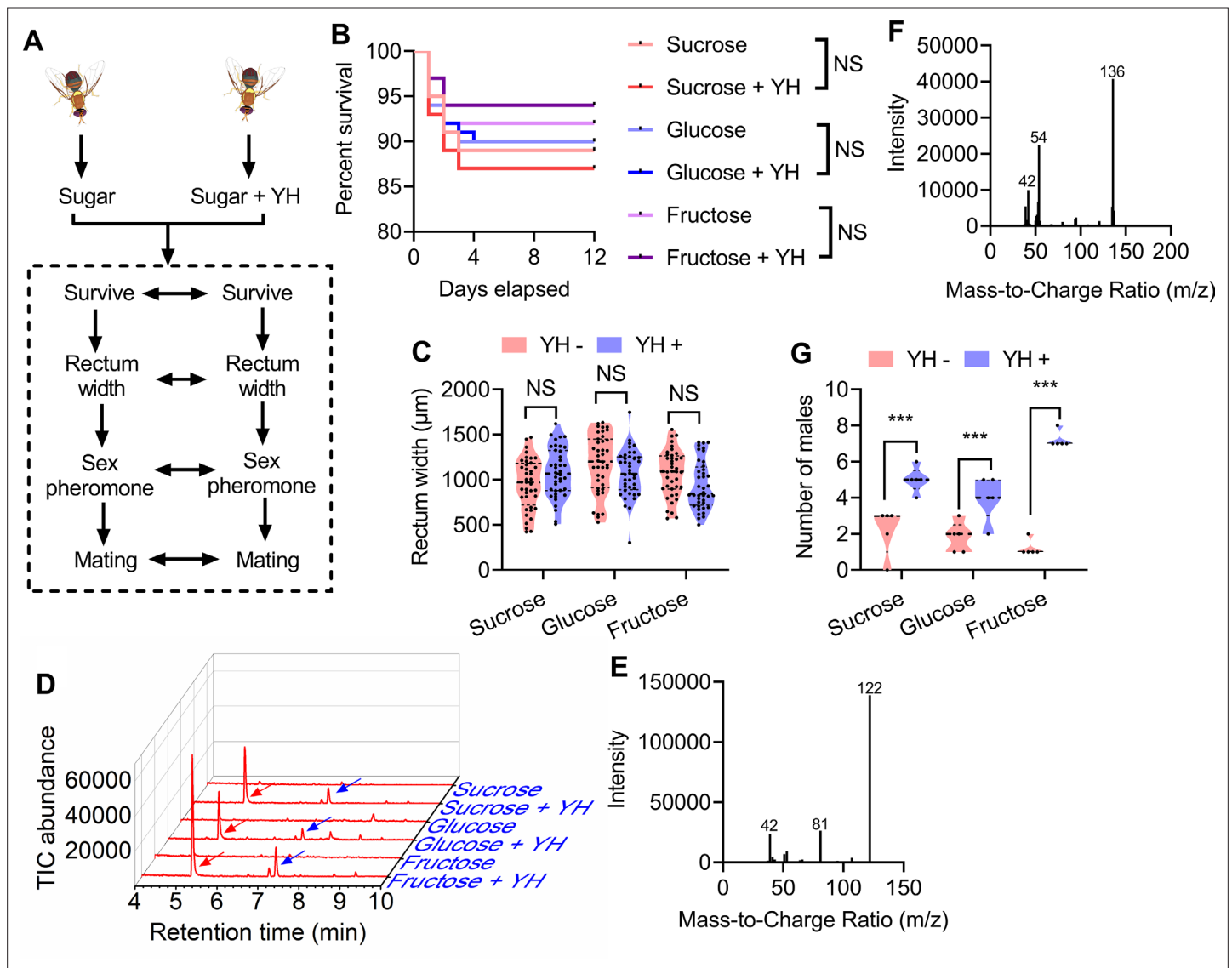


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