATTGTAAATCCGG	ATGGTCTAGAAAGCTTTATGTTGTTGAAGGTGCAACG
MLPL	Primer FW	Primer RV
NsMLPL1	AAGTTCTGTTTCAGGGCCCGGCTTCCAAGCTTGAAGTGG	ATGGTCTAGAAAGCTTTATGCCTTGAGAACATAATCAT
NsMLPL2	AAGTTCTGTTTCAGGGCCCGGCTTCAAAGATTGAAGTAGAAAT	ATGGTCTAGAAAGCTTTATGCCTTGAGAACATAATCAT
NsMLPL3	AAGTTCTGTTTCAGGGCCCGGCTTCAAAACTTGAAGTAGAAAT	ATGGTCTAGAAAGCTTTATTCCTTGAGAAGATAATCATCCA
ISY	Primer FW	Primer RV
NsISY	AAGTTCTGTTTCAGGGCCCGAGCTGGTGGTGGGCTG	ATGGTCTAGAAAGCTTCAAGGAACAATCTTGTAAGCCTT
NsP5ßR	AAGTTCTGTTTCAGGGCCCGAGCTGGTGGTGGGCTG	ATGGTCTAGAAAGCTTTAAGGAACAATCTTGAAAGCT

Supplementary Table 3: *Nepeta Sibirica* and *Lamium album* cloned genes

Gene	Sequence
NsNEPSL	ATGGCGAACAATTTCCTCATGCAATTGAAGAAGCTCGAAGGCAAAGTAGCCATTGTAACTGGCGGCGCCAGTG GCATCGGCGAGGCCACCGCCGCCTCTTCGCGAATCGCGGCGCCACGCCGTGGTTATAGCCGACATTCAGCA GGAGAAGGGCCGCTCCGTGGCGGAATCCATCGGGACGCAGCGGCCACGCGCGTGGACATCACCGCGCGCG
NsNEPS1A	ATGGCAAGCATTGTAAATCCGGTGCAGGTGATGAAGAAGAAGCTGGAAGGCAAAGTTGTGATAGTAACAGGC GGGGCGAGCGGCATCGGGGAGACGGCAGCGCGTGTGTTTGCGCAACATGGCGCGCGTGCAGTGGTGATCGCT GACATCCAATCTGAAGTTGGGAAGTCCGTGGCGGAGTCCATCGGGAAGCGGTGCAGCTACGTCCAGTGCGACG TCTCGGACGAGGAGCAGGTAAAGTCGATGATAGAATGGACGGCCAGCACGTACGGCGGGCTGGACGTGATGT TCTCCAATGTGGGCATCATGAGCAGCTCCGCTCAAACCGTGATGGACCTCGACCTTTCGGAGTACGATAAGGT GATGCGTGTGAACGCCGCGGGGCAGCGCCGCGTGCTTGAAGCAGGCGGCGCGCGAAGATGGTAGAGCTGGGAAC GAGAGGCACTATTATCTGCACGACCAGCGTGGTGGTGGCAGGGGGCGGGGCAAAGCCTGACGGACTAGGGCTGGAAC GCGTGTCGCGGTGGTGGGGCTGGTCGGCCGCGGCGAACGCTGGGGGCCACGGGGATAGGGTTAACT GCGTGCCGCGGTGGTGGTGCAGCGCCGCCGCCCCAAAGGATGGTGGGGGCCACGGGGCTGAGGGTTACCATACT CATTTTGGCAACTTCACTAGCCTCAAAGGAGTCTGCCTCACCGCCGACGACGACGCCGCCGAGGACTATCCATACT CGCTTCCGACGACGCCGCGGTTCATCACCGGGACATAATTTGGACGTCGATGGTGGACCGCCTTGTTTACCATTCG TTGCACCTTCAACAACATAA

NsNEPS1B	ATGGCAAGCATTGTAAATCCGGTGCAGGTGATGAAGAAGAAGCTGGAAGGCAAAGTTGTGATAGTAACGGGC GGGGCGAGCGGCATCGGGCAGACGGCAGCGCGTGTGTTTGCGCAACATGGCGCGCGTGCAGTGGTGATCGCTG ACATCCAATCTGAAGTTGGGAAGTCCGTGGCGGAGTCCATCGGGAAGCGGTGCAGCTACGTCCAGTGCGACGT CTCGGACGAGGAGCAGGTAAAGTCGATGATAGAATGGACGGCCAGCACGTACGGCGGGCTGGACGTGATGTT CTCCAATGTGGGCATCATGAGCAGTTCCGCTCAAACCGTAATGGACCTCAACCTTGGGGAGTTCGATAAGGTG ATGCGTGTGAACGCGCGCGGGGACGGCCGCGTGCTTGAAGCAGGCGGCGGCGCGAAGATGGTAGAGCTGGGAACG AGAGGCACTATTATCTGCACGACCAGCGCGGCGGCGAGCATGCAGGCGGGGCGGGC
NsNEPS2	ATGCACAAGAAGAAGCTCGAAGGCAAAGTAGCCATTGTAACCGGCGGCGCCAGCGGCATCGGCGAGACCGCC GCCCGCATATTCGCCGACCACGGCGCGCGCGCGGCGGTGGGGGGGCGCGGCGCAGCGAGGAATTGGGCCGGATGG TAGCGGAATCCATTGGGGCGAAGCGGTGCAGCTACGTGCAATGCGACATCGCCGACGAGGAGCAGGTTAAGT CCGCGGTAGAATGGACGGCCACCACCTACGGCGGCGCCCGACGTGGTGTTCTGCAACGCCGGCGCATCATGAGCCA CTCTGACTCCGGACAGACGGTGATGGAGCTCGATATGTCAAAGTTCGACGAGGTGATGCGTGGAACACGCGC GGGACGGCAGCGTGCGTGAAGCAGGCGGCGCGCATAATGTCAAAGTTCGACGAGGGGGGGG
NsNEPS4A	ATGGCAAGCATTGTAAATCCGGTGCAGGTGATGAAGAAGAAGCTGGAAGGCAAAGTTGTGATAGTAACAGGC GGGGCGAGCGGCATCGGGGAGACGGCAGCGCGTGTGTTTGCGCAACATGGCGCGCGTGCAGTGGTGATCGCT GACATCCAATCTGAAGTTGGGAAGTCCGTGGCGGGGGGCGCATCGGGGAGCGCGCGGCGGCGGCGGCGGCGGCGGCGGCGGCG
NsNEPS4B	ATGGCAAGCATTGTAAATCCGGTGCAGGTGATGAAGAAGAAGCTTGAAGGCAAAGTTGTGGTAGTAACGGGC GGGGCGAACGGCATCGGGGAGACGGCGGCGCGCGCGCGTGTTTGCGGAGCATGGCGCGCGTGCGGTGGTGATTGCT GACATCCAATCTGAAGTTGGGCAGTCCGTGGCGGAGGCCATCGGGGAGGGGGCGCAGCTACGTCCAGTGCGACA TCTCGGACGAGGAGCAGGTTAAGTCGATGATAGAATGGACGGCCAACACGTATGGCGGGCTGGACGTCATGTT CTGCAATGCGGGGCATCATTACCTACTCCCCTCAAACCATAATGCACCTCGACCTCTCGCAATTCGATAAGGTGA TGCGTGTGAACGCACACGGGACGGCCGCGTGCGTGCGTGAAGCAGGCGGCGCGTAAGATGGTGGAGCTGGGAACGA GAGGCACTATTATCTGCACGACTAGCGCGACAGCATCCAAGGGCGGCGCGTAAGATGGTGGAGCTGGGAACGA GAAGCACGCGGTGGTGGGGCTGGTCCGGTCAGCGACGACTCCAAGGGCGGACAAAACATGACGGACTATGCGATGTC GAAGCACGCGGTGGTGGGGCTGGTCCGGCCAGCGACGACTGCGGGGACTGGGGGCCCACGGGATTAGGGTTAACTG CGTGTCGCCCTCGGCGGTGCTCACGCCGCTCGCCCAAAGGATGGGGACTGCCACGCCTGATGATTATATACTC ATTTTGGCAACTTCACTAGCCTCAAAGGAGTCTACCTCACCGCCGACCAAGTCGCCGAAGCCGTCACCTTTCTC GCTTCCGACGACGCTGCTTTCATCACCGGACATAATTTGGACCTCGATGGTGGACTGCTTTGTTTACCATTCGTT GCACCTTCAACAACATAA
NsMLPL1	ATGGCTTCCAAGCTTGAAGTGGAGCTCGAGTTGAAATCTGATGTAGAAAAAATGTGGAAAAACTTTAAGGAAT TTACAAAATTATTCCCCAAGGCTTTGCCACATCTTTACGAAGGGATTGCCGTTGCCGAGGGCGATGGGATATCC GCCGGAACAATCTTCATAAGCACTCTTAAACCGACAGATCCGTCTAACCCGTGGTTTCGATCAACAAGGAGA GGATTGATTCTCTAGATGATGAAAAGAAAA
NsMLPL2	ATGGCTTCAAAACTTGAAGTAGAAATTGAGTTGAAAAACTCATTCAGAAAATCTGTGGAAAAACCTGAAAGAAT TCATCACTTTCTTCCCCCAAAGCATTGCCAAATATGTACGAAAGATCGATGTGATCGAAGGCGATGGAAGATC AGTTGGATCTGTCTTTGTGTCTACTCTAAAGCCATCAGAGTTAAACCCTGTGGGTTGAGGTCACAAAGGAGAGGA TCGAACTGTTGGATGAAGAGAAGA

