



Simultaneous Real-Time Measurement of Isoprene and 2-Methyl-3-Buten-2-ol Emissions From Trees Using SIFT-MS

Ann-Sophie Lehnert^{1,2*}, Erica Perreca^{3†}, Jonathan Gershenzon³, Georg Pohnert² and Susan E. Trumbore¹

¹ Department of Biogeochemical Processes, Max Planck Institute for Biogeochemistry, Jena, Germany, ² Institute for Inorganic and Analytical Chemistry, Bioorganic Analytics, Friedrich Schiller University, Jena, Germany, ³ Department of Biochemistry, Max Planck Institute for Chemical Ecology, Jena, Germany

OPEN ACCESS

Edited by:

Roger Deal,
Emory University, United States

Reviewed by:

Philippe Heynderickx,
Ghent University Global Campus,
South Korea
Murray McEwan,
University of Canterbury, New Zealand

*Correspondence:

Ann-Sophie Lehnert
alehnert@bgc-jena.mpg.de

† These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Technical Advances in Plant Science,
a section of the journal
Frontiers in Plant Science

Received: 30 June 2020

Accepted: 04 November 2020

Published: 27 November 2020

Citation:

Lehnert A-S, Perreca E,
Gershenzon J, Pohnert G and
Trumbore SE (2020) Simultaneous
Real-Time Measurement of Isoprene
and 2-Methyl-3-Buten-2-ol Emissions
From Trees Using SIFT-MS.
Front. Plant Sci. 11:578204.
doi: 10.3389/fpls.2020.578204

The C5 hemiterpenes isoprene and 2-methyl-3-buten-2-ol (MBO) are important biogenic volatiles emitted from terrestrial vegetation. Isoprene is emitted from many plant groups, especially trees such as *Populus*, while emission of MBO is restricted to certain North American conifers, including species of *Pinus*. MBO is also a pheromone emitted by several conifer bark beetles. Both isoprene and MBO have typically been measured by proton-transfer reaction mass spectrometry (PTR-MS), but this method cannot accurately distinguish between them because of their signal overlap. Our study developed a method for using selective ion flow tube mass spectrometry (SIFT-MS) that allows simultaneous on-line measurement of isoprene and MBO by employing different reagent ions. The use of $m/z(\text{NO}^+) = 68$ u for isoprene and $m/z(\text{O}_2^+) = 71$ u for MBO gave minimal interference between the compounds. We tested the suitability of the method by measuring the emission of young trees of *Populus*, *Picea*, and *Pinus*. Our results largely confirm previous findings that *Populus nigra*, *Picea glauca*, and *Picea abies* emit isoprene and *Pinus ponderosa* emits MBO, but we also found MBO to be emitted by *Picea abies*. Thus SIFT-MS provides a reliable, easy to use, on-line measuring tool to distinguish between isoprene and MBO. The method should be of use to atmospheric chemists, tree physiologists and forest entomologists, among others.

Keywords: conifers, *Picea*, poplar, *Pinus*, VOC, isoprene, MBO, SIFT-MS

INTRODUCTION

The C5-hemiterpene isoprene, or 2-methyl-1,3-butadiene, is the most abundant biogenic volatile compound emitted from vegetation. Its annual global emission is estimated to be 350 to 769 Tg yr^{-1} , approximately half of the total estimated emissions of biogenic volatile organic compounds (BVOC) (Guenther et al., 2012). Isoprene is emitted from mosses, ferns and higher plants, especially trees (Tingey et al., 1987; Hanson et al., 1999; Loreto, 2015). Angiosperms including species of *Populus* emit large amounts of isoprene, while in gymnosperms this hemiterpene is known to be emitted from species belonging to the genus *Picea*, including *Picea abies* and *Picea glauca*,

but not from species belonging to the genus *Pinus*. Instead, the related hemiterpene 2-methyl-3-buten-2-ol (MBO), is emitted by *Pinus* species native to Northern America, e.g., *Pinus ponderosa*, *Pinus lodgepole*, and *Pinus jeffreyi* (Goldan et al., 1993; Harley et al., 1998). Globally, MBO contributions represent only a minor component of total BVOC emissions (Guenther et al., 2012), but in Northern American pine forests, their levels can reach 4–7 times the level of isoprene (Goldan et al., 1993; Harley et al., 1998; Schade and Goldstein, 2001).

