PRIMATE CEREBELLAR SCALING IN CONNECTION TO THE CEREBRUM: A 34-SPECIES PHYLOGENETIC COMPARATIVE ANALYSIS

Neville Magielse¹⁻³, Roberto Toro⁴, Vanessa Steigauf⁵, Mahta Abbaspour^{6,7}, Simon B. Eickhoff^{1,3}, Katja Heuer^{4,8*} & Sofie L. Valk^{1-3*}

Corresponding author: Sofie L. Valk (s.valk@fz-juelich.de)

Address: Max Planck Institute for Human Cognitive and Brain Sciences, Dr. Sofie Valk,

Stephanstraße 1A, 04103 Leipzig, Sachsen, Germany.

Telephone: +49 341 9940-2658 | Fax: +49 341 9940-104

NM. Conceptualized the manuscript; wrote and adapted custom computer code; performed manual segmentations; performed analyses; wrote the manuscript; incorporated coauthor revisions. **RT.** Provided MRI data; wrote original custom code; wrote used software; provided revision comments manuscript. **VS and MA** provided manual segmentations of ansiform areas for 12 and 6 specimens, respectively. **SBE.** Provided revision comments manuscript; provided funding for the project. **KH*.** Provided MRI data; provided semi-automated segmentation and software support; wrote original custom code; wrote used software; provided revision comments manuscript; provided supervision. **SLV*.** Conceptualized the manuscript; provided a subset of primary segmentations; provided revision comments on manual segmentations, the manuscript, and computer code (multiple occasions); provided supervision; provided funding for the project.

¹ Institute of Neuroscience and Medicine, Brain and Behaviour (INM-7), Research Center Jülich, Wilhelm-Johnen-Straße, 52428 Jülich, Germany.

² Otto Hahn Cognitive Neurogenetics Group, Max Planck Institute for Human Cognitive and Brain Sciences, Stephanstraße 1A, 04103, Leipzig, Germany.

³ Institute of Systems Neuroscience, Medical Faculty, Heinrich Heine University Düsseldorf, Moorenstraße 5 40225 Düsseldorf, Germany.

⁴ Institut Pasteur, Unité de Neuroanatomie Appliquée et Théorique, Université Paris Cité, 25 rue du Dr. Roux, 75724 Paris, France.

⁵ Department of Biology, Northern Michigan University, 1401 Presque Isle Ave, MI 49855 Marguette, USA.

⁶ Berlin School of Mind and Brain, Humboldt-Universität zu Berlin, Luisenstraße 56, Haus 1, 10117 Berlin, Germany.

⁷ Department of Neurology, Charité – Universitätsmedizin Berlin, Bonhoefferweg 3, 10117 Berlin, Germany.

⁸ Department of Neuropsychology, Max Planck Institute for Human Cognitive and Brain Sciences, Stephanstraße 1A, 04103 Leipzig, Germany.

^{*} KH and SLV contributed to the manuscript equally and share last authorship.

Abstract

The cerebellum has increasingly been recognized for its role in diverse functional processes. The reciprocally connected cerebello-cerebral system may scaffold both brain size increase and advanced associative abilities, evolving highly coordinately in primates. In parallel, functional cerebello-cerebral modules have undergone reorganization and cerebellar lobules crura I-II (the ansiform area across mammals) have been reported to be specifically expanded in humans. Here we manually segmented 63 cerebella (34 primate species; 9 infraorders) and 30 crura I-II (13 species; 8 infraorders). We show that both constraints and reorganization may shape the evolution of the primate cerebello-cerebral system. Using phylogenetic generalized least squares, we find that the cerebellum scales isometrically with the cerebral cortex, whereas crura I-II scale hyper-allometrically versus both. Our phylogenetic analyses evidence primate-general crura I-II hyperscaling in contrast to virtually isometric cerebello-cerebral scaling. Crura I-II hyperscaling may be important for associative and cognitive brain functions in an evolutionary context.

Introduction

The cerebellum is implicated in abstract functions such as executive, social, and emotional processing^{1–4}. Lesion studies^{5,6} evidence behavioral deficits in these functional domains. The stereotyped structural organization of the cerebello-cerebral system, which has undergone marked modular reorganization in primates, likely supports its diverse functions^{7–10}. Human cerebellar involvement in cognitive modules^{11–13} may explain its activation in diverse cognitive tasks^{14,15} and its role in functional connectivity networks^{16–18}.

The cerebellum contains multiple output channels^{11,12,19,20}, which are organized into multiple reciprocal loops with the cerebral cortex^{12,21–24}. It likely forms internal models encoding mental representations from separate streams, returning updated predictions²⁵. Cerebellar structural stereotypy and phylogenetic conservation²⁶ argue for algorithmic^{27,28} uniformity across functional domains^{24,29–31}, and a direct link between function and cerebello-cerebral connectivity^{7,12}. Distinct cerebellar computations may still occur²⁹ and integration cannot be excluded based on overlapping connections³², fractured somatotopy³³, and transmodal integration in single granule cells³⁴.

Similar to humans^{21,22}, non-human primate (NHP) hemispheric cerebellar lobule VII, and especially crura I-II, connect reciprocally to prefrontal cortex (PFC)^{11,12,23,35}. A cross-species crura I-II homolog has been referred to as the ansiform area^{36,37}. We adopt this term here. Functionally, cerebellum-PFC connectivity modulates cognitive processing speed³⁸, congruent with reciprocal functional coherence between PFC and ansiform area³⁹. Human crura I-II additionally connect reciprocally to temporal and parietal associative cortices^{21,22}. Resting state fMRI (rsfMRI) studies illustrate widespread ansiform area involvement in functional networks, with extensive functional connectivity to parietal cortices including Brodmann area 7^{40,41}. Analyses ranging from winner-takes-all parcellations^{16,17,42}, to gradients⁴³ and lead-lag relationships⁴² indicate its unique involvement in associative networks such as the default mode (DMN), control, and frontoparietal networks, and its extreme position along the functional gradient separating DMN and language networks from motor functions. In humans, consistent with dominant contralateral cerebello-cerebral connectivity^{21,22} and conserved functional organizational asymmetry between macaques and humans⁴⁴, meta-analyses show left-lateralized executive and right-

lateralized language ansiform area activations¹⁴.

Primate brain size increase^{45–47} comes primarily from expansion of the cerebello-cerebral system⁴⁸. This system's structure and connectivity enable diverse functional specializations^{7,10,49}. Primate cerebellar and cerebral volumes^{50,51} and neuron numbers^{52,53} increase highly coordinately. Other vertebrates with abstract reasoning abilities^{54–56} have similar cerebello-cerebral anatomy and scaling^{57,58} as primates⁵⁷, suggesting strong conservation across phyla⁵⁹. Meanwhile, primate evolution seems to also involve reorganization of cerebello-cerebral areas and networks, exceeding volumetric increases⁶⁰ in their importance for cognitive abilities^{46,47,61}.

Previous work has revealed phylogenetic axes of functional organization 62,63. Structural modularity 64,65 within the primate cerebello-cerebral system may reflect functionally relevant reorganization with functionally and structurally connected subregions evolving at increased relative pace. Such disproportionate expansions of functionally-related structures may indicate behaviorally relevant changes, as they persist over millions of years 66 and against great metabolic costs 67–69. In great apes, cerebellar volumetric expansion may have outpaced that of the cerebrum 70. Similarly, cerebellar surface area expansion may have outpaced that of the cerebral cortex between macaque monkeys (33% of the cerebral cortex) and humans (78%) 71. Relatively large lateral cerebellar hemispheres characterize four vertebrate clades including primates 9 and especially hominoids 72. Specifically, the ansiform area volume fraction of the cerebellum is larger in humans than in macaques 13, and appears generally larger in most primates than in mice and rats 36. Expansion of lateral cerebellar areas alongside cerebral association areas in primates 7,73–75 putatively reflects selection on cerebello-cerebral networks 8,9,13,50,76

In the current study, we aimed to expand comparative analyses of the primate cerebello-cerebral system. We used a large (34 species, 63 specimens; Figure 1) MRI dataset alongside phylogenetic comparative methods to evidence isometric scaling between the cerebellum and cerebrum^{50,51,77} and assess ansiform area hyperscaling^{13,36}. Where possible, we explored intraspecific variability, which is substantial among brain traits^{72,78–82} and an inherent challenge for comparative primatology⁸³.

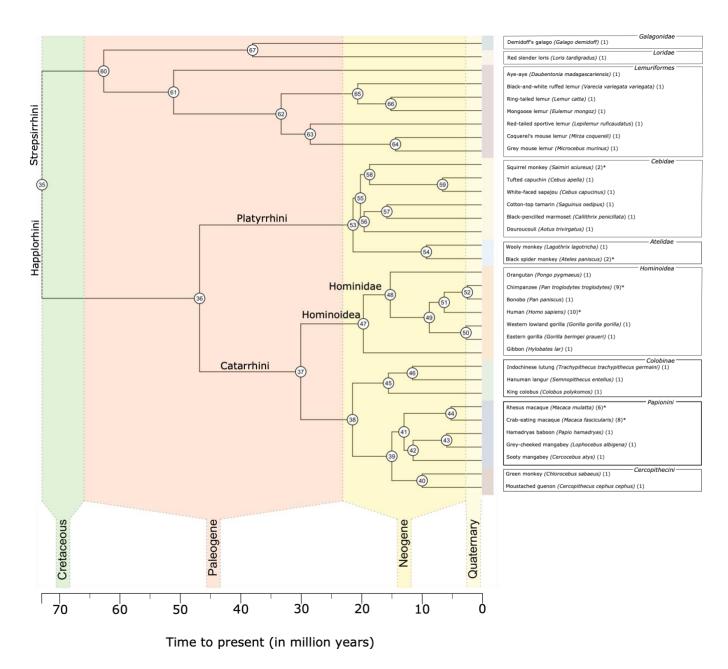


Figure 1: Consensus phylogenetic tree for the 34 primate species in this study. We obtained the consensus tree for the 34 species in our dataset from 10kTrees Arnold et al. (2010). Archeological time periods are superimposed over the tree to provide a temporal perspective of predicted species bifurcations. Internal node numbers are plotted and can be used to identify ancestral characters (Supplemental Table S1). Extant species included in the current study are given on the right and colored by their clade membership. Additionally, the numbers of specimens per species are given, and species with multiple specimens are marked with asterisks.

Our analyses confirmed isometric scaling between the cerebellum and cerebrum. Reanalysis of Stephan collection⁸⁴ data indicated this relation may be hypo-allometric. Conversely, the ansiform area scaled hyper-allometrically versus the cerebellum and

cerebrum. Contrary to our hypothesis based on previous work^{13,70,72}, hominoids did not exhibit exceptional scaling.

