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Supplemental Information

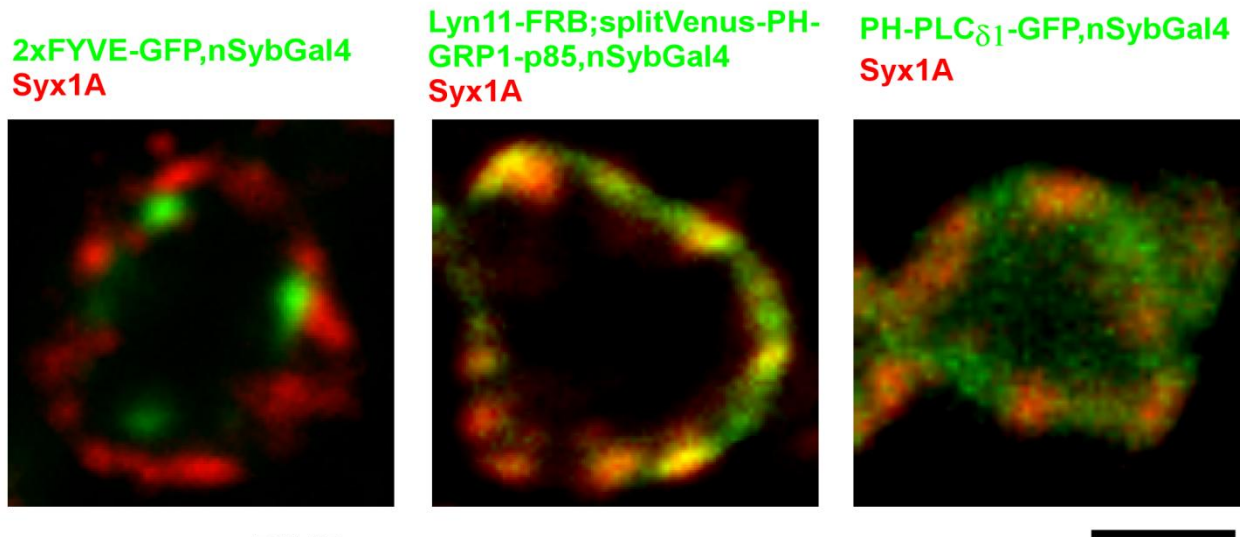
Synaptic PI(3,4,5)P₃ Is Required

for Syntaxin1A Clustering

and Neurotransmitter Release

Thang Manh Khuong, Ron L.P. Habets, Sabine Kuenen, Agata Witkowska, Jaroslaw Kasprowicz, Jef Swerts, Reinhard Jahn, Geert van den Bogaart, and Patrik Verstreken

A CONFOCAL



B PiMP

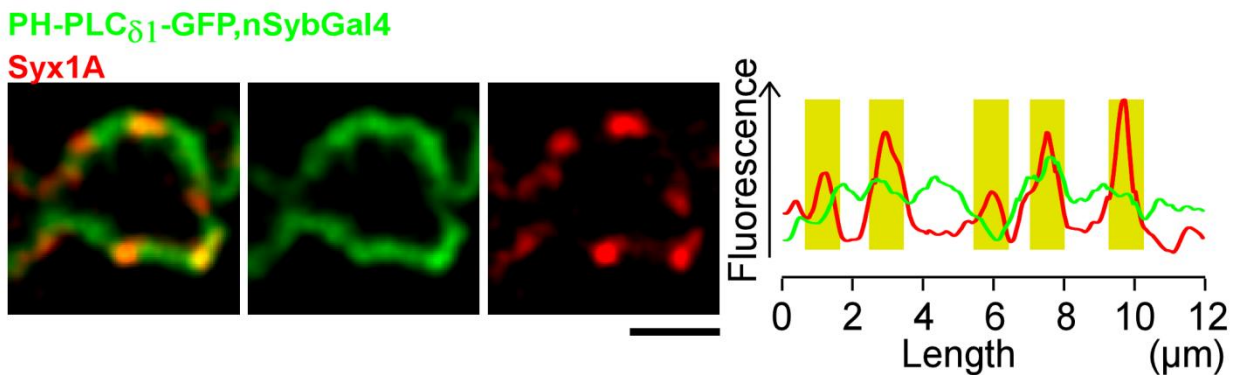


Figure S1, related to Figure 1. PH- PLC δ_1 -GFP that binds PI(4,5)P $_2$ does not extensively colocalize with Syntaxin1A