2-Methyl-3-buten-2-ol and isoprene are often measured together as a sum parameter due to the experimental restrictions outlined below. However, it is important to distinguish between the two compounds during research in several different fields.

Atmospheric Sciences

The oxidation of BVOCs, such as isoprene and MBO, in the atmosphere can produce tropospheric ozone in sufficiently NO-rich environments (Steiner et al., 2007), influencing air quality and, as ozone is a greenhouse gas, radiative warming. These compounds can also form secondary organic aerosols (Carlton et al., 2009), with both direct and indirect (as cloud condensation nuclei) impacts on radiative balance. Oxidation of MBO by OH-radicals represents one of the most important sources of acetone in those areas where it is emitted (Ferronato et al., 1998). However, the very different lifetimes of isoprene (2.8 h) and MBO (7 h) lead to different spatial and temporal distributions around areas of high emissions (Fantechi et al., 1998; Atkinson and Arey, 2003). Thus, the ability to measure these gases individually with a high time resolution would provide important insights into their relative roles in atmospheric chemistry and climate.

Plant Sciences

Isoprene is thought to protect plants against abiotic stress by its antioxidant properties and stabilization of thylakoid membranes at high temperature (Perreca et al., 2020). Recently this molecule has also been proposed to activate gene networks involved in abiotic stress tolerance (Zuo et al., 2019). Although the role of MBO in plants has not been well studied, it is expected to be similar to that of isoprene based on a similar response of emission rates to light and temperature changes (Schade et al., 2000) and biosynthesis from the same substrate, dimethylallyl diphosphate (Gray et al., 2003). However, due to the differences in chemical properties, the way the two compounds serve in plant protection might differ. Especially the antioxidant properties of MBO might differ from those of isoprene. MBO was detected in the bark extracts of some angiosperms (Zhang et al., 2012) that are known to emit isoprene. Thus care should be taken to distinguish between the two compounds in simultaneous measurement in order to assess if their roles are different.