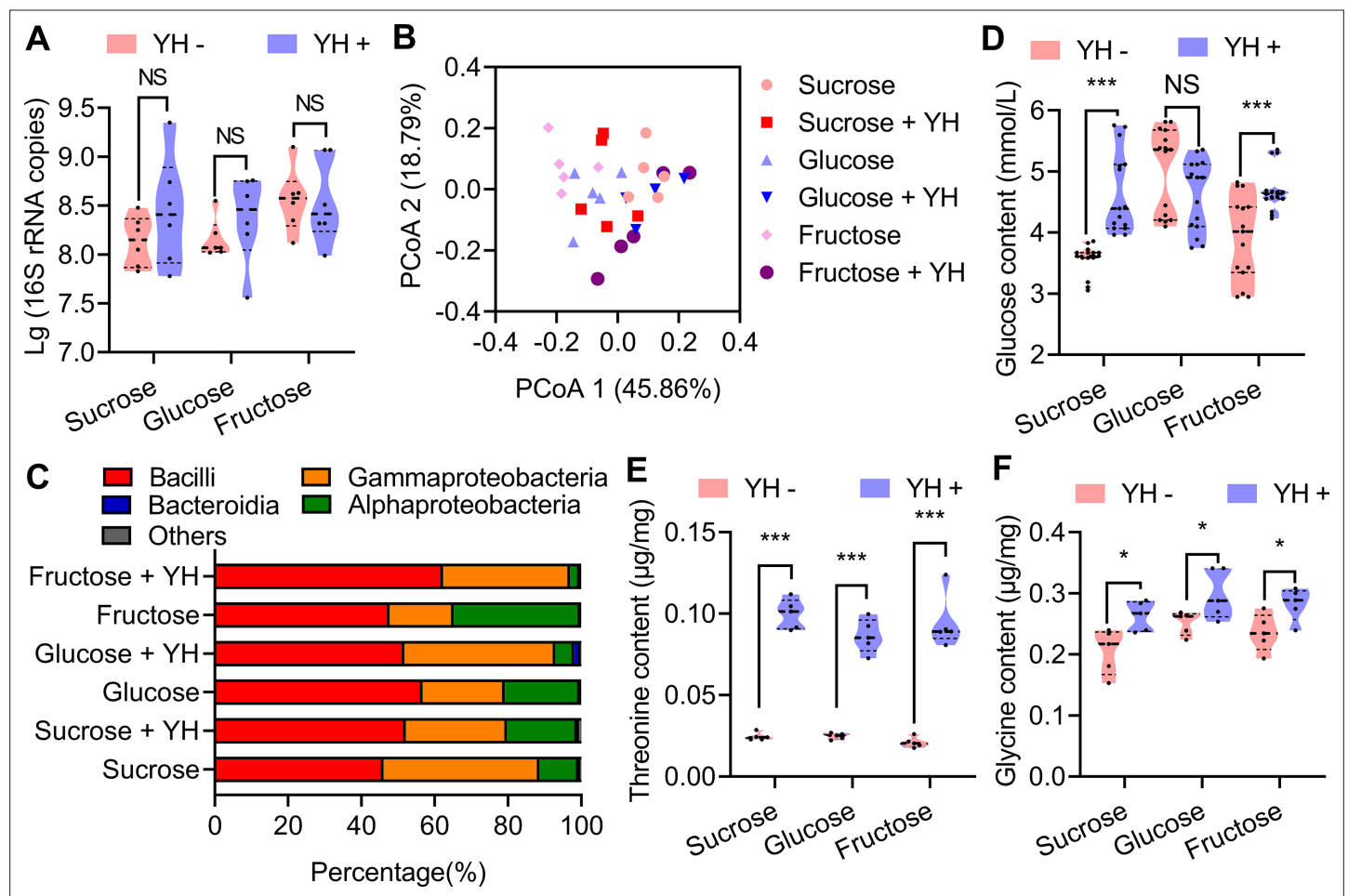
## Figures and figure supplements