(A) Single confocal section of third instar larval *Drosophila* NMJ boutons that express 2xFYVE-GFP (*yw; UAS-2xFYVE-GFP/+; nSybGal4/+*) of a bouton expressing FRB-Lyn11, FKBP-p85 and splitVenus PH-GRP $_1$ (*yw; UAS-Lyn11-FRB; UAS-VenusC-PH-GRP $_1$ UAS-VenusN-PH-GRP $_1$ UAS-FKBP-p85 nSybGal4*) on rapamycin, and of a bouton expressing PH-PLC δ_1 -GFP (*yw; UAS- PH-PLC δ_1 -GFP nSybGal4*) labeled with anti-Syntaxin1A^{8C3} (Syx1A, red). GFP or Venus fluorescence is in green.

(B) PiMP super resolution imaging of the bouton expressing PH-PLC $_{\delta 1}$ -GFP shown in (A) and fluorescence intensity plot (arbitrary units) along the circumference of the bouton on the left adjusted to the total bouton circumference of anti-Syntaxin labeling intensity and of PH-PLC $_{\delta 1}$ -GFP fluorescence intensity. Yellow highlighted sections mark peaks of labeling. Scale bar in A and B, 5 μ m.

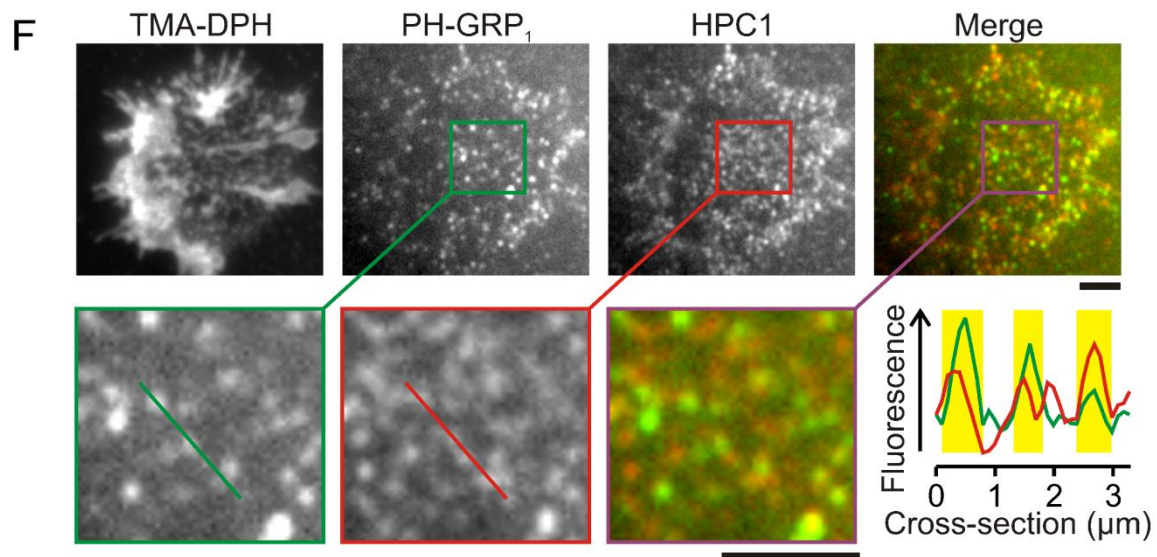
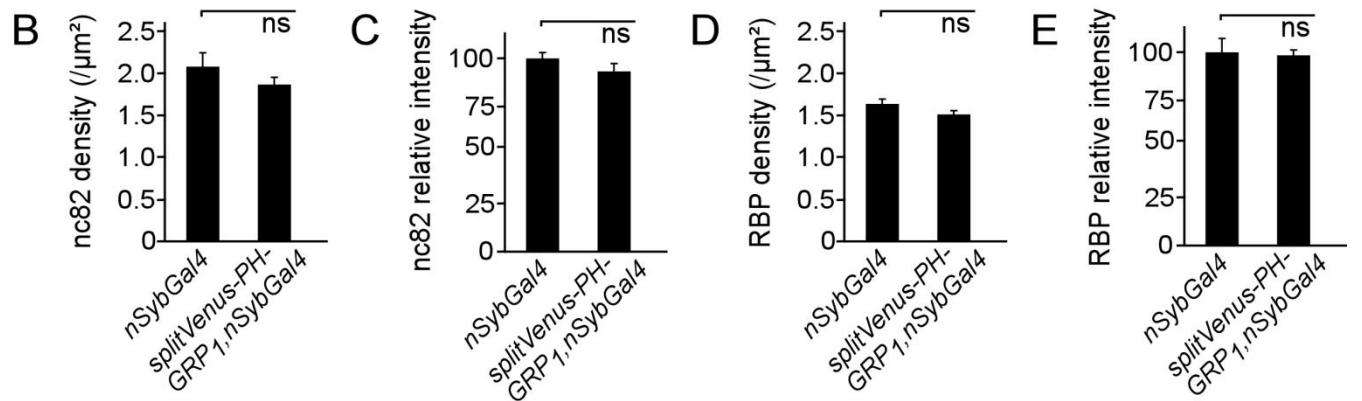
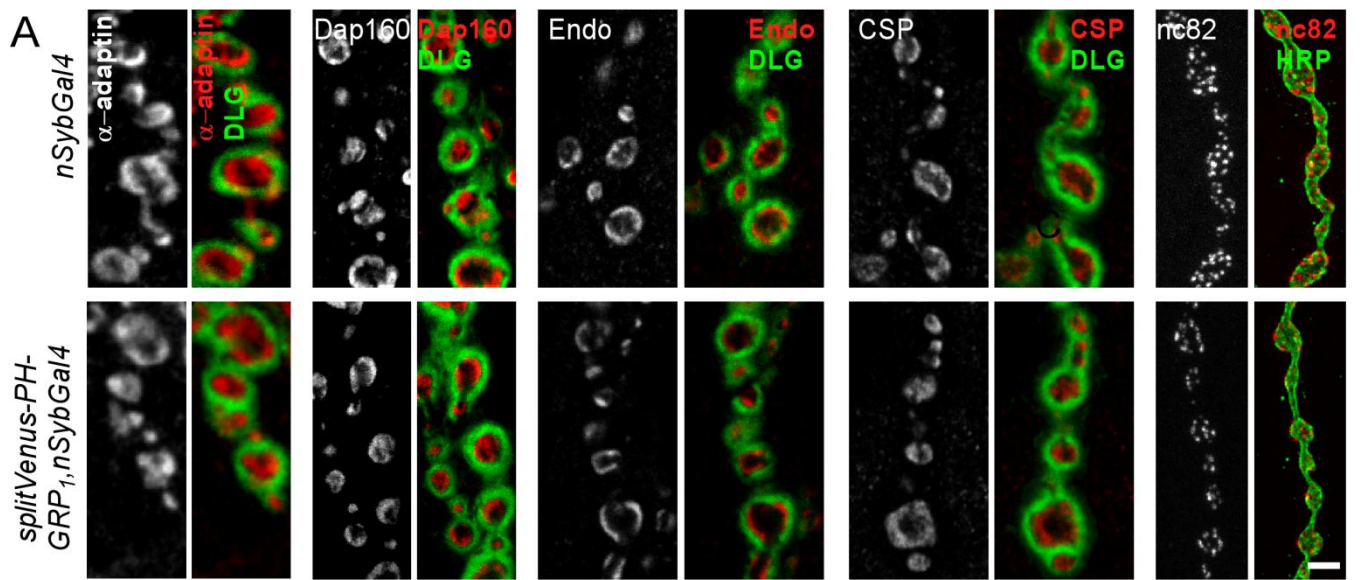


Figure S2, related to Figure 2. PH-GRP₁, labeling PI(3,4,5)P₃ colocalizes with Syntaxin1A in PC12 cells

(A) Single confocal sections of third instar larval *Drosophila* NMJ boutons of controls (top, *nSybGal4*) and of splitVenus-PH-GRP₁ expressing animals (bottom, *yw; UAS-VenusC-PH-GRP₁ UAS-VenusN-PH-GRP₁ nSybGal4*) labeled with anti-Alpha-adaptin, anti-Dap160, anti-Endophilin, anti-CSP and anti-BRP (nc82) (in Red) as well as with anti-DLG or anti-HRP (both synaptic markers, Green). Scale bar: 3 μm.