NsMLPL3	ATGGCTTCAAAGATTGAAGTAGAAATTGAGTTGAAAACTCCTTCAGATAAACTGTGGAAAAACCTGAAAGAAT TCGTTTTCTTCTTCCCCCAAAGCTTTGCCACATATGTTCGAGAAGAATGATGATGTGATAGAAGGCGATGGAAGATCA GTTGGATCTGTATTTGTGGCCACTGTTAAGCCATCAGAGTTATACCCGGTGGTTACCACAAAGGAGAGGATTGA AATGGTTGATGAAAAAAATAAGATGATGAGTTACAGTTTTGTTGAGGGGTGAAATGTTGAAAAAATAACAAGAAT TTCAAGGCCACAATGTGTGTGGAGCAGCAACAAAAATGATGGGTCTATAATCAAATATACAGCTGAATTTGAGA AGGCAAATGCAGTTCCAGATCCATATTTCGTTACGGATAATGCTGCTAAACTTTTACATGACGTGGATGATTAT CTTCTCAAGGCCATGA
NsISY	ATGAGCTGGTGGTGGGGCTGGAGCTACTGGCGCTGCCAAGAAAAGAATAGATGAAGAGAGGAGTCACTCCTAAAC CACCAATGCGTAGCTCTGATAGTCGGGGGTGACCGGACTCATCGGCAACAGCCTGGCGGAGATCCTGCCGCTCT CCGACACCCCCGGCGGCCCATGGAAGGTATACGGTGTGGCGCGCCGCCCCCGTCCCTCGGAACGAGGATCA CCCCATCACTACATCTCATGCGACGTAACCAACACAGCCGACGTGGAGGCCAAGCTATCCCCTCTCACCGAC GTAACACACATCTTCTACGCCACGTGGACCAGCCGATCCACCGAGGAGGAGAACTGCGAAGCCAACGGGAAA ATGCTGAAAAATGTGCTGGACGCAATGATCCCTAACTGCCCCAATTTGAAGCATATCTGCTTGCAGACCGGTA GATTCCACTACGTTGCTTCGGTTGTGGACTGGAAGACTGCGAAGCACGCCACCGACACCGGCCACGGAGAACTGCGAAGAACGAGGAGGCT CGATTGAACACGAAGAATTTCTACTATACGCAAGAGGATATACAGCCACGACACTCCGTTAACCGAGGAGGATTACCT CGATTGAACACGAAGAATTTCTACTATACGCAAGAGGAGGATATTCTGTTTGAGGAGGGTTAAGAAGAAGGAGGGGC TGACATGGTCCGTGCATCGGCCGGGGACTATCTTCGGGGGGTTCTGAGGTTCTCGGAGGAGATTTGGTTGG
NsP5βR	ATGAGCTGGTGGTGGGGCTGGAGCTATTGGCGCTGCCAAGAAAAGAATCGATGAAGATGAGGCACCGCGGAAC TACGAGAGCGTAGCTCTGATAGTGGGGGGGACCGGAATCGTAGGCAACAGCCTGGCGGAGATTCTCCCGCTCT CCGACACTCCCAGTGGCCCATGGAAGGTTTATGGGGTGGCCGCCCCCCGTCCCTCCTGGAACGACGATCA CCCCATTACCTACATCTCCTGCGATGTATTGGACTCCGTCGACGTGGAGGCGAAGCTATCCCCCTCACCGATG TAACACACATATTCTATGCCACATGGACCAAGGATCCACGGAGAGGAGGAAGCGCGAAGCTAATGGGAAAA TGCTGAAAAACGTGCTGAATGCAATGACCCTAATTGCCCCAATTTGAAGCATATCTGTTTGCAGAACTGGTAGG AAGCATTATGTTGGTGCATTTGAAATGGACCAAGAGATTAAAAGAAGTCACGATCCTCCGTTCACTGAGGAATTGCC TCGATTGGATTCCCAGAATTTCTATTATACACAAGAGGACATTCTGTTTGAGGAGGTTCAGAAGAAGGAGGGC TTGACATGGTCTGTGCATCGGCCTGGGAATATTTTCGGGTTCTCACCGTATAGCATGATGAAGAAGGAGGGC TTGACATGGTCTGTGCATCGGCCTGGGAATATTTTCGGGTGCCACGTATAGCATGATGAAGGAGGGCC TGGATACTCGGATTGCTCGGATGCAGACTGAACGAGGGGCGCGTGGATCCTTATGC GAAGAATGAGGCATTCAATGTGAGCAACGGCGATGTTTTCAAATGGAAGCATTCTGTAAGGGTGCCTGGG ATGGATACTCGGATTGCTCGGATGCAGACTGGCAGCATCAGAATATGGGCGGCCGTGGATCCTTATGC GAAGAATGAGGCATTCAATGTGAGCAACGGCGATGTTTTCAAATGGAAGCATTCTGGAAGGGTGTTGGCCGAA CAGTTTGGCGTGGAATGTGGGGAGAATGGGCAGGCAGGAAGTGAAGTTGCAGGATCTGATGAAGGATAAA GGTCCGATCTGGGACAAAATCGTGAGGGAGAATGGGTGTGCGGCTACGAAATTGGAGGATGTTGGGACTGGT GGTTTAGGACATTATTCTCGGGAATGAATGTTGGTTGGATACAATGAACAAAAGCAAGGAGCATGGATTGTC CGATTCAGGAATTCCAAGAATTCCTTCATTTCTTGGATTGACAAGGTGAAAGCTTCAAGATTGTTCCTTAA
LaISY	ATGCCGACCGAAACGATCATGAGTTGGTGGTATAAACGCAGCATTGGTGACATTGAACAGAAGAAACTTCAGT CCAATGGCCATGCACCGAGCTACAAATCGGTTGCGCTTATTGTGGGAGTTACGGGCATTGCGGGAACACTCGGGTACACAATCGGGTGCCGTGGGGGGATACTCCAGGAGGCCCGTGGAAAGTGTATGGGGTTGCACGGCGTCCGTGTC CAGAGTGGCTTACCACACTCCATGTCGACTATATCCAGTGTGACATTGCCAACACCGAAGAAACGAACTCCAA GCTGAGTCCGTTGAAAGATATTACCCATGTATTCTACGTGAGTTGGACAGGGAGTGAAGATGTTGCGCTGAAC ACGCTGATGTTCCGCAATATTCTCGACTCGGTGATCCCGAATGCCCCGAATCTGAAACATGTGGCCTGGCAAC CGGGATCAAATACTACTGGGGCAACATGGCCGAGATGGAAAGCACTAATCAGCCGCATGAATGCCCCTTCTAT GAGAATTTACCACGTCTGAAACAGGAAAACTTCTACTACAATCTGGAAGATTTGGTATATGAAGCAGGTTTGG GTCGCTCATCACTGACTTGGTCTGTGCACCGTCCTGCGGCTGATTTCGGGTTTTCTCCTTGTTCGATGATGAACG CCGTGAGCACCATGTGCGTCTATGCTGCGACCGTCTGCAAACATGAGAAACACCCTGGTCTATACCGGTACCGA AGTCAGCTGGACTTGTCGTGGGGATGCGGTAGATAGCGATCTGTAGCCGATCACTTTGTTTG