Methods

Overview

We perform phylogenetic comparative analyses on manually segmented cerebella from 34 primate species (N=63; Figure 1) and ansiform areas from 13 species (N=30). We provide here an overview of the methods. For a more detailed description see Supplemental Methods and for the full protocol see Heuer et al. (2019)⁸⁵.

Data

Primate comparative data are rare and sizable datasets are difficult to obtain. We used the collection of primate species collated from different sources in the Brain Catalogue Primates project⁸⁵, available on BrainBox⁸³. It included 35 species (N=66), which covered the primate phylogenetic tree relatively well, with 9 infraorders from Strepsirrhini and Haplorhini suborders. The red howler monkey was excluded due to irreparable damage. Cerebellar damage was also noted in other specimens, whose volumetric estimates need to be treated with due diligence. Cebidae (7 species), Hominoidea (7), Lemuriformes (6), and Papionini (5) were most extensively sampled (Figure 1). Scans included both fully abstracted brains and *in situ* brains, either full body scans or only including the skull. T1- or T2-weighted (T1w or T2w) MRI anatomical scans were obtained at either 1.5, 3T, 7T or 11.4T for up to 12 hours. See Supplemental Methods for basic data acquisition, provenance^{85–89} and damage description, and Supplemental File S1 for quality-control data including signal-to-noise ratios and resolutions.

Manual segmentation procedure

Manual segmentation provides ground truth for brain structure volumes⁹⁰. We obtained initial cerebellar masks semi-automatically, subtracting cerebral masks from whole brain masks with StereotaxicRAMON (github.com/neuroanatomy/StereotaxicRAMON) or using Thresholdmann (github.com/neuroanatomy/thresholdmann) to interpolate between thresholds at manually selected locations. Initial cerebellar masks were uploaded to BrainBox⁹¹ for

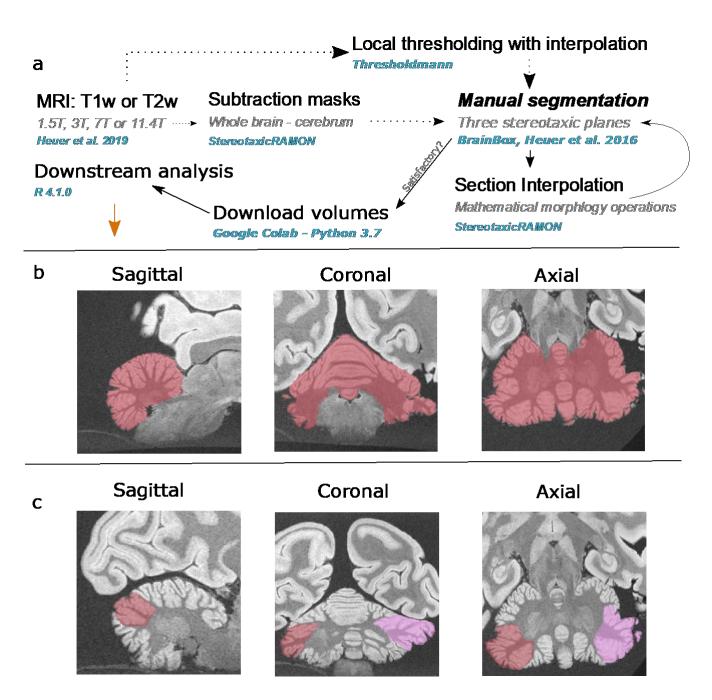


Figure 2: Manual segmentation method for primate brains. (a) A schematic representation of the manual segmentation pipeline. T1- and T2-weighed (T1w and T2w) MRI brain scans previously described in Heuer et al. (2019) were used to make initial cerebellar masks, by subtracting cerebral masks from whole brain masks with StereotaxicRAMON. Some initial cerebellar masks were made through local thresholding and interpolation with Thresholdmann. Cerebellar masks were then uploaded to BrainBox, where manual segmentation was performed. Manual segmentation was performed iteratively with interpolation between slices with mathematical morphology operations, until segmentations reached satisfactory quality. Volumes were then downloaded with a custom python script for subsequent analysis in R. (b-c) Example segmentations in stereotaxic planes for the cerebellum and ansiform area in the hamadryas baboon. (b) Cerebellar segmentations primarily involved removal of the brain stem, removing erroneously marked tissue and sulci, and reconstructing damaged tissue. (c) Ansiform area segmentations were slightly more challenging and additionally involved identification of superior posterior and ansoparamedian fissures, and segmentation between them. Segmentations were made so they did not enter vermal portions of lobule VII.

online collaborative segmentation of MRI images. Lastly, we used StereotaxicRAMON for mathematical morphology operations that preserve original mask topology (Figure 2a).

Cerebellum

Manual segmentations (Figure 2b) consisted of brainstem removal, cerebellar surface boundary determination, erroneously marked sulci erasure, and damaged tissue reconstruction. Segmentations were performed by NM, with SLV providing preliminary segmentations for a subset of specimens. Damaged tissue was reconstructed by carefully manually interpolating between non-damaged slices. Segmentations were performed to satisfaction using the three stereotaxic planes and alternating the manual segmentation procedure and mathematical morphology operations. All segmentations (N=65) were approved by KH and SLV.

Ansiform area

Segmentation (Figure 2c) relied on identification of superior posterior (SPF) and ansoparamedian (APMF) fissures^{13,92,93}. Since data quality was variable, segmentation was performed in a subset of specimens representative of the sample that allowed fissure identification with reasonable certainty: 5 apes (N=22; including 10 humans and 9 chimpanzees) and 8 non-apes. Segmentations were again iteratively performed to satisfaction. An interobserver strategy validated segmentations made by NM. For six species, a secondary observer (MA) provided blinded segmentations alongside NM. Additionally, a tertiary observer (VS) segmented the remaining human and chimpanzee ansiform areas in consultation with NM. All segmentations were checked by KH and SLV.

Reliability of segmentations

Reliability of ansiform area segmentations was assessed by ANOVA:

Human cerebellar and ansiform area manual segmentations were compared with automated cerebellar and lobular segmentations made using CERES⁹⁴ at volbrain.upv.es:

and

Ansiform area volume ~ method

Intraclass correlation (ICC) was additionally calculated⁹⁵. ICC of .75-.90 indicates good reliability and .90-1.00 excellent reliability⁹⁶.

Cerebellar volume was significantly related to method (Pr(>F)<.01). ICC was .26 (95% confidence interval (CI): -.38<ICC<.74). Inspection revealed that manual segmentations were on average 20.4 cm³ (16.2%) larger than CERES segmentations. Ansiform area volume was not related to method (Pr(>F)=.286), with ICC of .84 (95%CI: .51<ICC<.96). Manual volumes were only 2.7 cm³ (6.6%) larger. One specimen differed by 6.9 cm³ (18.2%). It represented an average-sized cerebellum, unlikely to alter the results significantly. Full results can be found in the GitHub repository.

Manual ansiform area volumes were significantly related to species (Pr(>F)<.001). No significant observer effect was detected (Pr(>F)=.468). ICC was .99 (95%CI: .92<ICC<1.00).

Neuroanatomical measurements

Phenotypes of interest included cerebellar, cerebral⁸⁵, and ansiform area volumes, and cerebellum-to-cerebrum and ansiform area-to-cerebellum ratios. Absolute measurements were log₁₀-transformed. Volumes were not corrected for shrinkage. Normality was examined for traits with multiple (>2) observations per species (Figure 1) by Shapiro-Wilk tests⁹⁷ and outliers were visualized on boxplots (±1.5 IQR from Q1 and Q3). This led to the exclusion from subsequent analyses of one rhesus and one crab-eating macaque (Supplemental Figure S1). Two different crab-eating and rhesus macaque specimens (Supplemental Figure S2) displayed cerebellum-to-cerebrum ratio outliers but were retained in analyses as cerebellar and cerebral volumes fell within the normal range.

Statistical analyses and reproducibility

Consensus phylogenetic trees for the 34- (Figure 1) and 13-species analyses were obtained from the 10kTrees⁶⁶ primate database (<u>10ktrees.nunn-lab.org</u>; version 3). They were constructed from 17 genes and 7 different loci⁶⁶ (<u>10ktrees.nunn-</u>

(3)

lab.org/downloads/10kTrees_Documentation.pdf).

We used R version R 4.1.0 for statistical analyses⁹⁸. Fit of extant phenotypes in the current and previous⁸⁵ studies were tested with three common evolutionary models of trait evolution using Rphylopars⁹⁹. Models included Brownian Motion (BM)¹⁰⁰, Ornstein-Uhlenbeck (OU)¹⁰¹ (with single alpha, alpha per phenotype, or full multivariate alpha matrix), and Early Burst (EB)102. BM describes traits varying randomly over time in direction and extent, leading to differences predictable by time since divergence¹⁰³. BM does not equal genetic drift but may represent weak selection within BM parameters or selection towards specific trait optima whose distribution is described by BM¹⁰⁴. OU processes behave like BM with a specific trait optimum, a phenotype with adaptive advantage^{105,106}. Lastly, EB processes¹⁰² – rapid change after adaptive radiations followed by comparative phenotypic stasis – may explain trait evolution. To assess phylogenetic signal within the data, we chi-squared tested BM with Pagel's λ =1.0 (full phylogenetic signal) versus λ =0.0 (star-model, no phylogenetic signal). Data fit was determined by Akaike information criterion (AIC)99,107, with lower values representing better fit. A difference of four to seven points represents significantly less support and a difference exceeding ten indicates virtually no support for the lower-performing model.

Ancestral character estimations (ACEs) were constructed by mapping extant trait values back in time following the best-supported evolutionary model, incorporating intraspecific variation. ACEs were mapped to consensus phylogenetic trees for visualization.