Entomology

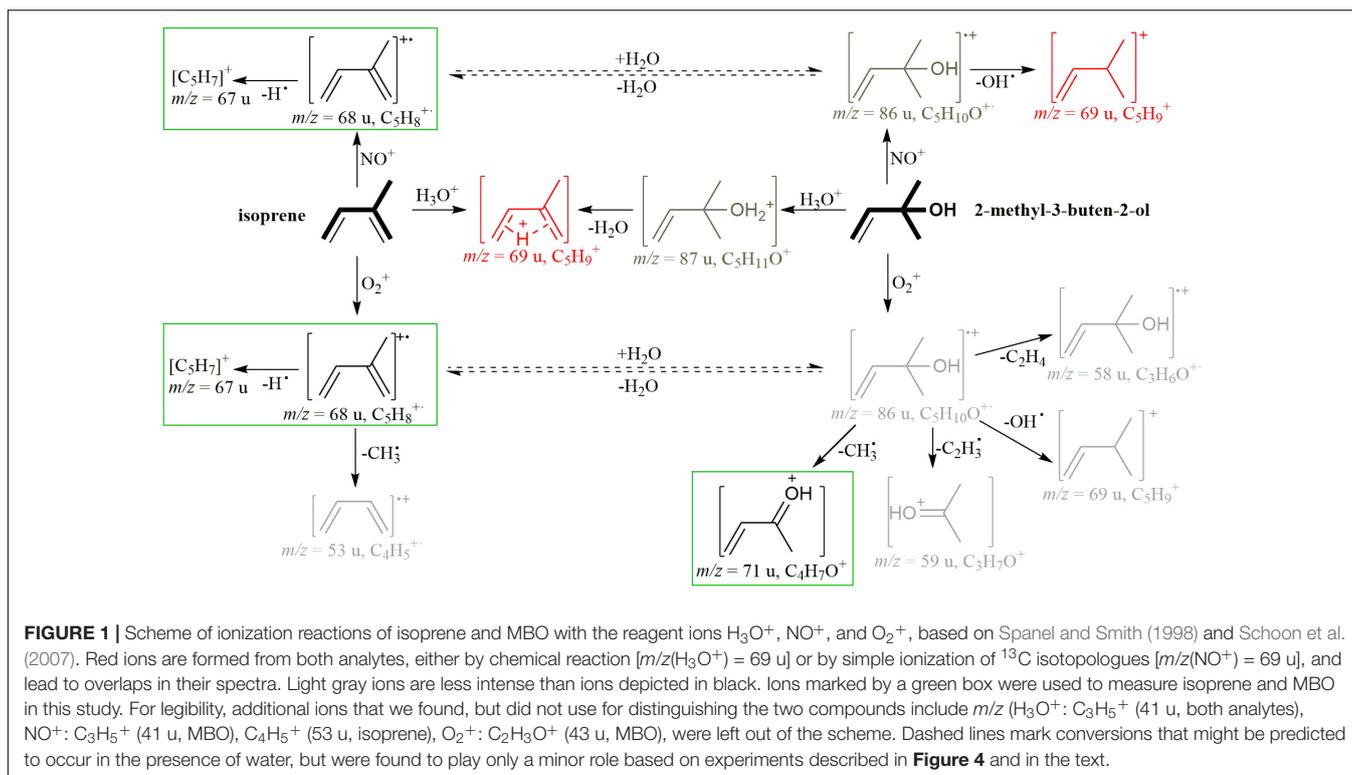
2-Methyl-3-buten-2-ol MBO is produced not only by trees, but also by tree pests. Several conifer bark beetles, e.g., the spruce bark beetle *Ips typographus*, produce MBO *de novo* as an aggregation pheromone (Bakke et al., 1977; Baader, 1989; Zhang et al., 2012). Thus simultaneous measurement of isoprene and MBO might

allow for distinguishing between abiotic stress and bark beetle infestation in field measurements.

Proton-transfer reaction mass spectrometry (PTR-MS) and gas chromatography-mass spectrometry (GC-MS) are the most widely used techniques for measuring BVOCs. PTR-MS ionizes gaseous analytes with H_3O^+ ions in a drift tube with a defined reaction time and detects them via mass spectrometry with a quadrupole or Time of Flight mass analyzer. This technique allows for online-measurements of gaseous analytes at low mixing ratios (Krechmer et al., 2018). However, with PTR-MS, the $[\text{M}+\text{H}-\text{H}_2\text{O}]^+$ fragment of MBO and the $[\text{M}+\text{H}]^+$ signal of isoprene have the same mass to charge ratio (cf. **Figure 1**, H_3O^+ pathway), so accurate distinction of the two compounds is difficult. GC-MS allows for a separation of the analytes based on differences in retention time and mass spectra, but is not suitable for online monitoring due to relatively long measurement times. In addition, GC-MS often requires pre-concentration on, e.g., cartridges before measurement. In practice, both techniques are usually employed side by side, using PTR-MS to acquire good time resolution and GC-MS for identification (Jardine et al., 2020). Another approach involves switchable reagent ion mass spectrometry (SRI-MS), a technique similar to PTR-MS, but including additional ion sources for NO^+ , O_2^+ , NH_4^+ , Kr^+ , and Xe^+ (Jordan et al., 2009). Using the NO^+ ion, isoprene can be detected at $m/z(\text{NO}^+) = 68$ u, and MBO at $m/z(\text{NO}^+) = 69$ u, as has been shown in a field study with SRI-MS (Karl et al., 2012), but these instruments are very costly and complex to operate. Because of the important role that isoprene and MBO play in different scientific fields, the possibility to distinguish between these compounds with accuracy using online measurements is desirable, but until now not realized technically with readily available instruments.