Supplementary Table 4: Summary of 7S stereoselective enzymatic activities

Figure	Substrate	ISY	NEPS/MLPL	7S iridodials	7S-trans- trans nepetalactol	7S-cis-trans nepetalactol	7S-trans-cis nepetalactol*	7S-cis-cis nepetalactol	7S-trans-trans nepetalactone	7S-cis-trans nepetalactone	7S-trans-cis nepetalactone	7S-cis-cis nepetalactone
2A	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
2A	8-oxogeranial	CrISY	NsNEPS2	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
2A	8-oxogeranial	CrISY	NsNEPS2-Y167F	tr.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
2B	8-oxogeranial	CrISY	NmNEPS3	+	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	tr.
2B	8-oxogeranial	CrISY	NcNEPS3A	+	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	++
2B	8-oxogeranial	CrISY	NcNEPS3A-V206Q	+	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	tr.
2B	8-oxogeranial	CrISY	NmNEPS3-Q206V	+	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	++
2D	8-oxogeranial	CrISY	NmNEPS1	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	tr.	tr.
2D	8-oxogeranial	CrISY	NmNEPS4	tr.	n.d.	n.d.	++	n.d.	n.d.	++	tr.	n.d.
2D	8-oxogeranial	CrISY	NmNEPS1 + NmNEPS4	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	++	+++	n.d.
2D	8-oxogeranial	CrISY	NmNEPS1-8 mutation graft	tr.	n.d.	n.d.	+	n.d.	n.d.	++	n.d.	n.d.
2D	8-oxogeranial	CrISY	NmNEPS1- 154SATA-S198L	tr.	n.d.	n.d.	++	n.d.	n.d.	++	n.d.	n.d.
2D	8-oxogeranial	CrISY	NmNEPS1- 154SATA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
2D	8-oxogeranial	CrISY	NmNEPS1- 154SVTA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	++	++	tr.
S6	8-oxogeranial	CrISY	NmNEPS1-S198L	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	tr.
S6	8-oxogeranial	CrISY	NmNEPS1 with NmNEPS4 loop	tr.	n.d.	n.d.	tr.	n.d.	n.d.	+++	+	tr.
S6	8-oxogeranial	CrISY	NmNEPS1-S198L with NmNEPS4 loop	tr.	n.d.	n.d.	tr.	n.d.	n.d.	+++	n.d.	n.d.
3C	8-oxogeranial	NsISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsNEPSL	n.d.	n.d.	n.d.	n.d.	n.d.	tr.	+++	tr.	tr.
3C	8-oxogeranial	NsISY	NsNEPS2	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsNEPS4A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsNEPS4B	+++	n.d.	+++	n.d.	n.d.	n.d.	tr.	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	tr.	tr.
3C	8-oxogeranial	NsISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	+	tr.
3C	8-oxogeranial	NsISY	NsMLPL1	tr.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsMLPL2	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3C	8-oxogeranial	NsISY	NsMLPL3	+++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	NsP5βR	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	NsISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	LaISY	N/A	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

