Bark beetles locate fungal symbionts by detecting volatile fungal metabolites of host tree resin monoterpenes

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Abstract

Outbreaks of the Eurasian spruce bark beetle (*Ips typographus*) have decimated millions of hectares of conifer forests in Europe in recent years. The ability of these 3-6 mm long insects to kill mature trees over a short period has been ascribed to two main factors: (1) mass attacks on the host tree to overcome tree defenses and (2) the presence of fungal symbionts that support successful beetle development in the tree. While the role of pheromones in coordinating mass attacks has been well studied, the role of chemical communication in maintaining the fungal symbiosis is poorly understood. We previously demonstrated that *I. typographus* can recognize beneficial fungal symbionts of the genera *Grosmannia*, *Endoconidiophora* and *Ophiostoma* by their *de novo* synthesized volatile compounds. Here, we hypothesize that the fungal symbionts of the bark beetles might metabolize spruce resin monoterpenes of the beetle's host tree, Norway spruce (*Picea abies*), and that the volatile products could be used as cues by beetles for locating breeding sites with beneficial symbionts. We show that *Grosmannia penicillata* and other fungal symbionts altered the profile of spruce bark volatiles by converting the major monoterpenes into an attractive blend of oxygenated derivatives. Bornyl acetate was metabolized to camphor, and α- and β-pinene to *trans*-4-thujanol and other oxygenated products. Other co-occurring non-beneficial fungi (*Trichoderma sp.)* also produce oxygenated monoterpenes, but in non-attractive ratios. Extensive electrophysiological measurements showed that the bark beetle possesses dedicated olfactory sensory neurons for oxygenated metabolites. Compounds such as camphor and *trans*-4-thujanol attracted beetles at specific doses in olfactory experiments, and the presence of symbiotic fungi enhanced attraction of

females to pheromones. Finally, we show that colonization of fungal symbionts on spruce bark diet stimulated beetles to make tunnels into the diet. Collectively, our study suggests that the blends of oxygenated metabolites of conifer monoterpenes produced by fungal symbionts are used by bark beetles as attractive cues to find breeding or feeding sites containing their essential microbial symbionts. The oxygenated metabolites may aid beetles in assessing the presence of the fungus, the defense status of the host tree and the density of conspecifics at potential feeding and breeding sites.

Introduction

Many interactions between insects and their host plants are known to be mediated by volatile organic compounds [1–4]. In contrast, volatile signals between herbivorous insects and their symbiotic microbes have been less studied, aside from a few well-known examples including ambrosia beetles, termites, and the vinegar fly *Drosophila melanogaster* [1–4]. Yet, such signals could be as critical for insect fitness as their response to host plant cues. In some insect-microbe symbioses, microbes transform host plant metabolites creating volatile signals that are used by insects for food or brood site selection [3–6]. For example, yeasts vectored by *D. melanogaster* metabolize dietary phenolic antioxidants and release volatile phenolics that attract both larvae and adults to feed on antioxidant-rich foods [7]. Nevertheless, there is still little information about how microbial transformation of host plant chemicals influences insect-microbe symbioses, and whether the resulting metabolites represent honest signals of partner benefits.

Microbial symbioses are especially characteristic of wood-boring insects such as bark and ambrosia beetles. Bark beetles have captured much attention recently because of their large-scale outbreaks in many parts of the world. In Europe, for example, the Eurasian spruce bark beetle (*Ips typographus*) has killed millions of hectares of spruce stands as a result of major population bursts, likely in response to global warming and management practices, which increase forest vulnerability to epidemic outbreaks [8–12]. This bark beetle feeds and raises broods in the phloem tissue of trees, which contains high levels of terpene and phenolic defense chemicals [8,13]. This insect

overcomes its unfavorable environment by mass attacks and by introducing a suite of microbes into the host tree, including ectosymbiotic ophiostomatoid fungi, primarily *Grosmannia penicillata*, *Leptographium europhioides*, *Endoconidiophora polonica* and *Ophiostoma bicolor* that cause blue staining of infected wood [14–18]. It is not yet well understood how these free-living fungal symbionts benefit *I. typographus*. Though they grow outside the insect, as necrotrophic fungi they may exhaust host tree defenses, metabolize host defense compounds, and provide nutritional benefits to larvae and adults [19–22].

Conifer oleoresins are a formidable defense against insects and pathogens, as they can poison and physically entrap invaders [23–26]. However, the volatile fraction of the resin, especially the monoterpenes, also plays a central role in the colonization of host trees by bark beetles [24,27,28]. After locating a suitable tree, pioneer male *I. typographus* oxidize the dominant host monoterpene α-pinene to *cis*-verbenol, which is used as an aggregation pheromone in combination with the *de novo* produced 2-methyl-3-buten-2-ol to attract conspecifics for a mass attack [29–31]. In addition to bark beetle pheromones, several other oxygenated monoterpenes such as terpinen-4-ol, camphor, *trans*-4-thujanol and borneol have also been detected at the entrance holes of *I. typographus* galleries [32–35]. Interestingly, the phloem colonized by ophiostomatoid fungi around these galleries also produces large amounts of oxygenated monoterpenes compared to galleries without evident fungal growth [33]. However, the ecological functions of these oxygenated monoterpenes remain poorly understood. In our previous work, we showed that *I. typographus* utilizes *de novo* synthesized fungal volatiles to maintain its association with specific beneficial symbionts and also to avoid saprophytes

[36]. However, it is unknown which volatiles are produced by these fungi when they colonize their native substrate i.e., the phloem and sapwood of the tree.

In this study, we investigated the volatile compounds emitted when fungal symbionts of *I. typographus* infect the bark of their Norway spruce (*Picea abies*) host trees. We show that these fungi dramatically alter the volatile monoterpene composition of spruce bark and demonstrate, using single sensillum recordings, that adult *I. typographus* can perceive the fungal-produced monoterpenes and are attracted to these compounds in behavioral bioassays. Our results indicate that bark beetles respond to symbiont biotransformation products of host tree metabolites and employ them to identify suitable sites for feeding and breeding.

Results

Ips typographus is attracted to volatiles from fungal symbionts on a spruce bark medium

Adult *I. typographus* beetles were strongly attracted to volatiles emitted by their symbiotic fungus *G. penicillata* grown on both potato dextrose agar (PDA) or spruce bark agar (SBA) compared to their respective fungus-free agar controls in laboratory trap bioassays (Fig 1) (PDA, $z = 3.34$, $p = 0.001$; SBA, $z = 2.83$, $p = 0.005$, Wilcoxon's test). Further, bark beetles showed a much stronger attraction towards *G. penicillata* grown on SBA over the same fungus grown on PDA (Fig 1D) ($z = 4.28$, $p < 0.001$, Wilcoxon's test). Volatiles from several other bark beetle primary and secondary fungal symbionts, such as *E. polonica* and *L. europhioides*, grown on SBA were also highly

attractive to adult beetles, although not all bark beetle-associated fungi, nor a non-beneficial saprophyte (*Trichoderma sp.*), tested in this way emitted attractive volatile blends (S1 Fig).

Symbiotic fungi alter the volatile profile of the bark

The headspace volatiles of *P. abies* bark inoculated with *I. typographus* fungal symbionts were distinct from those of fungus-free bark 4 d after inoculation when analyzed using gas chromatography with flame ionization detection (GC-FID) and gas chromatography-mass spectrometry (GC-MS) (S2-S7 Tables). For *G. penicillata*-inoculated bark, PCA analysis revealed that nearly 68% of the variation in the volatile profiles was explained by the first two principal components (Fig 2A).

Over a time course of 4, 8, 12 and 18 d post inoculation with *G. penicillata* on detached bark plugs (Fig 2B, S3 Fig, S2, S3, and S5 Tables) 79 compounds comprising host tree and fungal volatiles were detected and classified into different groups, namely aliphatic hydrocarbons (17 compounds), aromatics (2 compounds), monoterpene hydrocarbons (15 compounds), oxygenated monoterpenes) (26 compounds), sesquiterpenes (17 compounds) and spiroketals (3 compounds). The proportion of total oxygenated monoterpenes gradually increased to dominate the volatile profile of *G. penicillata*-infected bark reaching a maximum at 18 d post inoculation ($F_{(3,14)} = 3.54$, $p = 0.04$, ANOVA), while the proportion of total oxygenated monoterpenes was unchanged in mock-inoculated controls (Fig 2B). Camphor was the major contributor to the overall increase of oxygenated monoterpenes, with the highest relative abundance after 18 d and a significant difference between time points ($F_{(1,16)} = 13.06$, $p = 0.002$, ANOVA,

Tukey's test) (S5 Table). The proportion of monoterpene hydrocarbons gradually decreased over time in both treatments (control, $F_{(3,7)}$ = 11.6 , p = 0.004; *G. penicillata*, $F_{(3,15)}$ = 21.2, p < 0.001, ANOVA). The proportion of total sesquiterpenes also decreased significantly over time in *G. penicillata* infected bark ($F_{(3,15)}$ = 4.4, p = 0.02, ANOVA), but not in mock-inoculated control bark. When measuring emission rate (ng mg DW^{-1} h^{-1}), the emission of total monoterpene hydrocarbons in the control and the *G. penicillata* treated bark plugs was not significantly different at 4 d post inoculation (Fig 2C, S2 Table). However, there was a dramatic increase in the emission rate of total oxygenated monoterpenes at this time point in spruce bark inoculated with *G*. *penicillata* compared to the mock-inoculated control (Fig 2D) (9-fold increase, t(3.2) = 7.38, p = 0.004, Welch's t-test). Nineteen oxygenated monoterpenes were identified in the bark inoculated with *G. penicillata*, and a total of 15 compounds significantly increased in fungus infected bark compared to the control including camphor (Fig 2E) (S2 Table) (51-fold increase, t(3) = 7.7, p = 0.004, Welch's t-test), *endo*-borneol (18-fold increase, t(6) = 6.5, p < 0.001), isopinocamphone (3-fold increase, t(6) = 6.8, p < 0.001), verbenone (2-fold increase, t(6) = 4.3, p = 0.005) and bornyl acetate (3-fold increase, t(6) = 3.7, p = 0.01). Measurements conducted on four other *I. typographus* fungal symbionts also showed differences in the volatile composition of fungus-inoculated versus control bark (S2 Fig), with increases in the proportion of oxygenated monoterpenes in bark inoculated with *L. europhioides* and *O. bicolor* over time, but not for *E. polonica* (S3 Fig, S4, S6 and S7 Tables).

Symbiotic fungi produce oxygenated monoterpenes from spruce monoterpene hydrocarbons and bornyl acetate

The symbiotic fungus, *G. penicillata*, significantly decreased the amount of (-)-bornyl acetate added to PDA medium after 4 d compared to a fungus-free control (Fig 3A) (t = -3.38, p = 0.003). However, the amounts of two other monoterpenes, (-)-β-pinene and (-)-α-pinene, did not differ between *G. penicillata*-inoculated and fungus-free PDA.

The monoterpene metabolites of *G. penicillata* grown on either (-)-α-pinene- or (-)-β-pinene-enriched agar were similar with the oxygenated monoterpene terpinen-4-ol being the major product from both spruce precursors (Fig. 3B). Other oxygenated monoterpenes formed by *G. penicillata* from (-)-α-pinene and (-)-β-pinene include (+)-isopinocamphone, (+)-*trans*-4-thujanol and α-terpineol, with (-)-verbenone being detected only from (-)-α-pinene (S5, S6 Figs). From (-)-bornyl acetate, the dominant oxygenated monoterpenes of *G. penicillata* were camphor and *endo*-borneol (Fig 3B, bottom panel; S8 Fig), and their production coincided with the decrease of the precursor (Fig 3A). Similar results were obtained for the three other fungal symbionts tested (S4-S8 Figs).

The volatile blend of the co-occurring, but a non-beneficial fungus, *Trichoderma* sp., produced from a synthetic spruce monoterpene mixture (S8 Table) was distinct from that of *G. penicillata* (S9 Fig). *Trichoderma* sp. produced mainly the oxygenated monoterpenes, (-)-borneol and (-)-verbenone, while *G. penicillata* produced mainly (-)-camphor and terpinen-4-ol. These results collectively show that symbiotic fungi can dramatically alter the volatile profile of spruce bark by increasing the emission of oxygenated monoterpenes, and the emission profile of symbiotic fungi is distinct from that of a co-occurring fungal saprophyte.

