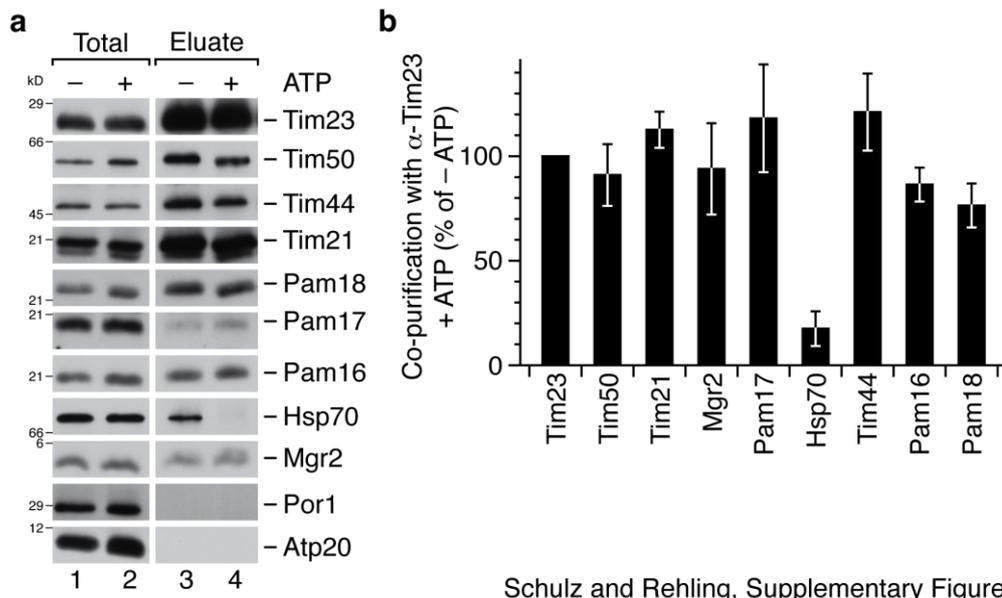
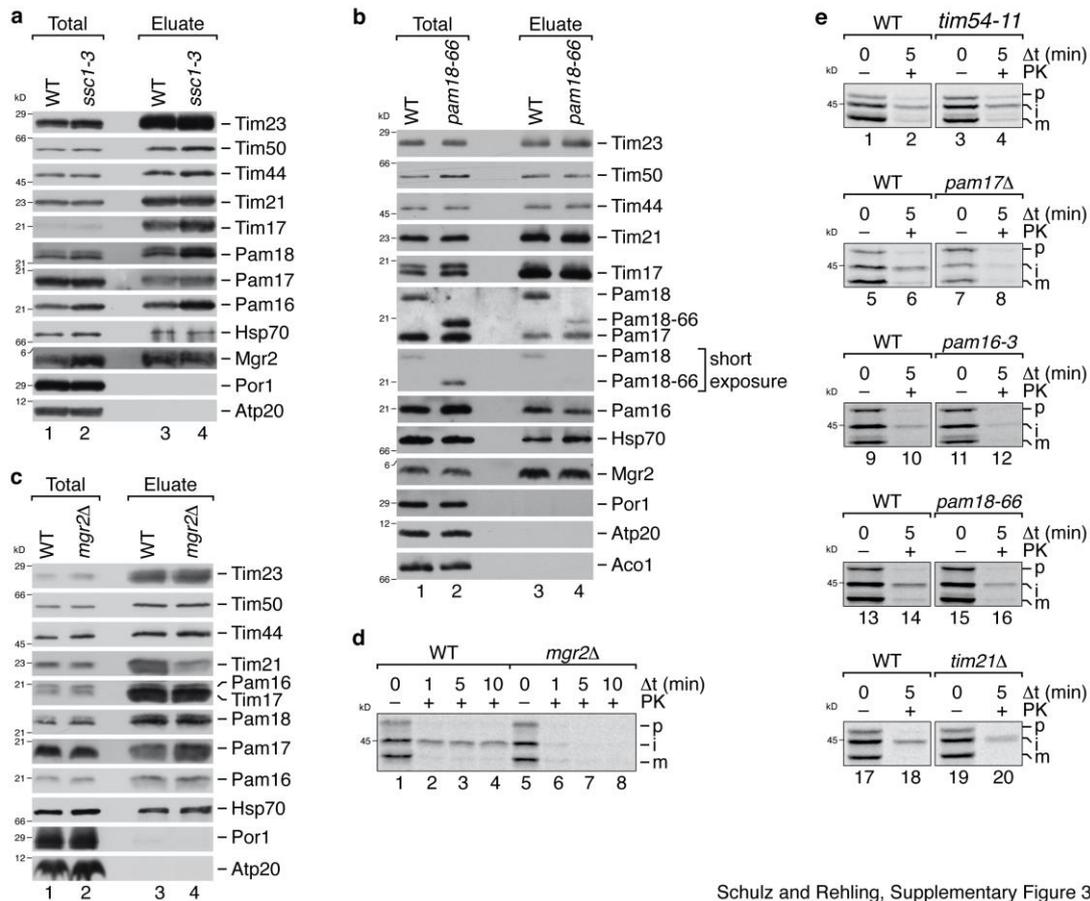