Extant primate traits are not statistically independent, but share variable amounts of evolutionary history¹⁰³. Substantial phylogenetic signal was detected, necessitating correction. Corroboratively, correlations between current and previously⁸⁵ recorded phenotypes illustrated severe collinearity (Figure 3). Hence, we performed regressions with phylogenetic generalized least squares (PGLS)^{103,108} implemented in *nlme*¹⁰⁹, which under BM¹⁰⁰ generalizes to phylogenetic independent contrast (PIC) regression forced through the origin¹¹⁰. PICs¹⁰³ were calculated with ape's¹¹¹ *pic.ortho* function, accounting for intraspecific variation. Regressions were performed for cerebellar volume regressed on cerebral volume for all species (1) and those with complete (including ansiform area) data (2), and ansiform area volume

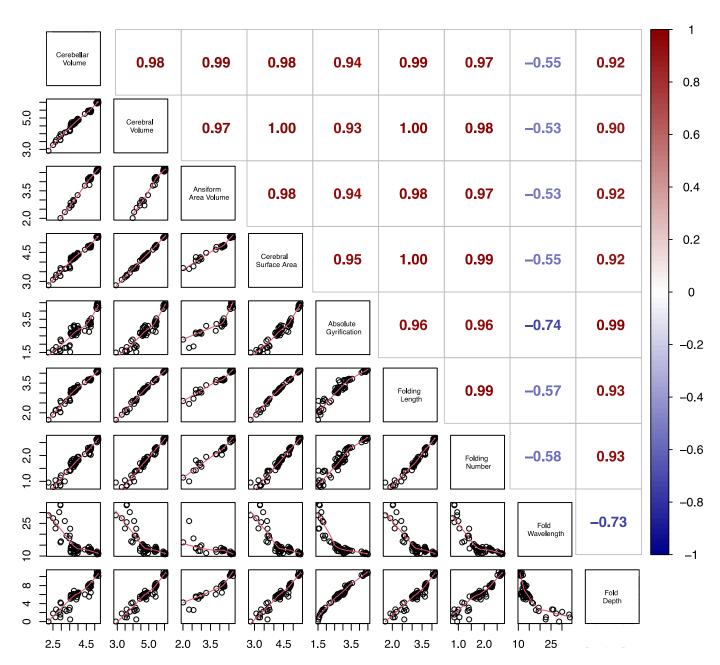


Figure 3: Correlations for neuroanatomical measurements. Volumes recorded in the current study were correlated with the measurements from **Heuer et al. (2019)**. The lower diagonal displays the correlations of log₁₀-transformed variables, which were not corrected for phylogeny and included all specimens. These correlations serve to illustrate the strong collinearity between the neuroanatomical variables, which are an expected outcome of allometric scaling. Corresponding R² coefficients for the correlations can be found on the upper diagonal. Cerebellar, cerebral, and ansiform area volumes correlated strongly and positively with all other variables, except fold wavelength, for which a negative correlation was observed.

regressed on rest of cerebellar volume (3) and cerebral volume (4), all taking species medians. ANOVA assessed if apes and non-apes showed different scaling in all regressions. We calculated R²_{likelihood}^{112,113} for full regressions, and apes or non-apes

separately. Fisher's R-to-Z transformation (<u>cran.r-project.org/web/packages/psych/index.html</u>) determined improvement of fit.

Lastly, we repeated analyses in the Stephan collection⁸⁴. Cerebellar and cerebral volumes were matched to primates in 10kTrees⁶⁶. This led to a partially overlapping (in species, not specimens) dataset of 34 species.

Results

Neuroanatomical measures

Median neuroanatomical measures are provided in Table 1. Intraspecific data spread is given as median absolute deviations (MADs). Cerebella and ansiform areas were largest in apes, with human volumes being two to three times larger than in any other species. Cerebellum-to-cerebrum ratios were relatively consistent within clades, but variable across them. They were comparatively high in lemurs and apes. Chimpanzee cerebella occupied a range of ratios comparable to other apes (median=18.36%; MAD=3.56), whereas human ratios were relatively low (median=14.26%; MAD=1.75). Ansiform area-to-cerebellum ratios were highest in great apes. Humans and chimpanzees displayed the highest ratios (30.76% and 27.25%). Several species, including the tufted capuchin (Cebus apella), hamadryas baboon (Papio hamadryas), and green monkey (Chlorocebus sabaeus) had high ansiform area-to-cerebellum ratios relative to cerebellum-to-cerebrum ratios. Other species including the aye aye (Daubentonia madagascariensis) and bonobo (Pan paniscus) showed the opposite relationship, suggesting that scaling of these traits may be partly dissociated.

Intraspecies variability is substantial

For several species, we analyzed intraspecific variability. Variability, quantified as median absolute deviations (MAD) as percentage of the median, ranged from 4.8 to 15.8% for cerebellar volume, and from 6.2 to 20.7% for cerebral volume. Intraspecific ansiform area observations were only available for humans (median:43.73±5.00 cm³; MAD=11.4%) and chimpanzees (median:15.54±1.93 cm³; 12.4%). Intraspecific variation in cerebellum-to-cerebrum ratio ranged from 1.7 to 20.7% of the median. Range was less substantial for ansiform area-to-cerebellum ratios for humans (median:30.76±1.91%; 6.2%) than for chimpanzees (median:27.25±3.69%; 13.5%). Volume and ratio varied relatively independently across species. Ordered bar graphs

Table 1: Neuroanatomical measurements. Species median measurements for previously reported cerebral volumes **Heuer et al. (2019)** are reported alongside cerebellar and ansiform area volumes (in mm³). Species ratios between median cerebellar and cerebral volumes, and ansiform area and cerebellar volumes are also given (in percentages). Species are ordered by the phylogenetic tree. For six species (*Ateles paniscus*, *Homo sapiens*, *Macaca fascicularis*, *Macaca mulatta*, *Pan troglodytes*, and *Saimiri sciureus*) several specimens were available. For these species, median absolute deviations are also reported. Some specimens were recorded as outliers and removed before subsequent analyses. These include a crab-eating and rhesus macaque, which were outliers in their cerebellar and cerebral volumes, as marked by the asterisks. Gal. = Galagonidae; Lor. = Loridae; Cerco. = Cercopithecini.

	Species	Cerebellar volume	Cerebral volume	Ansiform area volume	Cerebellum- cerebrum ratio	Ansiform area- cerebellum ratio
3	Demidoff's galago (1)	320	1,925		16.65	
9	Red slender loris (1)	447	3,071		14.54	
	Aye-aye (1)	3,032	17,151	374	17.68	12.33
	Black-and-white ruffed lemur (1)	3,377	16,032		21.06	
_	Ring-tailed lemur (1)	2,035	12,658		16.08	
Cellinationnes	Mongoose lemur (1)	3,224	14,495		22.24	
2	Red-tailed sportive lemur (1)	882	4,087		21.59	
	Coquerel's mouse lemur (1)	884	3,885		22.75	
í	Grey mouse lemur (1)	170	809		20.99	
	Squirrel monkey (2)	920 ± 55	8,540 ± 1,263	103	10.93 ± 2.26	11.20
	Tufted capuchin (1)	5,191	36,285	1,012	14.31	19.50
	White-faced sapajou (1)	4,274	4,005		10.67	
200	Cotton-top tamarin (1)	750	6,254		11.99	
n non	Black-pencilled marmoset (1)	485	3,938		12.31	
	Douroucouli (1)	1,627	11,746	217	13.85	13.31
	Wooly monkey (1)	5,595	49,385	874	11.33	15.62
	Black spider monkey (2)	8,903 ± 1,411	77,381 ± 10,948		11.49 ± 0.20	
1	Orangutan (1)	25,757	184,458	6,081	13.96	23.61
	Chimpanzee (9)	57,047 ± 6,915	284,482 ± 26,063	15,544 ± 1,932	18.36 ± 3.56	27.25 ± 3.69
	Bonobo (1)	48,408	243,036	6,627	19.92	13.69
99	Human (10)	142,171 ± 18,710	1,005,711 ± 179,368	43,726 ± 5,003	14.26 ± 1.75	30.76 ± 1.91
201111111111111111111111111111111111111	Western lowland gorilla (1)	58,402	237,690	13,107	24.57	22.44
	Eastern gorilla (1)	46,163	272,908		16.92	
	Gibbon (1)	8,681	60,685		14.31	
ì	Indochinese lutung (1)	3,756	37,988		9.89	
	Hanuman langur (1)	6,749	49,916		13.52	
3	King colobus (1)	5,787	40,468	842	14.30	14.55
	Rhesus macaque (6)*	10,456* ± 500	75,928* ± 4,689		13.54 ± 0.64	
	Crab-eating macaque (8)*	6,184* ± 955	46,286* ± 9,597		12.89 ± 0.98	
	Hamadryas baboon (1)	9,478	89,861	1,841	10.55	19.42
-	Grey-cheeked mangabey (1)	6,320	49,185		12.85	
Papionin	Sooty mangabey (1)	5,362	51,689		10.37	
Cerco.	Green monkey (1) Moustached guenon (1)	4,169 3,697	53,627 41,538	505	7.77 8.90	12.11

for phenotypes can be found in Supplemental Figure S3.

Evolutionary model testing

AIC ranking¹¹⁴ revealed the following support for evolutionary models: Brownian Motion (BM) (λ =1.0; AIC=6209.62) > Early Burst (EB) (AIC=6216.25) >> BM (λ =0.0 (star phylogeny); AIC=6267.87) >> Ornstein-Uhlenbeck (OU) (single α ; AIC=6609.29) >> OU (α per phenotype; AIC=7143.13) >> OU (full multivariate α ; AIC=8063.37). The BM was best-supported, and significantly more so than EB. Chi-squared testing revealed strong support for the λ =1.0 versus the λ =0.0 model (χ ²=53.68). We adopted BM for subsequent analyses.

Ancestral character estimations

We provided ancestral volumes for cerebellar, cerebral, and ansiform area volumes using BM (Supplemental Table S1). Full ancestral character estimations (ACEs) including those reported for neocortical measures⁸⁵ can be found in the GitHub repository.

Relative cerebello-cerebral traits differ across clades

We mapped ACEs to the phylogenetic tree, illustrating evolutionary dynamics of the cerebello-cerebral system. We plotted 95% confidence intervals for ACEs onto the trees (Figure 4), with uncertainty intuitively increasing with time to present (Supplemental Figure S4). Cerebellar and cerebral volumes showed virtually identical evolutionary dynamics (Figure 4a,b), with largest volumes in apes. Cerebellar volume at the ancestral node of the 34-species tree was estimated at 1856mm³, resembling the ring-tailed lemur in our data. When we mapped the cerebello-cerebral ratio (4c), relative cerebellar volume was increased in several clades specifically: Hominoidea, and Strepsirrhini clades including Galagonidae, Loridae, and Lemuriformes (Figure 1).