Elimination of symbiotic ophiostomatoid fungi from *I. typographus* reduces the production of oxygenated monoterpenes in bark beetle galleries

To determine the primary source of oxygenated monoterpenes in beetle galleries, unsterilized pupae were reared into fungus-free mature adults using a diet fortified with the fungistatic agent sodium benzoate. Sterility tests and scanning electron microscopy of beetle body parts revealed that beetles reared on this diet were free from fungi and the elytral pits were devoid of fungal spores and yeasts (Fig. 4A-iii, S9 Table), though bacteria were abundant. In comparison, elytral pits of beetles reared on diet without sodium benzoate contained abundant spore masses, with structural similarity to *O. bicolor* and yeast (Fig. 4A-ii, S9 Table) (Solheim 1996). Re-inoculation of fungus-free beetles with *G. penicillata* resulted in acquisition of *G. penicillata* spores in the elytral pits (Fig. 4A-iv).

Next, we quantified the amount of oxygenated monoterpenes present in the bark galleries constructed by untreated, fungus-free, and *G. penicillata*-reinoculated beetles separately using GC-MS analyses of gallery extracts. The concentrations of (-)-camphor and terpinen-4-ol were significantly higher in the galleries of reinoculated beetles than fungus-free beetles or in bark samples without beetles at all ((-)-camphor: $F\ (3,23) = 15.6$, $p < 0.001$, ANOVA; terpinen-4-ol: $H\ (3) = 18.6$, $p < 0.001$, Kruskal-Wallis) (Fig. 4B). On the other hand, the concentrations of these two oxygenated monoterpenes were similar when comparing galleries of *G. penicillata*-reinoculated beetles to those of untreated beetles. For (+)-isopinocamphone, the concentration in galleries constructed

by untreated beetles was significantly higher compared to bark samples without beetle galleries, but similar to that in galleries constructed by fungus-free and *G. penicillata*-reinoculated beetles ($F_{(3,23)}$ = 5.7, p = 0.005, ANOVA) (Fig. 4B). On the other hand, the concentration of (-)-borneol in fungus-free galleries was significantly higher compared to that in bark samples without galleries and was similar to that in the galleries of *G. penicillata*-reinoculated and untreated beetles ($F_{(3,23)}$ = 5.3, p = 0.006, ANOVA) (Fig. 4B). There were no significant differences in the concentrations of pinocamphone, α-terpineol and (-)-verbenone among treatments.

Analysis of the microbes present in each treatment revealed that galleries constructed by fungus-free beetles were indeed free from symbiotic ophiostomatoid fungi and yeasts but contained many saprophytes and (or) endophytes and bacteria (S10 Table). Microbes present in the galleries of untreated and *G. penicillata*-reinoculated beetles were similar to those found on beetles that were used to infest the bark (S9-10 Table).

Ips typographus detects oxygenated monoterpenes through specialized olfactory sensory neurons (OSN) in their antennae

To test if *I. typographus* antennal olfactory sensilla contain OSNs that detect the biotransformation products of monoterpenes, we challenged 231 olfactory sensilla with a test panel comprising 92 ecologically relevant compounds diluted in paraffin oil (1 µg μl^{-1}) using single cell recordings (S1 Table). Only 23 (~10%) of the sensilla housed neurons that did not respond to any of the compounds from the odor panel although their OSNs showed spontaneous firing. We obtained odor-evoked responses with strong excitation (>120 Hz) from 198 OSNs and weak excitation (<50 Hz) from 10

241 OSNs, allowing the grouping of these neurons into different classes based on their
242 response profile. From initial screening experiments at a 10 µg dose on filter paper (to
243 determine the maximum receptive range of OSNs), we identified 20 classes of OSNs.
244 Three OSN classes responded primarily to fungal-produced oxygenated monoterpenes.
245 We also identified neurons belonging to previously described OSN classes tuned to
246 pheromones, host tree volatiles and non-host odorants [37] that are not further
247 considered here.

248 OSN classes tuned to fungal-produced oxygenated monoterpenes were
249 identified in both the A_m and B_m regions on the antennae (Fig 5A). One of these OSN
250 classes responded most strongly to (+)-isopinocamphone, and this class was highly
251 specific for oxygenated monoterpenes, especially ketones (Fig 5C, left panel). Apart
252 from (+)-isopinocamphone, relatively strong responses were also elicited by (+)-
253 pinocamphone, (-)-isopinocamphone, (±)-pinocarvone, (±)-camphor, and (-)-
254 pinocamphone. (Fig 5C*,* left panel). Dose-response tests showed that this OSN class
255 was the most sensitive to (+)-isopinocamphone of all the compounds tested with
256 responses evident at a dose of 100 pg. The responses to (+)-pinocamphone, (-)-
257 isopinocamphone, (±)-pinocarvone and (±)-camphor all appeared between 1 ng and 10
258 ng doses (Fig 5D, left panel). Another OSN class with specific responses to fungal-
259 derived compounds responded most strongly to (+)-*trans*-4-thujanol and weakly to (±)-3-
260 octanol, (±)-1-octen-3-ol, (+)- and (-)-terpinen-4-ol, and (+)- and (-)-α-terpineol (Fig 5C,
261 middle panel). This OSN showed a 1000-fold lower response threshold to (+)-*trans*-4-
262 thujanol compared to the next best ligands, the C_8 alcohols (Fig 5D*,* middle panel).
263 Finally, an OSN class responding strongly to verbenone, α-isophorone and β-

264 isophorone, followed by weaker responses to (-)- and (+)-*trans*-verbenol, pinocarvone,
265 and (-)-*cis*-verbenol (Fig 5C, right panel) was also found. Dose- response tests revealed
266 that this neuron class responded the strongest to α-isophorone across most tested
267 doses, followed by slightly weaker and similarly strong responses to both verbenone
268 and β-isophorone (Fig 5D, right panel).

269 A few previously characterized OSN classes for host tree monoterpenes,
270 including the classes with primary responses to α-pinene, *p*-cymene, and Δ3-carene,
271 respectively [37], showed varying secondary responses to some of the fungal-derived
272 compounds tested here for the first time. For example, the α-pinene OSN class
273 responded also to (+)-isopinocamphone, (-)-isopinocamphone and (±)-pinocarvone and
274 weakly to (±)-camphor, (-)-myrtenol, *trans*-pinocarveol, carvone, borneol and (-)-
275 fenchone (S10A Fig). The *p*-cymene OSN class showed intermediate responses to (+)-
276 *trans*-4-thujanol and carvone (S10B Fig). Although the Δ3-carene OSN class showed
277 high specificity towards Δ3-carene, ligands such as camphor and (-)-isopinocamphone
278 also elicited weak responses from this neuron class (S10C Fig).

279 Oxygenated monoterpenes produced by fungal symbionts attract *I.* 280 *typographus*

281 *Grosmannia penicillata* growing on spruce bark agar with added 0.1 mg g^{-1} or 0.5 mg g^{-1}
282 (-)-β-pinene (within the range of natural concentrations in *P. abies* bark) was much more
283 attractive to adult *I. typographus* than a fungus-free control medium supplemented with
284 (-)-β-pinene (Fig 6A, left) (0.1 mg g^{-1}, $z = 2.22$, $p = 0.02$; 0.5 mg g^{-1}, $z = 2.54$, $p = 0.01$,
285 Wilcoxon's test). However, addition of 1 mg g^{-1} (-)-β-pinene completely abolished the

attraction to *G*. *penicillata* and adult beetles were unresponsive in the binary choice between fungus and fungus-free (-)-β-pinene-containing medium (Fig 6A, left). To further understand these concentration-dependent responses, individual electrophysiologically-active (-)-β-pinene biotransformation products were used in trap bioassays against a mineral oil control. At the highest dose tested (1 mg), both *trans*-4-thujanol and terpinen-4-ol were avoided by adult beetles (Fig 6B *trans*-4-thujanol, left panel; terpinene-4-ol, right panel) (1 mg *trans*-4-thujanol, z = -1.9, p = 0.05; 1 mg terpinen-4-ol, z = -1.7, p = 0.08, Wilcoxon's test). On the other hand, 100 µg *trans*-4-thujanol significantly attracted adult beetles (Fig 6B, left panel) (z = 2.78, p = 0.005, Wilcoxon's test). Adult beetles did not discriminate between *G*. *penicillata* grown on (-)-β-pinene enriched medium and *G*. *penicillata* grown on non-enriched medium (Fig 6E). Based on these results, we concluded that *I. typographus* shows concentration-specific responses to some fungal metabolites of (-)-β-pinene, but not to (-)-β-pinene itself.

Addition of another host tree monoterpene, (-)-bornyl acetate, to fungal growth medium at 0.05 mg g^{-1} (in the range of natural concentrations in *P. abies* bark) and 0.5 mg g^{-1} resulted in strong attraction of *I. typographus* adults towards *G*. *penicillata* when tested against a fungus-free control after 4 d incubation (Fig 6C, left panel) (0.05 mg g^{-1}, z = 3.31, p = 0.001; 0.5 mg g^{-1}, z = 3.21, p = 0.001, Wilcoxon's test). The major biotransformation product of (-)-bornyl acetate that formed in this period (Fig 6C), camphor, was significantly more attractive to adult beetles at a 100 µg dose than the mineral oil control (Fig 6D) (z = 2.58, p = 0.01, Wilcoxon's test). However, adult beetles preferred *G*. *penicillata* grown on unenriched medium against *G. penicillata* grown on high amount of (-)-bornyl acetate (0.5 mg g^{-1}) enriched medium (Fig 6F) (z = 2.12, p =

0.03, Wilcoxon's test) consistent with their preference for lower amounts of camphor. By contrast, in the absence of fungus beetles did not discriminate between diet enriched with monoterpenes and diet without monoterpenes (S11 Fig).

Volatiles of symbiotic fungi increase *I. typographus* attraction to pheromones

Ips typographus has been shown to display sex-specific responses to its aggregation pheromone [38,39], so the sexes were tested separately with pheromones and volatiles of their symbiotic fungi. Female beetles were significantly more attracted towards the individual aggregation pheromone components *cis*-verbenol and 2-methyl-3-buten-2-ol (*cis*-verbenol, $z = 2.98$, $p = 0.003$; 2-methyl-3-buten-2-ol, $z = 2$, $p = 0.046$, Wilcoxon's test), and towards a mixture of the two pheromone components ($z = 5.19$, $p < 0.001$, Wilcoxon's test) compared to the mineral oil control (Fig 7A). By contrast, adult males did not make a significant choice between these options in accordance with previous studies [42, 61]. However, when beetles had to choose between the pheromone mixture with or without *G. penicillata* volatiles, females significantly preferred the pheromone mixture together with *G*. *penicillata* volatiles (Fig 7B) ($z = 3.41$, $p = 0.001$, Wilcoxon's test).

Symbiotic fungi increase the tunneling of adult *I. typographus*

The presence of a symbiotic fungus increased the tunneling success of adult *I. typographus* beetles in medium after 48 h. Multiple logistic regression analysis revealed that successful tunneling odds in (-)-bornyl acetate-amended medium were significantly

influenced by the fungus when the beetle sex and the monoterpene treatment remained constant (Table 1) (β = 4.98, χ^2 = 20.99, p < 0.001). The presence of the fungus increased the tunneling odds by 145 units (percentage probability increased by 99%) compared to in the absence of the fungus. Additionally, males had significantly lower tunneling success compared to females (Table 1) (β = -1.72, χ^2 = 8.78, p = 0.003) with the tunneling odds for males being 0.18 units lower (percentage probability decreased by 15.2%) than for females. In (-)-β-pinene-amended medium, only the presence of the fungus significantly influenced the tunneling odds of beetles (Table 1) (β = 2.65, χ^2 = 20.45, p < 0.001), with an increase of 14.06 units (percentage probability increased by 93%) compared to in the absence of the fungus. Similarly, in (-)-α-pinene-amended medium only the presence of the fungus significantly influenced the tunneling odds of beetles (Table 1) (β = 2.02, χ^2 = 9.39, p = 0.002), with an increase of 7.51 units (percentage probability increased by 88%) compared to the absence of the fungus. Addition of the three monoterpenes without fungus did not have any effect on the tunneling behavior of adult beetles.

The growth of *G. penicillata* on monoterpene-enriched media resulted in significantly longer beetle tunnels than in the other treatments (Fig 8B-D) ((-)-β-pinene, $F_{(3,99)}$ = 4.95, p = 0.003; (-)-α-pinene, $F_{(3,92)}$ = 14.8, p < 0.001; (-)-bornyl acetate, $F_{(3,106)}$ = 6.6, p < 0.001, ANOVA, Tukey's test). However, there was no significant difference in tunnel lengths in treatments with fungus alone versus fungus plus monoterpenes except for the treatment with (-)-β-pinene (Fig 8B-D, Tukey's test). The sex of the beetle and the interaction of sex with other treatments had no effect on the tunnel length.