Protein feeding mediates sex pheromone biosynthesis in an insect

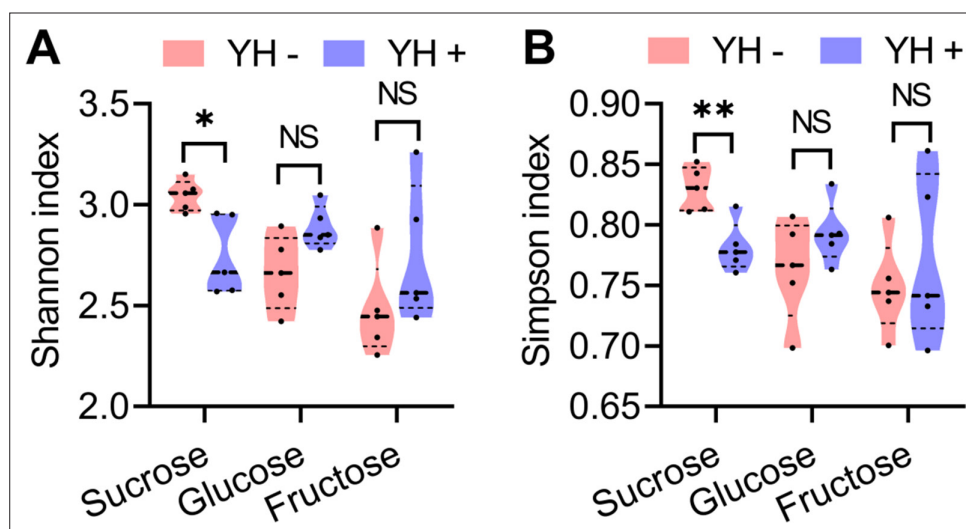
**Shiyu Gui *et al.***



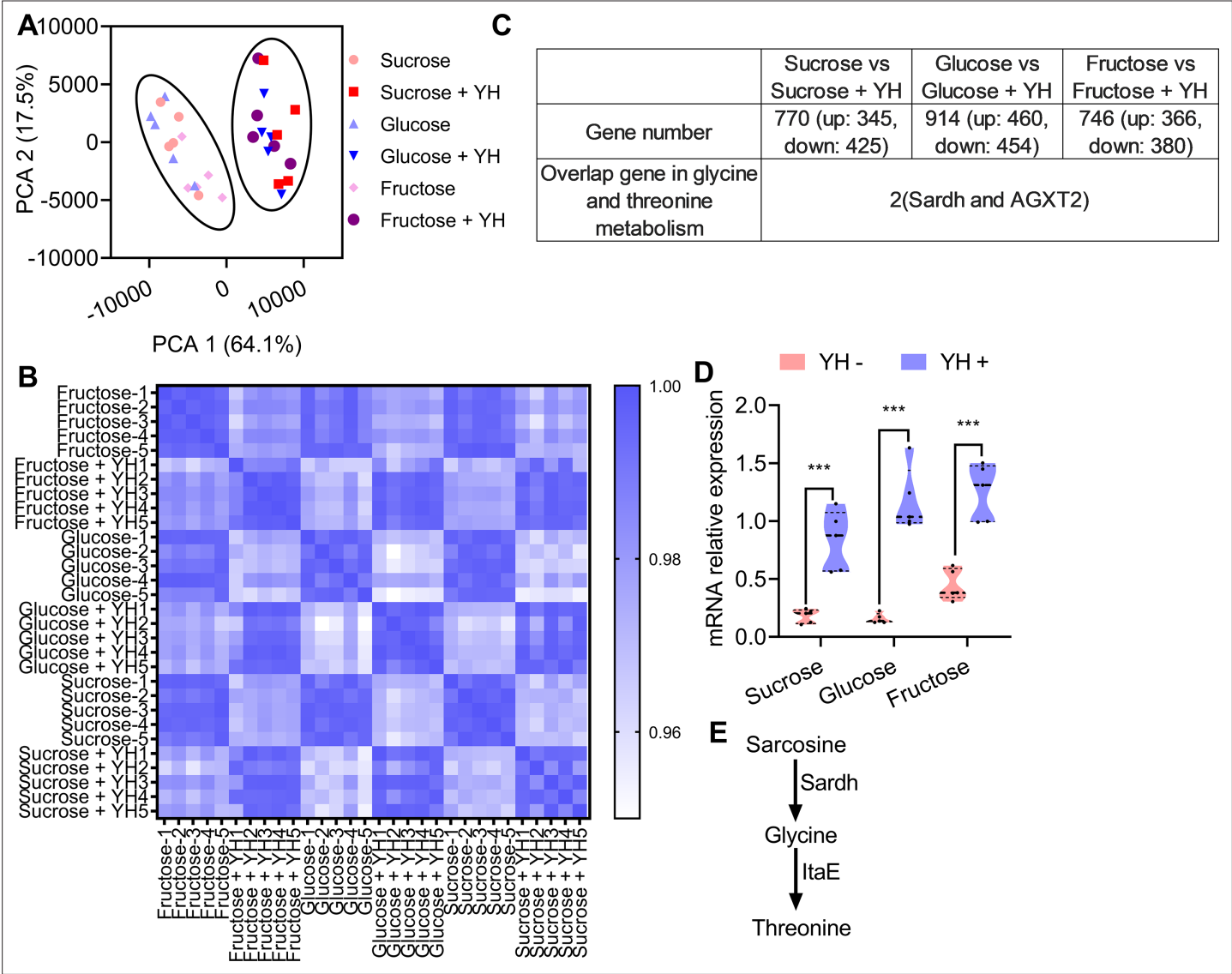
**Figure 1.** Influence of post-tenal protein and sugar feeding on male flies. **(A)** A schematic showing how the male flies were reared and the biological parameters compared. **(B)** Effect of post-tenal protein and sugar on survival ( $n=200$  individuals, Kaplan–Meier survival analysis was used, and NS: no significance). **(C)** Rectum size comparisons between yeast hydrolysate (YH)-deprived (YH-) and YH-fed (YH+) males ( $n=40$  individuals, Student's  $t$  test, and NS: no significance). **(D)** Gas chromatography–mass spectrometry (GC–MS) ion chromatograms of rectum extracts of males fed different types of food. Traces for the flies fed with YH expressing trimethylpyrazine (TMP; red arrow) and tetramethylpyrazine (TTMP; blue arrow) are shown. **(E)** and **(F)** GC–MS mass spectra of TMP and TTMP. **(G)** Mating ability comparisons between YH- and YH+ males ( $n=5$  replicates, Wilcoxon matched-pairs signed rank test, and \*\*\*  $p < 0.001$ ). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.



