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Supplemental Information

The *Dionaea muscipula* Ammonium Channel

***DmAMT1* Provides NH₄⁺ Uptake Associated**

with Venus Flytrap's Prey Digestion

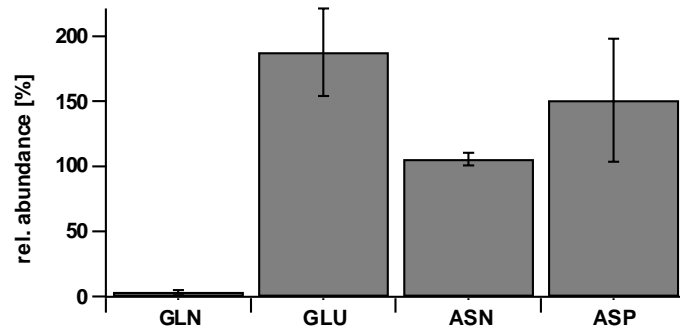
Sönke Scherzer, Elzbieta Krol, Ines Kreuzer, Jörg Kruse, Franziska Karl, Martin von Rüden, Maria Escalante-Perez, Thomas Müller, Heinz Rennenberg, Khaled A.S. Al-Rasheid, Erwin Neher, and Rainer Hedrich

A

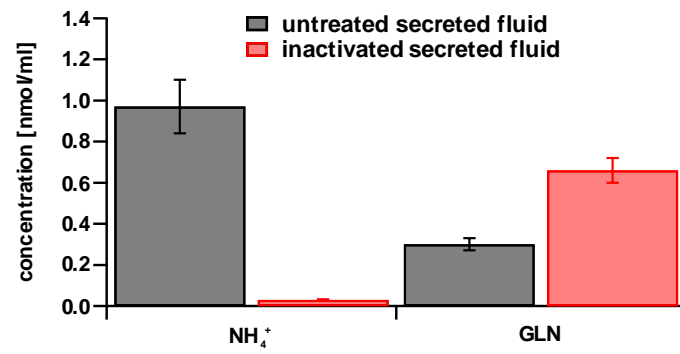
**Concentration of NH_4^+ [mM],
derived after digestion**

input	insect powder (<i>Cretonotos transiens</i>)	protein standard (casein)
5 mg	0.04 ± 0.01	0.61 ± 0.08
10 mg	0.09 ± 0.01	1.17 ± 0.09
20 mg	0.21 ± 0.07	2.65 ± 0.54