Discussion

The successful attack of bark beetles on their host trees is invariably associated with free-living fungal symbionts. These ascomycete symbionts may detoxify the terpene-rich defensive resin of the host tree, hasten host tree death, provide nutritional benefits, and increase resistance to pathogens, [16,17,22]. Here, we document the ability of *I. typographus* fungal symbionts to metabolize host tree monoterpenes to oxygenated derivatives that may assist adult beetles in locating suitable breeding and feeding sites [34,40]. Several oxygenated monoterpenes have been previously identified as volatiles released from trees that were attacked by *I. typographus* [32,33,35]. We showed, by removing symbiotic fungi from *I. typographus*, that these compounds were likely fungal metabolites that become dominant components of the volatile profile bark within 12 days following infection. Bark beetle olfactory sensory neurons can detect various oxygenated monoterpenes produced by symbiotic fungi, and these compounds attract adult beetles and stimulate tunneling on diets inoculated with fungal symbionts.

Oxygenated monoterpenes are widespread volatile cues in tree-feeding insects

Various forest insects that are associated with fungal symbionts, such as bark beetles, ambrosia beetles and wood wasps live in host trees producing large quantities of monoterpene volatiles. These insects are often attracted to their fungal symbionts through volatiles [2,36,41] and hence, fungus-produced monoterpene metabolites could be critical components of the attractive volatile blends. For bark beetles, oxygenated

374 monoterpenes derived from either the beetles, fungi or host trees are already known to
375 play important roles in their life history [42]. During mass attacks, the host-derived
376 oxygenated monoterpene *cis*-verbenol acts together with 2-methyl-3-buten-2-ol as an
377 aggregation pheromone for *I. typographus* to promote mass attack on individual trees
378 [28,43]. Beetles also utilize oxygenated monoterpenes to restrict the density of attack.
379 Microbes lining the gallery walls or living in the beetle gut oxidize *cis*- and *trans*-
380 verbenol into verbenone, which inhibits the attraction of both sexes to fully colonized
381 trees [44–46]. In addition, mated male *I. typographus* produce ipsenol and ipsdienol, of
382 which ipsenol acts as an anti-attractant [46]. Furthermore, an oxygenated monoterpene
383 from host trees, 1,8-cineole, which is produced in higher amounts in resistant or MeJA-
384 primed trees, inhibits attraction of beetles to their pheromones [47–49]. Oxygenated
385 monoterpenes are also used as reliable cues by parasitoids of bark beetles to locate
386 their prey [35,50].

387 Here we discovered that the oxygenated monoterpene blend emitted by bark beetle-
388 associated fungi growing on agar amended with spruce bark is distinct from the blend of
389 spruce monoterpene metabolites emitted by beetles alone [51] (S12 Fig) and attracted
390 adult *I. typographus*. Previously, we demonstrated that fungi grown on potato dextrose
391 agar without any spruce bark produced mixtures of non-terpene aliphatic and aromatic
392 volatiles that attract newly emerged (callow) adult beetles [36]. These compounds were
393 also detected here as major components of the volatile blend at later phases of fungal
394 growth (S3-7 Table). Although we focused principally on the symbiont *G. penicillata* in
395 this study, volatiles from other fungal symbionts were also investigated. *Leptographium*
396 *europhioides* and *E. polonica* also emitted volatile blends attractive to adult *I.*

typographus when growing on spruce bark agar (Fig. S1), but the volatiles of *O. bicolor* and saprophytes such as *O. piceae* and *Trichoderma sp.* were not attractive. The production of oxygenated monoterpenes by other co-inhabiting microbes including fungal saprophytes, yeasts and bacteria has been reported before, but these microorganisms produce different metabolites or the same metabolites in very different ratios than produced by symbiotic fungi [52–54]. Since all these species produce oxygenated derivatives of spruce bark monoterpenes (Tables S2-7) (S9 Fig), it is not the simple presence of oxygenated monoterpenes, but the composition of the entire volatile profile that determines its attractiveness. Notably, cross-attraction of *I. typographus* to fungi associated with the North American spruce bark beetle *D. rufipennis* attacking a different species of spruce has been reported recently. This attraction could be attributed to similar oxygenated monoterpene profiles as reported here, but it is unclear if an association with these fungi would be beneficial to *I. typographus* [55].

Oxygenated monoterpenes signal the presence of fungi to *I. typographus* and may modulate beetle colonization

The first chemical signals reported to mediate bark beetle colonization of their hosts were aggregation pheromones. Yet even in the presence of these pheromones, a large proportion of aggregating beetles that land on trees leave without tunneling into the bark [40,56,57]. This suggests that other cues may be needed to induce beetles to stay and bore into the bark. Indeed, bark beetles have been shown to respond to signals from host and non-host species when selecting trees for colonization [28,36,58–61]. Based

on our results, fungus-produced oxygenated monoterpenes might also belong to the list of colonization cues. For bark beetles, fungal metabolites can serve as indicators of host tree sectors where their fungal symbionts are already established. These compounds also provide evidence for the ongoing metabolism of host tree defenses, which could improve the success of bark beetle colonization.

Fungal volatiles also enhance the attraction of *I. typographus* to aggregation pheromones. Female *I. typographus* is known to use the aggregation pheromone (2-methyl-3-buten-2-ol and *cis*-verbenol) to locate trees suitable for mating and oviposition [40,62]. Here we showed that female *I. typographus* were more attracted at short range to a combination of pheromone plus fungal volatiles than to pheromone alone. Similarly, a recent study reported that the presence of fungal volatiles increased the attraction of dispersing beetles to their pheromones in the field compared to traps with pheromones only [63]. Oxygenated monoterpenes and other fungal volatiles provide information about the presence of fungal symbionts, which promote the successful development of their offspring. Similarly, the pheromone component *cis*-verbenol, itself an oxygenated monoterpene produced by *I. typographus* from the host tree precursor α-pinene [64], provides information about the presence of other beetles, especially males. The lack of response of males to pheromones in our experiments is not unexpected, as male *I. typographus* have been reported to be less responsive than females to high doses of pheromones in walking bioassays [39,62]. This behavior may help them avoid dense colonies of male conspecifics within a tree to reduce competition for mates and food. Depending on how fast symbiotic fungi colonize the tree bark, fungal cues could benefit both epidemic and endemic beetle populations. During the endemic phase, beetles

442 generally colonize stressed trees in a gradual fashion over months [56] and in this case,
443 volatiles emitted from fungal symbionts introduced into the tree by early colonizers, in
444 combination with beetle pheromones, could increase the probability for conspecifics to
445 locate attacked trees, present at low densities in spruce stands. In contrast, during
446 epidemics vigorous trees are typically colonized by large numbers of bark beetles within
447 a few days to overwhelm their defenses. Since ophiostomatoid fungi have been
448 reported to emit volatiles within one day following the infection of detached bark plugs in
449 the laboratory [65,66] these chemicals might function in attraction during natural attacks
450 upon epidemics as well. However, during rapid natural attacks it is unclear if fungi can
451 colonize a tree within few days and emit behaviorally active volatiles.

452 The oxygenated metabolites of host monoterpenes produced by fungal symbionts not
453 only attracted bark beetles, but also stimulated them to tunnel in a fungus-colonized
454 medium. Interestingly, both sexes showed increased tunneling in contrast to their
455 response to pheromones. The lack of sex-specific differences in tunneling could be due
456 to the fact that the nutritional advantage of feeding on fungus-colonized spruce bark is
457 beneficial to both sexes [17,18].

458 The proportion of oxygenated monoterpenes to total monoterpenes in the volatile blend
459 of *G. penicillata* increased over the time course studied to nearly 50% at 12 days and
460 nearly 70% at 18 days, a trend also observed for *L. europhioides* and *O. bicolor*. Thus,
461 higher proportions of oxygenated monoterpenes may indicate older fungal infection
462 sites and hence older beetle invasion sites that may be less attractive to newly arriving
463 beetles due to crowding. The lack of attraction and even repellency of higher
464 concentrations of individual oxygenated monoterpenes at higher concentrations, as

seen in laboratory bioassays in our own and in previous studies [67] is consistent with this interpretation. Among the oxygenated monoterpenes, already reported to inhibit *I. typographus* attraction, verbenone produced by microbial oxidation or auto-oxidation of the pheromone *cis*-verbenol, reduces aggregation of *I. typographus* in later phases of the attack on the tree [46,54,68]. The attractiveness or repellency of individual oxygenated monoterpenes to *I. typographus* depends on their structures as well as the stage of beetle attack cycle.

Oxygenated monoterpenes are signals not only for bark beetles, but also for their enemies. Both beetle predators and parasitoids employ these compounds and other volatiles to locate bark beetle larvae hidden under the bark [69]. Specifically, a three-component blend comprising camphor, isopinocamphone and terpinen-4-ol, all fungal metabolites of host tree monoterpene hydrocarbons, was reported to attract a coleopteran predator and several hymenopteran parasitoids of *I. typographus* in the presence of host tree background signals [50,70,71]. A similar mechanism is used by parasitoids of the wood wasp, *Sirex noctilio*, which locate their concealed host insect under the bark via the volatiles from the wood wasp fungal symbiont *Amylostereum areolatum* [72]. Furthermore, the bark beetle predator, *Thanasimus formicarius* possesses OSNs to detect oxygenated monoterpenes such as camphor and pinocamphone as well as several additional OSN classes detecting various oxygenated bark beetle pheromone compounds [73]. Thus, any benefit to the beetle arising from oxygenated monoterpene production by its symbiotic fungi may come at the cost of revealing its presence to natural enemies that employ these same volatiles to locate bark beetles.

Formation of oxygenated derivatives may reduce monoterpene toxicity for *I. typographus*

The conversion of host tree monoterpene defenses by symbiotic fungi to oxygenated products may alleviate toxicity to bark beetles. Terpene-rich resins are a general defense of *P. abies* and other conifers against herbivores and pathogens [74–76]. Thus, it is not surprising that monoterpenes have exhibited toxicity to bark beetles in many studies [77–79]. Monoterpene hydrocarbons, such as α-pinene, are typically more toxic to beetles than host tree-produced oxygenated monoterpenes, such as bornyl acetate [80]. Hence the oxidative transformations carried out by fungal symbionts described in this study could reduce toxicity to *I. typographus* through conversion to less poisonous derivatives. Such detoxification of host tree defenses could represent a significant benefit of fungal symbionts [81].

By contrast, oxygenated monoterpenes may be more toxic to fungi compared to monoterpene hydrocarbons [82,83]. Thus, the initial oxidation of monoterpene hydrocarbons may not constitute a detoxification unless it is a step towards further metabolism. The potential toxicity of oxygenated monoterpenes may explain why these substances are degraded further after initial oxidation by fungi specialized on conifers such as *G. penicillata*, *Heterobasidion parviporum* and *Seridium cardinale* [82,83]. The fungus *G. clavigera*, a symbiont of the bark beetle *Dendroctonus ponderosae,* possesses genes encoding cytochromes P450 and other oxidative enzymes that are up-regulated by dietary monoterpenes [84,85]. Another class of terpenes present in *P. abies* bark and other conifers host trees of bark beetles are diterpene resin acids, which

are reported to be toxic to associated fungi in another bark beetle system [86]. Whether bark beetles, their associated fungi or other microbes can detoxify diterpenes is not yet known.

Other sources of oxygenated monoterpenes in spruce-bark beetle interactions

Oxygenated monoterpenes emitted from trees attacked by *I. typographus* may arise from sources other than fungal symbionts. The host tree *P. abies* itself synthesizes large amounts of bornyl acetate [87] and small amounts of 1,8-cineole [88]. In these compounds, the oxygen functions are incorporated during biosynthesis from basic precursors, whereas the products from fungal symbionts are formed by oxidative modification of a previously formed monoterpene hydrocarbon skeleton. The compound *trans*-4-thujanol belongs to the latter group. We identified it as a *G. penicillata* metabolite of α- and β-pinene, but *trans*-4-thujanol may also be synthesized by the tree, although at low levels in *P. abies* bark [67]. As another alternative, this and other oxygenated monoterpenes could be produced via autoxidation of monoterpene hydrocarbons. The oxidation of monoterpenes upon exposure to air could explain the release of low but readily detectible amounts of these compounds from uninfected control bark plugs in our and other studies. In the field, oxygenated monoterpenes other than bornyl acetate and 1,8-cineole have been detected from damaged *P. abies* trees when monoterpenes were exposed to air [89,90]. However, in the present study the emission rate from uninfected bark plugs was much lower than from fungus-infected plugs, suggesting that microbial metabolism is a much more significant source of

oxygenated monoterpenes than autoxidation (S9 Fig) [91]. Nevertheless, the tree itself cannot be ruled out as a source of any of the detected oxygenated monoterpenes, since *P. abies* cell suspension cultures have been reported to oxidize added monoterpenes [91,92]. If *P. abies* produced monoterpenes as a defense response only when infected by fungi, it might be difficult to determine which organism was producing these compounds- the tree or the fungus- without identifying specific biosynthetic genes in one of the two genomes. However, our comparison of the volatiles produced by fungi infecting *P. abies* bark showed large differences among fungal species (Figs. S2, S8, S9). These seem more likely to occur if the compounds are of fungal origin. If the compounds were produced by the tree, one might expect a very similar response to each of these fungi, all ascomycetes.