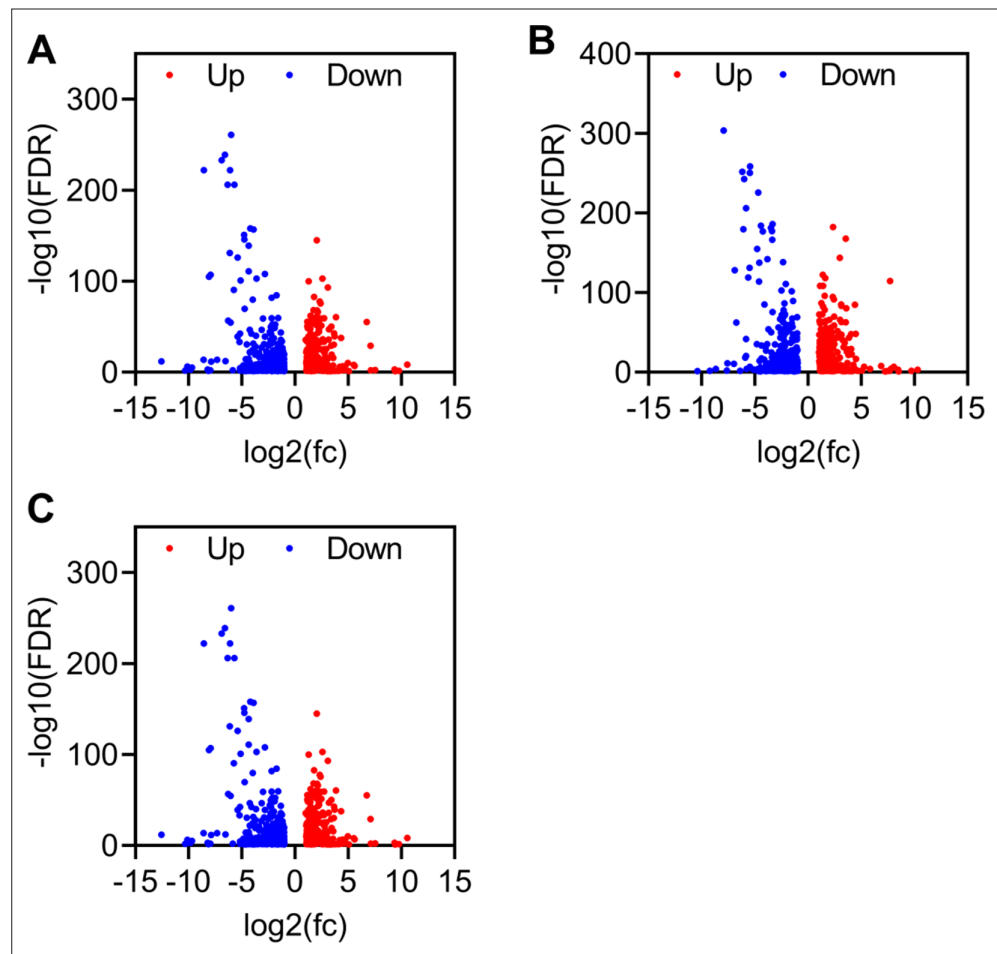
**Figure 2.** Influence of post-tenal protein on rectal bacteria and sex pheromone precursors. **(A)** Boxplot showing total bacteria in the male rectum estimated from 16S rRNA gene quantitative PCR (qPCR; n=6 replicates, Student's t test, and NS: no significance). **(B)** Principal coordinate analysis of the microbial community structure (beta diversity and class level) measured by the Bray–Curtis distance matrix of 16S rRNA gene amplicon sequences. **(C)** Class-level relative abundance of 16S rRNA gene amplicon sequences. Values are averaged according to yeast hydrolysate (YH)-deprived and YH-supplied males. **(D), (E), and (F)** Influence of YH supply on rectum glucose (n=15 replicates), threonine (n=5 replicates), and glycine (n=5 replicates) contents (Student's t test, NS: no significance, \* p<0.05, and \*\*\* p<0.0001). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.



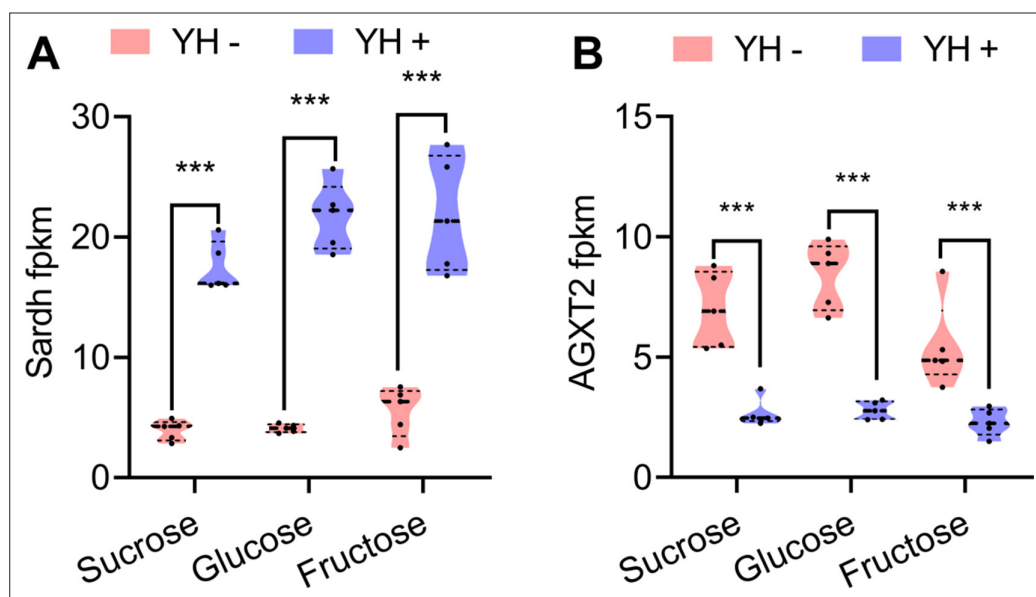
**Figure 2—figure supplement 1.** Boxplots showing the estimated diversity of the microbial community based on Shannon (A) and Inverse Simpson (B) indices of the 16S amplicon sequences (n=5 replicates, Student's *t* test, NS: no significance, \*  $p < 0.05$ , and \*\*  $p < 0.001$ ). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.