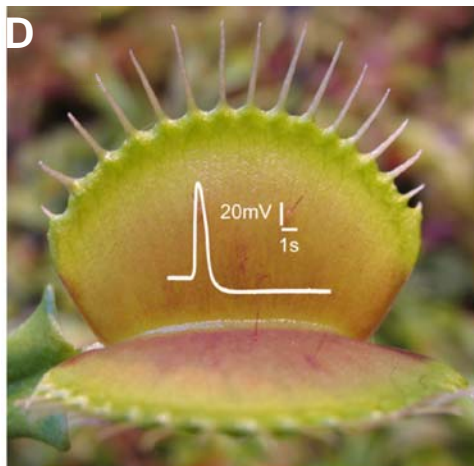
B



C



D



E

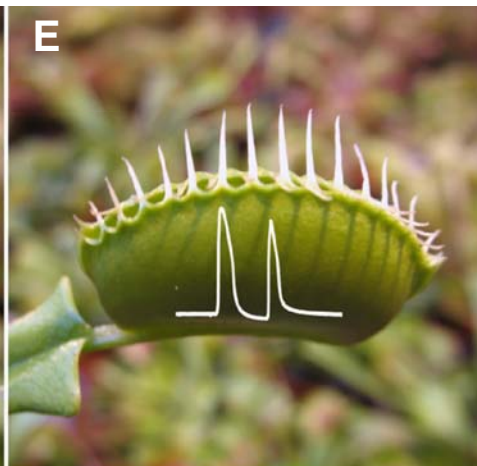
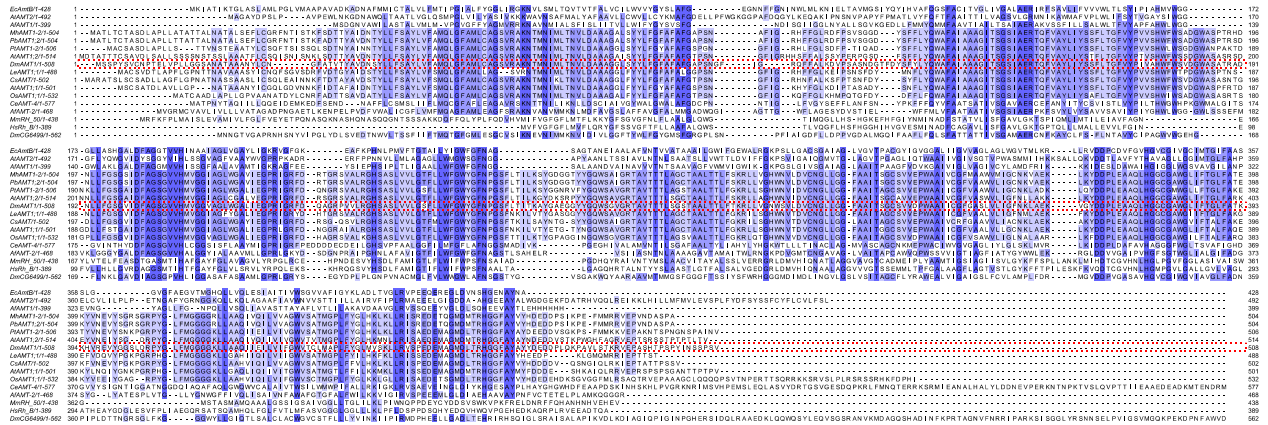


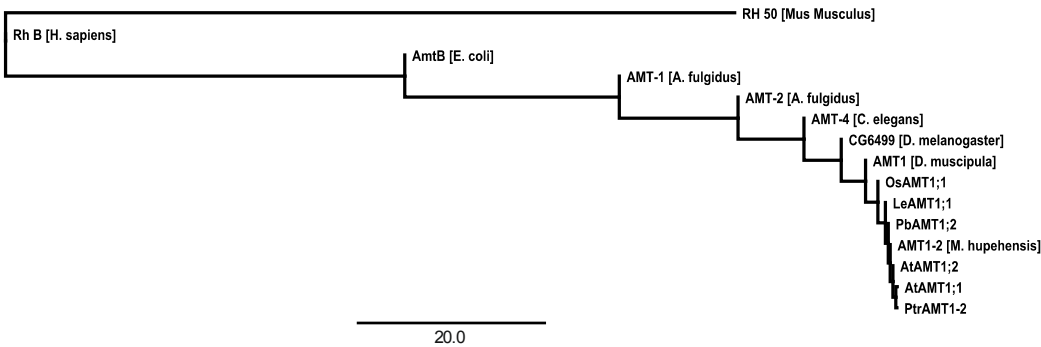
Figure S1. Related to Figure 1

(A): Production of NH_4^+ as a result of enzymatic reaction of insect powder or casein in the secreted fluid of *Dionaea muscipula* (n=4, mean \pm SD). **(B):** Detected AS level in the casein digest normalized to its abundance in the protein. While asparagine (ASN) and aspartate (ASP) could be detected with values close to the abundance in casein ($105.47 \pm 8.44\%$ and $150.72 \pm 81.81\%$), glutamine (GLN) was almost absent ($3.41 \pm 2.25\%$) and the level of glutamate (GLU) was increased ($187.76 \pm 58.09\%$) (n=12, mean \pm SD). **(C):** Ammonium and glutamine concentration in secreted fluid of *Dionaea muscipula*, after digestion of GLN (0.8 nmol/ml) in untreated or inactivated secretion fluid. Inactivation was performed by freezing (10 min/-20 °C) followed by addition of SDS (sodium dodecyl sulfate) (1 mM) and boiling (5 min) (n=3, mean \pm SD). **(D):** Mechano-stimulation of trigger hairs by a prey evokes an action potential (AP), which spreads over the entire leaf lobe that is densely packed with gland cell complexes. **(E):** Two action potentials close the Venus flytrap.