Among microbial sources of oxygenated monoterpenes are several yeast species including *Hansenula holstii, H. capsulata* and *Candida nitratophila*, which were isolated from *I. typographus,* and produce terpinen-4-ol, α-terpineol, borneol and *trans*-pinocarveol when grown in phloem medium or in α-pinene-supplemented medium [54]. In addition, several bacterial symbionts of bark beetles are capable of metabolizing monoterpenes into oxygenated derivatives, such as verbenols and verbenone [53]. Some bacteria co-inhabit the beetle gallery together with ophiostomatoid fungi, but it is not known if these bacteria induce the production of oxygenated monoterpenes, either by producing them themselves or by interacting with fungi to form these compounds [92] . Another bark beetle species, *Polygraphus poligraphus*, which is sometimes found together with *I. typographus*, was shown to emit large amounts of terpinen-4-ol [93,94]. Intermediate amounts of α-terpineol, *cis*- and *trans*-4-thujanol were also identified from

555 the hindgut as well as the entrance holes of this beetle's gallery and could be formed by
556 microorganisms associated with this beetle species from host tree monoterpenes, or by
557 the beetle itself. Our experiments with fungus-free *I. typographus*, however,
558 demonstrated that live beetles themselves did not produce high concentrations of any of
559 the fungal biotransformation products identified in this study. Instead, beetles formed a
560 range of other oxidation products (Fig. S12) that have been identified in previous
561 studies [51,95].

562 High selectivity of *I. typographus* olfactory neurons to oxygenated
563 monoterpenes suggest their role in detecting symbiotic fungi

564 *Ips typographus* possesses several classes of OSNs that were shown to detect the
565 oxygenated monoterpenes produced by their fungal symbionts with notable specificity.
566 For example, the isopinocamphone OSN class showed high specificity towards several
567 monoterpene ketones produced by fungal symbionts, including (+)- and (-)-
568 isopinocamphone, (+)- and (-)-pinocamphone, camphor, and pinocarvone, but not to
569 monoterpene alcohols such as borneol and *trans*-pinocarveol. The absence of any
570 response to monoterpene hydrocarbons indicates that this OSN is not tuned to detect
571 the host tree itself, but rather organisms metabolizing the major host monoterpenes.
572 The isopinocamphone OSN is similar to one recently reported OSN class from *I.*
573 *typographus* that responded best to pinocarvone and camphor (OSN class named Pcn;
574 isopinocamphone and pinocamphone were not tested) [90]. Our work shows that (+)-
575 isopinocamphone is the primary ligand of this OSN class, based on its greater activity
576 than the other active compounds. In addition, the response profile of this OSN class

577 matches very well with that of the odorant receptor (OR) ItypOR29, which recently was
578 characterized in *Xenopus laevis* oocytes [96].

579 Likewise, we showed that a previously described verbenone-sensitive OSN class [37]
580 also responds to *cis*- and *trans*-verbenol as well as α- and β-isophorone, compounds
581 that are believed to arise from bark beetle metabolism of host tree terpenes [51].
582 Verbenone is produced from the verbenols by microbes that colonize gallery walls and
583 beetle guts. Therefore, this OSN class appears to be tuned to signals from various
584 ecological sources providing information on bark beetle density as well as microbial
585 establishment [40,45,89]. Since, as discussed above, the individual oxygenated
586 monoterpenes arise from different organisms, including fungi, other microbes, the host
587 tree and the beetles themselves, the presence of OSNs responding to different
588 oxygenated monoterpenes may help the beetle identify the various life forms it
589 encounters.

590 Another OSN class responded most sensitively to the monoterpene alcohol *trans*-4-
591 thujanol, a fungal symbiont metabolite of α- and β-pinene. This OSN also responded to
592 the fungal metabolites terpinen-4-ol and α-terpineol, as well as C_8 alcohols, but only at
593 the highest doses tested, which extends prior results for this OSN [90] to other
594 compounds from our greatly expanded test odor panel. Strong electroantennographic
595 activity in *I. typographus* in response to these oxygenated monoterpenes has also been
596 reported [89,90], and the response spectrum of this OSN class matches well with that of
597 the receptor ItypOR23, which is evolutionarily related to the receptor (ItypOR29) that
598 detects isopinocamphone [96]. The specific responses of *I. typographus* bark beetles to
599 oxygenated monoterpenes may arise not only from the selectivity of the OSN classes,

but also due to further processing of the odor signal in the higher centers of the brain [97]. Moreover, since some of the oxygenated monoterpenes elicited secondary responses from other OSN classes, bark beetles like other insects may process olfactory information through combinatorial codes [98], which could lead to very specific responses to different compounds or mixtures.

Conclusion

We have shown that fungal ectosymbionts vectored by *I. typographus* metabolize host tree monoterpene hydrocarbons to oxygenated derivatives as they infect the bark. These oxygenated volatiles are detected by several classes of *I. typographus* olfactory sensory neurons and lead to beetle attraction to fungi and stimulation of tunneling at certain concentrations, but often to repulsion at higher concentrations. These compounds appear to signal the presence of established fungi, but their effects on beetles are context-dependent and could vary with the physiological status of the fungi, the age, sex and density of beetles and the stage of attack [36,66]. Moreover, the ecological roles we have proposed for these oxygenated monoterpenes are based on laboratory assays with walking beetles, so studies under natural conditions are necessary to confirm our findings. These compounds may also be useful in integrated pest management strategies as attractants or repellents of bark beetles perhaps in combination with pheromones [99–101]. In this way, oxygenated monoterpenes and other microbial volatiles represent a rich source of untapped insect semiochemicals that can be exploited for protecting forests from devastating pest species such as *I. typographus*.

1 Materials and methods

1.1 Fungal strains and growth medium

The fungal strains used in this study have been previously described [36] (listed in *Table S1*). In order to obtain spores from fungi, freshly inoculated PDA plates were incubated at room temperature for 15-20 d until the mycelium was old and dark. After 20 d, plates were kept briefly at 4˚C to induce sporulation. Four to six 1 cm diameter mycelium plugs were removed from each plate and inoculated into 20 mL potato dextrose broth and incubated at 25˚C at 150 rpm for 4 days. Once the broth was turbid, the spores were filtered using a 40 µm EASYstrainer™ (Greiner Bio-One, Frickenhausen, Germany), and the filtrate was spun down at 4200 rcf for 10 min to precipitate the spores. The supernatant was discarded and the spore suspension was washed three times with autoclaved water and then stored at 4˚C until used. The spore suspension prepared using this method was viable for several months when stored at 4˚C.

1.2 Bark beetle rearing

Bark beetles were reared and stored in the laboratory as described [36]. The starting beetle culture was obtained from an infested tree in October 2017 near Jena, Thuringia, Germany. Beetles were reared throughout the year in the laboratory in freshly cut spruce logs (ca. 30 cm diameter x 50 cm height) placed in an environmental chamber set at 25°C throughout the day, 65% relative humidity and a photoperiod of 20 h per day. Beetles emerged from breeding logs after ca. 35 days and were collected

643 manually. Emerged adults were sexed based on the bristle density on their pronotum
644 [102] and stored separately in Falcon tubes lined with moist paper at 4°C at least for a
645 week before using them in bioassays. Adult beetles were used only once in bioassays.

646 1.3 Spruce bark diet

647 The outer bark of a freshly cut mature tree was scraped off gently using a drawing knife
648 and the inner bark (phloem) was carefully peeled off using a chisel. The bark was cut
649 into small pieces and ground to a fine powder in vibratory micro mill (Pulverisette 0,
650 Fritsch GmbH, Idar-Oberstein, Germany). The instrument was pre-cooled with liquid
651 nitrogen and bark pieces were pulverized at an amplitude of 2.0 for ca. 10 minutes with
652 addition of liquid nitrogen every two minutes to prevent thawing. The ground powder
653 was stored in Falcon tubes at -80°C until used for diet preparation. For preparing spruce
654 bark diet, 7% powdered inner spruce phloem (w/v) was added to 4% Bactoagar (Roth)
655 and heat sterilized at 121°C for 20 minutes.

656 1.4 Identification and quantification of headspace volatiles of fungal
657 symbionts

658 Norway spruce bark plugs of approximately 28 mm diameter were removed from a
659 freshly felled tree in July, 2017 and a single bark plug was placed inside a 250 mL
660 volatile collection glass bottle. Before removing the bark plugs, the surface of the bark
661 and the cork borer were sterilized by thorough spraying with 70% ethanol in a laminar
662 hood. An 100 µL quantity of spore suspension ($1*10^6$ cells mL^{-1}), prepared as described
663 above, was added to the exposed section of the bark, and autoclaved water was added

to the control treatment. Each treatment was replicated four times including the control. The glass bottle was secured tightly and incubated at 25˚C for 4 days. After 4 d, activated charcoal-filtered air was passed into the bottle inlet at the rate of 50 mL min^{-1} and the outlet air was funneled through a SuperQ adsorbent filter (150 mg) for 4 hours. Afterwards, the filters were eluted with 200 µL dichloromethane spiked with 10 ng $µL^{-1}$ nonyl acetate (Sigma Aldrich) as an internal standard and stored at -20˚C. The spruce bark plugs were oven dried at 80˚C for 6 hours after the experiment and the dry weight was measured.

The eluted volatile samples were subjected to GC-MS and GC-FID analysis using an Agilent 6890 series GC (Agilent, Santa Clara, CA, USA) (injection, 1 µl splitless; flow, 2 ml min^{-1}; temperature, 45 to 180°C at 6°C min^{-1} and then to 300°C at 100°C min^{-1} for 10 min) coupled either to an Agilent 5973 quadrupole mass selective detector (interface temperature 270 °C, quadrupole temperature 150°C, source temperature 230 °C; electron energy 70 eV) or a flame ionization detector (FID, temp. 300 °C). The constituents were separated on a DB-5MS column (Agilent (30 m x 0.25 mm x 0.25 µm)), with He (MS) or H_2 (FID) as carrier gas. Peaks arising from the solvent and collection containers were identified by blank runs and excluded from the analysis. The identity of each peak was determined by comparing its mass spectra and retention times to those of reference libraries (NIST98 and Wiley275) and authentic standards. The amount of each compound was calculated from the peak area obtained from the FID detector relative to the internal standard and standardized to the spruce bark dry weight.

1.5 Time series headspace volatile collection

For time series volatile analysis, spruce bark plugs (10 mm diameter) were removed using a cork borer from a freshly felled spruce tree in October, 2016. Each spruce bark plug was placed in a 15 mL clear glass vial (Supelco-Sigma-Aldrich) and 50 µL spore suspension ($1*10^6$ cells mL^{-1}), prepared as described above, was added to treatment plugs while control plugs received sterile water. The headspace volatiles were captured on three polydimethylsiloxane (PDMS) sorbent silicone tubes (0.5 cm), which were hung in each glass vial using a manually crafted metal hook attached to the bottom of PTFE/silicone septa in the screw cap [103]. The headspace volatiles were collected from each treatment for 2 h at 4, 8, 12 and 18 d after inoculation. After sampling, silicone tubes were placed in 1.5 mL brown glass vials and stored at -20°C until analysis.

Volatiles collected on PDMS tubes were analyzed using a GC-2010 plus gas chromatograph coupled to a MS-QP2010 quadrupole mass spectrometer equipped with a TD-20 thermal desorption unit (Shimadzu, Japan) and a GC Cryo-Trap filled with Tenax. A single tube was placed in a 89 mm glass thermal desorption tube and desorbed at a flow rate of 60 mL min^{-1} for 8 min at 200 °C under a stream of N_2 gas. The desorbed substances were focused on a cryogenic trap at -20 °C. The Tenax adsorbent was heated to 230 °C and the analytes were injected using split mode (1:100) onto a Rtx-5MS GC column (30 m x 0.25 mm x 0.25 µm) with helium as carrier gas. Compounds were identified as above (1.5) from authentic standards and libraries, and quantified from the area of each peak obtained using GC-MS post run analysis software from Shimadzu. Peaks arising from contaminants from the solvent, medium or bark plug container were identified by blank runs and excluded from the

710 analysis. The PLS-DA plot in Fig 2A was generated by using MetaboAnalyst 3.0
711 software with normalized GC-MS data (both log transformed, and range scaled) [104].