Schulz and Rehling, Supplementary Figure 1

**Supplementary Figure 1: Import defects in *ssc1* temperature sensitive mutants.** Isolated WT, *ssc1-2* and *ssc1-3* mitochondria were subjected to 15 min heat shock. Subsequently, [ $^{35}\text{S}$ ]Atp2 was imported for the indicated times. The reaction was stopped by dissipation of  $\Delta\psi$  and proteinase K treatment. Samples were analyzed by SDS-PAGE and digital autoradiography (left). Quantification of the amount of mature Atp2 is shown (right, 100 %: amount imported in WT after 15 min). p, precursor; m, mature.



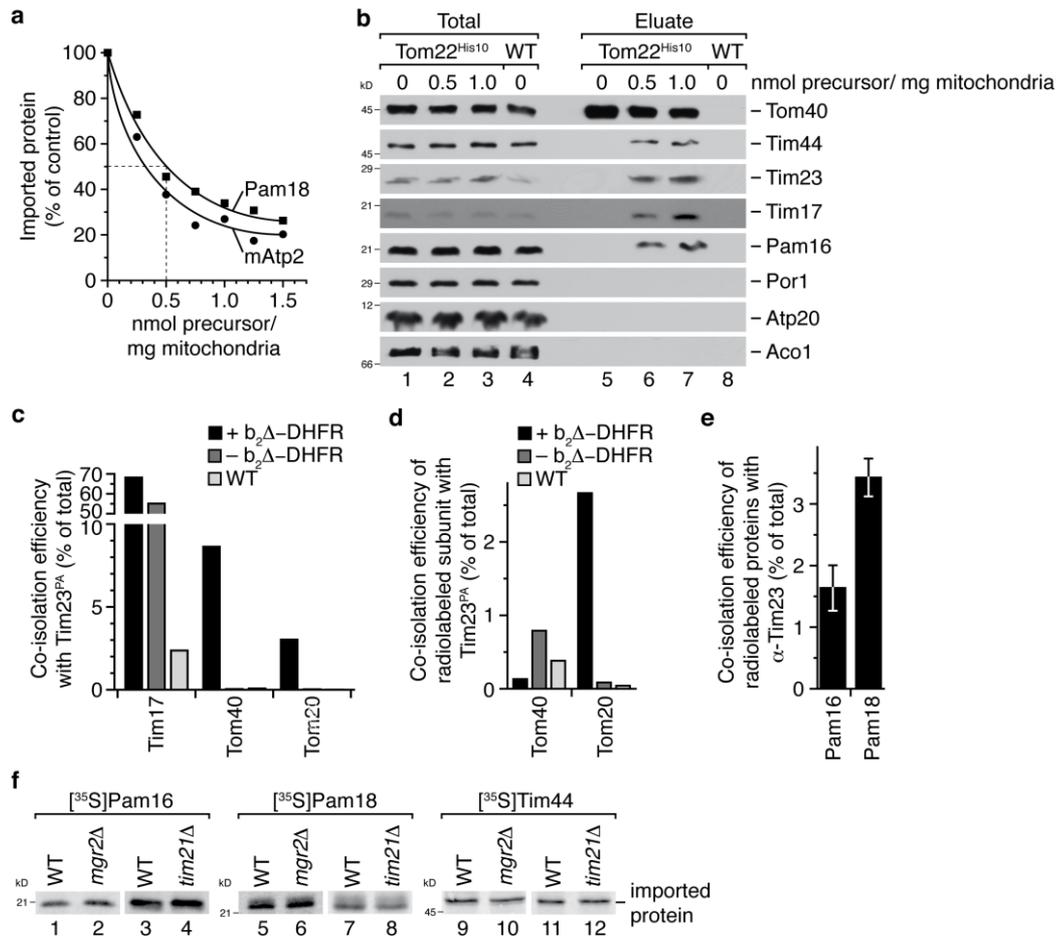
**Supplementary Figure 2: Recovery of the TIM23 complex by  $\alpha$ -Tim23 co-immunoprecipitations.** (a)  $\alpha$ -Tim23 co-immunoprecipitations were performed from WT mitochondria solubilised with digitonin in the absence or presence of ATP. Samples were analyzed by SDS-PAGE and Western-blotting. Total, 5%; eluate, 100%. (b) Quantification of experiments performed as described in (a). The co-purification with Tim23 and +ATP is shown as % of -ATP sample (normalized to Tim23, mean  $\pm$  SEM, n=3).



