

## Article Addendum

# ELF4 as a Central Gene in the Circadian Clock

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circadian clock, entrainment, *ELF4*, *CCA1*, *LHY*, *TOC1*, feedback loop

### Addendum to:

#### *ELF4 is Required for Oscillatory Properties of the Circadian Clock*

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### ABSTRACT

The light-dark cycle of the environment serves as one of the major *Zeitgebers* in entrainment of the circadian clock. The circadian system consists of interconnected feedback loops in which the *CCA1/LHY-TOC1* loop has a central position. Genetic analyses of the *elf4* mutant suggested that it is a positive regulator of *CCA1* and *LHY* expression. Recently, we refined the mode-of-action of *ELF4* in entrainment of the clock, and here hypothesize that *ELF4* expression is interlocked with the *CCA1/LHY-TOC1* loop.

The circadian clock is an internal pacemaker. Primary qualities of a circadian clock include anticipation of daily events at dawn and dusk and a free-running rhythmicity of approximately 24 hours. In addition, circadian organisms can set internal rhythms to match the environmental light-dark cycle. This adjustment occurs daily and is known as entrainment.

Various groups have arranged the molecular components of the *Arabidopsis thaliana* circadian system into a model of transcription/translation feedback loops. Each loop consists of elements specific to certain time points (phases) of the 24 h day. In the most central loop, the expression peak times of the two MYB-like transcription factors *CCA1* and *LHY* are in the morning, and the pseudo response regulator *TOC1* peaks in the evening.<sup>1-3</sup> This loop is required for sustaining rhythms.<sup>4</sup> Several other clock genes—*GI*, *ELF3*, *ELF4*, *PRR7* and *PRR9*—are known to be important for proper periodicity.<sup>5-11</sup> Recently, mathematical modeling has assisted expression analyses and expanded the understanding of the circadian system.<sup>12</sup> Two additional feedback loops containing *GI* and *PRR7-PRR9*, respectively, were proposed to interlock with the *CCA1/LHY-TOC1* feedback loop.<sup>13,14</sup> In these studies, the position of the clock-controlling gene *ELF4* was not fully defined. In previous studies, it was found that *ELF4* genetically acts as a positive regulator of *CCA1*.<sup>5,15</sup> It remained a challenge to further define the mode-of-action of *ELF4* in connection to the performance of the circadian system.

We previously showed that *ELF4* is a clock gene whose expression peaks in the evening.<sup>5</sup> *ELF4* loss-of-function leads to an impaired photoperiod response, circadian dysfunction (imprecision and arrhythmicity) under free-running conditions, and very low amplitude of the light-induced clock gene *CCA1*. Thus, *ELF4* function was suggested to be important for input of light signaling to the clock, which is the process of clock resetting termed entrainment. Two other papers supported this conclusion.<sup>15,16</sup> What was missing from these efforts was a definition of *ELF4*'s placement within the clock network. Accordingly, we sought to expand *ELF4* genetic analyses to address the role of *ELF4* in sensing the photoperiodic signals of the environment and its connection to core clock genes.

In the *Plant Physiology* paper, to which we make this Addendum, we assessed clock performance of genotypes altered in *ELF4* expression and connected this to the expression phenotypes of the core clock genes *CCA1*, *LHY*, and *TOC1* in relation to the light-dark cycle. We found that the *elf4* mutant had elevated levels of *TOC1* expression implicating *ELF4* as a negative regulator of *TOC1*. In contrast, *ELF4* promotes the expression of the morning genes *CCA1* and *LHY*, because the expression of these two genes was markedly attenuated in the *elf4* loss-of-function mutant. A reciprocal effect was seen in *ELF4* overexpression lines: *TOC1* levels were low and *CCA1* and *LHY* levels were high. In addition, it was previously reported that *ELF4* expression was increased in the *cca1 lhy* double mutant.<sup>15</sup> Collectively, these expression data leads to a model where *ELF4* has a central position in the interconnected feedback loops (Fig. 1). Our conclusion is thus that *ELF4* is a core-clock gene working in the evening phase of the circadian cycle to drive morning expression of *CCA1* and *LHY*.