Similarly, ansiform area volumes (4d) showed largest values in great apes as expected due to larger body size. Ansiform area-to-cerebellum ratios mapped to the tree (4e) were more interesting. In contrast to cerebellum-to-cerebrum ratios (4c), ansiform area-to-cerebellum ratios were highest in great apes, but not in the one Strepsirrhini species we measured (a lemur: aye aye; *Daubentonia madagascariensis*). Within great apes, humans (30.76%) and chimpanzees (27.25%) had the highest ratios. The tufted capuchin (*Cebus apella*; 19.50%) showed a high

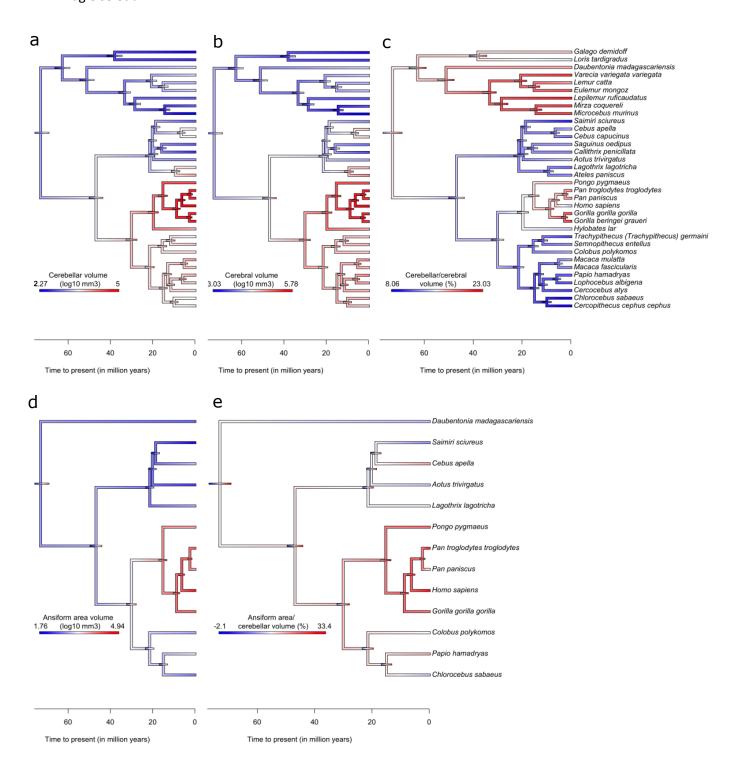


Figure 4: Ancestral character estimations for neuroanatomical phenotypes. Ancestral character estimations (ACEs) based on the Brownian Motion model of trait evolution are provided for absolute (a, b, d) and relative (c, e) volumes, alongside nodewise confidence intervals (colored bars on tree nodes). ACEs for cerebellar (a), cerebral (b), and relative cerebellar-to-cerebral (c) volumes were calculated from and mapped to the full 34-species tree, whereas ansiform area (d) and relative ansiform area-to-cerebellar (e) volumes were calculated from and mapped to the 13-species tree. The ancestral node was estimated at 73 million years to present.

ratio for its phylogenetic position (versus the squirrel monkey; *Saimiri sciureus*; 10.93%±2.26). For all ancestral character estimations, we provide additional

visualization in Supplemental Figure S5, which highlights the relationship between trait values and evolutionary time more directly.

Allometric scaling relationships

Accelerated ansiform area scaling in primates

We employed PGLS analysis in the context of BM evolution (Figure 5). Median cerebellar volumes were regressed on median cerebral volume for full 34-species data (5a) and for the 13 species with complete data (5b). The scaling relationships were slightly below 1:1 in both cases (slope₃₄=.955 and slope₁₃=.940). Although trending towards hypo-allometry, cerebellar and cerebral volumes evolved isometrically. Ansiform area volume scaled hyper-allometrically to rest of cerebellar (ROC; 5c) and cerebral (5d) volumes (slope_{ROC}=1.297; slope_{cerebrum}=1.245), with lower intercept for cerebrum (intercept_{cerebrum}=-2.784) than for ROC (intercept_{ROC}=-1.833). Together, the ansiform area scaled hyper-allometrically to both ROC and cerebral volume.

No distinct allometries in apes

We split the data into ape and non-ape subsets for cerebellum (7 apes (N=24); 27 non-apes (N=39)) and ansiform area (5 apes (N=22); 8 non-apes). We did not find significant interaction with ape membership for cerebellum regressed on cerebrum: F(Df=1, Df_{denom.}=30)=1.17, p=.288. Fisher's R-to-Z transformation indicated significantly worse fit for apes (r=.75, Z=4.37, p<.0001.) and non-apes (r=.87, Z=3.00, p<.01) versus the full model (r=.97). Evidence for difference between groups became even weaker in the 13-species data: F(Df=1, Dfdenom.=9)=.52, p=.49. The full model again better described the data (r=.94) than apes (r=.43, Z=2.86, p<.01) or non-apes (r=.76, Z=1.70, p=.089) separately. We also found no support in the ansiform area regressions on rest of cerebellar (F(Df=1, Df_{denom.}=9)=1.87, p=.204), or cerebral volume (F(Df=1, Df_{denom.}=9)=.045, p=.84). The full model outperformed partial models for cerebellar regression (full: r=.85; ape: r=.61, Z=1.19, p=.23; non-ape: r=.90, Z=.53, p=.596) and cerebral regression (full: r=.87; ape: r=.56, Z=1.57, p=.116; non-ape: r=.68, Z=1.14, p=.254). Models fit better in non-apes than in apes for rest of cerebellar (rapes=.61, rnon-apes=.90, Z=1.72 p=.085) and cerebral regressions (rapes=.56, rnonapes=.68, Z=.43, p=.667), with only a trend towards significance observed for the cerebellar regression.

Considering this primate-general scaling, we explored if ratio differences within

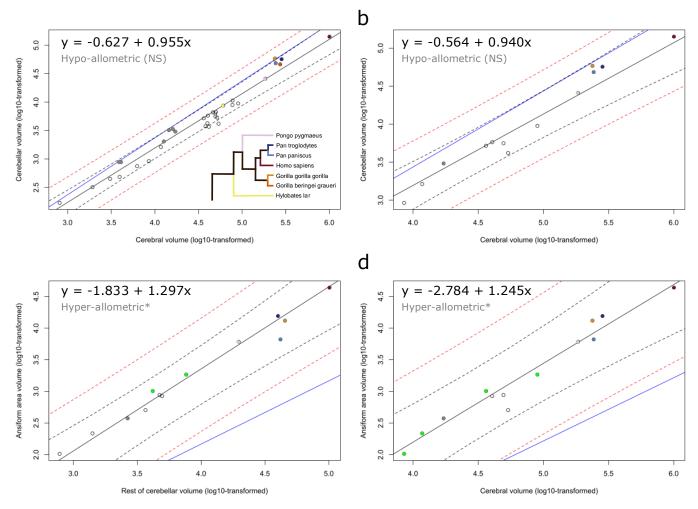


Figure 5: Allometric scaling relationships of cerebellar and ansiform area volumes. Phylogenetic generalized least squares (PGLS) regressions for cerebellar volume regressed on cerebral volume (a,b), and ansiform area regressed on rest of cerebellar and cerebral volumes (c,d). Median volumetric data were taken from 34-species (a) and 13-species (b-d) data. Log₁₀-transformed neuroanatomical measures were plotted and overlaid with regression lines obtained in respective PGLS models. 95% confidence (black dotted line) and prediction (red dotted line) intervals are provided alongside a fictive isometric scaling relationship with the same intercept (blue line). Exclusion of the isometric scaling relationship from the confidence intervals was taken to indicate significant allometry, as indicated by the asterisks. (a,b) Cerebellar volumes regressed on cerebral volumes for full data (a) and for the 13 species with complete data (b) both illustrate isometric scaling trending towards hypo-allometry. Lemuriformes and Hominidea were the two clades with most impressive cerebellar-to-cerebral volume ratios (Figure 4c) and were thus specifically colored here (Lemuriformes colored in a bold gray; a-d) to highlight their phylogenetic scaling relationships. All species belonging to these infraorders displayed higher cerebellum-to-cerebrum scaling than the primate sample as a whole (a). Zooming in, (a) illustrates that while most Hominoidea, including Homo sapiens, Pongo pygmaeus, and Hylobates lar fall on the regression line of the sample, all but one of the Lemuriformes (Lemur catta; on the line) exceed the general trend. Additionally, in both clades, several species approach isometry, with one member of each falling on the isometric line (Gorilla gorilla gorilla and Mirza coquereli). (b) Illustrates shallowing of the PGLS slope, connected to the smaller sample of volumes. (c,d) Ansiform area volume regressed on rest of cerebellar volume (c) and cerebral volume (d) illustrate hyper-allometric scaling relationships in both cases. Because of the strong positive allometry, species with larger cerebella and cerebra are expected to have larger relative ansiform areas. (c,d) Both illustrate that given this observed scaling, Hominoidea do not have unique hyper-scaling of the ansiform area necessarily. Although some Hominoidea lie above this steep regression line, so do several other species (as colored in green; Mirza coquereli, Aotus trivirgatus, Cebus apella, and Papio hamadryas). NS = nonsignificant.

and between clades (Figure 4) could be driven by body size. Linear regressions on body mass¹¹⁵ revealed association with ansiform area-to-cerebellum ratios ($R^2_{adj.}$ =.44; p<.01) (Supplemental Figure S6a) but not cerebellum-to-cerebrum ratios ($R^2_{adj.}$ =-.03; p=.88) (S6b).

Replication in the Stephan dataset

BM was again the best-supported model (AIC=-19.13) \approx EB (AIC=-17.51) \approx OU (diagonal α ; AIC=-16.03). It significantly outperformed the star-model (χ^2 =84.60). Pagel's λ likelihood distributions showed higher likelihoods for higher λ . We adopted BM to map the Stephan 34-species ancestral character estimations (ACEs) to the tree. Cerebellar and cerebral volume ACEs displayed only slightly larger spread and cladewise distribution was similar. Cerebellum-to-cerebrum ratio evolved similarly as in the main analysis. A notable exception was that Hominoidea had relatively less impressive ratios. Here, relatively large cerebella appeared to be a distinct Strepsirrhini trait (Supplementary Figure S7a). PGLS for cerebellar volume regressed on cerebral volume indicated significant hypo-allometry, with its slope of approximately .92 slightly shallower than in the main analysis (S7b).

Discussion

Overview

In the current work, we studied cerebellar volumes for 34 primate species (N=63) and ansiform area volumes for 13 species (N=30). The cerebellum, and the ansiform area specifically, may be part of important cerebello-cerebral loops and networks contributing to associate functions^{7,10,13,16,20–23,42,43,50,57,58}. Manual segmentations of the cerebellum and ansiform area were reliable as shown by inter-observer reliability, and comparison with automated⁹⁴ segmentations. The cerebellum scaled isometrically versus the cerebrum in PGLS (coefficient: ~.95). Replication in the Stephan data⁸⁴ argued that the primate cerebellum may in fact scale hypo-allometrically versus the cerebrum (~.92). Conversely, the ansiform area displayed strong positive allometry versus rest of cerebellar (~1.29) and cerebral (~1.25) volumes. Given this scaling across the primate sample, species with larger cerebella and cerebra i.e., hominoids, are expected to have larger relative ansiform areas. Therefore, although we find the largest relative ansiform areas in humans (30.76%) and chimpanzees (27.25%), these

ratios are expected from primate-general scaling rules, as illustrated by explorative body mass correlations. We did not find statistical support for distinct allometries in apes.