Figure	Substrate	ISY	NEPS/MLPL	7S iridodials	7S-trans- trans nepetalactol	7S-cis-trans nepetalactol	7S-trans-cis nepetalactol*	7S-cis-cis nepetalactol	7S-trans-trans nepetalactone	7S-cis-trans nepetalactone	7S-trans-cis nepetalactone	7S-cis-cis nepetalactone
4B	N/A	N. sibiri	ca leaf tissue extract	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+	n.d.	n.d.
4B	8-oxogeranial	CrISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	n.d.
4B	8-oxogeranial	CrISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	++	n.d.
4B	8-oxogeranial	LaISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
4B	8-oxogeranial	LaISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S3	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S3	8-oxogeranial	CrISY	NsNEPS2-Y163F	tr.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
\$3	8-oxogeranial	CrISY	NsNEPS1A-Y167F	++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
\$3	8-oxogeranial	CrISY	NsNEPS1B-Y164F	++	n.d.	+	n.d.	n.d.	n.d.	++	n.d.	n.d.
\$3	8-oxogeranial	CrISY	NcNEPS3A-Y165F	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S3	8-oxogeranial	CrISY	NmNEPS4-Y168F	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
\$3	8-oxogeranial	CrISY	NmNEPS1-Y167F	+	n.d.	n.d.	n.d.	n.d.	n.d.	++	n.d.	n.d.
S4	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S4	8-oxogeranial	CrISY	NcNEPS3A	n.d.	n.d.	n.d.	n.d.	++	n.d.	n.d.	n.d.	+++
S4	8-oxogeranial	CrISY	NcNEPS3A-V206M	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.	n.d.	+
S4	8-oxogeranial	CrISY	NcNEPS3A-V206E	++	n.d.	++	n.d.	++	n.d.	n.d.	n.d.	n.d.
S4	8-oxogeranial	CrISY	NcNEPS3A-V206N	tr.	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	tr.
S4	8-oxogeranial	CrISY	NcNEPS3A-V206G	tr.	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	tr.
S4	8-oxogeranial	CrISY	NcNEPS3A-V206L	tr.	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	tr.
S4	8-oxogeranial	CrISY	NcNEPS3A-V206A	tr.	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	+
S4	8-oxogeranial	CrISY	NcNEPS3A-V206I	tr.	n.d.	+	n.d.	+++	n.d.	n.d.	n.d.	+
S5	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S5	8-oxogeranial	CrISY	NcNEPS3A	n.d.	n.d.	n.d.	n.d.	++	n.d.	n.d.	n.d.	+++
S5	8-oxogeranial	CrISY	NcNEPS3A with NmNEPS1 loop	+	n.d.	+++	n.d.	n.d.	n.d.	+	n.d.	tr.
S5	8-oxogeranial	CrISY	NcNEPS3A with NmNEPS4 loop	++	n.d.	+++	n.d.	n.d.	n.d.	++	n.d.	+
S5	8-oxogeranial	CrISY	NcNEPS3A with NmNEPS5 loop	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SATA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SATS	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SSTA	tr.	n.d.	n.d.	tr.	n.d.	n.d.	+++	+++	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SALA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154AATA	tr.	n.d.	n.d.	+++	n.d.	n.d.	++	+	n.d.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SATG	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.

Figure	Substrate	ISY	NEPS/MLPL	7S iridodials	7S-trans- trans nepetalactol	7S-cis-trans nepetalactol	7S-trans-cis nepetalactol*	7S-cis-cis nepetalactol	7S-trans-trans nepetalactone	7S-cis-trans nepetalactone	7S-trans-cis nepetalactone	7S-cis-cis nepetalactone
S7	8-oxogeranial	CrISY	NmNEPS1- 154SGTG	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SVTA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+++	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154TASA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SAGA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	tr.	tr.
S7	8-oxogeranial	CrISY	NmNEPS1- 154SPTA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+++	tr.
S8	8-oxogeranial	CrISY	NmNEPS1- 154SATA-S198M	+	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S8	8-oxogeranial	CrISY	NmNEPS1- 154SATA-S198L	tr.	n.d.	n.d.	+++	n.d.	n.d.	+++	n.d.	n.d.
S8	8-oxogeranial	CrISY	NmNEPS1- 154SATA-S198P	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S8	8-oxogeranial	CrISY	NmNEPS1- 154SATA-S198V	+++	n.d.	tr	+	n.d.	n.d.	+++	n.d.	n.d.
S8	8-oxogeranial	CrISY	NmNEPS1- 15454TA-5198G	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S8	8-oxogeranial	CrISY	NmNEPS1-	+	n.d.	n.d.	tr.	n.d.	n.d.	+++	+	tr.
S8	8-oxogeranial	CrISY	NmNEPS1-	+	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S8	8-oxogeranial	CrISY	NmNEPS1-	+	n.d.	tr	tr.	n.d.	n.d.	+++	tr.	n.d.
60	0 overeniel	CHEV	154SATA-S198C									
59	8-oxogeranial	CrISY	NSNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	+	tr.
S9	8-oxogeranial	CrISY	NsNEPS1B- 151GAMS	tr.	n.d.	+++	n.d.	n.d.	n.d.	++	++	tr.
S9	8-oxogeranial	CrISY	NSNEPS1B-	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	++	tr.
S9	8-oxogeranial	CrISY	NSNEPS1B- 15154TA	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S9	8-oxogeranial	CrISY	NsNEPS1B-S195L	+	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	tr.
S9	8-oxogeranial	CrISY	NsNEPS1B- 151GSSA	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S9	8-oxogeranial	CrISY	NsNEPS1B- 151AAMS	+	n.d.	+++	n.d.	n.d.	n.d.	++	++	tr.
S9	8-oxogeranial	CrISY	NsNEPS1B- 151ASTA	+	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	tr.
S9	8-oxogeranial	CrISY	NsNEPS1B- 151ASMA	+	n.d.	+++	n.d.	n.d.	n.d.	++	++	tr.
S11	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

Figure	Substrate	ISY	NEPS/MLPL	7S iridodials	7S-trans- trans nepetalactol	7S-cis-trans nepetalactol	7S-trans-cis nepetalactol*	7S-cis-cis nepetalactol	7S-trans-trans nepetalactone	7S-cis-trans nepetalactone	7S-trans-cis nepetalactone	7S-cis-cis nepetalactone
S11	8-oxogeranial	LaISY	N/A	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxogeranial	NsISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxogeranial	NsP5βR	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	LaISY	N/A	++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	NsISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	NsP5βR	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NcMLPLA	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NcMLPLB	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NcNEPS2	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NcNEPS5	n.d.	n.d.	n.d.	n.d.	n.d.	tr.	+++	tr.	n.d.
S13	8-oxogeranial	CrISY	NmNEPSL1	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	CrISY	NmNEPSL2	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	CrISY	NmNEPS2	n.d.	n.d.	+++	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	CrISY	NmNEPS5	n.d.	n.d.	n.d.	n.d.	n.d.	tr.	+++	tr.	n.d.
S13	8-oxogeranial	CrISY	NmMLPL1	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NmMLPL2	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NmMLPL3	+++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	NcMLPL4	+	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	CrISY	HoNEPSLB	n.d.	n.d.	n.d.	n.d.	n.d.	tr.	+++	tr.	n.d.
S13	8-oxogeranial	CrISY	HoNEPSLA	n.d.	n.d.	n.d.	n.d.	n.d.	tr.	+++	tr.	n.d.

Note: The prescence of a particular chemical was graded in the following relative scale: not detected (n.d.), traces (tr.) and three levels of detection beyond traces, (+, ++, and +++), where each "+" sign indicates increasing amounts.