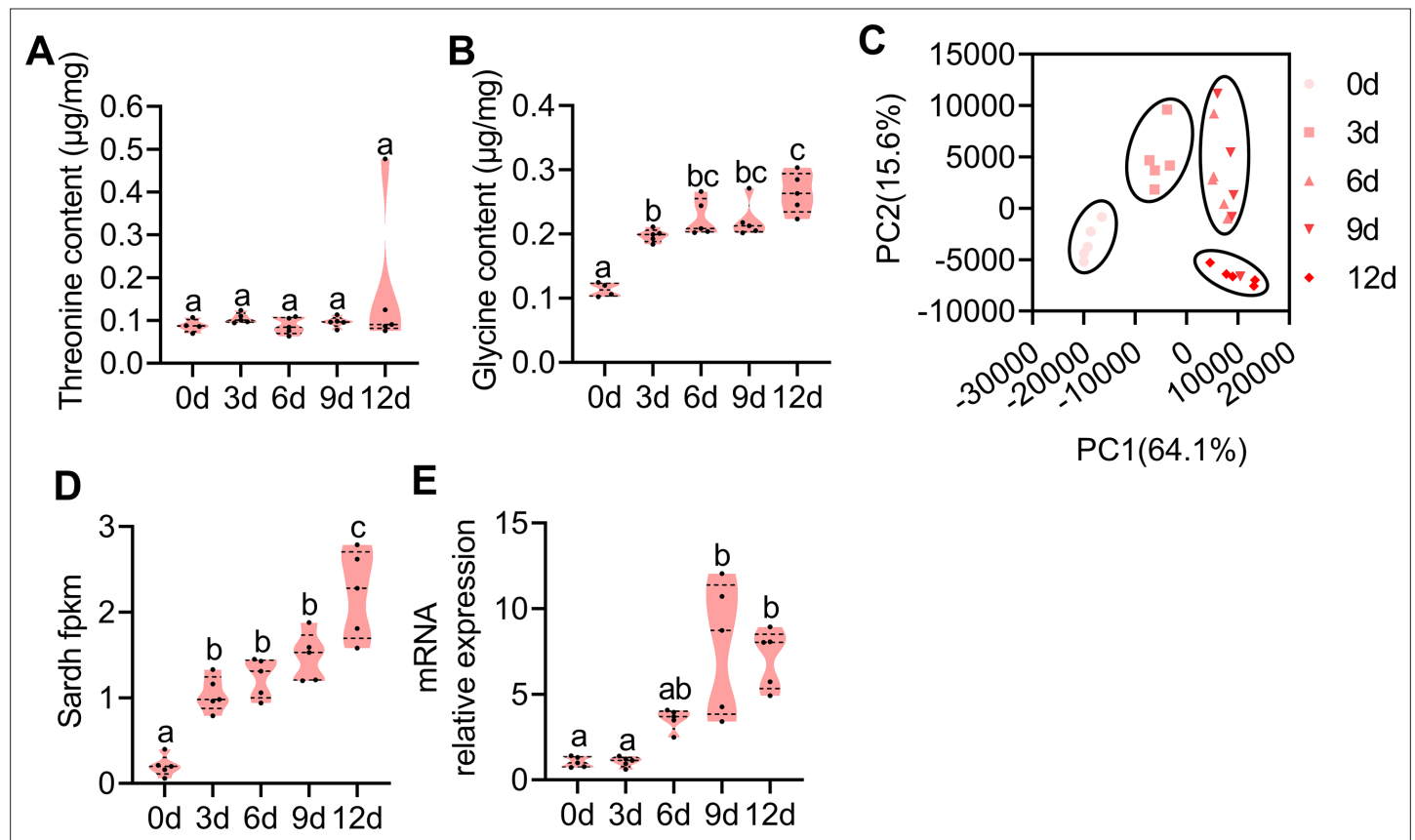
**Figure 3.** Transcriptome comparisons between yeast hydrolysate (YH)-fed and YH-deprived males. **(A)** Principal component analysis (PCA) obtained from gene expression profiles showing differences between YH-fed and YH-deprived males. Flies are clustered according to YH fed or not. **(B)** Pearson correlation coefficient showing the similarity between the samples. Higher similarity of the transcriptome is shown by a darker blue color (higher correlation coefficient). **(C)** Table showing the number of genes found in any given category and the genes involved in the threonine metabolism pathway between comparisons. **(D)** Quantitative PCR (qPCR) verifying the expression of *Sardh* in YH-fed and YH-deprived males ( $n=5$  replicates, Student's  $t$  test, and \*\*\*  $p<0.001$ ). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median. **(E)** Proposed model of the threonine metabolism pathway in insects.