712 1.6 Biotransformation of host tree compounds

713 Experiments were conducted in 9 cm Petri dishes containing 2% potato dextrose agar
714 (PDA) supplemented with test solutions. The tested compounds were (-)-α-pinene, (+)-
715 α-pinene, (-)-β-pinene, myrcene, γ-terpinene, terpinolene, sabinene, camphene, *p*-
716 cymene, and (-)-bornyl acetate. Sources of these compounds are given in Table S1.
717 Compounds were added after dissolving in dimethyl sulfoxide (DMSO). These were
718 then added into PDA to reach a final concentration of 0.5 mg mL^{-1} before pouring into
719 Petri dishes. A 5 mm agar plug containing a fungal colony was placed in the center of
720 each dish and incubated at 25˚C in darkness for 6 d. Each treatment was replicated four
721 times and for the control the PDA contained only DMSO plus monoterpene. The
722 headspace volatiles were collected after 4d using three PDMS tubes, which were
723 mounted on sterile metal wires and imbedded in PDA for one hour and stored at -20˚C.
724 The identification and quantification of compounds were conducted in the same way as
725 reported for the time series (section 1.5). The headspace volatiles from fungus grown
726 on PDA enriched with myrcene, γ-terpinene, terpinolene, camphene and *p*-cymene did
727 not yield detectable amounts of monoterpene transformation products on analysis.

728 To identify if symbiotic fungi can reduce the amount of monoterpenes in their substrate,
729 fungi were grown on PDA enriched with 0.5 mg g^{-1} (-)-α-pinene, (+)-α-pinene, (-)-β-
730 pinene, and (-)-bornyl acetate as described above. Control plates contained only DMSO
731 and the tested monoterpene. After 4 d, three plugs of 6 mm diameter were removed,

732 weighed and transferred to 1.5 ml sterile glass vials. Agar plugs were homogenized
733 using sterile plastic pestles and 1 ml hexane (extraction solvent) spiked with 10 ng μL^{-1}
734 nonyl acetate was added and samples were vortexed for 30 s. Supernatants were
735 transferred to new vials and stored at -20°C until identification and quantification by GC-
736 MS and GC-FID (1.4). Data analysis was identical to that reported above.

737 ## 1.7 Chemical synthesis of (+)-isopinocamphone and β-isophorone

738 **(+)-isopinocamphone** ((1*R*,2*R*,5*S*)-2,6,6-Trimethylbicyclo[3.1.1]heptan-3-one) was
739 synthesized as described in [105,106]. A mixture of (1*R*,2*R*,3*R*,5*S*)-(-)-isopinocampheol
740 (200 mg, 1.30 mmol, Sigma-Aldrich) and Dess-Martin-periodinane (825 mg, 1.95 mmol)
741 in anhydrous CH_2Cl_2 (15 mL) was stirred at room temperature for 1 hour, followed by
742 the addition of water and sat. aq. $NaHCO_3$ solution. The mixture was extracted twice
743 with methyl *t*-butyl ether. The organic phase was washed with brine, dried over
744 anhydrous Na_2SO_4 and concentrated *in vacuo*. The residue was purified by short-path
745 chromatography using an SPE cartridge (Chromabond SiOH, 6 mL, 500 mg, Macherey-
746 Nagel, *n*-hexane:EtOAc = 10:1) to yield (+)-isopinocamphone (159 mg, 1.04 mmol,
747 80%). NMR measurements were carried out on a Bruker Avance AV-500HD
748 spectrometer, equipped with a TCI cryoprobe using standard pulse sequences as
749 implemented in Bruker Topspin ver. 3.6.1. (Bruker Biospin GmbH, Rheinstetten,
750 Germany). Chemical shifts were referenced to the residual solvent signals of acetone-d_6
751 (δ_H 2.05/ δ_C 29.84) or $CDCl_3$ (δ_H 7.26/ δ_C 77.16), respectively.

752 ^{1}H-NMR (500 MHz, $CDCl_3$) δ ppm: 2.64 (*ddd*, *J*=18.6/3.0/3.0 Hz, 1H), 2.62 (*dddd,*
753 *J*=10.1/6.2/6.1/3.0 Hz, 1H), 2.52 (*bd*, *J*=18.6 Hz, 1H), 2.46 (*dq*, *J*=7.3/1.9 Hz, 1H), 2.12

(*ddd*, *J*=9.1/6.1/3.0 Hz, 1H), 2.06 (*ddd*, *J*=6.2/6.2/1.9 Hz, 1H), 1.31 (*s*, 3H), 1.21 (*d*, *J*=7.3 Hz, 3H), 1.19 (*bd*, *J*=10.1 Hz, 1H), 0.88 (*s*, 3H). ^{13}C-NMR (125 MHz, $CDCl_3$) δ ppm: 215.9, 51.7, 45.3, 45.1, 39.4, 39.1, 34.8, 27.4, 22.2, 17.3. GC-MS t_R: 13.6 min. EI-MS (70 eV): *m/z* (%) 152 (14), 110 (15), 95 (44), 83 (89), 69 (97), 55 (100), 41(64) (S13 Fig).

β-isophorone (3,5,5-Trimethyl-3-cyclohexen-1-one). β-Isophorone was synthesized from α-isophorone (Acros Organics, Fair Lawn, NJ, USA) following published methods [107]. Since β-isophorone was very unstable during column chromatography [108], the compound was used for the bioassay without purification. The purity of β-Isophorone was 91% with 6% of α-isophorone as assessed by NMR analysis. ^{1}H-NMR (500 MHz, acetone-d_6) δ ppm: 5.44 (*s*, 1H), 2.69 (*bs*, 2H), 2.27 (*s*, 2H), 1.70 (*bs*, 3H), 1.00 (*s*, 6H). ^{13}C-NMR (125 MHz, acetone-d_6) δ ppm: 209.3, 133.0, 130.6, 53.6, 44.2, 36.9, 29.8, 22.8. GC-MS t_R: 10.3 min. EI-MS (70 eV): *m/z* (%) 138 (69), 123 (68), 96 (99), 95 (84), 81 (100) (S14 Fig).

Purity of the synthesized compounds was also determined using the following GC-MS program: injection, 1 µl splitless; flow, 2 ml min^{-1}; temperature, 45 °C (held for 2 min) to 250 °C with 6 °C min^{-1}.

1.8 Electrophysiology

Laboratory reared adult beetles from the same German culture that were used in bioassays were used for electrophysiological single sensillum recordings (SSR) using tungsten microelectrodes according to established methodology [36,37], using the SSR set-up (Syntech, Kirchzarten, Germany) and odor delivery system previously described

[109]. The odor panel comprising 92 compounds consisted of beetle pheromones, host tree, non-host tree and fungal compounds ([36], Table S2). Both major and minor fungal volatiles identified during the chemical analysis were included in the odor panel. All odors were dissolved in odorless paraffin oil (w/v). SSR traces were analyzed as described [100] using Autospike 3.0 (Syntech). Males and females were initially screened for responses to the odor panel using a high stimulus dose (10 µg on filter paper placed inside capped standard Pasteur pipette odor cartridges [109]. OSN classes shown to primarily respond to fungus-derived oxygenated monoterpenes were subsequently studied in dose-response experiments with active stimuli diluted in ten-fold steps and tested from lowest to highest dose with the least active ligands tested first at each dose. To reduce variation due to odor depletion, stimulus cartridges were used for a maximum of 8 stimulations during screening and 2 stimulations during dose-response tests [110].

1.9 Trap bioassay

The arena [36] was designed so that adult beetles had to make their choice through olfaction and not by contact cues. Fungi were inoculated on spruce bark agar-based diet and incubated at 25˚C for 4 days. With the help of a cork borer (10 mm diameter), bark plugs with or without fungus were inserted into circular cups (1.8 cm height * 1.8 cm diameter) facing each other. Two beetles were placed inside each arena and the olfactometer was placed inside a laminar flow cabinet in darkness. Each experiment was replicated at least 25 times with 2 beetles per replicate. The choice of beetles was determined periodically for up to six hours by counting the number of beetles trapped inside the cups and represented as percentage choice (percentage of insects

responding to either control traps or treatment traps or no response). When two spruce bark agar control plugs were tested simultaneously, adult beetles showed no preference for traps on one side of the arena versus the other side. Preliminary experiments showed that the sex of the beetle did not influence the olfactory response towards fungus grown either alone or in the diet enriched with monoterpenes. Therefore, two beetles were randomly chosen for trap bioassays.

For bioassays using terpenes, stock solutions were prepared by dissolving the compounds in DMSO, which were then added to 7% spruce bark agar to a final concentration of 0.05 to 1 mg g^{-1}. To determine the response of adult beetles to (-)-β-pinene and (-)-bornyl acetate amended diet containing the fungus, *G. penicillata* was used as this species emitted higher amounts of biotransformation products compared to other fungi. Controls were treated with DMSO plus monoterpene (no fungus) or DMSO plus *G. penicillata* (no monoterpene). 7% spruce bark agar plugs (10 mm) supplemented with monoterpenes or plugs containing *G. penicillata* were placed in the control cups, and *G. penicillata* colonized plugs from monoterpene-enriched medium were placed in the treatment cups. The volatile emission from each control and treatment plug used in the bioassays was determined using PDMS tubes as adsorbents and analyzed as described previously (section 1.5). For bioassays with synthetic compounds, stock solutions of authentic standards were prepared by dissolving them in mineral oil (w/v) and further diluted in $\log_{10}$ steps by dissolving in mineral oil. 10 µL was applied to 10 mm Whatmann filter paper laid on the top of spruce bark agar plugs placed inside the cups. Control traps were treated with 10 µL paraffin oil. For the experiment with pheromone blend in the presence of *G. penicillata* volatiles, *G.*

penicillata colonized spruce bark plugs were placed in treatment cups and 10 µL of a pheromone mixture (*cis*-verbenol:2-methyl-3-buten-2-ol in the ratio of 1:50 diluted 1:100 in paraffin oil) was applied to filter paper as described above. Control cups were treated with 10 µL of the pheromone mixture. Males and females were tested separately in this experiment as *I. typographus* bark beetles show sex-specific responses to their pheromones.

1.10 Tunneling behavior bioassay

Ips typographus tunneling behavior was studied using a protocol modified from [86] We tested beetles in 35 ×10 mm Petri dishes (Greiner Bio-one, Frickenhausen, Germany) filled with ca. 3 ml of spruce bark diet. The spruce bark diet was prepared as before with some modifications: 7% (w/v) spruce inner bark powder was mixed with 1% fibrous cellulose (Sigma), 2% glucose (Roth), and 4% Bactoagar (Roth) in water and autoclaved for 20 minutes at 121 ºC. Before pouring the medium into the Petri dishes, the medium was mixed with 2% solvent (DMSO: ethanol, 1:1) with 1 mg g^{-1} of various monoterpenes ((-)-α-pinene, (-)-β-pinene and (-)-bornyl acetate) and solvent only as a control. For treatment with fungus, 5 µl spore suspension of *G. penicillata* (1 x 10^6 cells mL^{-1}) was added to the center of Petri dishes containing monoterpene-enriched media or solvent controls and incubated at 25°C for 4 days. A single beetle was introduced per plate, and the plates were sealed with Parafilm and kept in the environmental chamber for 48 h under conditions described above (section 1.2). The beetles were monitored for their tunneling activity after 2, 4, 6, 24 and 48 h with tunneling recorded as a binary event. If beetles were inside the media, it was noted as 1 and if outside, noted as 0. After 48 h, tunnel lengths made by

beetles in each plate were measured using Image J software. Each treatment was replicated with 15 male and female beetles.

1.11 Data analysis

IBM SPSS Statistics V25.0 was used to analyze the volatile differences between treatments (*E. polonica*-, *G. penicillata*-, *L. europhioides*-, *O. bicolor*-, and *O. piceae* treated bark samples and untreated control). Data were log-transformed to meet the assumptions of normal distribution, as needed. All individual compounds assigned to monoterpenes (MTs) or oxygenated monoterpenes were combined and their concentrations (in dry weight (ng h^{-1} mg^{-1})) were subjected to a t-test or Welch's for estimating differences between control and *G. penicillata* (Fig. 2). Additionally, separate ANOVAs for all individual compounds in each group were also performed (Table S2). For volatile time course samples, a separate ANOVA test was performed for all individual compounds and compound groups from each fungus with time intervals as an independent factor (Table S3-7). All ANOVA tests were followed by Tukey's *post-hoc* tests to test for differences among treatment combinations. For behavioral bioassays, the CI values from each experimental group were analyzed by Wilcoxon's singed ranked test to compare the differences between control and treatment samples. Binary data from bark beetle tunneling assays were subjected to multiple logistic regression to analyze independent variables such as monoterpene, sex, and fungus that influence the tunneling activity of beetles (dependent variable) in the medium. During data analysis, the male was coded as 1 and female as 0, the presence of fungus coded as 1 and absence of fungus as 0, tunneling inside the medium coded as 1 and not tunneling or staying outside the medium as 0. After

testing all possible independent variables and their interactions among them, the following best-fitted logistic regression model was created to predict the odds of beetles tunneling in the different media.

$$\text{Ln [odds] (tunneling odds)} = \beta 0 + \beta 1 * \text{compound} + \beta 2 * \text{sex} + \beta 3 * \text{fungus}$$

Here, β0 is constant whereas β1, β2, and β3 are logistic coefficients or estimates for the parameters for compound, sex and fungus respectively. The strength of association between beetle tunneling odds and effect of monoterpenes or sex or fungus is expressed as odds ratios ($OR=exp^{\beta \cdot}$) where OR<1 indicates a negative relationship between the two events, i.e., the tunneling event is less likely to happen in response to a selected independent variable (coded as 1) in comparison with its base group (coded as 0), OR=1 indicates no relationship between two events, OR>1 shows positive relationship between two events.