Together, our findings show that while the cerebellum and cerebrum scale isometrically in primates^{50,51,59}, the ansiform area expands relative to the system in a modular and accelerated fashion. Our findings further previous literature, showing that although the ansiform area has become relatively large in humans, this may result from general primate brain architecture and scaling rules.

Differences between manual and automated segmentations

Manual segmentations were globally similar to automated segmentations in CERES⁹⁴. However, mean manual volumes were systematically higher for the cerebellum. Visual inspection by NM, KH & SLV confirmed accuracy of segmentations. We observed two main differences. First, manual segmentation identified the edges of the cerebellum more exhaustively, whereas automated segmentation was too conservative. Secondly, manual segmentation may have included white matter at the pontine-cerebellar border to separate these structures similarly across all species. Universal guidelines for manual segmentation might help decrease differences, essential for relatively small-sized comparative studies.

Relative volumes provide advantages

Body mass is strongly coupled to brain size across distant classes such as reptiles, birds¹¹⁶, and mammals¹¹⁷ including primates^{46,69}. The relation between (relative) brain size and intelligence 'remains one of the thorniest issues in comparative neurobiology'¹¹⁶, and simply regressing out body mass may lose important signal. We know that humans have large brains for their bodies⁴⁶. Principal component analysis of cerebello-cerebral phenotypes across mammals separates humans from even primates, a separation that is greatly diminished when body mass is taken out of analysis¹¹⁸. We instead believe focusing on reorganization of brain structures themselves may be more revealing for relative importance of brain structures^{60,119,120}. Relative volumes provide conceptual and methodological advantages, illustrated by remapping studies that map specific functional areas against a reference area^{119,120}. It removes some systematic differences across datasets, species, and specimens, such as acquisition- and shrinkage-related biases

(although not tissue-specific shrinkage)^{36,84}. Additionally, previously used^{70,84,115,121} compilations report body mass – which is a highly volatile trait – for only a low number of unrelated specimens. Altogether, we did not regress out body mass in analyses.

The cerebello-cerebral system and Brownian Motion

Our analyses of cerebellar-cerebral system evolution showed strongest support for the Brownian Motion (BM) model. This is in line with previous findings for the evolution of neocortical phenotypes⁸⁵ and with our additional analyses of data from the Stephan collection⁸⁴. High estimates of Pagel's λ^{122} also supported strong phylogenetic signal. BM argues for trait evolution that is random in extent and direction over time. Such a model of incremental changes, predictable by time since divergence, fits with constrained scaling of the cerebello-cerebral system observed in the current study. At first glance, it clashes with notions of adaptive evolution, as departures from BM have been interpreted to indicate adaptive variation 102,123,124. The two are not mutually exclusive, as BM models can be supported if selection to specific trait optima is distributed following BM parameters¹⁰⁴. Recently, generalization of the BM model has been shown to represent both neutral drift and rapid adaptive evolutionary change, the latter of which BM does not do well¹²⁵. It remains open whether homogeneous models (a single regimen per tree, with useful statistical properties¹⁰⁴) or heterogeneous models (different regimes on subtrees) will best elucidate primate evolutionary complexity¹²⁵.

The cerebellum and cerebrum scale isometrically

Our study supports primate isometric cerebello-cerebral^{50,51,89,118,126} evolution, tending towards hypo-allometry. A recent study reported similar and significant hypo-allometric cerebello-cerebral scaling across mammals¹¹⁸. When we reran PGLS in the Stephan sample, we observed the same cerebello-cerebral scaling coefficient as this study did (.92)¹¹⁸. We found no support for distinct scaling patterns in apes.

Lemurs and apes displayed the highest cerebellum-to-cerebrum ratios. Corroboratively, lemurs may have large brains for their bodies¹¹⁸, a trait shared with hominoids and especially humans. High relative volumes (Figure 4c) and positive deviations above the slope of the primate dataset (Figure 5a and Supplemental Figure S6b) evidence relatively large lemur and hominoid cerebella. We were unable to detect

statistical support for unique ape scaling and did not test lemur uniqueness to prevent hypothesizing after results are known (HARKing)¹²⁷. Although ratios vary markedly across and within clades, median species traits can be closely predicted by general scaling rules. True uniqueness may be found as deviation from this scaling, potentially regressing out appropriately sampled body mass data.

Previous work assessing branch-wise evolutionary rates through the Bayesian reversible jump variable rates model shows that cerebella scale significantly more rapidly than neocortices in great apes⁷⁰. Our approach, examining ancestral characters based on the Brownian Motion model, shows that relatively high cerebellar-to-cerebral scaling may characterize hominoid evolution, with the notable exception of humans. We did not find greater cerebellar scaling versus the greatly expanded neocortex ^{89,128}. Accelerated cerebellar volumetric evolution⁷⁰ in great apes may be a response to cerebral expansion. Previous findings of relatively large cerebella – versus what is expected – in contemporary humans and apes, and much less volumetrically pronounced cerebella in recent human ancestors, support this theory¹²⁹, although our data do not. Larger ape samples may shed light on these discrepancies. Alternatively, expansion of the lateral cerebellum may have been a hominoid adaptation⁷², with disproportionate neocortical-to-whole brain expansion⁸⁹ in the lineage leading to humans. A study examining cerebellar⁷⁰ (and ansiform area) evolutionary rates in the current data may be warranted.

Primate ansiform areas scale hyper-allometrically

The ansiform area scaled to the rest of cerebellum at an approximate 1.3:1 ratio, without unique scaling in apes. This indicates that ansiform area hyperscaling may be a general feature within the examined primates. We offer the cautionary note that we were only able to examine one Strepsirrhini species and could not preclude that the trend may be Haplorhini-specific. Humans and chimpanzees, or hominoids in general, did not differ from allometry more extensively than the supposed lower primates. The strong positive allometry may directly account for high relative ansiform area volumes found in chimpanzees and humans previously 13,130. Humans may have unique cognitive abilities and exceptional relative brain size 46, but evolution has not acted to make humans exceptional (the *scale naturæ*). General primate scaling rules logically lead to great trait values and emergent abilities in large-brained humans, supported by increased neuron and connection numbers 131. Species-specific specializations and

reorganizations can occur at any stage of evolution to compound these general changes 132,133.

Developing structures may become disproportionally large because conserved neurodevelopmental mechanisms scaffold the development of the brain¹³⁴. Small timing differences of cell division cycles may explain the hyperscaling of (neo)cortical surface area in primates¹³⁵ and deviations from isometric cerebello-cerebral scaling observed across mammals⁵⁸ including primates^{50,51,53,74}, as well as birds⁵⁷. Primategeneral hyperscaling across structures is a common occurrence: predictable relative increases in neocortical volume⁸⁹, white-to-grey matter scaling¹³⁶, cerebellar-to-cerebral neuron numbers⁵³, and prefrontal volume^{137,138} have been reported. Hyperscaling of cerebral cortices occurs in simian monkeys¹³⁹ and between macaques and humans⁷⁴ similarly. Truly disproportionate increases within primates may be rare and much of brain organization may be subject to developmental⁵⁹ or functional⁶⁵ constraints. Simultaneously, reorganization has consistently occurred with primate brains generally scaling up – apparently functionally important – connected components. Cognitive modules of the cerebello-cerebral system are primary suspects of such adaptive evolution^{7,12,13,140}.

Small sample sizes warrant caution

Range of absolute and relative volumes across even the 13-species data was substantial, as expected^{80–82}. The ansiform area was observed to consistently take up a disproportionately high percentage of the cerebellum in humans and chimpanzees, fitting well with the 1.3:1 scaling rule. Our data generally resemble prior observations for mean ansiform area volumes in humans (current study: 43.94 cm³; Balsters et al.: 53.65 cm³; Makris et al.: 43.01 cm³), intraspecific variability (5.00 cm³; 8.01 cm³; 6.38cm³), and relative ansiform area-to-cerebellum ratio (30.76%; 36.51%; 29.58%)^{13,130}, but illustrate how intraspecific variability cannot be ignored. Reanalysis of ansiform area scaling in larger datasets will be necessary to confirm our observations as they may be partially driven by an underpowered sample and influenced by specimen-specific idiosyncrasies^{83,141,142}.

Community-wide sharing accelerates comparative neurosciences

Reanalysis in the Stephan dataset⁸⁴ offered the following: 1) there is potential for sizable intraspecific samples; 2) but integrating datasets comes with great challenges

due to methodological differences, incomplete descriptions, lack of individual observations, or missing provenances; and 3) intraspecific variability is substantial and likely one of the primary drivers of spurious results^{80–83,141}. Notable discrepancies in volumetric data between the current study and previous datasets^{13,72,84,126,143} underline the pressing need for intraspecific data in comparative analyses.

Community-wide guidelines for data collection and statistical handling need to be created to facilitate integration of data from primate studies^{7,144–146}. Data must be shared openly and completely. Most studies^{9,50,70,72,126,143} contain data provenance but studies describing specimen-specific observations are still rare⁷². Online databases and catalogs for primate brain data – such as PRIMatE Data Exchange (PRIME-DE)¹⁴⁷ and BrainBox⁹¹ – can be used to earmark all primate brains with a unique identification number, linked to relevant metadata including sex, age, body weight, captivity status, data availability, and inclusion in specific studies. This would help interpret contradictory results¹⁴¹, and test their statistical significance^{148,149}. Access to individual observations allows incorporation of intraspecific variability. There is a pressing need to incorporate this statistical uncertainty^{80,83}, and statistical methods to do this exist^{99,111,121}.

Summary and outlook

We report cerebellar volumetric data for 34 species (N=63) and ansiform areas (crura I-II) for 13 species (N=30). Our phylogenetic analyses provide corroborating evidence for isometry between cerebellar and cerebral volumes^{50,51,118} and evidence primategeneral ansiform area hyperscaling^{13,130}. A combination of constrained⁵⁹ and modular⁶⁴ evolution may shape the primate cerebello-cerebral system. The consistent finding of cerebello-cerebral coevolution argues for their structure and function being intimately linked. The strong positive allometry of the ansiform area in primates, potentially alongside that of structurally connected cerebral association cortices^{74,75,137,138}, shows detachment from isometric cerebello-cerebral scaling. Exploring associations between ansiform area volume and cognitive ability in relation to intraspecific variability, development, and primate evolution is of prime interest. Larger comparative datasets are necessary: exchange platforms have been initiated. Such datasets in combination with relevant metadata and sizable intraspecific samples will facilitate evolutionary analyses sensitive to intraspecific variability.