Selected ion flow tube mass spectrometry (SIFT-MS) is a cheaper (though still rather costly) and easy-to-use alternative to SRI-MS. Like PTR-MS and SRI-MS, SIFT-MS is a chemical ionization mass spectrometry technique for measuring gaseous analytes. Unlike PTR-MS, it utilizes multiple reagent ions that react differently with gaseous analytes, so one can obtain more structural information from the respective spectra. By measuring one analyte with more than one reagent ion, multiple spectra are generated. Comparison of these spectra allows identification of ions with the least interference from other VOCs and thus specific quantification of the target analyte.

In SIFT-MS, reagent ions are generated by a moist air plasma and then selected by a quadrupole. Reagent ions and gaseous analytes are mixed in a flow tube that is flushed continuously with a carrier gas. They travel together, and their reaction time is determined by the time they need to cross the flow tube (Smith and Spänel, 2011). During this time, the analytes are ionized during collision with the reagent ions. In our case, the reagent ions used were H_3O^+ , NO^+ , and O_2^+ , but it is also possible to use the negative ions OH^- , O^- , O_2^- , NO_2^- , and NO_3^- . With H_3O^+ , mostly proton transfer reactions occur, with NO^+ electrons are transferred or NO^+ adducts are formed and with O_2^+ , electrons are transferred and sometimes fragmentation reactions occur. The product ions and remaining reagent ions are detected via a quadrupole mass analyzer. A library is



implemented in the software of the manufacturer that can be used to calculate their mixing ratio directly from the measured counts.

When measuring isoprene and MBO with SIFT-MS, H_3O^+ generates $m/z(\text{H}_3\text{O}^+) = 69$ u for both analytes, like in PTR-MS (cf. **Figure 1** for a reaction scheme). However, with NO^+ , isoprene generates $m/z(\text{NO}^+) = 68$ u, and MBO generates $m/z(\text{O}_2^+) = 69$ u. With this difference, one can measure isoprene well, but the ^{13}C isotopologue of isoprene interferes with MBO measurement. With O_2^+ , isoprene forms two product ions, $m/z(\text{O}_2^+) = 67$ u and 68 u, at similar intensities, whereas MBO mostly forms $m/z(\text{O}_2^+) = 71$ u (Spanel and Smith, 1998; Schoon et al., 2007).

In this study, we demonstrate the use of SIFT-MS for simultaneous measurement of isoprene and MBO by monitoring isoprene with $m/z(\text{NO}^+) = 68$ u and MBO with $m/z(\text{O}_2^+) = 71$ u. To validate our method, we performed measurements on three different isoprene-emitting tree species, *Populus nigra*, *Picea abies*, and *Picea glauca*, and on *Pinus ponderosa*, which is reported to emit MBO.

MATERIALS AND METHODS

Supplies and General Remarks

Isoprene, MBO, and dodecane were purchased from Sigma Aldrich (Darmstadt, Germany). Deuterated water was purchased from TCI (Eschborn, Germany). Distilled water was generated by a Enviro FALK GEO + EDI 200 electrode ionization cell (Enviro FALK, Westerborg). The tubing used for the tree chamber experiment was opaque black 1/4" PFA-tubing, the tubing used

for the standard measurements was opaque black 1/8" PFA-tubing. Connectors for the tree experiment were Galtex PFA fittings (Entegris, United States), for the standard measurements and calibrations Swagelok stainless steel fittings (Swagelok, United States). All setups were built such that an overflow line to room air ensured ambient pressure in the chamber and at the SIFT-MS inlet.

