https://doi.org/10.1186/s12864-023-09856-7

Köllner et al. BMC Genomics



The terpene synthase gene family in maize – a clarification of existing community nomenclature



Main text

Terpenes are important natural products functioning in both the primary and specialized metabolism of plants, bacteria, fungi, and other life forms. Core structural diversity is mainly determined by terpene synthases (TPS), enzymes that convert ubiquitous prenyl diphosphates such as geranyl diphosphate, farnesyl diphosphate, and geranylgeranyl diphosphate into the various terpene backbones.

(2023) 24:744

Terpene synthases in the crop plant maize (*Zea mays*) have been the subject of active research since the 1990s. A majority of maize TPS enzymes have already been functionally described and characterized, many of them in the laboratories of the authors of this clarification effort (reviewed in [1, 2]; Table 1). A comprehensive analysis of the *TPS* genes in the maize inbred lines B73 and W22 showed that both contain about 40 *TPS* genes, although the number varies between the different lines (43 in B73 versus 38 in W22) [3, 4]. These numbers include apparent pseudogenes, as it has been shown

*Correspondence:

that a pseudogene in one maize line can be functional in another line [3, 5, 6].

The amazing quantitative and qualitative plasticity of the maize *TPS* gene family was confirmed in a recent paper by Sun and coworkers, who analyzed *TPS* genes in the genomes of 26 inbred lines [7]. However, only 31 gene loci were included in this analysis, resulting in one-third of the already characterized *TPS* genes being omitted by the authors. Furthermore, the authors did not address the extensive pre-existing literature on maize terpene synthases prior to proposing a new nomenclature that was both incomplete and inconsistent with previously published names. Our concern with this approach is that it could lead to massive confusion in this field as readers will be unable to compare the new names with the original names without extensive sequence comparisons.

With the goal of minimizing confusion, we provide an overview of the existing maize nomenclature(s) and cite the primary literature in which the maize TPSs were first described and enzyme products characterized (Table 1). In addition, following the previously published TPS names, we propose to designate all mono- and sesquiterpene synthase genes with the abbreviation "ZmTPS" and a sequential numbering (ZmTPS1 - ZmTPS36). Further we propose the continued designation of the class I diterpene synthase genes, namely kaurene synthase-like (KSL) genes, as ZmTPS42/KSL1 to ZmTPS47/KSL6. Similarly, the class II diterpene synthase genes, namely the five copalyl diphosphate synthase (CPS) genes, are abbreviated as ZmTPS37/CPS1 to ZmTPS41/CPS5 (Table 1). Those involved in biosynthesis of the gibberellin hormone also have been designated by the original mutant names - i.e., an1/2 and d5, with the latter further modified as



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Tobias G. Köllner

koellner@ice.mpg.de

¹Department of Natural Product Biosynthesis, Max Planck Institute for Chemical Ecology, D-07745 Jena, Germany

²Department of Biochemistry, Max Planck Institute for Chemical Ecology, D-07745 Jena, Germany

³Roy J. Carver Department of Biochemistry, Biophysics & Molecular Biology, Iowa State University, Ames, IA 50011, USA

⁴Department of Plant Biology, University of California-Davis, Davis, CA 95616, USA

⁵Section of Cell and Developmental Biology, University of California, San Diego, La Jolla, CA 92093-0380, USA

Table 1 Current existing nomenclature of the terpene synthase gene family in maize. The first column contains the names already assigned to functionally characterized maize TPS
genes in the literature (for citations see last column). To fill the remaining gaps in the TPS numbering and to unify the nomenclature, we propose the names listed in the second
column, mainly following the original names or the names already given in Ding et al. [4] (third column). Please note that this table also includes obvious pseudogenes, because it has
been shown that a pseudogene in one maize line can be functional in another line [3, 5, 6]