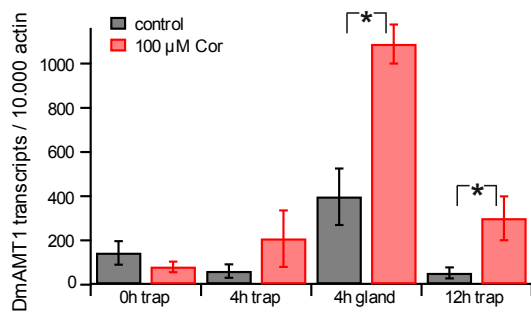
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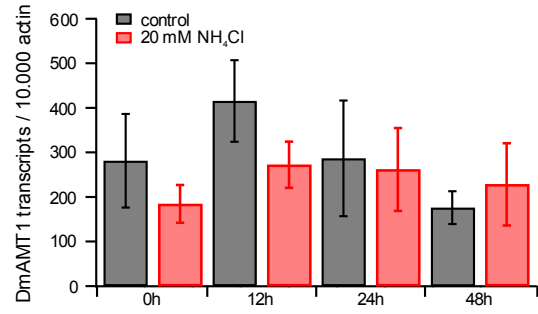


Figure S2. Related to Figure 2

(A) Sequences of AMT/MEP/Rh homologs. Alignment of 17 AMT/MEP/Rh proteins of various origins by clustalw2. AMT1 from *Dionaea muscipula* is boxed in red. Identical amino acids were highlighted in blue (jalview 2.7). EcAmtB: *E. coli* AmtB; AtAMT2: *A. thaliana* AMT2; AfAMT1: *A. fulgidus* AMT-1; MhAMT1-2: *M. hupehensis* AMT1-2; PbAMT1;2: *P. betulifolia* AMT1;2; PtAMT1-2: *P. trichocarpa* AMT1-2; AtAMT1;2: *A. thaliana* AMT1;2; DmAMT1: *Dionaea muscipula* AMT1;

LeAMT1;1: *L. esculentum* AMT1;1; CsAMT: *C. sinensis* ammonium transporter; AtAMT1;1: *A. thaliana* AMT1;1; OsAMT1;1: *O. sativa* AMT1;1; CeAMT-4: *C. elegans* AMT-4; AfAMT-2: *A. fulgidus* AMT-2; MmRH_50: *M. musculus* RH 50; HsRh_B: *H. sapiens* Rh B; DmCG6499: *D. melanogaster* ammonium transporter. **(B)** Phylogeny of ammonium transporters. This phylogenetic tree bases on an alignment with selected protein sequences of mammalian, plant, bacterial and fungal ammonium permeases of the AMT/MEP/Rh family. The tree was constructed with Geneious using Neighbor-Joining [S1]. Scale bar indicates the genetic distance. Among the ammonium permeases DmAMT1 closes the gap between the plant field and other members of this family in different kingdoms. Although it shares a high identity-score (71) to OsAMT1;1 whereas its homolog in *Drosophila* has only a score of 28 (calculated by ClustalW2). **(C)** DmAMT1 expression was upregulated in isolated glands already 4h after application of 100 μ M coronatine. Transcripts of DmAMT1 were normalized to 10.000 molecules of DmACT. Asterisks indicate statistically significant upregulation by coronatine ($P < 0.01$ by one-way ANOVA)($n \geq 3$, mean \pm SD). **(D)** Quantification of DmAMT1 transcript levels in response to ammonium. DmAMT1 transcripts in traps of *Dionaëa muscipula* were normalized to 10.000 molecules of DmACT and expression was measured after application of H₂O (control) or 200 μ l NH₄Cl at indicated time points. No significant differences could be observed between water and ammonium treatment ($P > 0.9$ by one-way ANOVA)($n \geq 5$, mean \pm SE).