The interrelationship between *ELF4* and *TOC1* is still unclear, but as both are evening genes, they appear to act coordinately to control the clock. Under light-dark cycles, we found an early phase of *ELF4* expression in the *toc1* mutant, in agreement with the short-period phenotype of *toc1*. The mean expression level of *ELF4* under a diurnal cycle was unaltered in *toc1*. We hypothesize that *ELF4* expression would eventually dampen in *toc1* lines subjected to an extension of light period, as was seen for *ELF4* expression in the *cca1 lhy* mutant.<sup>15</sup> Thus, it is likely that core-oscillator genes are all codependent. Here, we suggest that *ELF4* is a “full” component of the central-clock machinery. That said, reporter levels driven by the *ELF4* promoter in the *elf4* mutant lead to mean levels of expression. Why *ELF4* is not in an autoregulatory loop to regulate its own overall expression level is an open question. As well, the transcripts of *ELF4* and *TOC1* both dampen in constant darkness and thus how evening genes are required for rhythm persistence in the absence of their transcript accumulation is unknown. We note that the *TOC1* profile has a bimodal peak, but *ELF4* does not.<sup>2,5</sup> Therefore, we wonder if circadian-controlling differences exist in the fine structure of *TOC1* and *ELF4* rhythmic expression.

The importance of *ELF4* activity in the circadian clock was corroborated by expression phenotypes under light-dark cycles (diurnal conditions). In contrast to wild type, the promoter: luciferase profiles in *elf4* were driven by “lights on” and “lights off.” Furthermore, *elf4* lost anticipation of the phase of the environmental light-dark cycle and displayed an unusual, fast-resetting behavior after a simulated “jet lag.” The light-induced peaks of *CCA1* and *LHY* expression can directly explain these diurnal patterns of gene expression in *elf4* confirming the position of *ELF4* in the central loop of the clock. In addition, the rhythm of gene expression of several clock markers in an *ELF4*-overexpression genotype displayed a long period. This suggests to us that *ELF4* functions in a dose-dependent manner as a repressor of periodicity. This overexpression-phenotype is remarkably different from that seen in other clock-gene overexpression lines (e.g., *CCA1-ox*, *lhy-1* and *TOC1-ox*),<sup>13,17,18</sup> and might indicate differences in post-translational performance among the central clock elements.

Analysis of overexpression lines has been supplemented by loss-of-function studies and it was found that mutations in the clock-oscillator genes differ in phenotype (arrhythmicity vs. short period).<sup>2,19</sup> Notably, when released into continuous light, the clock does not arrest in the *cca1 lhy* double, and in the *elf4* single mutant, there is no arrest until after a half-full circadian cycle.<sup>5,19</sup> This can be interpreted as that other “cogwheels” of the clock continue for a limited period of time when one wheel is blocked. Our idea is that *ELF4* function is analogous to an hourglass, where the oscillator stops after a fixed amount of time (after 12 h, subjective dusk) in absence of *ELF4* activity. Subsequently, the clock enters an arrhythmic state where the amplitude of output rhythms is lost, and this is probably due to the low levels of *CCA1* and *LHY* expression in *elf4*. An unknown factor (*X*) was modeled to function as an inducer of *CCA1* and *LHY* expression.<sup>12,13</sup> Further studies are needed to elucidate whether *ELF4* possess some features of *X* (e.g., the effect of *TOC1* on *ELF4*).