(B-E) Quantification of anti-BRP (nc82) and anti-RBP labeling intensity (C, E) and the number of dots per area (B, D) at third instar synaptic boutons of controls and of animals neuronally expressing splitVenus-PH-GRP₁. Data is mean ± SEM; n (the number of animals) >5; t-test: ns: not significant.

(F) PC12 membrane sheet immunostained with an Atto647N-labeled primary antibody raised against Syntaxin1A (HPC1) and stained with PH-GRP₁ genetically fused to mCherry. Sheets were imaged with TMA-DPH, a generic membrane dye. A representative image is shown from 16 sheets of 2 independent preparations. Bottom right, fluorescence intensity plot of PH-GRP₁-Cherry fluorescence (green) and Syntaxin1A labeling intensity (red) along the line shown. Yellow highlighted sections mark peaks of labeling. Scale bar, 3 μm.

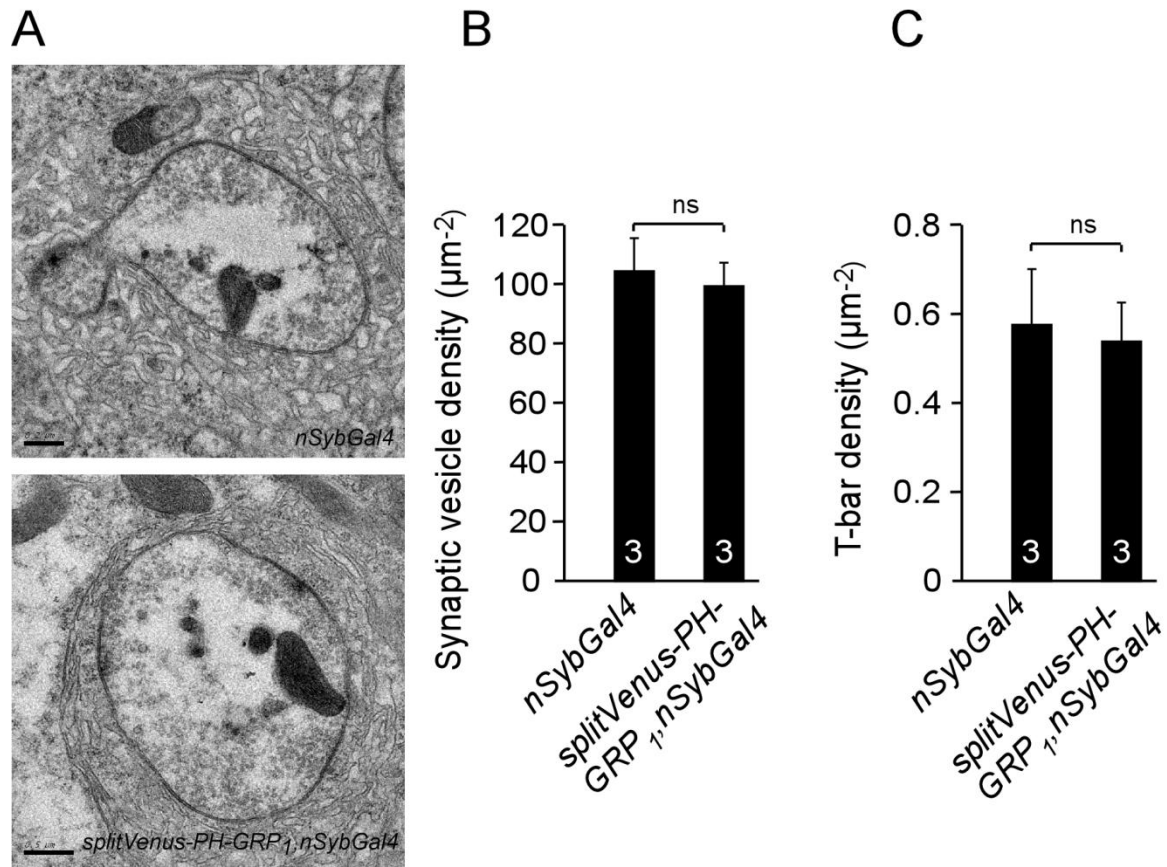


Figure S3, related to Figure 5. Normal ultrastructure of synaptic boutons in flies neuronally expressing splitVenus-PH-GRP₁.

(A) Electron micrographs of type Ib boutons of control (*nSybGal4*) (top) and animals expressing splitVenus-PH-GRP₁ (*yw; UAS-VenusC-PH-GRP₁ UAS-VenusN-PH-GRP₁ nSybGal4*, bottom).