The primate cerebello-cerebral system is a strong functional neuroanatomical scaffold⁷. Within it, the ansiform area is characterized by strong positive allometry^{13,130}, unique functional activations^{14,15} and involvement in functional networks^{16–18,42,43}. It furthermore has a distinct developmental trajectory¹⁵⁰ and structural properties^{150,151}, and a characteristic role in disorders⁷. Concludingly, the ansiform area should be a primary area of interest for the study of human and comparative cognitive correlates in the future⁷.

Declarations

Author contributions

NM. Conceptualized the manuscript; wrote and adapted custom computer code; performed manual segmentations; performed analyses; wrote the manuscript; incorporated coauthor revisions. **RT.** Provided MRI data; wrote original custom code; wrote used software; provided revision comments manuscript. **VS and MA** provided manual segmentations of ansiform areas for 12 and 6 specimens, respectively. **SBE.** Provided revision comments manuscript; provided funding for the project. **KH*.** Provided MRI data; provided semi-automated segmentation and software support; wrote original custom code; wrote used software; provided revision comments manuscript; provided supervision. **SLV*.** Conceptualized the manuscript; provided a subset of primary segmentations; provided revision comments on manual segmentations, the manuscript, and computer code (multiple occasions); provided supervision; provided funding for the project.

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Competing interests

The authors have no conflicts of interest to declare.

Data availability

Data obtained in the current project, including volumes and ratios, but also

intermediate text files and figures, are uploaded to the repository accompanying this project: github.com/NevMagi/34primates-evo-cerebellum. Input files necessary to replicate our results can also be found here, alongside full-size figures to aid legibility. MRI data have been collated from different sources, and published Open Access. The Stephan collection (DOI: 10.1159/000155963) was not published Open Access. Therefore, access to the data was requested by the authors through the Copy Clearance Center Marketplace and facilitated through the Central Library of Forschungszentrum Jülich GmbH. Reuse permission for these data may be requested from the publisher.

Code availability

Custom computer code was used at all stages of analysis. Some of the scripts used are adapted scripts⁸⁵. Full custom code can be obtained from the GitHub repository accompanying this project: github.com/NevMagi/34primates-evo-cerebellum. Full accreditation of R software used in the project can be found in Supplemental Table \$2.

References

- Van Overwalle, F. et al. Consensus Paper: Cerebellum and Social Cognition. The Cerebellum 19, 833–868 (2020).
- Koziol, L. F. et al. Consensus paper: The cerebellum's role in movement and cognition. The Cerebellum 13, 151–177 (2014).
- Mariën, P. et al. Consensus paper: Language and the cerebellum: An ongoing enigma.
 Cerebellum 13, 386–410 (2014).
- 4. Adamaszek, M. *et al.* Consensus Paper: Cerebellum and Emotion. *Cerebellum* **16**, 552–576 (2017).
- 5. Botez, M. I., Gravel, J., Attig, E. & Vézina, J. L. Reversible chronic cerebellar ataxia after phenytoin intoxication: Possible role of cerebellum in cognitive thought. *Neurology* **35**, 1152–1157 (1985).
- 6. Schmahmann, J. D. & Sherman, J. C. The cerebellar cognitive affective syndrome. *Brain* 121,

561-79 (1998).

- 7. Magielse, N., Heuer, K., Toro, R., Schutter, D. J. L. G. & Valk, S. L. A comparative perspective on the cerebello-cerebral system and its link to cognition. *The Cerebellum* (2022) doi:10.1007/s12311-022-01495-0.
- 8. Smaers, J. B. Modeling the Evolution of the Cerebellum. From Macroevolution to Function.

 *Progress in Brain Research 210, 193–216 (2014).
- Smaers, J. B., Turner, A. H., Gómez-Robles, A. & Sherwood, C. C. A cerebellar substrate for cognition evolved multiple times independently in mammals. *eLife* 7, e35696 (2018).
- Shine, J. M. & Shine, R. Delegation to automaticity: The driving force for cognitive evolution?
 Frontiers in Neuroscience 8, 90 (2014).
- 11. Kelly, R. M. & Strick, P. L. Cerebellar loops with motor cortex and prefrontal cortex of a nonhuman primate. *Journal of Neuroscience* **23**, 8432–8444 (2003).
- 12. Ramnani, N. The primate cortico-cerebellar system: Anatomy and function. *Nature Reviews Neuroscience* **7**, 511–523 (2006).
- 13. Balsters, J. H. *et al.* Evolution of the cerebellar cortex: The selective expansion of prefrontal-projecting cerebellar lobules. *NeuroImage* **49**, 2045–2052 (2010).
- 14. Stoodley, C. J. & Schmahmann, J. D. Functional topography in the human cerebellum: A metaanalysis of neuroimaging studies. *NeuroImage* **44**, 489–501 (2009).
- 15. King, M., Hernandez-Castillo, C. R., Poldrack, R. A., Ivry, R. B. & Diedrichsen, J. Functional boundaries in the human cerebellum revealed by a multi-domain task battery. *Nature Neuroscience* **22**, 1371–1378 (2019).
- Xue, A. et al. The Detailed Organization of the Human Cerebellum Estimated by Intrinsic
 Functional Connectivity Within the Individual. *Journal of Neurophysiology* 125, 358–384 (2020).
- 17. Buckner, R. L., Krienen, F. M., Castellanos, A., Diaz, J. C. & Thomas Yeo, B. T. The organization of the human cerebellum estimated by intrinsic functional connectivity. *Journal of Neurophysiology* **106**, 2322–2345 (2011).

- 18. Habas, C. Functional Connectivity of the Cognitive Cerebellum. *Frontiers in Systems*Neuroscience **15**, 642225 (2021).
- Dum, R. P. & Strick, P. L. An unfolded map of the cerebellar dentate nucleus and its projections to the cerebral cortex. *Journal of Neurophysiology* 89, 634–639 (2003).
- 20. Ramnani, N. *et al.* The evolution of prefrontal inputs to the cortico-pontine system: Diffusion imaging evidence from macaque monkeys and humans. *Cerebral Cortex* **16**, 811–818 (2006).
- 21. Palesi, F. *et al.* Contralateral cortico-ponto-cerebellar pathways reconstruction in humans in vivo: Implications for reciprocal cerebro-cerebellar structural connectivity in motor and non-motor areas. *Scientific Reports* **7**, 1–13 (2017).
- 22. Palesi, F. *et al.* Contralateral cerebello-thalamo-cortical pathways with prominent involvement of associative areas in humans in vivo. *Brain Structure and Function* **220**, 3369–3384 (2015).
- 23. Strick, P. L., Dum, R. P. & Fiez, J. A. Cerebellum and Nonmotor Function. *Annual Review of Neuroscience* **32**, 413–434 (2009).
- 24. Leiner, H., Leiner, A. & Dow, R. Cerebro-cerebellar learning loops in apes and humans. *The Italian Journal of Neurological Sciences* **8**, 423–436 (1987).
- 25. Ito, M. Control of mental activities by internal models in the cerebellum. *Nat Rev Neurosci* **9**, 304–313 (2008).
- Eccles, J. C., Ito, M. & Szentágothai, J. The Cerebellum as a Neuronal Machine. The Cerebellum as a Neuronal Machine (Springer, 1967). doi:10.1007/978-3-662-13147-3.
- 27. Marr, D. & Poggio, T. From Understanding Computation to Understanding Neural Circuitry. *A.I. memo* **357**, 1–22 (1976).
- 28. Marr, D. Vision: A Computational Investigation of Visual Representation in Man. vol. 8 (Freeman and Company, 1982).
- 29. Diedrichsen, J., King, M., Hernandez-Castillo, C., Sereno, M. & Ivry, R. B. Universal Transform or Multiple Functionality? Understanding the Contribution of the Human Cerebellum across Task Domains. *Neuron* **102**, 918–928 (2019).

- 30. Schmahmann, J. D. Disorders of the cerebellum: Ataxia, dysmetria of thought, and the cerebellar cognitive affective syndrome. *Journal of Neuropsychiatry and Clinical Neurosciences*16, 367–378 (2004).
- 31. Schmahmann, J. D. From movement to thought: Anatomic substrates of the cerebellar contribution to cognitive processing. *Human Brain Mapping* **4**, 174–198 (1996).
- 32. Lu, X., Miyachi, S., Ito, Y., Nambu, A. & Takada, M. Topographic distribution of output neurons in cerebellar nuclei and cortex to somatotopic map of primary motor cortex. *Eur J Neurosci* **25**, 2374–2382 (2007).
- 33. Shambes, G. M., Gibson, J. M. & Welker, W. Fractured somatotopy in granule cell tactile areas of rat cerebellar hemispheres revealed by micromapping. *Brain, Behavior and Evolution* **15**, 94–140 (1978).
- 34. Ishikawa, T., Shimuta, M. & Häuser, M. Multimodal sensory integration in single cerebellar granule cells in vivo. *eLife* **4**, e12916 (2015).
- 35. Middleton, F. A. & Strick, P. L. Cerebellar projections to the prefrontal cortex of the primate. *Journal of Neuroscience* **21**, 700–712 (2001).
- 36. Luo, Y. *et al.* Lobular homology in cerebellar hemispheres of humans, non-human primates and rodents: a structural, axonal tracing and molecular expression analysis. *Brain Structure and Function* **222**, 2449–2472 (2017).
- 37. Sugihara, I. Crus I in the Rodent Cerebellum: Its Homology to Crus I and II in the Primate

 Cerebellum and Its Anatomical Uniqueness Among Neighboring Lobules. *The Cerebellum* 17,

 49–55 (2018).
- Wong, C. H. Y. et al. Fronto-cerebellar connectivity mediating cognitive processing speed.
 NeuroImage 226, 117556 (2021).
- 39. Krienen, F. M. & Buckner, R. L. Segregated fronto-cerebellar circuits revealed by intrinsic functional connectivity. *Cerebral cortex* **19**, 2485–2497 (2009).
- 40. Habas, C. et al. Distinct cerebellar contributions to intrinsic connectivity networks. The Journal

- of neuroscience: the official journal of the Society for Neuroscience 29, 8586–8594 (2009).
- 41. O'Reilly, J. X., Beckmann, C. F., Tomassini, V., Ramnani, N. & Johansen-Berg, H. Distinct and overlapping functional zones in the cerebellum defined by resting state functional connectivity.

 *Cerebral Cortex 20, 953–965 (2010).
- 42. Marek, S. *et al.* Spatial and Temporal Organization of the Individual Human Cerebellum.