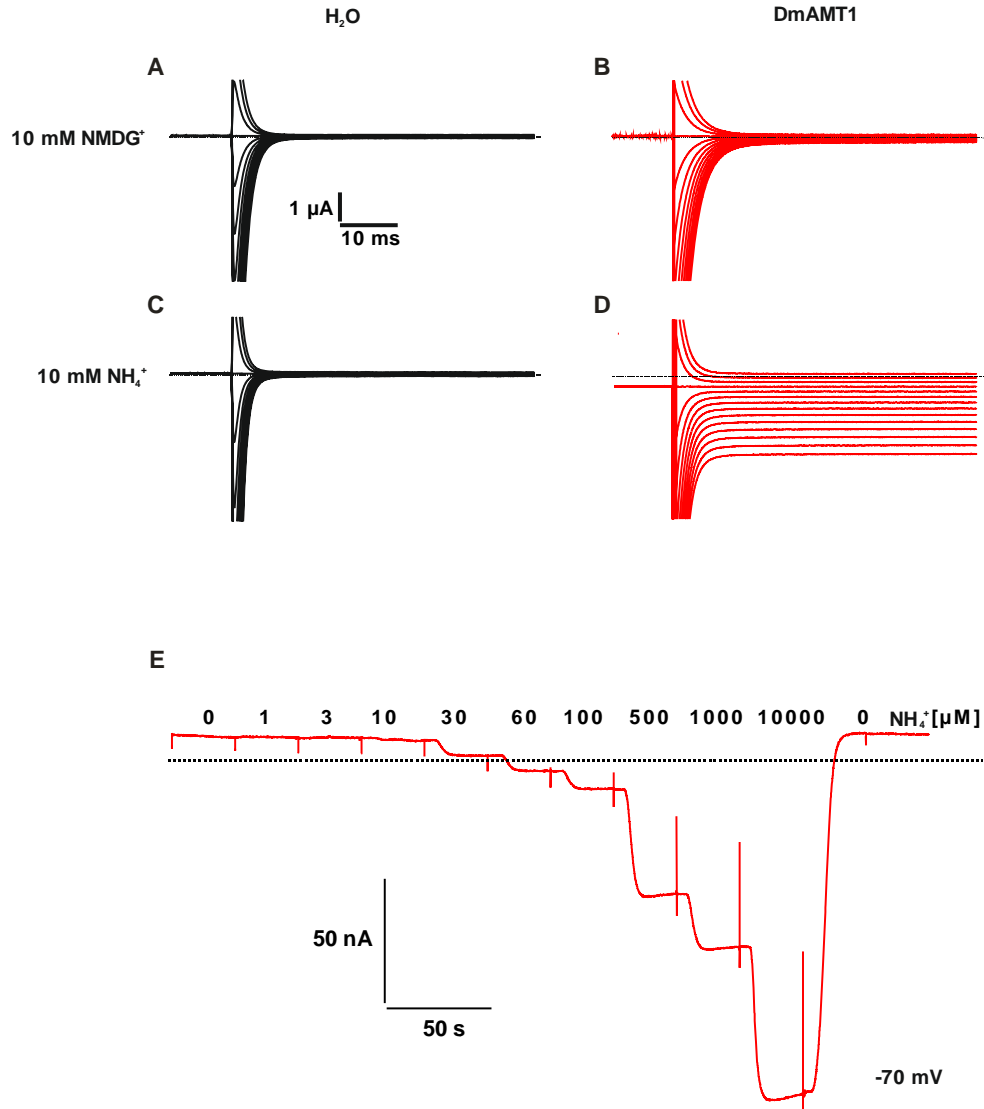


Figure S3. Related to Figure 3

Raw currents recorded at a typical voltage protocol changing the membrane potential from +60 to -200 mV in 20 mV decrements. Holding potential was 0 mV and dotted lines indicate 0 μA. No macroscopic currents are detectable with water-injected oocytes (A) as well as in DmAMT1 expressing oocytes without ammonium available (B). Addition of 10 mM NH₄⁺ leads neither to macroscopic currents in water-injected oocytes (C), but an ammonium influx of up to 3 μA was detectable in DmAMT1 expressing oocytes (D). (E) DmAMT1 mediated currents in *Xenopus* oocytes at the clamped membrane potential of -70 mV. NH₄⁺ concentration in the bath medium was exchanged as indicated and the monovalent cation concentration was complemented with K⁺ to 10mM. Dotted line represents 0 mV. Currents were recorded in standard bath solution.