Name in the primary literature	Name proposed in this study	Uing et al. [4]	sun et al. [7]	b/3 reference GRAMENE 4.0	b/3 reference NAM 5.0	ט היו או או שרו שרו שרו שרו שרו שרו שרו שרו שרו שר	Liter- ature
tps2	ZmTPS2	tps2	ZmTPS15	Zm00001d015053	Zm00001eb230410		6
tps3	ZmTPS3	tps3		Zm00001d015054	Zm00001eb230440		6
tps4	ZmTPS4	tps4		Zm00001d024478	Zm00001eb415080		[2]
tps5	ZmTPS5	tps5		Zm00001d024481	Zm00001eb415090		[2]
tps6/Zx1	ZmTPS6/Zx1	tps6	ZmTPS21	Zm00001d024207	Zm00001eb412960		[10]
tps7	ZmTPS7	tps7	ZmTPS4	Zm00001d032230	Zm00001eb041770		[11]
ZmTPS8	ZmTPS8	tps8	ZmTPS2	Zm00001d029195	Zm00001eb017120		[12]
	ZmTPS9	tps9	ZmTPS24	Zm00001d024477	Zm00001eb415070		
tps10	ZmTPS10	tps 10		Zm00001d024486	Zm00001eb415160		[13]
tps11/Zx3	ZmTPS 1 1/Zx3	tps 11		Zm00001d024210	Zm00001eb412960		[10]
TPS 1 2/Zx2	ZmTPS12/Zx2	tps 12		Zm00001d024208	Zm00001eb412960		[4]
TPS 1 3/Zx4	ZmTPS13/Zx4	tps 13		Zm00001d024211	Zm00001eb412990		[4]
	ZmTPS 14		ZmTPS9	Zm00001d004484	Zm00001eb089110		
tps15	ZmTPS 15	tps 15	ZmTPS16	Zm00001d035682	Zm00001eb267020		[14]
	ZmTPS 16		ZmTPS26	Zm00001d024667	Zm00001eb416710		
ZmEDS	ZmTPS 1 7/ZmEDS	tps 17	ZmTP510	Zm00001d004509	Zm00001eb089360		[15]
	ZmTPS18	tps 18	ZmTPS8	Zm00001d004279	Zm00001eb087570		
tps19/stc1	ZmTPS19/STC1	tps19/stc1	ZmTPS19	Zm00001d045054	Zm00001eb374210		[16]
tps20	ZmTPS20			Zm00001d024669	Zm00001eb416720		[14]
tps21	ZmTPS21	tps21	ZmTPS20	Zm00001d047440	Zm00001eb394330		[1]
tps22	ZmTPS22	tps22	ZmTPS23	Zm00001d024359	Zm00001eb414190		[14]
tps23	ZmTPS23	tps23	ZmTPS22	Zm00001d024234	Zm00001eb413120		[9]
	ZmTPS24		ZmTPS14	Zm00001d053916	Zm00001eb208380		
	ZmTPS25			Zm00001d053918	Zm00001eb208400		
tps26	ZmTPS26	tps26	ZmTPS17	Zm00001d037092	Zm00001eb278400		[16]
ZmTPS27	ZmTPS27	tps30	ZmTPS1	Zm00001d029139	Zm00001eb016730		[12]
	ZmTPS28		ZmTP518	Zm00001d018611	Zm00001eb298110		
	ZmTPS29		ZmTPS27	Zm00001d029523	Zm00031ab020550		
	ZmTPS30			Zm00001d024479	Zm00001eb415090		
	ZmTPS31			Zm00001d024480	Zm00001eb415100		
	ZmTPS32		ZmTPS29			Zm00038ab090090	
	ZmTPS33		ZmTPS30			Zm00038ab090300	
	ZmTPS34		ZmTPS31			Zm00026ab417460	
	ZmTPS35		ZmTPS28			Zm00034ab064260	
	ZmTPS36			Zm00001d000337			

Table 1 (continued)							
Name in the primary literature	Name proposed in this study	Ding et al. [4]	Sun et al. [7]	B73 reference GRAMENE 4.0	B73 reference NAM 5.0	Gene identifier (other maize lines)	Liter- ature
an 1/ZmCPS 1	ZmTPS37/CPS1/AN1	an1/cps1	ZmTPS6	Zm00001d032961	Zm00001eb048020		[18]
an2/ZmCPS2	ZmTPS38/CPS2/AN2	an2/cps2	ZmTPS3	Zm00001d029648	Zm00001eb021200		[19]
ZmCPS3	ZmTPS39/CPS3	cps3		Zm00001d024512	Zm00001eb415420		[20]
ZmCPS4	ZmTPS40/CPS4	cps4	ZmTPS12	Zm00001d048874	Zm00001eb167120		[20]
	ZmTPS41/CPS5			Zm00001d048867			
ZmKSL1	ZmTPS42/KSL1	ks/1	ZmTPS13	Zm00001d049957	Zm00001eb176190		
ZmKSL2	ZmTPS43/KSL2	ksl2	ZmTPS 1 1	Zm00001d041082	Zm00001eb133200		[21]
d5/ZmKSL3	ZmTPS44/KS(L3)/D5	ksl3	ZmTPS7	Zm00001d002349	Zm00001eb071070		[22]
ZmKSL4	ZmTPS45/KSL4	ks/4	ZmTPS5	Zm00001d032858	Zm00001eb047160		[23]
ZmKSL5	ZmTPS46/KSL5	ksl5		Zm00001d002350	Zm00001eb071080		[22]
	ZmTPS47/KSL6	ksl6	ZmTPS25	Zm00001d024514	Zm00001eb415430		
tps 1	ZmTPS 1/KSL7	tps1		Zm00001d002351	Zm00001eb071090		[8, 22]

KS(L3)/D5 to highlight its activity as an *ent*-kaurene synthase. Note that this nomenclature includes not only the 43 *TPS* gene loci found in the B73 reference genomes GRAMENE 4.0 and NAM 5.0 (www.maizegdb.org), but also four additional *TPS* genes not present in B73 but identified in other maize lines by Sun and coworkers [7]. The improvement of the already sequenced genomes and the sequencing of additional maize lines will lead to continued changes in the absolute number of known maize *TPS* genes in the future. Therefore, the nomenclature proposed here is itself evolving and merits periodic revision that builds upon existing knowledge.