**Figure 3—figure supplement 1.** Differential expressed genes in sucrose (A), glucose (B), and fructose (C) groups.

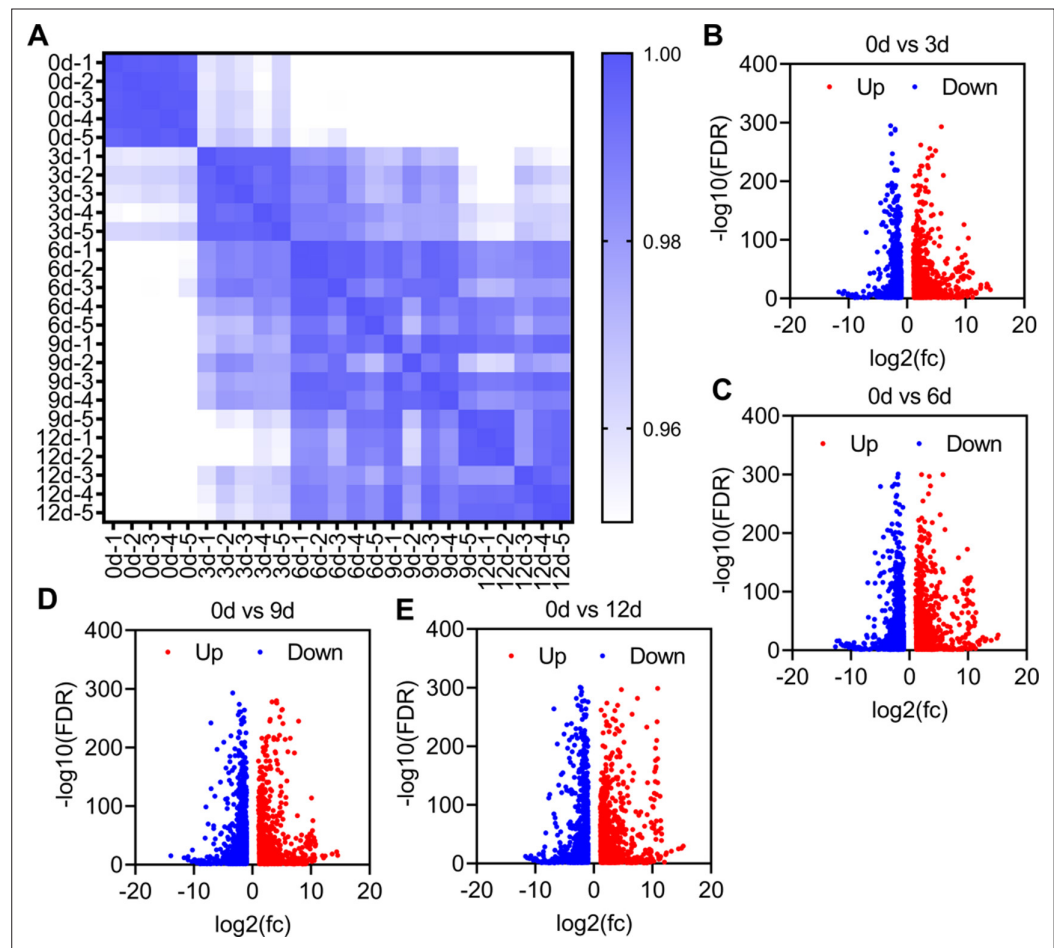


**Figure 3—figure supplement 2.** Fragments Kilobase of exon model per million mapped reads(fpkm) values of the differentially expressed genes in glycine, serine, and threonine metabolism pathway (n=5 replicates, Student's *t* test, and \*\*\* *p*<0.0001). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.