 *Neuron 100, 977-993.e7 (2018).
- 43. Guell, X., Schmahmann, J. D., Gabrieli, J. D. E. & Ghosh, S. S. Functional gradients of the cerebellum. *eLife* **7**, e36652 (2018).
- 44. Wan, B. *et al.* Heritability and cross-species comparisons of human cortical functional organization asymmetry. *eLife* **11**, e77215 (2022).
- 45. Deaner, R. O., Isler, K., Burkart, J. & Van Schaik, C. Overall brain size, and not encephalization quotient, best predicts cognitive ability across non-human primates. *Brain, Behavior and Evolution* **70**, 115–124 (2007).
- 46. Jerison, H. J. Evolution of the brain and intelligence. (Academic Press, 1973).
- 47. Jerison, H. J. The theory of encephalization. *Annals of the New York Academy of Sciences* **299**, 146–160 (1977).
- 48. Smaers, J. B. & Vanier, D. R. Brain size expansion in primates and humans is explained by a selective modular expansion of the cortico-cerebellar system. *Cortex* **118**, 292–305 (2019).
- 49. Doya, K. What are the computations of the cerebellum, the basal ganglia and the cerebral cortex? *Neural Networks* **12**, 961–974 (1999).
- 50. Smaers, J. B., Steele, J. & Zilles, K. Modeling the evolution of cortico-cerebellar systems in primates. *Annals of the New York Academy of Sciences* **1225**, 176–190 (2011).
- 51. Whiting, B. A. & Barton, R. A. The evolution of the cortico-cerebellar complex in primates: anatomical connections predict patterns of correlated evolution. *Journal of Human Evolution* **44**, 3–10 (2003).
- 52. Herculano-Houzel, S. The remarkable, yet not extraordinary, human brain as a scaled-up

- primate brain and its associated cost. PNAS 109, 10661–10668 (2012).
- 53. Azevedo, F. A. C. *et al.* Equal numbers of neuronal and nonneuronal cells make the human brain an isometrically scaled-up primate brain. *Journal of Comparative Neurology* **513**, 532–541 (2009).
- 54. Van Horik, J. O., Clayton, N. S. & Emery, N. J. Convergent Evolution of Cognition in Corvids,

 Apes and Other Animals. in *The Oxford Handbook of Comparative Evolutionary Psychology*(eds. Vonk, J. & Shackleford, T.) 80–101 (University Press, 2012).
- 55. Highfill, L., Spencer, J. M., Fad, O. & Arnold, A. M. Performance on a means end task by Asian elephants (Elephas maximus) in a positive reinforcement-based protected contact setting.

 **International Journal of Comparative Psychology 29, 32061 (2016).
- 56. Highfill, L., Burns, M., Przystawik, K. & Vincent, J. Performance on a means-end task by African elephants (Loxondonta africana): A replication study. *International Journal of Comparative Psychology* (2018) doi:10.46867/ijcp.2018.31.00.04.
- 57. Gutiérrez-Ibáñez, C., Iwaniuk, A. N. & Wylie, D. R. Parrots have evolved a primate-like telencephalic-midbrain-cerebellar circuit. *Scientific Reports* **8**, 9960 (2018).
- 58. Muller, A. S. & Montgomery, S. H. Co-evolution of cerebral and cerebellar expansion in cetaceans. *Journal of Evolutionary Biology* **32**, 1418–1431 (2019).
- 59. Finlay, B. L. & Darlington, R. B. Linked regularities in the development and evolution of mammalian brains. *Science* **268**, 1578–1584 (1995).
- 60. Smaers, J. B. & Soligo, C. Brain reorganization, not relative brain size, primarily characterizes anthropoid brain evolution. *Proceedings of the Royal Society B: Biological Sciences* **280**, 20130269 (2013).
- 61. Marino, L. A Comparison of Encephalization between Odontocete Cetaceans and Anthropoid Primates. *Brain, Behavior and Evolution* **51**, 230–238 (1998).
- 62. Valk, S. L. *et al.* Shaping brain structure: Genetic and phylogenetic axes of macroscale organization of cortical thickness. *Science Advances* **6**, eabb3417 (2020).

- 63. Valk, S. L. *et al.* Genetic and phylogenetic uncoupling of structure and function in human transmodal cortex. *Nature Communications* **13**, 2341 (2022).
- 64. Barton, R. A. & Harvey, P. H. Mosaic evolution of brain structure in mammals. *Nature* **405**, 1055–1058 (2000).
- 65. Montgomery, S. H., Mundy, N. I. & Barton, R. A. Brain evolution and development: Adaptation, allometry and constraint. *Proceedings of the Royal Society B: Biological Sciences* **283**, 20160433 (2016).
- 66. Arnold, C., Matthews, L. J. & Nunn, C. L. The 10kTrees website: A new online resource for primate phylogeny. *Evolutionary Anthropology* **19**, 114–118 (2010).
- 67. Huang, C. H., Yu, X. & Liao, W. B. The Expensive-Tissue Hypothesis in Vertebrates: Gut Microbiota Effect, a Review. *International Journal of Molecular Sciences* **19**, 1792 (2018).
- 68. Isler, K. & van Schaik, C. P. The Expensive Brain: a framework for explaining evolutionary changes in brain size. *Journal of human evolution* **57**, 392–400 (2009).
- 69. Schoenemann, P. T. Evolution of the size and functional areas of the human brain. *Annual Review of Anthropology* **35**, 379–406 (2006).
- 70. Barton, R. A. & Venditti, C. Rapid evolution of the cerebellum in humans and other great apes.

 *Current Biology 24, 2440–2444 (2014).
- 71. Sereno, M. I. *et al.* The human cerebellum has almost 80% of the surface area of the neocortex.

 *Proceedings of the National Academy of Sciences of the United States of America 117, 19538–19543 (2020).
- 72. MacLeod, C. E., Zilles, K., Schleicher, A., Rilling, J. K. & Gibson, K. R. Expansion of the neocerebellum in Hominoidea. *Journal of Human Evolution* **44**, 401–429 (2003).
- 73. Sherwood, C. C., Bauernfeind, A. L., Bianchi, S., Raghanti, M. A. & Hof, P. R. Human brain evolution writ large and small. *Progress in brain research* **195**, 237–254 (2012).
- 74. Hill, J. et al. Similar patterns of cortical expansion during human development and evolution.

 Proceedings of the National Academy of Sciences of the United States of America 107, 13135—

- 13140 (2010).
- 75. Buckner, R. L. & Krienen, F. M. The evolution of distributed association networks in the human brain. *Trends Cogn. Sci.* **17**, 648–665 (2013).
- 76. Smaers, J. B., Steele, J., Case, C. R. & Amunts, K. Laterality and the evolution of the prefronto-cerebellar system in anthropoids. *Annals of the New York Academy of Sciences* **1288**, 59–69 (2013).
- 77. Rilling, J. K. Human and NonHuman Primate Brains: Are They Allometrically Scaled Versions of the Same Design? *Evolutionary Anthropology* **15**, 65–77 (2006).
- 78. Schoenemann, P. T., Budinger, T. F., Sarich, V. M. & Wang, W. S. Y. Brain size does not predict general cognitive ability within families. *Proceedings of the National Academy of Sciences of the United States of America* **97**, 4932–4937 (2000).
- 79. Giedd, J. N. *et al.* Child Psychiatry Branch of the National Institute of Mental Health

 Longitudinal Structural Magnetic Resonance Imaging Study of Human Brain Development. *Neuropsychopharmacology* **40**, 43–49 (2015).
- 80. Charvet, C. J., Darlington, R. B. & Finlay, B. L. Variation in Human Brains May Facilitate

 Evolutionary Change toward a Limited Range of Phenotypes. *Brain Behav Evol* 81, 74–85

 (2013).
- 81. Reardon, P. K. *et al.* Normative brain size variation and brain shape diversity in humans. *Science* **360**, 1222–1227 (2018).
- 82. Croxson, P. L., Forkel, S. J., Cerliani, L. & Thiebaut de Schotten, M. Structural Variability Across the Primate Brain: A Cross-Species Comparison. *Cerebral Cortex* **28**, 3829–3841 (2018).
- 83. Harmon, L. J. & Losos, J. B. The effect of intraspecific sample size on type I and type II error rates in comparative studies. *Evolution; international journal of organic evolution* **59**, 2705–10 (2005).
- 84. Stephan, H., Frahm, H. & Baron, G. New and revised data on volumes of brain structures in insectivores and primates. *Folia Primatologica* **35**, 1–29 (1981).

- 85. Heuer, K. *et al.* Evolution of neocortical folding: A phylogenetic comparative analysis of MRI from 34 primate species. *Cortex* **118**, 275–291 (2019).
- 86. Arbuckle, S. A., Diedrichsen, J. & Pruszynski, J. A. Non-human primate anatomicals. (2018) doi:10.5281/ZENODO.1319671.
- 87. Craddock, C. *et al.* The Neuro Bureau Preprocessing Initiative: open sharing of preprocessed neuroimaging data and derivatives. *Frontiers in Neuroinformatics* **7**, (2013).
- 88. Milham, M. P. *et al.* An Open Resource for Non-human Primate Imaging. *Neuron* **100**, 61-74.e2 (2018).
- 89. Rilling, J. K. & Insel, T. R. The primate neocortex in comparative perspective using magnetic resonance imaging. *Journal of Human Evolution* **37**, 191–223 (1999).
- 90. Morey, R. A. *et al.* A comparison of automated segmentation and manual tracing for quantifying hippocampal and amygdala volumes. *Neuroimage* **45**, 855–866 (2009).
- 91. Heuer, K., Ghosh, S., Robinson Sterling, A. & Toro, R. Open Neuroimaging Laboratory. *Research Ideas and Outcomes* **2**, e9113 (2016).
- 92. Faber, J. et al. Manual sub-segmentation of the cererbellum. 2022.05.09.22274814 Preprint at https://doi.org/10.1101/2022.05.09.22274814 (2022).
- 93. Larsell, O. & Jansen, J. *The Comparative Anatomy and Histology of the Cerebellum: Vol. 2. From Monotremes through Apes.* vol. 2 (University of Minnesota Press, 1970).
- 94. Romero, J. E. *et al.* CERES: A new cerebellum lobule segmentation method. *NeuroImage* **147**, 916–924 (2017).
- 95. Gamer, M., Lemon, J. & Singh, I. *irr: Various Coefficients of Interrater Reliability and Agreement*. (2010).
- 96. Koo, T. K. & Li, M. Y. A Guideline of Selecting and Reporting Intraclass Correlation Coefficients for Reliability Research. *Journal of Chiropractic Medicine* **15**, 155–163 (2016).
- 97. Shapiro, S. S. & Wilk, M. B. An Analysis of Variance Test for Normality (Complete Samples) on JSTOR. *Biometrika* **52**, 591–611 (1965).