The overview of the maize *TPS* gene family presented in this paper, together with the proposed nomenclature that includes all previously published names, is intended to help to minimize confusion about maize *TPS* names. In addition, the list of uncharacterized *TPS* genes presented in Table 1 can serve as a reference point to motivate future research on TPSs and their biological roles in maize.

Abbreviations

- TPS Terpene synthase
- KSL Kaurene synthase/kaurene synthase-like
- CPS Copalyl diphosphate synthase

Acknowledgements

Not applicable.

Author contribution

TGK, JG, RJP, PZ, and EAS analyzed data and literature. TGK wrote the manuscript. All authors read and approved the final manuscript.

Funding

This research was supported by the Max Planck Society, grants from the NIH (GM131885) and USDA-NIFA (2020-67013-32557) to RJP, and grants from NSF (1758976) and the DOE-JGI (CSP2568) to PZ and EAS. The funders had no role in the experimental design, data collection and analysis or preparation of the manuscript.

Data Availability

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 3 July 2023 / Accepted: 28 November 2023 Published online: 06 December 2023

References

 Degenhardt J, Köllner TG, Gershenzon J. Monoterpene and sesquiterpene synthases and the origin of terpene skeletal diversity in plants. Phytochemistry. 2009;70(15–16):1621–37.

- Block AK, Vaughan MM, Schmelz EA, Christensen SA. Biosynthesis and function of terpenoid defense compounds in maize (*Zea mays*). Planta. 2019;249(1):21–30.
- 3. Springer NM, Anderson SN, Andorf CM, Ahern KR, Bai F, Barad O, Barbazuk WB, Bass HW, Baruch K, Ben-Zvi G, Buckler ES, Bukowski R, Campbell MS, Cannon EKS, Chomet P, Dawe RK, Davenport R, Dooner HK, Du LH, Du CG, Easterling KA, Gault C, Guan JC, Hunter CT, Jander G, Jiao YP, Koch KE, Kol G, Köllner TG, Kudo T, Li Q, Lu F, Mayfield-Jones D, Mei WB, McCarty DR, Noshay JM, Portwood JL, Ronen G, Settles AM, Shem-Tov D, Shi JH, Soifer I, Stein JC, Stitzer MC, Suzuki M, Vera DL, Vollbrecht E, Vrebalov JT, Ware D, Wei SR, Wimalanathan K, Woodhouse MR, Xiong WW, Brutnell TP. The maize W22 genome provides a foundation for functional genomics and transposon biology. Nat Genet. 2018;50(9):1282–.
- 4. Ding YZ, Weckwerth PR, Poretsky E, Murphy KM, Sims J, Saldivar E, Christensen SA, Char SN, Yang B, Tong AD, Shen ZX, Kremling KA, Buckler ES, Kono T, Nelson DR, Bohlmann J, Bakker MG, Vaughan MM, Khalil AS, Betsiashvili M, Dressano K, Kollner TG, Briggs SP, Zerbe P, Schmelz EA, Huffaker A. Genetic elucidation of interconnected antibiotic pathways mediating maize innate immunity. Nat Plants. 2020;6(11):1375–88.
- Köllner TG, Schnee C, Gershenzon J, Degenhardt J. The variability of sesquiterpenes emitted from two *Zea mays* cultivars is controlled by allelic variation of two terpene synthase genes encoding stereoselective multiple product enzymes. Plant Cell. 2004;16(5):1115–31.
- Köllner TG, Held M, Lenk C, Hiltpold I, Turlings TCJ, Gershenzon J, Degenhardt J. A maize (E)-beta-caryophyllene synthase implicated in indirect defense responses against herbivores is not expressed in most American maize varieties. Plant Cell. 2008;20(2):482–94.
- Sun Y, Xiao WQ, Wang QN, Wang J, Kong XD, Ma WH, Liu SX, Ren P, Xu LN, Zhang YJ. Multiple variation patterns of terpene synthases in 26 maize genomes. BMC Genomics. 2023;24(1):46.
- Schnee C, Köllner TG, Gershenzon J, Degenhardt J. The maize gene terpene synthase 1 encodes a sesquiterpene synthase catalyzing the formation of (*E*)-beta-farnesene, (*E*)-nerolidol, and (*E*,*E*)-farnesol after herbivore damage. Plant Physiol. 2002;130(4):2049–60.
- Richter A, Schaff C, Zhang ZW, Lipka AE, Tian F, Köllner TG, Schnee C, Preiss S, Irmisch S, Jander G, Boland W, Gershenzon J, Buckler ES, Degenhardt J. Characterization of Biosynthetic pathways for the production of the volatile Homoterpenes DMNT and TMTT in Zea mays. Plant Cell. 2016;28(10):2651–65.
- Köllner TG, Schnee C, Li S, Svatos A, Schneider B, Gershenzon J, Degenhardt J. Protonation of a Neutral (S)-beta-bisabolene intermediate is involved in (S)-beta-macrocarpene formation by the maize sesquiterpene synthases TPS6 and TPS11. J Biol Chem. 2008;283(30):20779–88.
- Ren F, Mao HJ, Liang J, Liu J, Shu K, Wang Q. Functional characterization of ZmTPS7 reveals a maize tau-cadinol synthase involved in stress response. Planta. 2016;244(5):1065–74.
- Saldivar EV, Ding YZ, Poretsky E, Bird S, Block AK, Huffaker A, Schmelz EA. Maize Terpene Synthase 8 (ZmTPS8) Contributes to a Complex Blend of Fungal-Elicited Antibiotics. *Plants* 2023, 12, (5).