Supplementary Table 5: Summary of *7R* stereoselective enzymatic activities

Figure	Substrate	ISY	NEPS/MLPL	7R iridodials	7R-trans- trans nepetalactol	7R-cis-trans nepetalactol	7R-trans-cis nepetalactol	7R-cis-cis nepetalactol	7R-trans-trans nepetalactone	7R-cis-trans nepetalactone	7R-trans-cis nepetalactone	7R-cis-cis nepetalactone
S12	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPSL	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPS2	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPS4A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPS4B	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	++	+++	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+++	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsMLPL1	+++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsMLPL2	+++	n.d.	+	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S12	8-oxogeranial	LaISY	NsMLPL3	+++	n.d.	+	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	NsP5βR	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	NsISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	CrISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
3D	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
4B	N/A	N. sib	<i>irica</i> leaf tissue extract	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	n.d.	n.d.
4B	8-oxogeranial	CrISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
4B	8-oxogeranial	CrISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
4B	8-oxogeranial	LaISY	NsNEPS1A	n.d.	n.d.	n.d.	n.d.	n.d.	++	++	n.d.	n.d.
4B	8-oxogeranial	LaISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	n.d.	n.d.
5B	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
5B	8-oxogeranial	LaISY	NmNEPS3- Q206V	+	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
5B	8-oxogeranial	LaISY	NsNEPS2-Y167F	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
5B	8-oxogeranial	LaISY	NsNEPS2	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
5B	8-oxogeranial	LaISY	NsMLPL1 + NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	++	+++	n.d.	+
5B	8-oxogeranial	LaISY	NmNEPS1- 154SVTA	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+	n.d.	n.d.
S9	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B	n.d.	n.d.	n.d.	n.d.	n.d.	++	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151GAMS	++	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151SATS	n.d.	n.d.	n.d.	n.d.	n.d.	+++	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151SATA	++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- S195L	n.d.	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.

Figure	Substrate	ISY	NEPS/MLPL	7R iridodials	7R-trans- trans nepetalactol	7R-cis-trans nepetalactol	7R-trans-cis nepetalactol	7R-cis-cis nepetalactol	7R-trans-trans nepetalactone	7R-cis-trans nepetalactone	7R-trans-cis nepetalactone	7R-cis-cis nepetalactone
S9	8-oxogeranial	LaISY	NsNEPS1B- 151GSSA	+	n.d.	n.d.	n.d.	n.d.	tr	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151AAMS	++	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151ASTA	++	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.
S9	8-oxogeranial	LaISY	NsNEPS1B- 151ASMA	+++	n.d.	n.d.	n.d.	n.d.	+	+++	n.d.	n.d.
S11	8-oxogeranial	CrISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxogeranial	NsISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxogeranial	NsP5βR	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	CrISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	NsISY	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S11	8-oxoneral	NsP5βR	N/A	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	N/A	+++	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NcMLPLA	++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NcMLPLB	++	n.d.	++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NcNEPS2	n.d.	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NcNEPS5	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmNEPSL1	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmNEPSL2	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmNEPS2	n.d.	n.d.	+++	n.d.	n.d.	n.d.	+	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmNEPS5	n.d.	n.d.	n.d.	n.d.	n.d.	+++	++	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmMLPL1	+++	n.d.	+	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmMLPL2	+++	n.d.	+	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NmMLPL3	+++	n.d.	tr.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	NcMLPL4	+	n.d.	+++	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
S13	8-oxogeranial	LaISY	HoNEPSLB	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.
S13	8-oxogeranial	LaISY	HoNEPSLA	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	+++	n.d.	n.d.

Note: The presence of a particular chemical was graded in the following relative scale: not detected (n.d.), traces (tr.) and three levels of detection beyond traces, (+, ++, and +++), where each "+" sign indicates increasing amounts.

Supplementary Table 6: Primers used for generating mutants in this study.

figure number	Gene description	FW primer	RV primer
2	NsNEPS2-Y163F	CGGTCATGTAGACACTGATTTTGTTATGTCC AAACACGC	AATCAGTGTCTACATGACC
2	NcNEPS3A_M2_SG	GTGGTGACGCCACTCACCCGGAACCAGGGG	CGGGTGAGTGGCGTCACCACGGCCATCGGCG
2	V206Q NmNEPS3_M2_SG	ATTICGTCGCCGGCTGATGTACAGAATGTT GCCGTGGCGACGCCGCTCACCCGGAACGTT	ACACGCIGITAA GGTGAGCGGCGTCGCCACGGCCATCGGCGAC
2	Q206V NmNEPS1 154SATA	GGCATTTCGACGCCGGATGATGTACAGAAA ACCACCTCTGCTACAGCAAGCCGTGGCGGG	ACGCTGTTAACCCT CTTGCTGTAGCAGAGGTGGTGCAAATAATGG
3	loop NmNEBS1 154S4T4	CAAAGTATGACCGATTATGCGATGAGC	TACCACG
3	loop + S198L	GCTGGCGCAGCGTATGG	TAATGGCGTCACGCAGCTGAGCACCAC
3	SVTA loop	ACCACCTCTGTTACAGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCTGTAACAGAGGTGGTGCAAATAATGG TACCACG
S 3	NcNEPS3A-Y163F	CATAACGTTACAGATTTTGTAATGTCCAAAC ATGCGGT	TACAAAATCTGTAACGTTATGCG
S 3	NmNEPS4-Y163F	CAAAATATGACTGACTTTGCGATGAGTAAG CACGCC	TCGCAAAGTCAGTCATATTTTGGC
S3	NmNEPS1-Y163F	CAAAGTATGACCGATTTTGCGATGAGCAAA CATGC	TCGCAAAATCGGTCATACTTTGCC
S3	NsNEPS1A-Y163F	AGAGCCTGACCGACTTTGTGATGAGCAAGC ATG	ATCACAAAGTCGGTCAGGCTCTGTC
S3	NsNEPS1B-Y163F	CAAAGCATGACGGACTTTGTGATGTCGAAG CACG	TCACAAAGTCCGTCATGCTTTGCC
S4	NcNEPS3A-V206M	GATCTCCAGTCCAGCTGAT	TCAGCTGGACTGGAGATCCCCATATTGCGAG TAAGAGGAGTTAC
S4	NcNEPS3A-V206E	GATCTCCAGTCCAGCTGAT	TCAGCTGGACTGGAGATCCCTTCATTGCGAGT AAGAGGAGTTAC
S4	NcNEPS3A-V206N	GATCTCCAGTCCAGCTGAT	TCAGCTGGACTGGAGATCCCGTTATTGCGAG TAAGAGGAGTTAC
S4	NcNEPS3A-V206G	GATCTCCAGTCCAGCTGAT	ATCAGCTGGACTGGAGATCCCACCATTGCGA GTAAGAGGAGTTA
S4	NcNEPS3A-V206L	GATCTCCAGTCCAGCTGAT	ATCAGCTGGACTGGAGATCCCTAAATTGCGA GTAAGAGGAGTTA
S4	NcNEPS3A-V206A	GATCTCCAGTCCAGCTGAT	ATCAGCTGGACTGGAGATCCCAGCATTGCGA GTAAGAGGAGTTA
S4	NcNEPS3A-V206I	GATCTCCAGTCCAGCTGAT	ATCAGCTGGACTGGAGATCCCAATATTGCGA GTAAGAGGAGTTA
S5	NcNEPS3A_NmNEPS 1 150-162 loop	ACTACACCCCTGTCGAGCCGTGGTGGGCAAT CTATGACAGATTACGTAATGTCCAAACAT	GCTCGACAGGGGTGTAGTTGTGCAGATGATG CTACCTCCA
S5	NcNEPS3A_NmNEPS 4 150-162 loop	ACGTCGGCCACGGCAAGCAAGGGCGGCCAA	GCTTGCCGTGGCCGACGTGGTGCAGATGATG
S5	NcNEPS3A_NmNEPS 5 150-162 loop	ACAAGCCCGGCAAGCACTATGGGCGGCCAC	AGTGCTTGCCGGGCTTGTCGTGCAGATGATG CTACCTCCA
S6	NmNEPS1_NmNEPS4	ACCACGTCGGCCACGGCAAGCAAGGGCGGC	CTTGCCGTGGCCGACGTGGTGCAAATAATGG
S6	150-162 100p NmNEPS1-S198L	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGGTCAGCACCAC
	NmNEPS1-		
S6	S198L_NmNEPS4 150-162 loop	CAAAACATGACCGATTATGCGATGAGC	TACCACG
S7	NmNEPS1-154SATS	ACCACCTCTGCTACATCTAGCCGTGGCGGGC AAAGTATGACCGATTATGCGATGAGC	CTAGATGTAGCAGAGGTGGTGCAAATAATGG TACCACG
S7	NmNEPS1-154SSTA	ACCACCTCTTCAACAGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCTGTTGAAGAGGTGGTGCAAATAATGG TACCACG
S 7	NmNEPS1-154SALA	ACCACCTCTGCTTTAGCAAGCCGTGGCGGGC AAAGTATGACCGATTATGCGATGAGC	CTTGCTAAAGCAGAGGTGGTGCAAATAATGG TACCACG
S 7	NmNEPS1-154AATA	ACCACCGCAGCTACAGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCTGTAGCTGCGGTGGTGCAAATAATGG TACCACG
S 7	NmNEPS1-154SATG	ACCACCTCTGCTACAGGTAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTACCTGTAGCAGAGGTGGTGCAAATAATGG TACCACG
S7	NmNEPS1-154SGTG	ACCACCTCTGGTACAGGTAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTACCTGTACCAGAGGTGGTGCAAATAATGG TACCACG
S 7	NmNEPS1-154SVTA	ACCACCTCTGTTACAGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCTGTAACAGAGGTGGTGCAAATAATGG TACCACG
S7	NmNEPS1-154TASA	ACCACCACAGCTTCTGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCAGAAGCTGTGGTGGTGCAAATAATGG TACCACG
S7	NmNEPS1-154SAGA	ACCACCTCTGCTGGTGCAAGCCGTGGCGGG CAAAGTATGACCGATTATGCGATGAGC	CTTGCACCAGCAGAGGTGGTGCAAATAATGG TACCACG