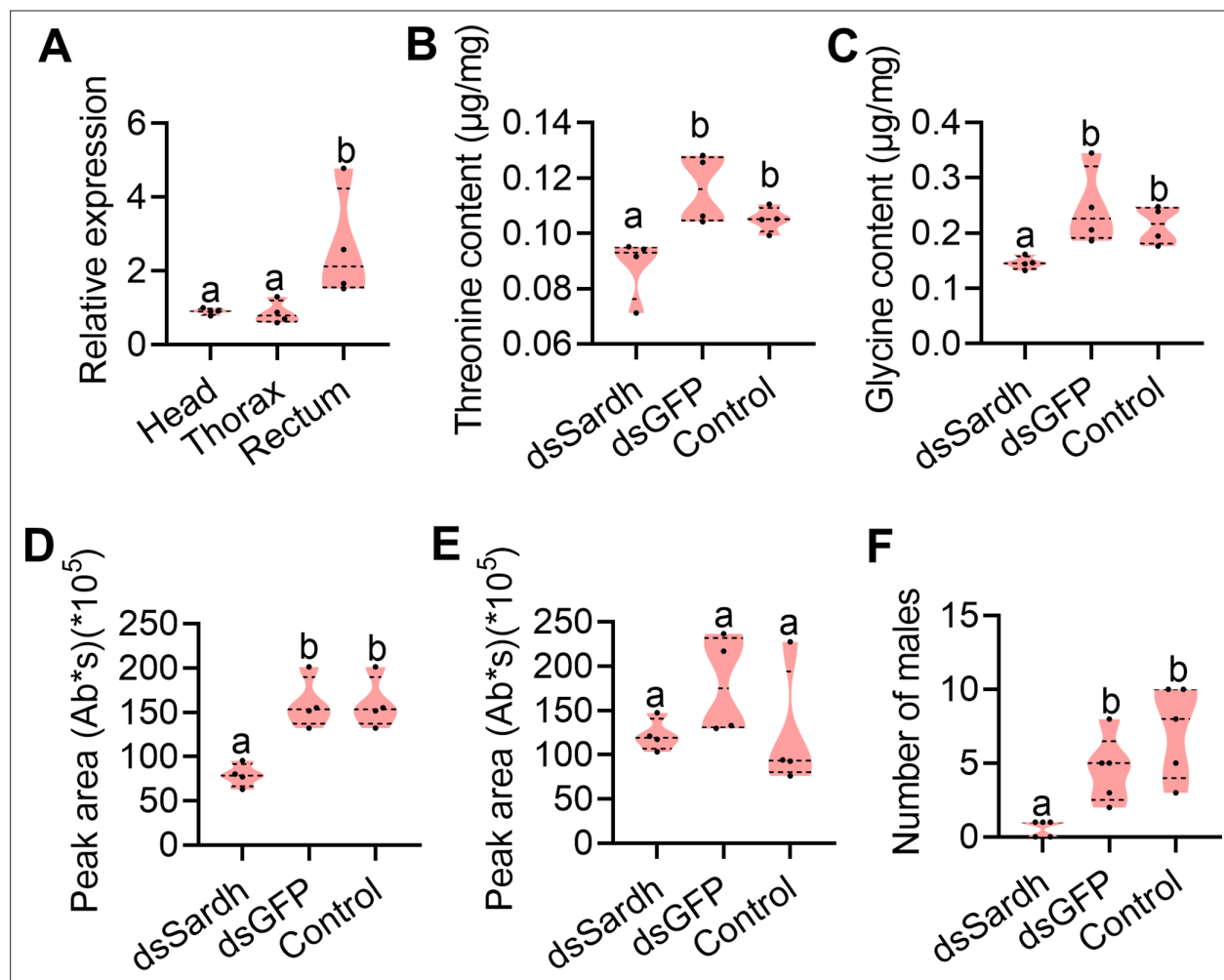


**Figure 4.** Amino acid contents and transcriptome investigation of male rectums at different developmental stages. **(A and B)** Threonine (n=5 replicates) and glycine (n=5 replicates) contents in the rectum at different developmental stages (different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). **(C)** Principal component analysis (PCA) using differential expression (DE) genes obtained from pairwise comparisons between different developmental stages. **(D and E)** Expression profiles of *Sardh* obtained by RNA-seq and quantitative PCR (qPCR; n=5 replicates, different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.