- Schnee C, Köllner TG, Held M, Turlings TCJ, Gershenzon J, Degenhardt J. The products of a single maize sesquiterpene synthase form a volatile defense signal that attracts natural enemies of maize herbivores. Proc Natl Acad Sci USA. 2006;103(4):1129–34.
- Luck K, Chen XL, Norris AM, Chen F, Gershenzon J, Köllner TG. The reconstruction and biochemical characterization of ancestral genes furnish insights into the evolution of terpene synthase function in the Poaceae. Plant Mol Biol. 2020;104(1–2):203–15.
- Liang J, Liu J, Brown R, Jia MR, Zhou K, Peters RJ, Wang Q. Direct production of dihydroxylated sesquiterpenoids by a maize terpene synthase. Plant J. 2018;94(5):847–56.
- Lin CF, Shen BZ, Xu ZN, Köllner TG, Degenhardt J, Dooner HK. Characterization of the monoterpene synthase gene *tps26*, the ortholog of a gene induced by insect herbivory in maize. Plant Physiol. 2008;146(3):940–51.
- Ding YZ, Huffaker A, Köllner TG, Weckwerth P, Robert CAM, Spencer JL, Lipka AE, Schmelz EA. Selinene volatiles are essential precursors for Maize Defense promoting Fungal Pathogen Resistance. Plant Physiol. 2017;175(3):1455–68.
- Bensen RJ, Johal GS, Crane VC, Tossberg JT, Schnable PS, Meeley RB, Briggs SP. Cloning and characterization of the maize *an1* gene. Plant Cell. 1995;7(1):75–84.
- Harris LJ, Saparno A, Johnston A, Prisic S, Xu M, Allard S, Kathiresan A, Ouellet T, Peters RJ. The maize *An2* gene is induced by *Fusarium* Attack and encodes an ent-copalyl diphosphate synthase. Plant Mol Biol. 2005;59(6):881–94.
- Murphy KM, Ma LT, Ding YZ, Schmelz EA, Zerbe P. Functional characterization of two class II Diterpene synthases indicates additional Specialized Diterpenoid pathways in Maize (Zea mays). Front Plant Sci 2018, 9.
- Ding Y, Murphy KM, Poretsky E, Mafu S, Yang B, Char SN, Christensen SA, Saldivar E, Wu M, Wang Q, Ji L, Schmitz RJ, Kremling KA, Buckler ES, Shen Z, Briggs SP, Bohlmann J, Sher A, Castro-Falcon G, Hughes CC, Huffaker A, Zerbe P, Schmelz EA. Multiple genes recruited from hormone pathways partition maize diterpenoid defences. *Nature Plants* 2019, 5, 1043–1056.
- Fu JY, Ren F, Lu X, Mao HJ, Xu MM, Degenhardt J, Peters RJ, Wang Q. A Tandem array of ent-kaurene synthases in Maize with roles in Gibberellin and more specialized metabolism. Plant Physiol. 2016;170(2):742–51.
- Mafu S, Ding Y, Murphy KM, Yaacoobi O, Bennett Addison J, Wang Q, Shen Z, Briggs SP, Bohlmann J, Castro-Falcon G, Hughes CC, Betsiashvili M, Huffaker A, Schmelz EA. Discovery, biosynthesis and stress-related Accumulation of Dolabradiene-Derived defenses in Maize. Plant Physiol. 2018;176:2677–90.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.