Various studies implicate *ELF4* in distinct input nodes of light signaling to the oscillator. In an earlier report, increased *ELF4* expression was found in the *elf3* mutant and therefore *ELF3* was defined as a negative regulator of *ELF4* expression.<sup>15</sup> We found that the *elf4* mutant had a minor role in gating the acute induction of *CAB* expression, and this was around subjective dusk. This means that

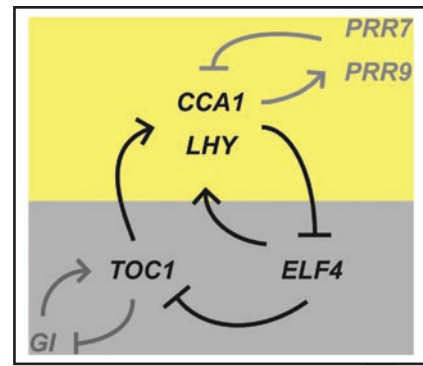


Figure 1. *ELF4* in the three-loop model of the circadian clock. *ELF4* expression is interlocked with the *CCA1/LHY-TOC1* loop in the three-loop model of the central circadian network.<sup>13,14</sup>

both *ELF3* and *ELF4* have roles in controlling light input to the clock.<sup>20</sup> Similarly, *elf4* arrhythmicity is evident under all wavelengths of light (McWatters H, unpublished), supporting the concept of *ELF4* in both light signaling and in entrainment. Furthermore, *ELF4* acts predominantly in response to the light-dark *Zeitgeber*, as *elf4* mutants can entrain (albeit weakly) to temperature cycles. Temperature as an entrainment cue remains to be tested for many of the clock genes. *PRR7* and *PRR9* have been found to mediate in temperature signaling to the clock,<sup>21</sup> and in another aspect of clock buffering, *CCA1* and *LHY* were found to participate in specific roles of temperature compensation.<sup>22</sup> The poor free-running rhythm of *elf4* after entrainment to temperature cycles indicates that more is yet to be learned about the role of *ELF4* in perception of environmental *Zeitgeber* signals.

The timing of dawn and dusk, which predictably fluctuates in most environments over the year, represents the fixed points for clock entrainment. We provided results supporting the idea that *ELF4* has a central function in the circadian clock and in day-length perception. In future studies, the post-translational relationship among and within the *CCA1/LHY-TOC1/ELF4* loops should be explored in order to gain a more detailed view of the feedback mechanism of the circadian clock.

#### References

- Schaffer R, Ramsay N, Samach A, Corden S, Putterill J, Carre IA, Coupland G. The *late elongated hypocotyl* mutation of *Arabidopsis* disrupts circadian rhythms and the photoperiodic control of flowering. *Cell* 1998; 93:1219-29.
- Strayer C, Oyama T, Schultz TF, Raman R, Somers DE, Mas P, Panda S, Kreps JA, Kay SA. Cloning of the *Arabidopsis* clock gene *TOC1*, an autoregulatory response regulator homolog. *Science* 2000; 289:768-71.
- Wang ZY, Tobin EM. Constitutive expression of the *CIRCADIAN CLOCK ASSOCIATED 1* (*CCA1*) gene disrupts circadian rhythms and suppresses its own expression. *Cell* 1998; 93:1207-17.
- Ding Z, Davis SJ. A complex genetic interaction between *Arabidopsis thaliana* *TOC1* and *CCA1/LHY* in driving the circadian clock and in output regulation. *Genetics* 2007; doi: 10.1534/genetics.107.072769.
- Doyle MR, Davis SJ, Bastow RM, McWatters HG, Kozma-Bognar L, Nagy F, Millar AJ, Amasino RM. The *ELF4* gene controls circadian rhythms and flowering time in *Arabidopsis thaliana*. *Nature* 2002; 419:74-7.
- Fowler S, Lee K, Onouchi H, Samach A, Richardson K, Morris B, Coupland G, Putterill J. *GIGANTEA*: A circadian clock-controlled gene that regulates photoperiodic flowering in *Arabidopsis* and encodes a protein with several possible membrane-spanning domains. *EMBO J* 1999; 18:4679-88.
- Hicks KA, Albertson TM, Wagner DR. *EARLY FLOWERING3* encodes a novel protein that regulates circadian clock function and flowering in *Arabidopsis*. *Plant Cell* 2001; 13:1281-92.
- Hicks KA, Millar AJ, Carre IA, Somers DE, Straume M, Meeks-Wagner DR, Kay SA. Conditional circadian dysfunction of the *Arabidopsis* *early-flowering 3* mutant. *Science* 1996; 274:790-2.