- 98. R Core Team. R: A Language and Environment for Statistical Computing. (2021).
- 99. Goolsby, E. W., Bruggeman, J. & Ané, C. Rphylopars: fast multivariate phylogenetic comparative methods for missing data and within-species variation. *Methods in Ecology and Evolution* **8**, 22–27 (2017).
- 100. Freckleton, R. P., Harvey, P. H. & Pagel, M. Phylogenetic analysis and comparative data: a test and review of evidence. *Am Nat* **160**, 712–726 (2002).
- 101. Uhlenbeck, G. E. & Ornstein, L. S. On the Theory of the Brownian Motion. *Physical Review* **36**, 823–841 (1930).
- 102. Harmon, L. J. et al. Early bursts of body size and shape evolution are rare in comparative data.

 Evolution; international journal of organic evolution 64, 2385–2396 (2010).
- 103. Felsenstein, J. Phylogenies and the comparative method. Am. Nat 125, 1–15 (1985).
- 104. Harmon, L. Phylogenetic Comparative Methods: Learning From Trees. Preprint at https://doi.org/10.32942/OSF.IO/E3XNR (2019).
- Lande, R. Natural Selection and Random Genetic Drift in Phenotypic Evolution. *Evolution* 30, 314–334 (1976).
- 106. Hansen, T. F. Stabilizing Selection and the Comparative Analysis of Adaptation. *Evolution* 51, 1341–1351 (1997).
- 107. Akaike, H. A new look at the statistical model identification. *IEEE Transactions on Automatic Control* **19**, 716–723 (1974).
- 108. Symonds, M. R. E. & Blomberg, S. P. A primer on phylogenetic generalised least squares. in Modern Phylogenetic Comparative Methods and their Application in Evolutionary Biology 105– 130 (Springer Berlin Heidelberg, 2014). doi:10.1007/978-3-662-43550-2_5.
- 109. Pinheiro, J., Bates, D. & R Core Team. nlme: Linear and Nonlinear Mixed Effects Models. (2022).
- 110. Blomberg, S. P., Lefevre, J. G., Wells, J. A. & Waterhouse, M. Independent Contrasts and PGLS Regression Estimators Are Equivalent. Systematic Biology 61, 382–391 (2012).
- 111. Paradis, E. & Schliep, K. ape 5.0: an environment for modern phylogenetics and evolutionary

- analyses in R. Bioinformatics 35, 526-528 (2019).
- 112. Ives, A. R. R2s for Correlated Data: Phylogenetic Models, LMMs, and GLMMs. *Systematic Biology* **68**, 234–251 (2019).
- 113. Ives, A. & Li, D. rr2: An R package to calculate R2s for regression models. *Journal of Open Source Software* **3**, 1028 (2018).
- 114. Burnham, K. P. & Anderson, D. R. Multimodel inference: Understanding AIC and BIC in model selection. *Sociological Methods and Research* **33**, 261–304 (2004).
- 115. Isler, K. *et al.* Endocranial volumes of primate species: scaling analyses using a comprehensive and reliable data set. *Journal of Human Evolution* **55**, 967–978 (2008).
- 116. Font, E., García-Roa, R., Pincheira-Donoso, D. & Carazo, P. Rethinking the Effects of Body Size on the Study of Brain Size Evolution. *BBE* **93**, 182–195 (2019).
- 117. Schoenemann, P. T. Brain size scaling and body composition in mammals. *Brain Behav Evol* **63**, 47–60 (2004).
- 118. Heuer, K., Traut, N., Sousa, A. A. de, Valk, S. & Toro, R. Diversity and evolution of cerebellar folding in mammals. 2022.12.30.522292 Preprint at https://doi.org/10.1101/2022.12.30.522292 (2022).
- 119. Passingham, R. E. & Smaers, J. B. Is the Prefrontal Cortex Especially Enlarged in the Human Brain? Allometric Relations and Remapping Factors. *Brain, Behavior and Evolution* **84**, 156–166 (2014).
- 120. Halley, A. C. & Krubitzer, L. Not all cortical expansions are the same: the coevolution of the neocortex and the dorsal thalamus in mammals. *Current Opinion in Neurobiology* 56, 78–86 (2019).
- 121. Powell, L. E., Isler, K. & Barton, R. A. Re-evaluating the link between brain size and behavioural ecology in primates. *Proceedings of the Royal Society B: Biological Sciences* **284**, 20171765 (2017).
- 122. Pagel, M. Inferring the historical patterns of biological evolution. *Nature* **401**, 877–884 (1999).

- 123. O'Meara, B. C., Ané, C., Sanderson, M. J. & Wainwright, P. C. Testing for different rates of continuous trait evolution using likelihood. *Evolution* **60**, 922–933 (2006).
- 124. Harmon, L. J., Schulte, J. A., Larson, A. & Losos, J. B. Tempo and Mode of Evolutionary Radiation in Iguanian Lizards. *Science* **301**, 961–964 (2003).
- 125. Elliot, M. G. & Mooers, A. Ø. Inferring ancestral states without assuming neutrality or gradualism using a stable model of continuous character evolution. *BMC Evolutionary Biology* 14, 226 (2014).
- 126. Rilling, J. K. & Insel, T. R. Evolution of the Cerebellum in Primates: Differences in Relative

 Volume among Monkeys, Apes and Humans. *Brain, Behavior and Evolution* **52**, 308–314 (1998).
- 127. Kerr, N. L. HARKing: hypothesizing after the results are known. *Pers Soc Psychol Rev* **2**, 196–217 (1998).
- 128. Frahm, H. D., Stephan, H. & Stephan, M. Comparison of brain structure volumes in Insectivora and Primates. I. Neocortex. *Journal fur Hirnforschung* **23**, 375–389 (1982).
- 129. Weaver, A. H. Reciprocal evolution of the cerebellum and neocortex in fossil humans.

 *Proceedings of the National Academy of Sciences of the United States of America 102, 3576—3580 (2005).
- 130. Makris, N. *et al.* MRI-based surface-assisted parcellation of human cerebellar cortex: an anatomically specified method with estimate of reliability. *Neuroimage* **25**, 1146–1160 (2005).
- 131. Herculano-Houzel, S. The Evolution of Human Capabilities and Abilities. *Cerebrum: the Dana forum on brain science* **2018**, cer-05-18 (2018).
- 132. Semendeferi, K., Armstrong, E., Schleicher, A., Zilles, K. & Van Hoesen, G. W. Prefrontal Cortex in Humans and Apes: A Comparative Study of Area 10. American Journal of Physical Anthropology 114, 224–241 (2001).
- 133. Eichert, N. *et al.* Cross-species cortical alignment identifies different types of anatomical reorganization in the primate temporal lobe. *eLife* **9**, e53232 (2020).
- 134. Charvet, C. J., Striedter, G. F. & Finlay, B. L. Evo-Devo and Brain Scaling: Candidate

- Developmental Mechanisms for Variation and Constancy in Vertebrate Brain Evolution. *Brain Behav Evol* **78**, 248–257 (2011).
- 135. Rakic, P. A small step for the cell, a giant leap for mankind: a hypothesis of neocortical expansion during evolution. *Trends Neurosci* **18**, 383–388 (1995).
- 136. Bush, E. C. & Allman, J. M. The scaling of white matter to gray matter in cerebellum and neocortex. *Brain, Behavior and Evolution* **61**, 1–5 (2003).
- 137. Bush, E. C. & Allman, J. M. The scaling of frontal cortex in primates and carnivores. *Proceedings* of the National Academy of Sciences of the United States of America **101**, 3962–3966 (2004).
- 138. Donahue, C. J., Glasser, M. F., Preuss, T. M., Rilling, J. K. & Van Essen, D. C. Quantitative assessment of prefrontal cortex in humans relative to nonhuman primates. *Proceedings of the National Academy of Sciences of the United States of America* **115**, E5183–E5192 (2018).
- 139. Chaplin, T. A., Yu, H. H., Soares, J. G. M., Gattass, R. & Rosa, M. G. P. A Conserved Pattern of Differential Expansion of Cortical Areas in Simian Primates. *Journal of Neuroscience* **33**, 15120–15125 (2013).
- 140. Willemet, R. Allometry unleashed: an adaptationist approach of brain scaling in mammalian evolution. Preprint at https://doi.org/10.7287/PEERJ.PREPRINTS.27872V1 (2019).
- 141. Wartel, A., Lindenfors, P. & Lind, J. Whatever you want: Inconsistent results are the rule, not the exception, in the study of primate brain evolution. *PLoS ONE* **14**, e0218655 (2019).
- 142. Hooper, R., Brett, B. & Thornton, A. Problems with using comparative analyses of avian brain size to test hypotheses of cognitive evolution. *PLOS ONE* **17**, e0270771 (2022).
- 143. Maseko, B. C., Spocter, M. A., Haagensen, M. & Manger, P. R. Elephants Have Relatively the Largest Cerebellum Size of Mammals. *The Anatomical Record: Advances in Integrative Anatomy and Evolutionary Biology* **295**, 661–672 (2012).
- 144. Bakker, R., Wachtler, T. & Diesmann, M. CoCoMac 2.0 and the future of tract-tracing databases. *Frontiers in Neuroinformatics* **6**, 30 (2012).
- 145. Mars, R. B. et al. Primate comparative neuroscience using magnetic resonance imaging:

- Promises and challenges. Frontiers in Neuroscience 8, 298 (2014).
- 146. Friedrich, P. *et al.* Imaging evolution of the primate brain: the next frontier? *NeuroImage* **228**, 117685 (2021).
- 147. Milham, M. *et al.* Accelerating the Evolution of Nonhuman Primate Neuroimaging. in *Neuron* vol. 105 600–603 (Cell Press, 2020).
- 148. Barton, R. A. & Venditti, C. Reply to Smaers: Getting human frontal lobes in proportion.

 *Proceedings of the National Academy of Sciences 110, E3683–E3684 (2013).
- 149. Sherwood, C. C. & Smaers, J. B. What's the fuss over human frontal lobe evolution? *Trends in Cognitive Sciences* 17, 432–433 (2013).
- 150. Liu, X. *et al.* A multifaceted gradient in human cerebellum of structural and functional development. *Nature Neuroscience* **25**, 1129–1133 (2022).
- 151. Zheng, J. *et al.* Three-Dimensional Digital Reconstruction of the Cerebellar Cortex: Lobule Thickness, Surface Area Measurements, and Layer Architecture. *Cerebellum* (2022) doi:10.1007/s12311-022-01390-8.