Schulz and Rehling, Supplementary Figure 3

**Supplementary Figure 3: TIM23 complex composition in mutants and inward driving activity.** (a) Isolated wild type (WT) and *ssc1-3* mitochondria were treated at 37°C prior to solubilisation with digitonin buffer and subjected to  $\alpha$ -Tim23 co-immunoprecipitation. Samples were analyzed by SDS-PAGE and Western-blotting. Total, 5%; eluate, 100%. The co-isolation efficiency for Pam16 in mutant mitochondria was reproducibly similar to the wild type sample upon quantification. (b) Experiment as described in (a) performed with WT and *pam18-66* mitochondria.  $\alpha$ -Pam18 efficiently recognizes the mutant form of Pam18 (lane 2). (c) Experiment performed as in (a) with WT and *mgr2Δ* mitochondria except that the heat-shock was omitted. (d) Inward driving activity using *b<sub>2</sub>(220)*-DHFR as described in Fig. 1g. Samples were analyzed by SDS-PAGE and autoradiography. (e) Inward driving activity using

b<sub>2</sub>(220)-DHFR as described in Fig. 1h. Samples were analyzed by SDS-PAGE and autoradiography.



Schulz and Rehling, Supplementary Figure 4

**Supplementary Figure 4: Determination of semi-saturating amounts of b<sub>2</sub>(167)Δ-DHFR for TIM23/TOM co-isolation efficiency.** (a) The indicated amounts of b<sub>2</sub>(167)Δ-DHFR were arrested in mitochondria using MTX for 15 min at 25°C. After reisolation, [<sup>35</sup>S]Atp2 (circle) and [<sup>35</sup>S]Pam18 (square) were imported for 30 min at 25°C and Δψ was dissipated. The reaction was proteinase K (PK) treated and analyzed by SDS-PAGE and digital autoradiography. The amount of imported protein is shown (100%, amount imported in the absence of arrested b<sub>2</sub>(167)Δ-DHFR). (b) 0, 0.5 or 1.5 nmol b<sub>2</sub>(167)Δ-DHFR per mg mitochondria were arrested using MTX for 15 min at 25°C. After Δψ dissipation the reaction was chased for 5 min at 25°C and washed. Samples were solubilised with digitonin and Tom22<sup>His10</sup> was isolated.