9. Ito S, Matsushika A, Yamada H, Sato S, Kato T, Tabata S, Yamashino T, Mizuno T. Characterization of the *APRR9* pseudo-response regulator belonging to the *APRR1/TOC1* quintet in *Arabidopsis thaliana*. *Plant Cell Physiol* 2003; 44:1237-45.
10. Kaczorowski KA, Quail PH. *Arabidopsis PSEUDO-RESPONSE REGULATOR7* is a signaling intermediate in phytochrome-regulated seedling deetiolation and phasing of the circadian clock. *Plant Cell* 2003; 15:2654-65.
11. Park DH, Somers DE, Kim YS, Choy YH, Lim HK, Soh MS, Kim HJ, Kay SA, Nam HG. Control of circadian rhythms and photoperiodic flowering by the *Arabidopsis GIGANTEA* gene. *Science* 1999; 285:1579-82.
12. Locke JCW, Southern MM, Kozma-Bognar L, Hibberd V, Brown PE, Turner MS, Millar AJ. Extension of a genetic network model by iterative experimentation and mathematical analysis. *Mol Syst Biol* 2005; 1:0013.
13. Locke JCW, Kozma-Bognar L, Gould PD, Feher B, Kevei E, Nagy F, Turner MS, Hall A, Millar AJ. Experimental validation of a predicted feedback loop in the multi-oscillator clock of *Arabidopsis thaliana*. *Mol Syst Biol* 2006; 2:59.
14. Zeilinger MN, Farre EM, Taylor SR, Kay SA, Doyle FJ. A novel computational model of the circadian clock in *Arabidopsis* that incorporates *PRR7* and *PRR9*. *Mol Syst Biol* 2006; 2:60.
15. Kikis EA, Khanna R, Quail PH. *ELF4* is a phytochrome-regulated component of a negative-feedback loop involving the central oscillator components *CCA1* and *LHY*. *Plant J* 2005; 44:300-13.
16. Khanna R, Kikis EA, Quail PH. *EARLY FLOWERING 4* functions in phytochrome B-regulated seedling de-etiolation. *Plant Physiol* 2003; 133:1530-8.
17. Makino S, Matsushika A, Kojima M, Yamashino T, Mizuno T. The *APRR1/TOC1* quintet implicated in circadian rhythms of *Arabidopsis thaliana*: I. Characterization with *APRR1*-overexpressing plants. *Plant Cell Physiol* 2002; 43:58-69.
18. Mas P, Alabadi D, Yanovsky MJ, Oyama T, Kay SA. Dual role of *TOC1* in the control of circadian and photomorphogenic responses in *Arabidopsis*. *Plant Cell* 2003; 15:223-36.
19. Alabadi D, Yanovsky MJ, Mas P, Harmer SL, Kay SA. Critical role for *CCA1* and *LHY* in maintaining circadian rhythmicity in *Arabidopsis*. *Curr Biol* 2002; 12:757-61.
20. McWatters HG, Bastow RM, Hall A, Millar AJ. The *ELF3* zeitnehmer regulates light signaling to the circadian clock. *Nature* 2000; 408:716-20.
21. Salome PA, McClung CR. *PSEUDO-RESPONSE REGULATOR 7* and *9* are partially redundant genes essential for the temperature responsiveness of the *Arabidopsis* circadian clock. *Plant Cell* 2005; 17:791-803.
22. Gould PD, Locke JC, Larue C, Southern MM, Davis SJ, Hanano S, Moyle R, Milich R, Putterill J, Millar AJ, Hall A. The molecular basis of temperature compensation in the *Arabidopsis* circadian clock. *Plant Cell* 2006; 18:1177-87.