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Competing Interests

I have read the journal's policy and I declare that the authors of this manuscript have no competing interests.

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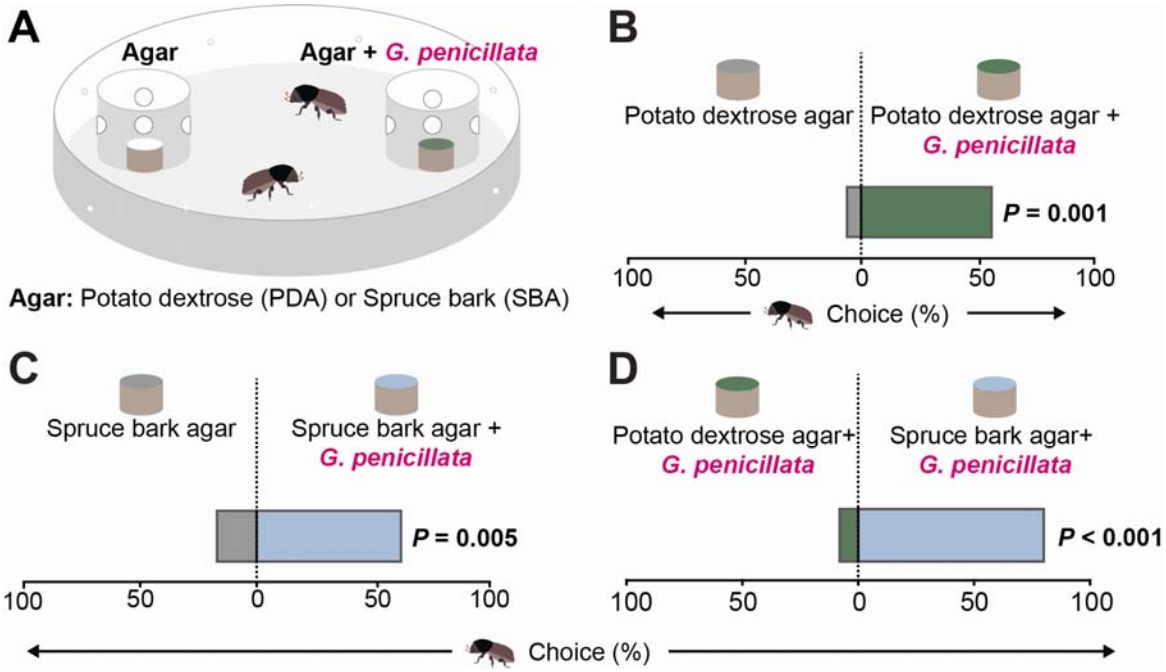

Figure 1: ***Ips typographus* uses volatile cues from spruce bark inoculated with *G. penicillata (Gp)* to detect this symbiotic fungus.**

(A) Arena used for trap bioassays to study the behavior of *I. typographus* to volatiles emitted by a symbiotic fungus. Cups containing agar (with and without fungus) were offered to beetles as odor samples. Holes on each side of the cup allowed the beetles to smell, but not touch the agar until they entered the cup and then could no longer escape. **(B), (C)** Adult beetles chose *Gp*-colonized agar medium over fungus-free medium (n = 25, with two beetles per replicate). **(D)** Adult beetles chose *Gp*-inoculated spruce bark agar over *Gp*-inoculated agar without spruce bark (n = 25). **(B), (C), (D)** Deviation of response indices against zero was tested using Wilcoxon's test. Asterisks denotes significant differences, *P < 0.01, ***P < 0.001.

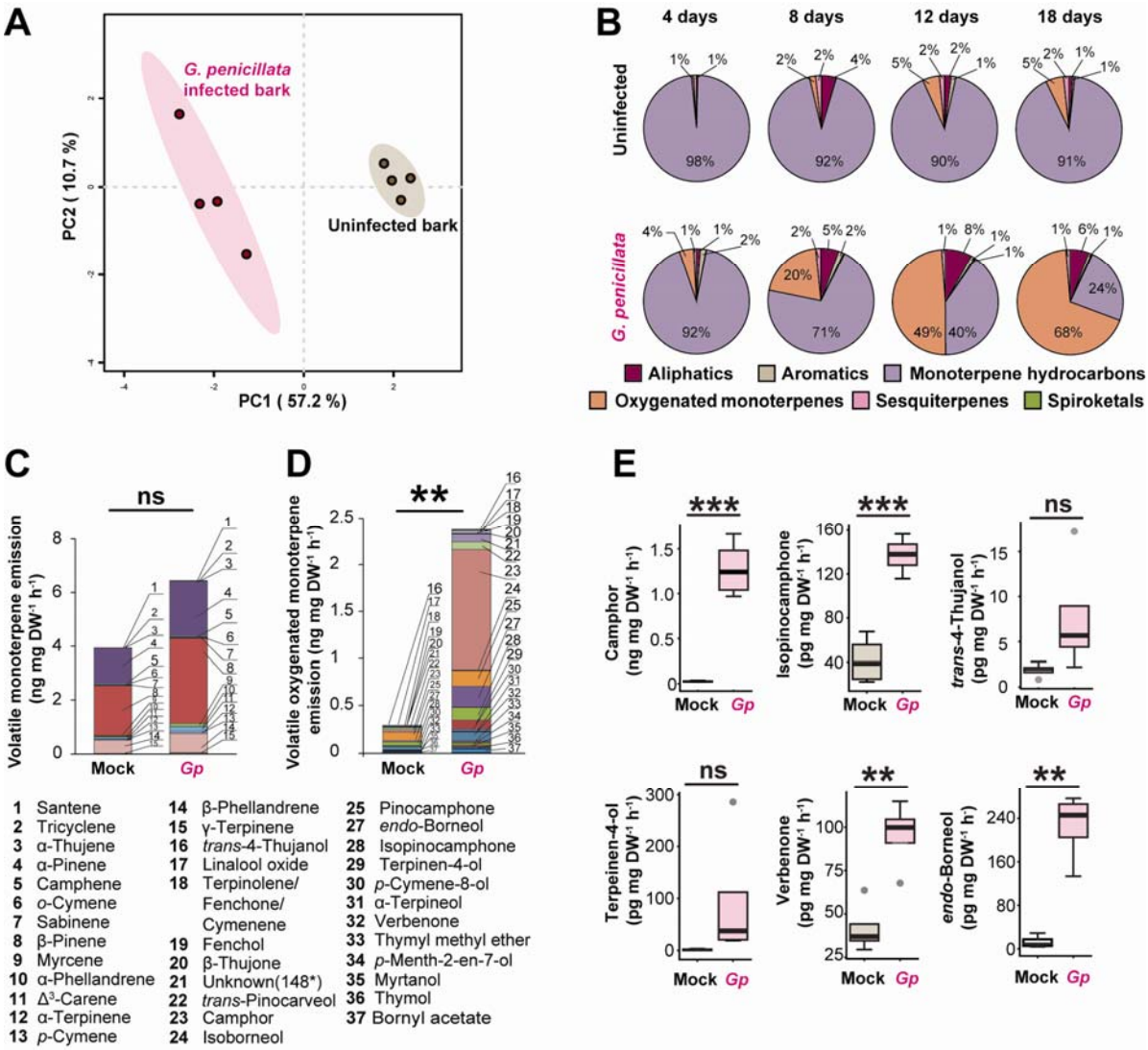

Figure 2: **Growth of *Ips typographus* symbiotic fungi on spruce bark induces increased emission of oxygenated monoterpenes.**

(A) Volatile emission pattern differed between spruce bark inoculated with *G. penicillata* (*Gp*) and uninfected bark 4 days after inoculation, as depicted in a partial least squares discriminant analysis (PLS-DA). Principal components (PC1 and PC2) explain 57.2% and 10.7% of total variation, respectively, and ellipses denote 95% confidence intervals around each treatment. Complete volatile emission data by compound for *G. penicillata* and other *I. typographus* fungal symbionts are given in Table S2. **(B)** Changes in volatile emission profiles of spruce bark due to *G. penicillata* infection over an 18 d time course. Compounds are classified into six groups by their chemical structures (n = 5). Complete volatile emission data by compound and time point for *G. penicillata* and other symbionts are given in Tables S3-S6. (**C**), (**D**) Emission of specific monoterpenes from fresh spruce bark inoculated with *G. penicillata* at four days post inoculation. Identified compounds were classified into monoterpene hydrocarbons **(C)** and oxygenated monoterpenes **(D).** The individual compounds are stacked within a single bar representing the total emission. Significant differences in the total emission levels induced by *G. penicillata* are denoted by asterisks

above the bars (*n* = 4, Welch's t-test with ns= not significant **P* < 0.05, ***P* < 0.01, ****P* < 0.001). The numbers denote the identities of the compounds in the stacked bars. Complete volatile emission data are given in Table S2**. (E)** Emission rate of major oxygenated monoterpenes from spruce bark inoculated with *G. penicillata* four days post inoculation. Asterisks indicate significant differences between the spruce bark-inoculated *G. penicillata* and the fungus-free control (Welch's t-test).

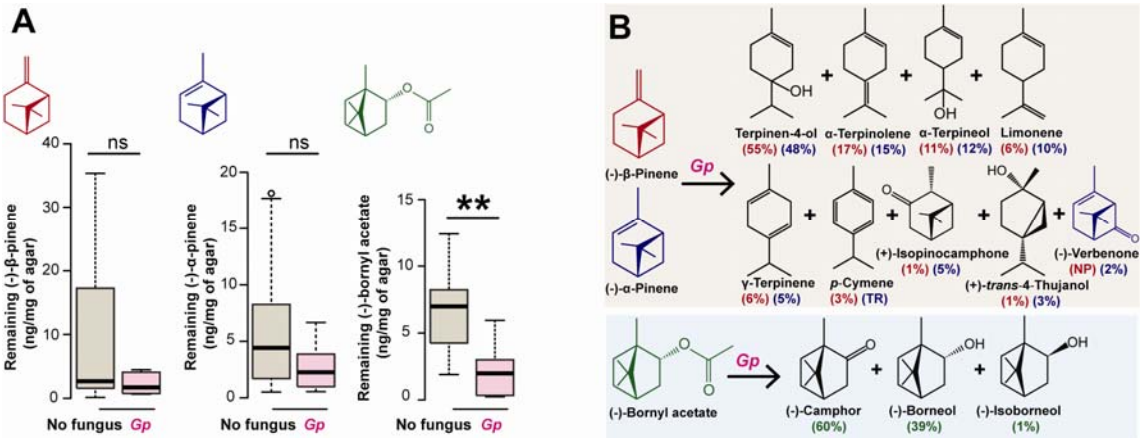

Figure 3: **Metabolism of major spruce monoterpenes by a symbiotic fungus produces a variety of oxygenated monoterpenes.**

(A-B) Metabolism of major spruce monoterpenes by *G. penicillata* after fungal-infected vs. uninfected PDA was supplemented with 0.5 mg/g of (-)-β-pinene, (-)-α-pinene, and (-)-bornyl acetate. **(A)** The amounts of starting monoterpenes remaining after four days. Error bars represent SEM (n = 5). Asterisks indicate a significant difference between the PDA-inoculated *G. penicillata* and the fungus-free control (Welch's t-test) with ns = not significant, ***P* < 0.01. **(B)** The most abundant metabolites of administered monoterpenes are depicted with their percentages relative to the total amounts of metabolites detected for each compound (derived from (-)-β-pinene in red; derived from (-)-α-pinene in purple; derived from (-)-bornyl acetate in green) (*n* = 4). Amounts were determined by headspace collection of volatiles from the agar (*n* = 3).