Analysis by SDS-PAGE and Western-blotting. Total, 5%; eluate, 100%. **(c)** Quantification of experiments shown in Figure 3b and c. The co-isolation efficiency of the authentic subunits is shown for the isolation with Tim23<sup>PA</sup> arrested with b<sub>2</sub>(167)<sub>Δ</sub>-DHFR (black bar), without b<sub>2</sub>(167)<sub>Δ</sub>-DHFR (dark gray bar) or in wild type (WT) without b<sub>2</sub>(167)<sub>Δ</sub>-DHFR (light gray bar) (% of total). **(d)** Quantification of experiments shown in Figure 3b and c. The co-isolation efficiency of the radiolabeled subunits with Tim23<sup>PA</sup> is shown (% of total). Bars as in **(c)**. **(e)** Radiolabeled Pam16 and Pam18 were imported into WT mitochondria for 20 min. After dissipation of the  $\Delta\psi$  a part of the reaction was proteinase K treated (imported fraction), the remainder was solubilized and subjected to  $\alpha$ -Tim23 co-immunoprecipitation. Samples were analyzed by SDS-PAGE, autoradiography and quantification. The co-isolation efficiency is shown as % of imported (n=3, SEM). **(f)** Radiolabeled Pam16, Pam18 and Tim44 were imported into *mgr2Δ* or *tim21Δ* mitochondria (and the respective WT) for 20 min to saturation of the import reaction. After dissipation of the  $\Delta\psi$  a part of the reaction was proteinase K treated and analyzed by SDS-PAGE and autoradiography. The remaining reaction was processed as described in Fig. 4b.