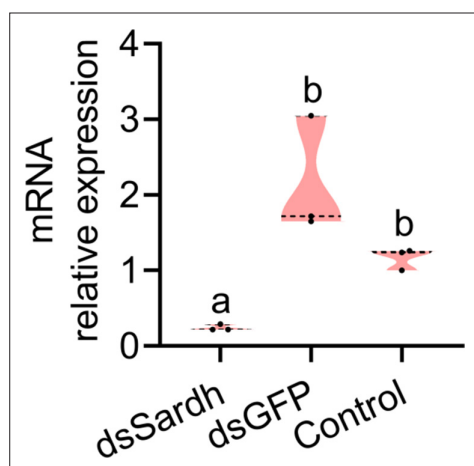




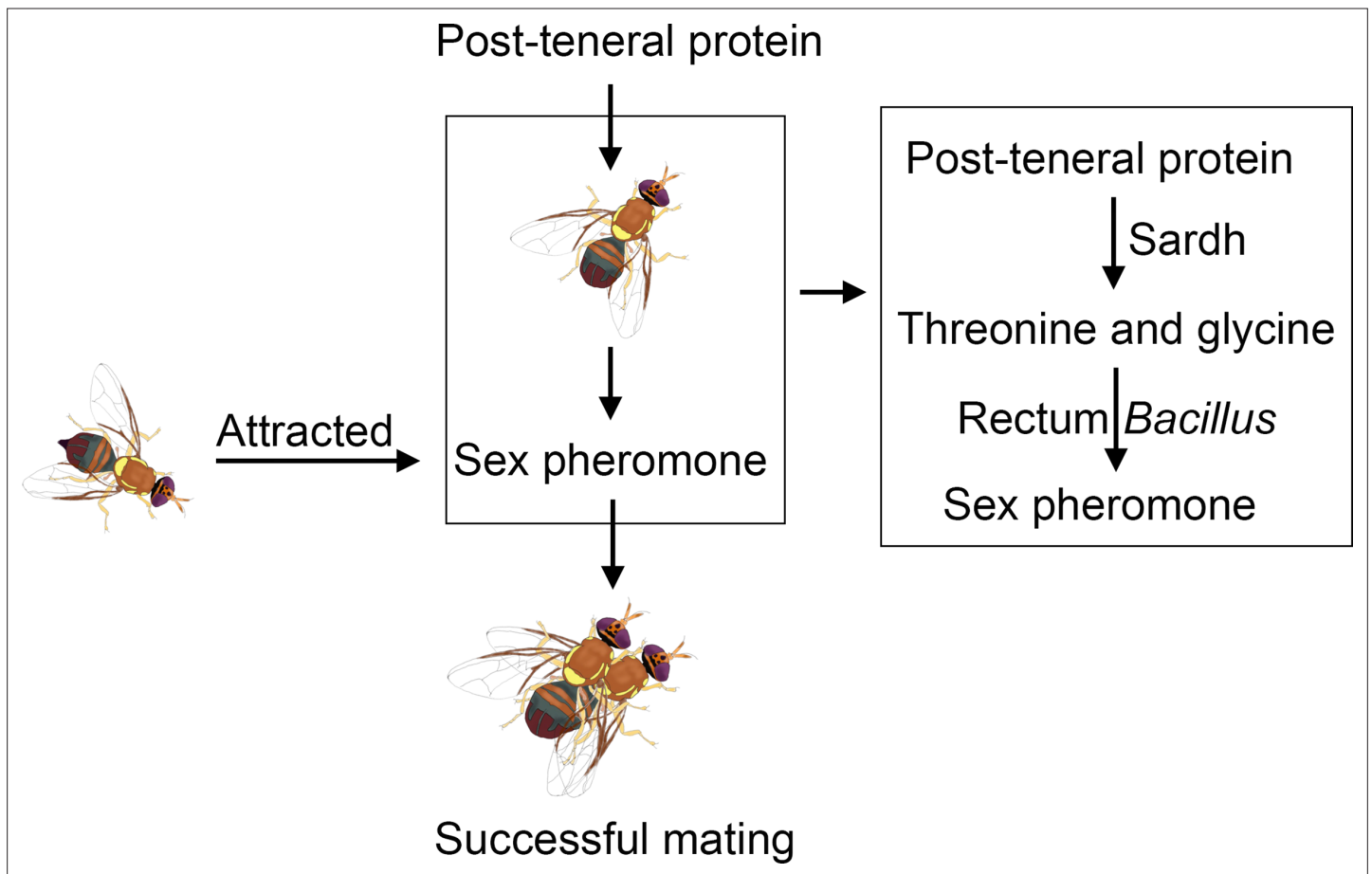
**Figure 4—figure supplement 1.** Transcriptome comparisons between different development stages. (A) Pearson correlation coefficient showing the similarity between the samples. Higher similarity of the transcriptome was shown by darker blue color (higher correlation coefficient). (B–E) Differential expressed genes between different development stages.



**Figure 5.** Functional verification of *Sardh* in sex pheromone biosynthesis. **(A)** Expression of *Sardh* in different tissues with quantitative PCR (qPCR;  $n=5$  replicates, different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). **(B and C)** Threonine ( $n=5$  replicates) and glycine ( $n=5$  replicates) contents in the rectum with *Sardh* knockdown (different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). **(D and E)** Sex pheromone (trimethylpyrazine [TMP] and tetramethylpyrazine [TTMP]) quantification in the rectum with *Sardh* knockdown ( $n=4$  replicates, different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). **(F)** Mating ability comparisons between males with *Sardh* knockdown and controls ( $n=5$  replicates, different letters above the error bars indicate significant differences at the 0.05 level analyzed by Kruskal–Wallis test followed by Dunn's multiple comparisons test). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.



**Figure 5—figure supplement 1.** RNAi efficiency of Sardh after 48 hr (n=3 replicates, different letters above the error bars indicate significant differences at the 0.05 level analyzed by ANOVA followed by Tukey's test). In violin plots, where the violin encompass the first to the third quartiles, inside the violin the horizontal line shows the median.



**Figure 6.** Schematic illustrating the sex pheromone biosynthesis hypothesis that *B. dorsalis* rectum. *Bacillus* uses threonine and glycine, which are converted by Sardh with post-teneral protein feeding by *B. dorsalis*, as precursor substances to synthesize the sex pheromone.