figure number	Gene description	FW primer	RV primer						
S 7	NmNEPS1-154SPTA	ACCACCTCTCCAACAGCAAGCCGTGGCGGG	CTTGCTGTTGGAGAGGGGGGGGGGGGAAATAATGG TACCACG						
	NmNEPS1 154SATA								
S8	S198M	CAACACCGGATGATTTTCAT	GCGTCACGCAGTTAA						
S 8	NmNEPS1-154SATA- S198L	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC TAATGGCGTCACGCAGTTAA						
S 8	NmNEPS1-154SATA- S198P	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC AGGTGGCGTCACGCAGTTAA						
S 8	NmNEPS1-154SATA- S198V	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC AACTGGCGTCACGCAGTTAA						
S8	NmNEPS1-154SATA- S198G	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC TCCTGGCGTCACGCAGTTAA						
S 8	NmNEPS1-154SATA- S198A	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC TGCTGGCGTCACGCAGTTAA						
S8	NmNEPS1-154SATA- S198T	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC TGTTGGCGTCACGCAGTTAA						
S8	NmNEPS1-154SATA- S198C	GCTGGCGCAGCGTATGG	CCATACGCTGCGCCAGCGGGGTCAGCACCAC ACATGGCGTCACGCAGTTAA						
S9	NsNEPS1B-151GAMS	ACGACCGGTGCGATGTCGTCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGACGACATCGCACCGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-151SATS	ACGACCAGCGCGACATCGTCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGACGATGTCGCGCTGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-151SATA	ACGACCAGCGCGACAGCTTCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGAAGCTGTCGCGCTGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-S195L	TCACGCCGCTCGCCCAAAGGATGGGGTTTTC CACGCCCGATGATTTCCATACTCATTTTG	CCTTTGGGCGAGCGGCGTGATCACCAATAAC GGCGACACGCAGTTAACCCTAA						
S9	NsNEPS1B-151GSSA	ACGACCGGTTCTTCAGCTTCCAGGGGCGGGC AAAGCATGACGGACTATGTGATGTCGA	GGAAGCTGAAGAACCGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-151AAMS	ACGACCGCTGCGATGTCGTCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGACGACATCGCAGCGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-151ASTA	ACGACCGCGTCTACAGCATCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGATGCTGTAGACGCGGTCGTGCAGATAATA GTGCCTCTC						
S9	NsNEPS1B-151ASMA	ACGACCGCAAGCATGGCTTCCAGGGGCGGG CAAAGCATGACGGACTATGTGATGTCGA	GGAAGCCATGCTTGCGGTCGTGCAGATAATA GTGCCTCTC						

NEPS	NcL1	NmL1	NsL	Nc3A	Nc3B	Nm3	Ns2	Nm2	Nc2	Nm5	Nc5A	Ns4A	Ns4B	Nm4	Nc4	Nc1	Nm1	Ns1A	Ns1B
MpIDPH	73	70	70	66	65	66	72	70	69	64	64	67	66	63	65	63	62	66	65
NcNEPSL1	100	89	78	69	68	69	71	69	68	67	67	70	68	65	66	68	67	72	70
NmNEPSL1		100	77	68	68	68	69	67	66	65	65	68	66	64	64	66	65	69	67
NsNEPSL			100	66	66	67	68	67	66	63	63	67	65	64	63	66	64	68	66
NcNEPS3A				100	92	93	72	69	68	65	64	68	66	64	65	64	64	68	67
NcNEPS3B					100	95	72	68	67	64	62	66	65	63	65	64	64	67	66
NmNEPS3						100	72	69	68	65	63	68	66	64	66	65	65	68	68
NsNEPS2							100	90	89	72	71	75	73	70	72	72	71	75	74
NmNEPS2								100	95	70	68	73	71	69	72	72	71	73	73
NcNEPS2									100	70	68	72	71	68	71	71	71	72	71
NmNEPS5										100	95	80	80	77	79	81	81	80	79
NcNEPS5A											100	78	79	75	77	79	79	80	79
NsNEPS4A												100	96	88	89	87	83	88	86
NsNEPS4B													100	88	89	85	81	84	83
NmNEPS4														100	92	83	80	81	80
NcNEPS4															100	85	82	84	82
NcNEPS1																100	93	89	90
NmNEPS1																	100	86	87
NsNEPS1A																		100	94
NsNEPS1B																			100

Supplementary Figure 1: Pairwise comparison of amino acid identities of NEPS sequences. Abbreviated sequences at the top consist of the same names of each enzyme in the left column excluding the abbreviation "NEPS". Lighter shades of red correspond to lower protein sequence identity.