(B-C) Quantification of vesicle density (B) and T-bar density (C) in control and animals expressing splitVenus-PH-GRP₁. Data is mean \pm SEM; n (the number of animals) is indicated in the bars; Tukey's test: ns: not significant.

Table S1, related to Figure 3. Conservation of positively charged juxtamembrane domain residues in Syntaxin1A

<i>Species</i>	<i>juxtamembrane domain sequence and basic stretch (highlight)</i>
<i>Drosophila melanogaster</i>	KKALKYQS KARRKK IMILICLT
<i>Xenopus (Silurana) tropicalis</i>	KKAVKYQS KARRKK IMIIIICCV
<i>Rattus norvegicus</i>	KKAVKYQS KARRKK IMIIIICCV
<i>Homo sapiens</i>	KKAVKYQS KARRKK IMIIIICCV

Table S2, related to the experimental procedures. Primers used

Primer	Sequence
VenusN	
VenusN-F	CTCC <u>CGGCCCGC</u> GTTCGACACAAAATGGTGAGCAAGGGCGAGG
VenusN-R	CTCG <u>AGATCT</u> GAGTCCGGAGGCGGTGATATAGACGTTG
VenusC	
VenusC-F	CTCCGGATCC <u>CGGCCCGC</u> GTTCGACACAAAATGGACAAGCAGA AGAACGGCATC
VenusC-R	CTCG <u>AGATCT</u> GAGTCCGGACTTGTACAGCTCGTCCATGC
Lyn11-FRB	
Lyn11-F	CTCC <u>CGGCCCGC</u> CACAAAATGGGATGTATAAAATCAAAG
Lyn11-R	CTTAGCGGCCG <u>CGGTACC</u> CTTATGCGTAGTCTGGTACG
FKBP-p85	
p85-F	CTCC <u>CGGCCCGC</u> GTTCGACACAAAATGGTGAGCAAGGGCGAGG
p85-R	CTTAGCGGCCG <u>CGGTACC</u> CTCACGTGCGCTCCTCGTGG

HA-syntaxin1A^{WT}	
pFL44S AscI Syx1A F	AGCGTCAGCGGGTTCTCGACGGTCACGGCGGGCATGTCGAAGG <u>CGCGCCGGTGAAAACGTGCTGATTTG</u>
Syx1A Part1 R	GTTCTCCTGTTGAAAACACACAAAAGTAGCCTCATCACTCAC ACACTCACAACACCGATGAAGAAGAG
Syx1A Part2 F	CTCTTCTTCATCGGTGTTGTGAGTGTGTGAGTGATGAGGCTACT TTTGTGTGAGTTTTCAACAGGAGAAC
HA syx1A Part2 R	GATAATACCCCGACACAAAGATGTACCCCTACGACGTGCCCGA <u>CTACGCCACTAAAGACAGATTAGCCGC</u>
HA syx1A Part3 F	GCGGCTAATCTGTCTTTAGTGGCGTAGTCGGGCACGTCGTAGG <u>GGTACATCTTTGTGTCGGGGTATTATC</u>
Syx1A Part3 R	TGCGTCCCTCTCAAACACACACCCTGAATCGGCGCCGACGACG CGTACGCAACAAACGAAAATAAAACGC
Syx1A Part4 F	GCGTTTTATTTTCGTTTGTGCGTACGCGTCGTCGGCGCCGATT CAGGGTGTGTGTTTGAGAGGGACGCA
pFL44S PacI Syx1A R	CAAAAATGGGTTTTATTA ACTTACATACTAG AATTCACCT <u>TAATTA ACTCCAATAGGACCAGTGTTG</u>
HA-syntaxin1A^{KARRAA}	
pFL44S AscI Syx1A F	AGCGTCAGCGGGTTCTCGACGGTCACGGCGGGCATGTCGAAGG <u>CGCGCCGGTGAAAACGTGCTGATTTG</u>
Syx1A Part1 R	GTTCTCCTGTTGAAAACACACAAAAGTAGCCTCATCACTCAC ACACTCACAACACCGATGAAGAAGAG
Syx1A Part2 F	CTCTTCTTCATCGGTGTTGTGAGTGTGTGAGTGATGAGGCTACT