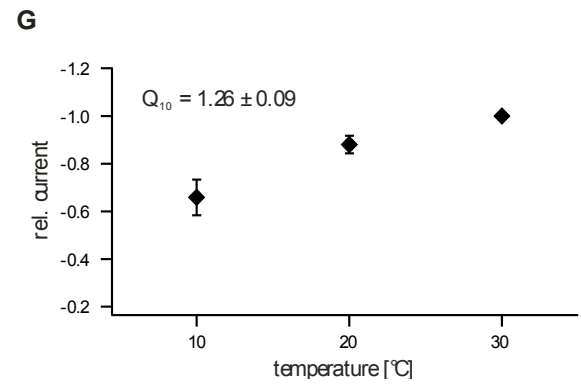
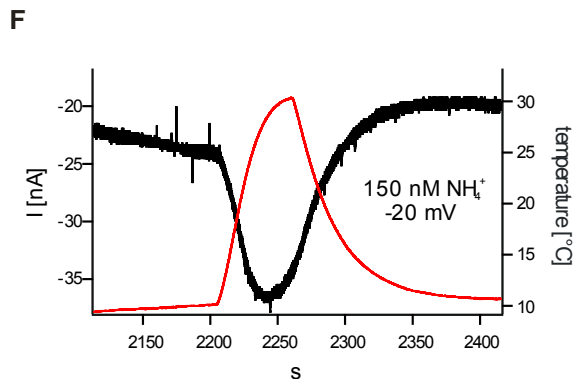
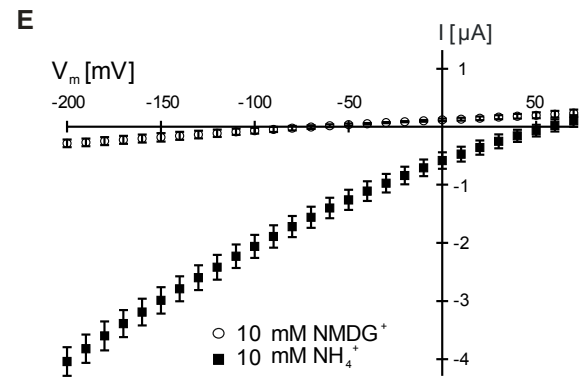
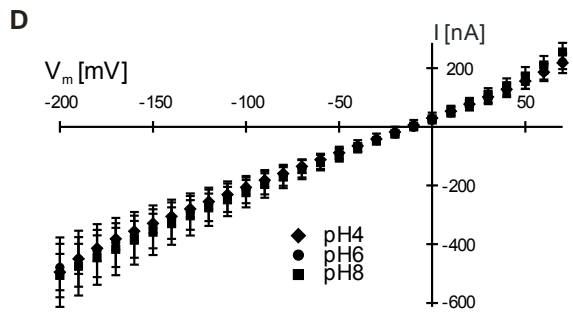
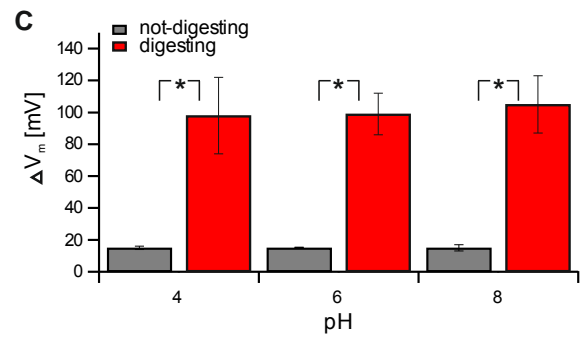
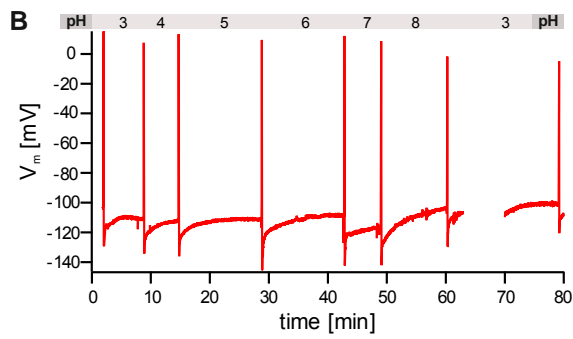
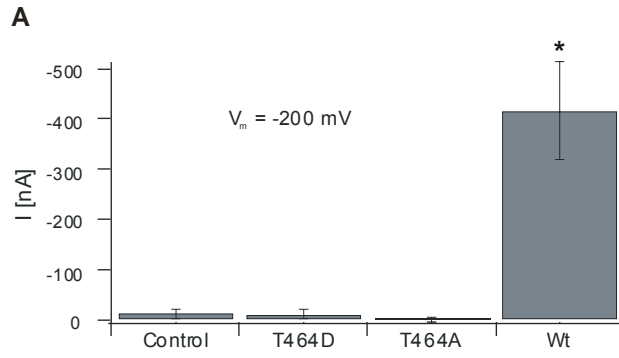