Supplementary Figure 2: Sequence alignment of all NEPS enzymes from three species: *Nepeta sibirica* (Ns), *Nepeta cataria* (Nc), and *Nepeta mussinii* (Nm). MpIDPH is a relative enzyme from *Mentha piperita* (spearmint). Positions with identical residues are highlighted in red, whereas similar residues are highlighted in yellow. Additional categories related to this study are described in the legend.



Supplementary Figure 3: NEPS catalytic tyrosine mutations to phenylalanine leads to various degrees of disruption of oxidation. NsNEPS2 is shown completely abolishing oxidation to 7*S*-*cis*-*trans* nepetalactone **4a** while maintaining cyclization activity (as can be seen by the disappearance of iridodials). NsNEPS1A has similarly lost oxidation activity and maintained cyclization. On the other hand, NsNEPS1B and NmNEPS1 still are able to oxidize to 7*S*-*cis*-*trans* nepetalactone **4a**, while NcNEPS3A and NmNEPS4 appear inactive for both cyclization and oxidation. Results were repeated twice independently with similar results.



Supplementary Figure 4: Mutagenesis of NcNEPS3A residue V206. A. NcNEPS3A and NmNEPS3 native activities. B. Crystal structure model showing V206 residue (highlighted in green, annotated as V206) and its location relative to NAD+ and a manually docked *7S-cis-cis* nepetalactol **3b** molecule. C. Oxidation activity to **4b** can be changed with various V206 point mutations. Highlighted parts of chromatograms represent the molecular structure highlighted with the same color. Results were repeated twice independently with similar results.



Supplementary Figure 5: Loop 150-162 swap variants generated in NcNEPS3A. A. Amino acid sequence alignment showing the 150-162 loop region to be swapped. B. Crystal structure model showing the 150-162 loop (highlighted in green, annotated as A151, G152, S153, S154, V156, R157, G158, A159, H160, N161, V162) and its location relative to NAD+ and a manually docked 7*S*-*cis*-*cis* nepetalactol **3b** molecule. C. Cyclization and oxidation activities in NcNEPS3A change when the 150-162 loop is replaced with those from NmNEPS1 and NmNEPS4. Results were repeated twice independently with similar results.



Supplementary Figure 6: Engineering 7S-trans-cis **3c** cyclization in NmNEPS1. A. Partial sequence alignment of the active site regions with highlighted residue differences (blue asterisks). B. Crystal structure models of NmNEPS1 and NmNEPS4 based on NsNEPS2 with 7S-trans-cis nepetalactol **3c** manually docked into the active site (light blue), NAD+ highlighted (green, annotated as NAD+) and residues that are different between the two enzymes (violet). C. Enzyme roles in cyclization (NmNEPS4) and oxidation (NmNEPS1) of 7S-trans-cis nepetalactol **3c** and enzymatic assays of NmNEPS1 variants coupled with 8-oxogeranial **1** and CrISY. Complete graft of all 8 active site residue differences from NmNEPS4 into NmNEPS1 shows some 7S-trans-cis nepetalactol **3c** cyclization gained (which spontaneously opens into 7S-trans-cis iridodial **5c**) but oxidation activity was lost. Loop swap of the 152-164 region shows some gained activity for 7S-trans-cis nepetalactone **4c** but remains a minor product. Adding S198L mutation to the loop swap disrupts cyclization of 7S-trans-cis nepetalactol **3c** gained. When only the 154SATA residues are grafted, 7S-trans-cis nepetalactol **3c** production is improved and its subsequent oxidation to 7S-trans-cis nepetalactone **4c** is maintained. Highlighted parts of chromatograms

represent the molecular structure highlighted with the same color. Results were repeated twice independently with similar results.



Supplementary Figure 7: 154SATA region variants generated in NmNEPS1. A. Enzyme roles in cyclization (NmNEPS4) and oxidation (NmNEPS1) of 7S-trans-cis nepetalactol 3c. B. Crystal structure model of NmNEPS1 154SATA variant based on NsNEPS2 with 7S-trans-cis nepetalactol 3c docked into the active site (light blue), NAD+ highlighted (green, annotated as NAD+) and 154SATA residues highlighted (violet). C. Variations in the 154SATA loop region have direct impact in cyclization and oxidation of 7S-trans-cis nepetalactol 3c. S154A mutation does not appear to disrupt cyclization but oxidation to 7S-trans-cis nepetalactone 4c was disrupted. S156T on the other hand, did not change the profile suggesting that a polar group in 156 position is needed for stabilization of 7S-trans-cis nepetalactol **3c** for oxidation. Position 155 changes made did not disrupt cyclization or oxidation but A155S, A155V and A155P had positive impact towards production of 7S-trans-cis nepetalactone 4c. Changing position 156 to non-polar Gly and Leu had a large impact on 7S-trans-cis nepetalactol 3c cyclization, suggesting that this residue is involved in cyclization activity. Finally, although position 157 does not appear to be directly in contact with the substrate (part B) it seems to have an impact on the overall loop stability, given that mutations A157G and A157S do have detrimental impact on 7S-trans-cis nepetalactone 4c production. Highlighted parts of chromatograms represent the molecular structure highlighted with the same color. Results discussed in the text (such as 154SATA, and 154SVTA mutants) were repeated at least twice times independently with similar results.



Supplementary Figure 8: Variations of the residue S198 in the NmNEPS1-154SATA variant enzyme. A. Enzyme roles in cyclization (NmNEPS4) and oxidation (NmNEPS1) of 7*S*-trans-cis nepetalactol **3c**. B. Crystal structure model of NmNEPS1 154SATA variant based on NsNEPS2 with 7S-trans-cis nepetalactol **3c** docked into the active site (light blue), NAD+ highlighted (green, annotated as NAD+) and S198 residue highlighted (yellow). C. Variations of S198 residue of NmNEPS1-154SATA variant have an impact in 7*S*-trans-cis nepetalactone **4c** production. Highlighted parts of chromatograms represent the molecular structure highlighted with the same color. Results discussed in the text (such as S198L mutant) were repeated three times independently with similar results.