	TTTGTGTGAGTTTTCAACAGGAGAAC
20bp before AA 20bp after R	ACCAGAGT <u>AAAGCCCGACGAGCCGCC</u> ATCATGATACTGATCTG CCT
20bp before AA 20bp after F	AGGCAGATCAGTATCATGAT <u>GGCGGCTCGTCGGGCTTT</u> ACTCT GGT
HA syx1A Part2 R	GATAATACCCCGACACAAAGATGTACCCCTACGACGTGCCCGA <u>CTACGCCACTAAAGACAGATTAGCCGC</u>
HA syx1A Part3 F	GCGGCTAATCTGTCTTTAGTGGCGTAGTCGGGCACGTCGTAGG <u>GGTACATCTTTGTGTCGGGGTATTATC</u>
Syx1A Part3 R	TGCGTCCCTCTCAAACACACACCCTGAATCGGCGCCGACGACG CGTACGCAACAAACGAAAATAAAACGC
Syx1A Part4 F	GCGTTTTATTTTCGTTTGTTGCGTACGCGTCGTCGGCGCCGATT CAGGGTGTGTGTTTGAGAGGGACGCA
pFL44S PacI Syx1A R	CAAAAATGGGTTTTATTA ACTTACATACATACTAGAATTCACCT <u>TAATTA</u> ACTCCAATAGGACCAGTGTTG

Table S3, related to the experimental procedures. Genotypes of flies used

Abbreviation	Genotype
<i>nSybGal4</i>	<i>yw; nSybGal4</i>
<i>yv¹/+; nSybGal4/+</i>	<i>yv¹/yw; nSybGal4/+</i>
<i>w¹¹¹⁸/+; nSybGal4 > DCR-2/+</i>	<i>w¹¹¹⁸/yw UAS-DCR-2; nSybGal4/+</i>
<i>splitVenus-PH-GRP₁ nSybGal4</i>	<i>yw; UAS-VenusC-PH-GRP₁ UAS-VenusN-PH-GRP₁</i> <i>nSybGal4</i>

<i>Lyn11-FRB ; splitVenus-PH-GRP₁</i> <i>nSybGal4,</i>	<i>yw; UAS-Lyn11-FRB/+ ; UAS-VenusC-PH-GRP₁, UAS-</i> <i>VenusN-PH-GRP₁ nSybGal4</i>
<i>Lyn11-FRB ; splitVenus-PH-GRP₁</i> <i>FKBP-p85 nSybGal4</i>	<i>yw; UAS-Lyn11-FRB/+ ; UAS-VenusC-PH-GRP₁ UAS-</i> <i>VenusN-PH-GRP₁ UAS-FKBP-p85 nSybGal4/ UAS-VenusC-</i> <i>PH-GRP₁ UAS-VenusN-PH-GRP₁ nSybGal4</i>
<i>nSybGal4 > TRIP PI3K92E^{35798 or 27690}</i>	<i>yv¹/yw; UAS-RNAi(TRIP PI3K92E^{35798 or 27690} /nSybGal4</i>
<i>nSybGal4 > DCR-2, RNAi PI4P5K⁴⁷⁰²⁷</i>	<i>w¹¹¹⁸/yw UAS-DCR-2; UAS-RNAi (PI4P5K⁴⁷⁰²⁷)/nSybGal4</i>
<i>yv¹/+; nSybGal4 > DCR-2/+</i>	<i>yv¹/yw UAS-DCR-2; nSybGal4/+</i>
<i>nSybGal4 > DCR-2, TRIP Syx1A²⁵⁸¹¹</i>	<i>yv¹/yw UAS-DCR-2; UAS-RNAi(TRIP Syx1A²⁵⁸¹¹ /nSybGal4</i>
<i>HA-syx1A^{WT}</i>	<i>yw; P{CaryP}attP40 (syx1A^{WT})</i>
<i>HA-syx1A^{KARRAA}</i>	<i>yw; P{CaryP}attP40 (syx1A^{KARRAA})</i>
<i>HA-syx1A^{WT}; syx1A^{Δ229}/+</i>	<i>yw/+; P{CaryP}attP40 (syx1A^{WT}); syx1A^{Δ229}/+</i>
<i>HA-syx1A^{KARRAA}; syx1A^{Δ229}/+</i>	<i>yw/+; P{CaryP}attP40 (syx1A^{KARRAA}); syx1A^{Δ229}/+</i>