Figure S4. Related to Figure 4

(A) Point mutation at T464 leads to inactive DmAMT1. Oocytes were injected with water, DmAMT1 T464D, T464A, and wild type RNA. Macroscopic currents were only recorded with wild type DmAMT1 expressing oocytes in standard bath solution containing 10 mM NH₄Cl. Statistically significant differences were only observed for wt- against mutant- and control-currents ($P < 0.01$ by one-way ANOVA)($n=6$, mean \pm SD). **(B):** Membrane voltage responses of unstimulated *Dionaëa muscipula* gland cells to different proton concentrations (as indicated). Just before each pH change an AP was mechanically evoked to confirm intracellular localization of measuring electrode. Changing the proton concentration of up to 100.000-fold did only weakly affect the resting potential. **(C)** pH dependency of ammonium-induced depolarization in glands. Significant differences were only observed between digesting and not-digesting plants, whereas depolarization within this two groups is not pH dependent ($P < 0.01$ by one-way ANOVA)($n=3$, mean \pm SD). **(D)** Current/voltage dependency under different pH values. Varying H⁺ concentration by a factor of 10.000 didn't affect the voltage independent currents of DmAMT1. The reversal potentials ($V_{revpH4} = 13.9 \pm 0.88$, $V_{revpH6} = 11.8 \pm 0.6$, and $V_{revpH8} = 14.2 \pm 0.8$ mV) as well as the positive outward currents at membrane potentials positive from V_{rev} suggest that the oocytes harbored contents of ammonium. Experiments were performed in standard bath solution containing 10 mM NH₄Cl ($n=5$, mean \pm SD). **(E)** Ammonium influx in DmAMT1 expressing oocytes. Macroscopic currents of up to 4 μ A were only detectable with 10 mM NH₄⁺ in the bath solution. If NMDG⁺ is presented instead only small ohmic leakage currents were investigated ($n=5$, mean \pm SD). **(F)** Temperature dependency of DmAMT1 mediated ammonium currents. To minimize changes in the oocyte internal NH₄⁺ concentration during measurement caused by large ammonium currents, low external concentration (150 μ M) and a small electrical gradient ($V_m = -20$ mV) was used. Increasing the bath temperature from 10 to 30 °C (red line) resulted in rapid increasing inward-currents from 24 to 37 nA. This behavior was reversible by lowering the temperature again. **(G)** DmAMT1 functions as channel. Relative currents were recorded at $V_m = -200$ mV at 3 different temperatures. Q_{10} value was calculated at membrane potentials which resulted in distinct currents ($V_m = -100 - -200$ mV) and as mean factor between currents recorded at 10°C - 20°C, 20°C – 30°C and $\sqrt{10^\circ\text{C} - 30^\circ\text{C}}$. Experiments were performed in standard bath solution containing 150 μ M NH₄Cl ($n=5$, mean \pm SD).

Supplemental References

- S1. Ko JH, Yang SH, Han KH (2006) Upregulation of an *Arabidopsis* RING-H2 gene, *XERICO*, confers drought tolerance through increased abscisic acid biosynthesis. Plant J 47: 343-355.