Supplementary Figure 9: NsNEPS1B variants tested with stereo-divergent iridoid synthases (ISY). Achiral GC-MS traces showing the impact of various mutations in NsNEPS1B on the product profile for 7*S* (left) and 7*R* (right) isomers. Highlighted parts of chromatograms represent the standard peaks highlighted with the same color. This was an initial screen to engineer *trans,trans* activity. Since all results were negative, the assays were only performed once and were not investigated further.



Supplementary Figure 10: Chiral column chromatograms and mass spectra of nepetalactone standards. These are standards and have been measured more than three times over many months.



Supplementary Figure 11: Influence of 8-oxoneral as a substrate to ISY in the resulting product profile. 8-oxogeranial and 8-oxoneral were assayed side by side with *N. sibirica* ISY and P5 β R, as well as CrISY and LaISY in order to see the impact of the substrate stereochemistry in the product profile. For CrISY, NsISY and NsP5 β R, both substrates result in 7*S cis-trans* nepetalactol **3a** and iridodials (asterisks), indicating no change in profile. In the case of LaISY, while 8-oxogeranial results in 7*R* products, 8-oxoneral leads to both 7*S* and 7*R* products. Highlighted parts of chromatograms represent 7*S* products (light yellow) and 7*R* products (light blue). LaISY and CrISY were each performed twice with independent results.



Supplementary Figure 12: Achiral GC-MS data of *N. sibirica* NEPS and MLPL assayed in combination with 8-oxogeranial **1** and *7R*-specific iridoid synthase from *Lamium album*, LaISY. Highlighted parts of chromatograms represent the molecular structure highlighted with the same color. Results were repeated twice independently with similar results.



Supplementary Figure 13: Chiral GC-MS survey of *N. mussinii*, *N. cataria* and *H. officinalis* selected NEPS and MLPL assayed in combination with 8-oxogeranial and both, 7*S*-specific iridoid synthase CrISY and 7*R*-specific iridoid synthase LaISY. Highlighted parts of chromatograms represent 7*S* products (light yellow) and 7*R* products (light blue). Asterisks indicate iridodials. Results were repeated twice independently with similar results.

Supplementary method: Nepetalactone epimerization and purification

The base 1,8-diaza-bicyclo[5.4.0]undec-7-ene (DBU) was added (126 mg, 0.846 mmol) to a stirred solution of 7S-cis-cis nepetalactone (126 mg, 0.758 mmol) in toluene (6 mL) at room temperature and the resulting mixture was refluxed for 24h. After cooling to room temperature, the reaction was purified by silica gel column chromatography (PE/ethyl acetate = 20/1) to afford 7*S*-trans-trans 4d nepetalactone (9.5 mg, 8%) and recover 7S-cis-cis nepetalactone 4b (100 mg).



DBU (8.9 uL mg, 0.0596 mmol) was added to a stirred solution of 7*R*-trans-trans nepetalactone 4d' (9 mg, 0.0542 mmol) in toluene (1 mL) at room temperature and the resulting mixture was refluxed for 12h. After cooling to room temperature, the reaction was purified by silica gel column chromatography (PE/ethyl acetate = 20/1) to afford 7*R*-*cis*-*cis* nepetalactone **4b**' (4.5 mg, 47%).



NMR

NMR spectra were measured on a 400 MHz Bruker Avance III HD (Bruker Biospin GmbH, Rheinstetten, Germany) (Supplementary Figures 13-17). CDCl₃ was used as solvent. NMR spectra were referenced to the residual solvent signals at $\delta_{\rm H}$ 7.26 ppm and $\delta_{\rm C}$ 77.0 ppm. For spectrometer control and data processing Bruker TopSpin ver. 3.6.1 was used. Mass spectral data for these compounds are also provided in Supplementary Figure 10.

Data of 7S-trans-trans nepetalactone 4d: ¹H NMR (400 MHz, CDCl₃) δ 6.25 (dg, J = 3.1, 1.6 Hz, 1H), 2.58-2.48(m, 1H), 2.30-2.17 (m, 1H), 2.12-2.01 (m, 1H), 1.98-1.83 (m, 2H), 1.69 (t, J = 1.5 Hz, 3H), 1.51-1.36 (m, 2H), 1.19 (d, J = 6.6 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ171.5, 136.3, 120.6, 52.4, 41.8, 32.5, 31.6, 25.6, 20.4, 14.0.

Data of 7*R*-*cis*-*cis* nepetalactone **4b**': ¹H NMR (400 MHz, CDCl₃) δ 6.19-6.16 (m, 1H), 3.10 (t, *J* = 9.6 Hz, 1H), 2.85 - 2.75 (m, 1H), 2.68 - 2.56 (m, 1H), 1.96 - 1.74 (m, 3H), 1.60 (t, J = 1.2 Hz, 3H), 1.39 - 1.28 (m, 1H), 1.00 (d, J = 7.2 Hz, 3H);¹³C NMR (100 MHz, CDCl₃) δ 170.2, 134.3, 115.5, 46.2, 39.4, 38.4, 32.7, 30.4, 17.2, 14.8.

Data of 7S-cis-cis nepetalactone **4b**: ¹H NMR (400 MHz, CDCl₃) δ 6.21–6.14 (m, 1H), 3.10 (t, J = 9.6Hz, 1H), 2.85–2.75 (m, 1H), 2.67–2.56 (m, 1H), 1.93–1.76 (m, 3H), 1.60 (t, J = 1.3 Hz, 3H), 1.37-1.31 (m, 1H), 1.00 (d, J = 7.2 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃) δ 170.2, 134.2, 115.5, 46.2, 39.4, 38.4, 32.7, 30.4, 17.2, 14.8.

Data of 7*R*-*trans*-trans nepetalactone **4d**': ¹H NMR (400 MHz, CDCl₃) δ 6.25 (dq, J = 3.2, 1.6 Hz, 1H), 2.59–2.47 (m, 1H), 2.28-2.19 (m, 1H), 2.12–2.01 (m, 1H), 1.98–1.84 (m, 2H), 1.69 (t, J = 1.6 Hz, 3H), 1.51–1.36 (m, 2H), 1.19 (d, J = 6.6 Hz, 3H);

 ^{13}C NMR (100 MHz, CDCl_3) δ 171.5, 136.3, 120.6, 52.5, 41.8, 32.5, 31.6, 25.7, 20.4, 14.0.



Supplementary Figure 14: Proton (A) and Carbon (B) NMR of 7S-trans-trans nepetalactone 4d.



Supplementary Figure 15: Proton (A) and Carbon (B) NMR of 7*R trans-trans* nepetalactone 4d'.



Supplementary Figure 16: Proton (A) and Carbon (B) NMR of 7R cis-cis nepetalactone 4b'.



Supplementary Figure 17: Proton (A) and Carbon (B) NMR of 7S-cis-cis nepetalactone 4b.



Supplementary Figure 18: Representative SDS-PAGE gel of proteins purified for Figures 2D and S6. The major band at 32 kDa represents the protein of interest. These proteins were expressed in E. coli and analyzed by SDS-PAGE at least twice.