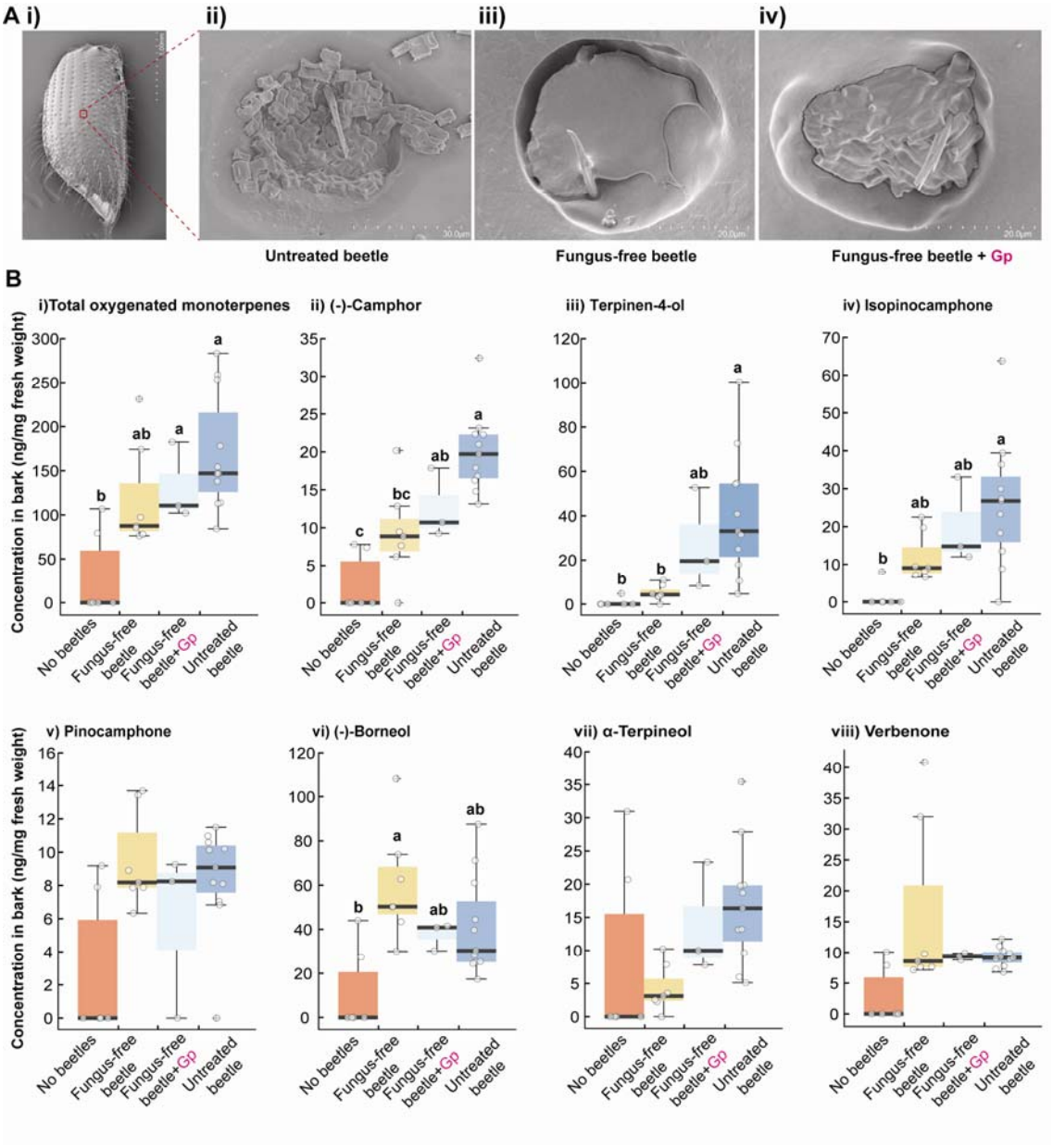

Figure 4: Oxygenated monoterpenes identified in *I. typographus*-infested spruce bark are produced by fungi associated with bark beetles

(A) Scanning electron micrographs of: i) an elytron of an untreated bark beetle showing ii) spores of an ophiostomatoid fungus in the elytral pit, iii) an empty elytral pit of a fungus-free beetle and iv) spore mass of *G. penicillata* in the elytral pit of a fungus-free beetle reinoculated with this fungal species. **(B)** Concentration of total oxygenated monoterpenes (i), camphor (ii), terpienen-4-ol (iii), isopinocamphone (iv), pinocamphone (v), borneol (vi), α-

terpineol (vii), and verbenone (viii) in bark without beetles, infested with fungus-free beetles, infested with fungus-free beetles reinoculated with *G. penicillata,* and untreated beetles. Different lowercase letters indicate significant differences between treatments (ANOVA, Tukey's test, $P < 0.05$).

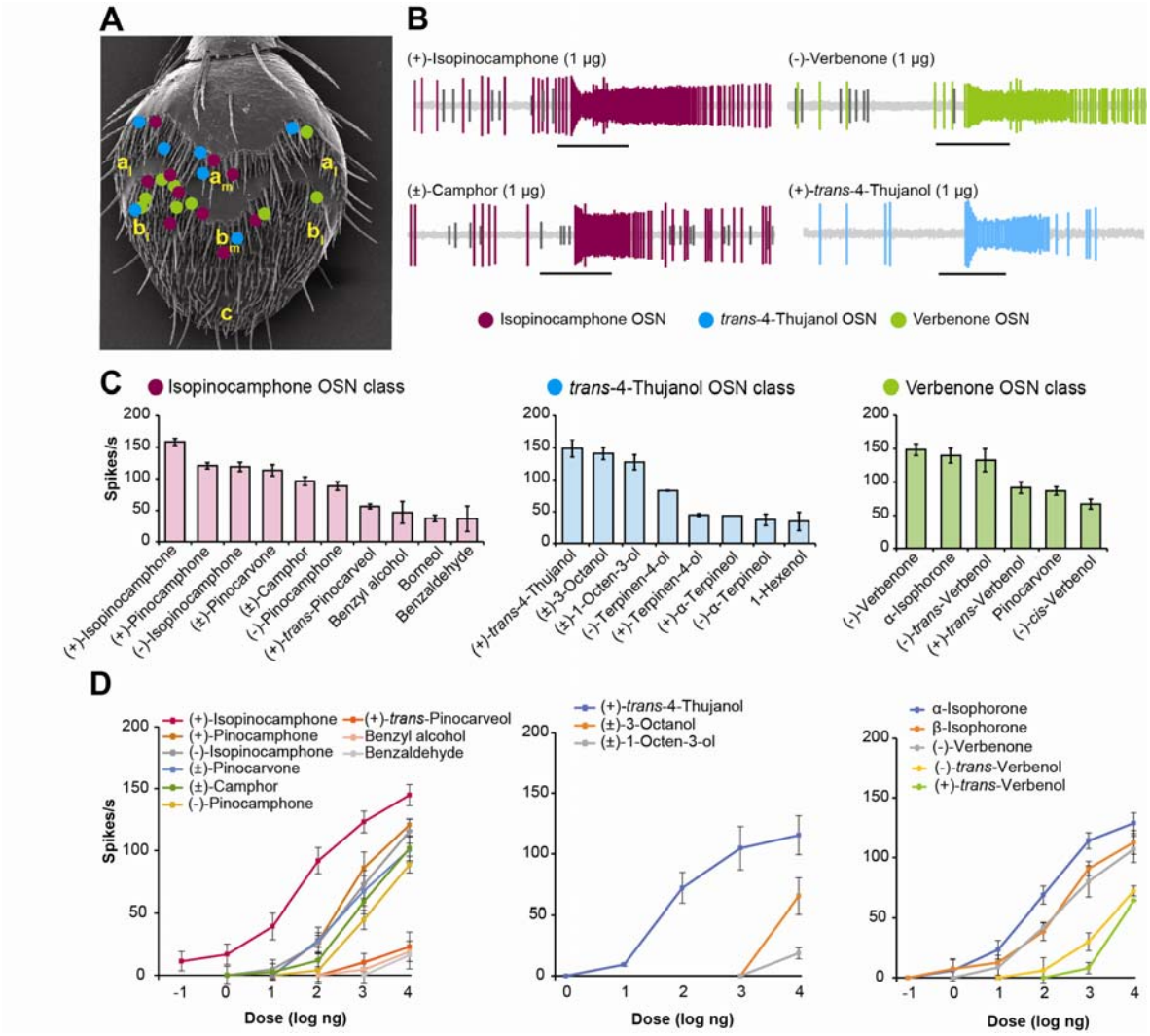

Figure 5: **Oxygenated monoterpenes derived from fungal metabolism of host tree monoterpene hydrocarbons are detected by specialized olfactory neurons in the *I. typographus* antenna**

(A) Mapping of three classes of olfactory sensory neurons (OSN) selective for oxygenated monoterpenes (isopinocamphone; verbenone; (-)-*trans*-4-thujanol) on the antenna. Olfactory sensilla housing these OSN classes are distributed in medial (subscript "m") and lateral (subscript "l") regions of sensillum areas a and b (but not in area c) of the antenna. **(B)** Representative spike trains from an isopinocamphone-responsive neuron stimulated with 1 µg of (+)-isopinocamphone (top left) and (±)-camphor (bottom left); a verbenone-responsive neuron stimulated with 1 µg (-)-verbenone (top right); a (+)-*trans*-4-thujanol-responsive neuron stimulated with 1 µg (+)-*trans*-4-thujanol (bottom right). Black horizontal bars indicate the 0.5 s odor puffs. **(C)** Response spectra of OSN classes responding predominantly to

oxygenated monoterpenes produced by fungi at the 10 µg screening dose. The average number of spikes/second was recorded from the isopinocamphone-tuned OSN class (left) (*n* = 6 except (+) and (-)-pinocamphone (*n* = 3)), *trans*-4-thujanol-tuned OSN class (middle) (*n* = 5) and verbenone and isophorone-tuned OSN class (right) (*n* = 4) after neurons were stimulated with a panel of 97 odors. Error bars represent SEM. **(D)** Dose-response curves of the OSNs stimulated with their most active ligands: isopinocamphone-tuned OSN class (left) ((*n* = 9) except for (+) and (-)-pinocamphone (*n* = 3)), *trans*-4-thujanol-tuned OSN class (middle) (*n* = 3), and the OSN class tuned to isophorone and verbenone (right) (*n* = 3). Error bars represent SEM.

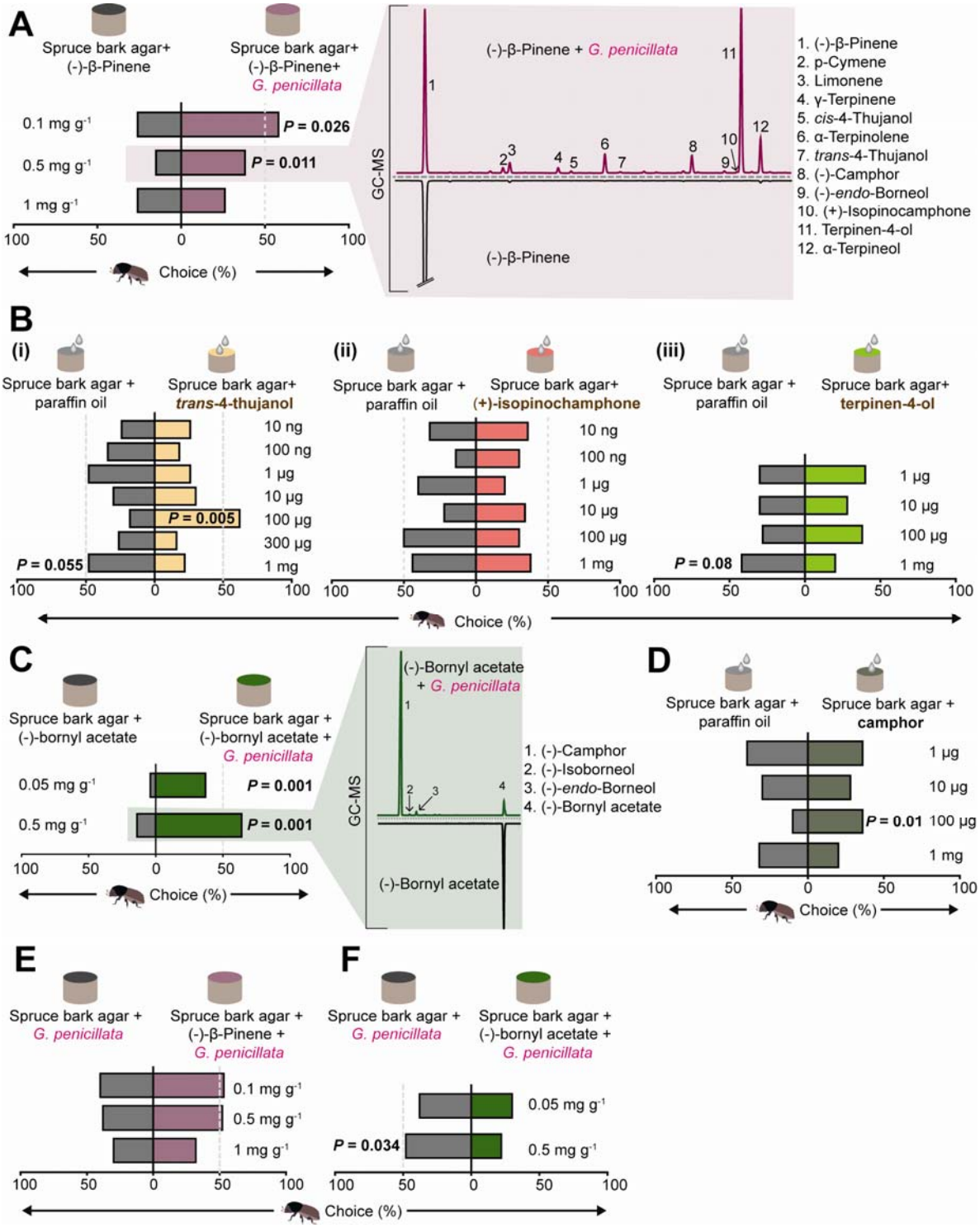

Figure 6: ***Ips typographus* is attracted to oxygenated monoterpenes produced by symbiotic fungi.**