## Supplementary Figure 5

Original scans of key Western blots and gels presented in the paper

Figure 1d

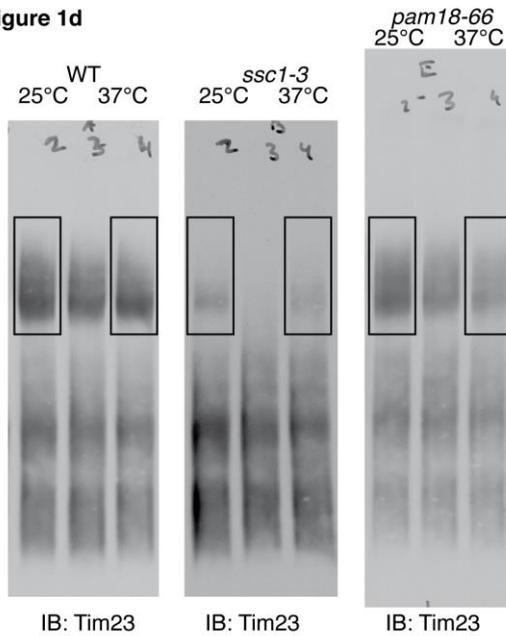


Figure 2c-f

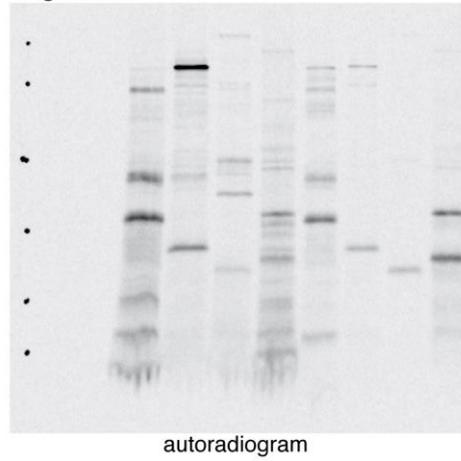


Figure 2g

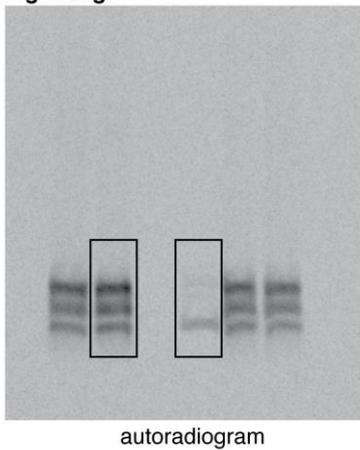


Figure 2h

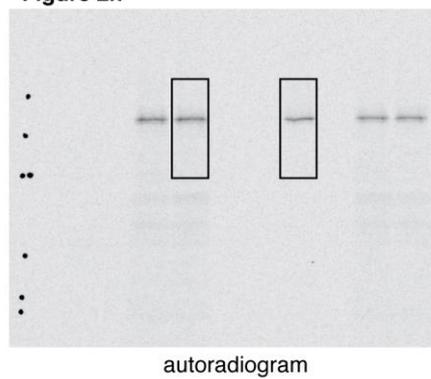


Figure 3b

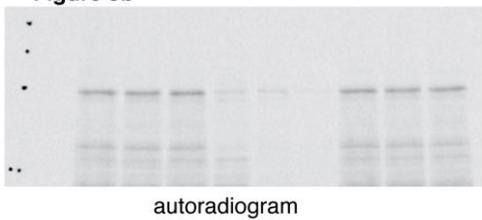


Figure 3b

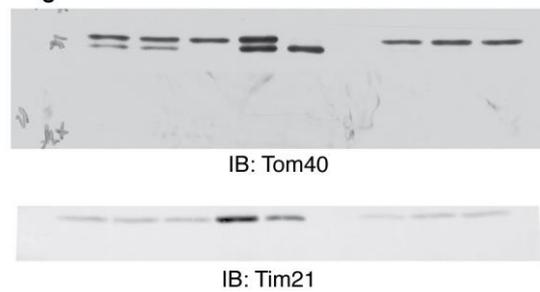


Figure 3c



Figure 3d

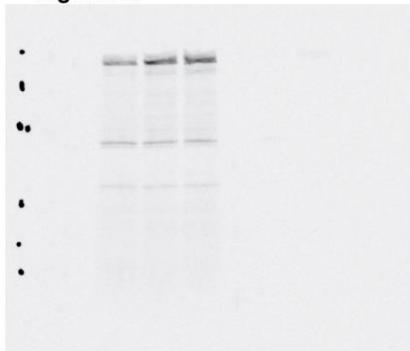


Figure 3e

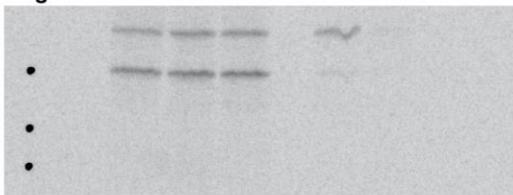


Figure 3f

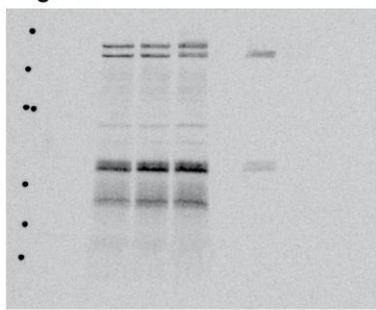


Figure 3c



Figure 3c



Figure 3d



Figure 3d



Figure 3e

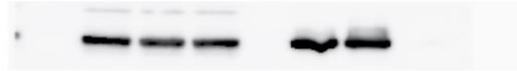


Figure 3e

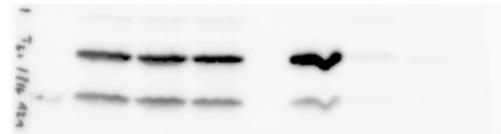


Figure 3f

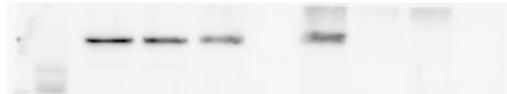


Figure 3f



Figure 3f