(A) Adult beetles preferred spruce bark agar enriched with 0.1 mg/g, and 0.5 mg/g of (-)-β-pinene inoculated with *G. penicillata* for 4 days over spruce bark agar enriched with 0.1 mg/g, and 0.5 mg/g (-)-β-pinene but without fungus (left). GC-MS traces of the headspace volatiles of (-)-β-pinene-enriched agar with and without *G. penicillata* for 4 days (right) showing the oxygenated monoterpenes produced by the fungus from (-)-β-

pinene. Numbers refer to the identities of compounds. **(B)** Adult beetles chose *trans*-4-thujanol (left) at a 100 µg dose but avoided *trans*-4-thujanol (left) at a 1 mg dose diluted in mineral oil, when tested against a mineral oil control. Adult beetles showed indifferent responses to (+)-isopinocamphone (middle), and terpinen-4-ol (right), applied in various doses in mineral oil. **(C)** Adult beetles preferred spruce bark agar enriched with various amounts of (-)-bornyl acetate inoculated with *G. penicillata* for 4 days over spruce bark agar enriched with (-)-bornyl acetate but without fungus (left). GC-MS traces of the headspace volatiles of 0.5 mg/g (-)-bornyl acetate-enriched agar with and without *G. penicillata* (right) showing the oxygenated monoterpenes produced by the fungus from (-)-bornyl acetate. Numbers refer to the identities of compounds. **(D)** Adult beetles preferred (±)-camphor at a 100 µg dose against a mineral oil control, but not at other doses. **(E)** Adult beetles did not discriminate between *G. penicillata* on agar with three different concentrations of (-)-β-pinene and *G. penicillata* without (-)-β-pinene. **(F)** Adult beetles chose *G. penicillata* on agar without any administrated (-)-bornyl acetate vs. *G. penicillata* on agar with 0.5 mg/g (-)-bornyl acetate. **(A), (B), (C), (D), (E), (F)** Deviation of response indices against zero was tested using Wilcoxon's test. Asterisks denotes significant differences, *P<0.05, **P<0.01, ***P< 0.001. n = 25 for each trial.

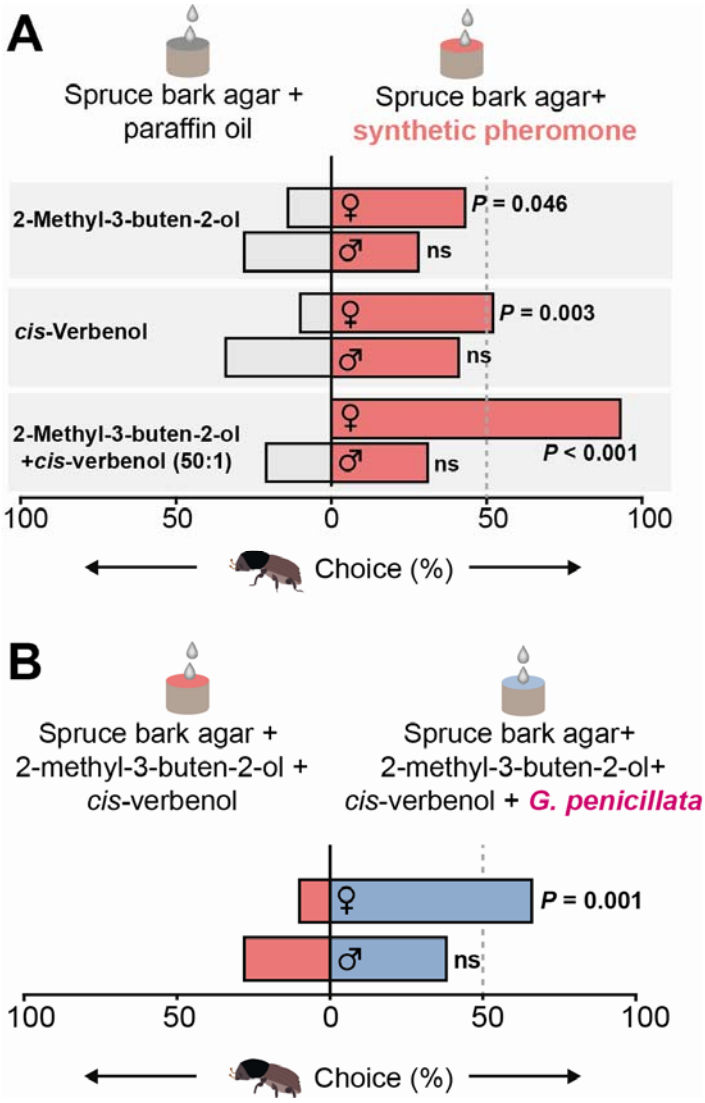

Figure 7: Female adult *I. typographus* are attracted towards a pheromone mixture in the presence of *G. penicillata* volatiles

(A) Adult females chose traps containing 2-methyl-3-buten-2-ol (MB) and *cis*-verbenol at 10^{-2} concentration diluted in mineral oil over control traps containing mineral oil. Females strongly preferred traps containing a binary pheromone blend (*cis*-verbenol: 2-methyl-3-buten-2-ol, 50:1) diluted in mineral oil over the mineral oil control (bottom). Adult males were unresponsive to these concentrations of individual pheromones and their blend. **(B)** Adult females preferred pheromone blend in the presence of *G. penicillata* volatiles over pheromone blend without fungus. **(A), (B)**. Deviation of response indices against zero was tested using Wilcoxon's test. Asterisks denotes significant differences, *P<0.05, **P<0.01, ***P< 0.001 (n = 28 for each experiment).

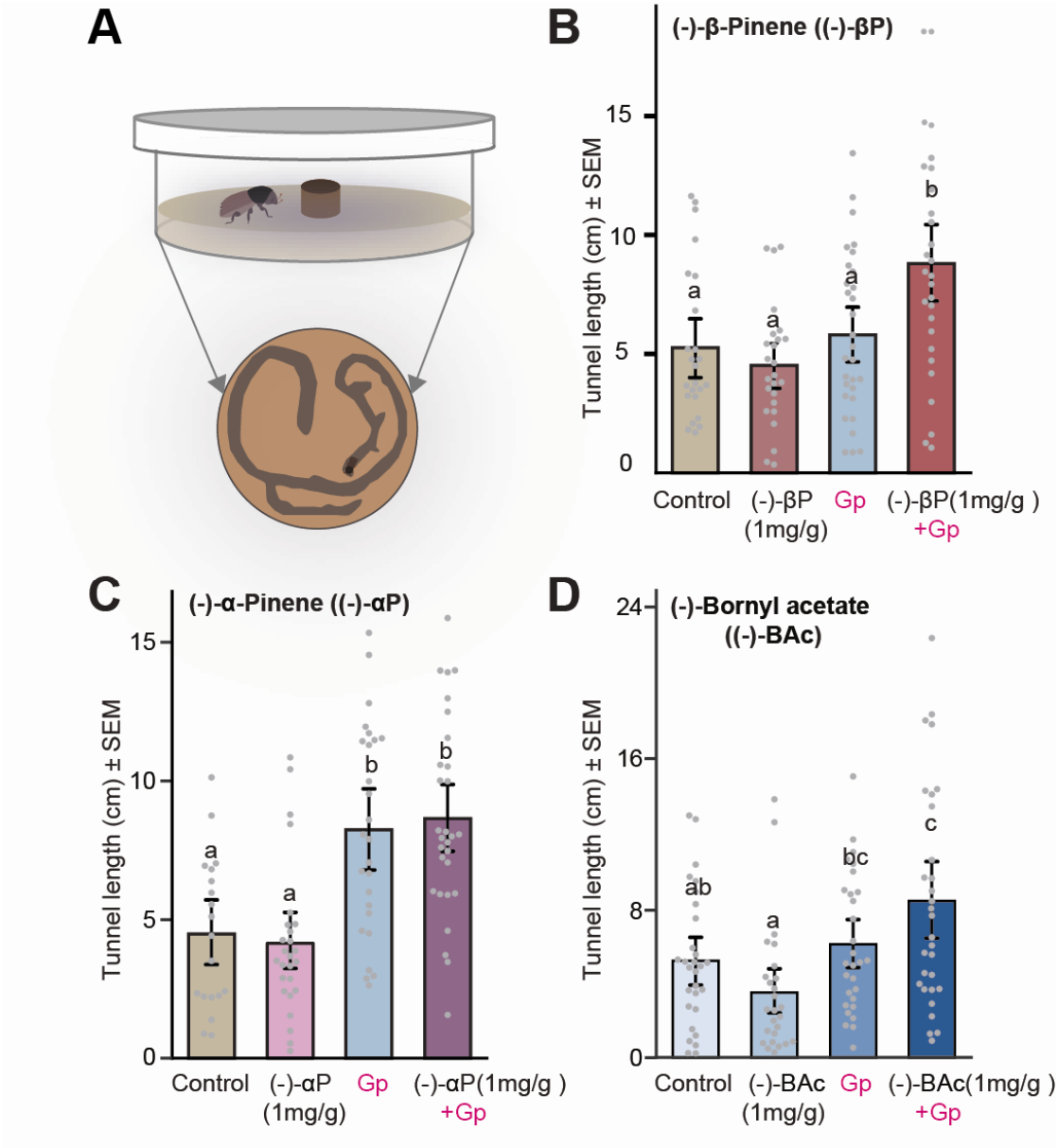

Figure 8: ***Ips typographus* tunnels more in monoterpene-enriched diet in the presence of a symbiotic fungus.**

(A) Schematic drawing of a Petri dish arena used for the no-choice tunneling assay. The dish was filled with monoterpene-enriched spruce bark agar and inoculated with *G. penicillata* (Gp) (top). Example of the tunneling pattern of an adult beetle within the fungus-colonized diet as pictured from the bottom side of Petri dish (bottom). **(B), (C), (D)** Tunnel lengths (cm) made by adult beetles after 48 hours in diet containing *G. penicillata* only, monoterpenes only, *G. penicillata* and monoterpenes, or controls with neither *G. penicillata* nor monoterpenes. Error bars represent SEM (n = 30 (15 ♂, 15 ♀) for each trial). Monoterpenes: (-)-β-pinene **(B)**, (-)-α-pinene **(C)**, (-)-bornyl acetate **(D)**. Different lowercase letters indicate significant differences between treatments (ANOVA, Tukey's test, P< 0.05).

Table 1: Multiple logistic regression analysis predicting the odds of adult *I. typographus* tunneling into media enriched in different monoterpenes with and without *G. penicillata* in a no-choice assay (see also Fig 6A).

Successful tunneling in diets with[a]	Predictors	β	SE[b]	Wald χ^2	*P* value	Exp(β)	95% CI for Exp(β)	
							Lower	Upper
(-)-Bornyl acetate	Monoterpene	-0.31	0.56	0.31	ns	0.73	0.25	2.19
	Sex	-1.72	0.58	8.78	**0.003**	0.18	0.06	0.56
	Fungus	4.98	1.09	20.99	**<0.001**	145.30	17.27	1222.52
(-)-β-Pinene	Monoterpene	-0.87	0.54	2.57	ns	0.42	0.14	1.22
	Sex	0.25	0.52	0.24	ns	1.29	0.47	3.55
	Fungus	2.65	0.58	20.45	**<0.001**	14.09	4.48	44.36
(-)-α-Pinene	Monoterpene	0.00	0.52	0.00	ns	1.00	0.36	2.75
	Sex	0.00	0.52	0.00	ns	1.00	0.36	2.75
	Fungus	2.02	0.66	9.39	**0.002**	7.51	2.07	27.28

[a]The reference category is unsuccessful tunneling
[b]Standard error of *β*
ns=not significant