SIFT-MS Settings

Measurements were conducted with a Voice 300 ultra SIFT-MS (Syft Technologies Ltd., Christchurch, New Zealand) with a positive ion source that was customized as described by Lehnert et al. (2019). 40 cm³/min sample gas flow, 156 cm³/min helium carrier gas flow, 50 V flow tube voltage, 120°C flow tube temperature, and 105°C sample plate and sampling line heater temperature were used. To suppress dimer formation at high mixing ratios, the larger trees (*P. nigra*, *P. abies*, and *P. glauca* #1) were measured at 390 cm³/min carrier gas flow.

The ratio of the reagent ions NO^+ and O_2^+ varied between the different experiments. However, this did not affect our results significantly since for the interference calculation, measurements from the same ions were used, and for the calculation of mixing ratios and release rates, the ratios of product ion to reagent ion were used.

SIFT-MS Measurements of Isoprene and MBO Standards

Full mass spectra were measured for both standards using a diffusion cell flushed with VOC-free air from a pure air generator

(PAG 003, Ecophysics, Dürnten, Switzerland). An 1.5 mL vial with Teflon septum was filled with 50 μL isoprene or MBO. A thin needle (23 G \times 1") was pierced through the septum, and then the vial was placed in a 40 mL headspace vial that was flushed with 0.5 L/min pure air humidified to 0, 25%, 50%, 75%, and 100% relative humidity at 25°C by a GCU gas calibration unit (IONICON Analytik GmbH, Innsbruck, Austria). The method captured ion counts between 10 and 250 u for all three reagent ions. The dwell time was 100 ms, and the count limit 10,000. 10 scans of each substance were measured and averaged.

For distinguishing the two standards, a selected ion monitoring (SIM) scan was set up for 10 min, with 500 ms dwell time/scan time and 100,000 cps count limit (36 scans, first and last omitted for averaging). The masses used are listed in **Supplementary Table S1**. The increased count limit and scan time compared to standard SIFT-MS settings was used to decrease variability in the measured reagent ion counts. The maximum ratio of product to reagent ions was 3%, so the assumption that the reagent ion counts remain unchanged in the flow tube is still valid.

Ionization of Isoprene and MBO in the Presence of Deuterated Water

We humidified pure air by bubbling it through deuterated water at room temperature. This moist air was mixed with pure air that was enriched in isoprene or MBO, respectively, by passing it over water with 1 μL isoprene or MBO in the diffusion cell. Both flows were 400 mL/min. Mass spectra were recorded between $m/z = 15$ and 150 u, with a scan time limit 1 s, count limit 100,000 counts, and four repeats per measurement. As a control, the experiment was repeated with normal, non-deuterated water.

Tree Cultivation

Populus nigra trees were grown from stem cuttings obtained from trees grown in a common garden of *P. nigra* accessions in Isserstedt, Germany. The 1-year-old trees were grown in the greenhouse of the Max Planck Institute of Chemical Ecology (MPICÖ) Jena, Germany under the following conditions: 20/18°C (day/night), relative humidity 60%, natural light with 9–14 h photoperiod, and supplemental light for 12 h, with SON-T Agro lamps (Philips, Andover, MA, United States).

Three-year-old *Picea abies* trees were planted originally from seeds in 2016 and were grown outdoors in the garden of the MPICÖ, until the experiment was performed. Trees were irrigated every day. One-year-old *Pinus ponderosa* trees were obtained from a local nursery in Thuringia. Four-year-old *Picea glauca* trees (accession #1) were obtained from a local nursery in Thuringia in 2017 and grown prior to the experiment outdoors in the garden of the MPICÖ. Trees were irrigated every day. Three-year-old *Picea glauca* (accession #2) trees were obtained as seedlings from the Laurentian Forestry Centre, Quebec, Canada, in 2016, and grown under controlled environmental conditions in a growing chamber in the MPICÖ until the start of the experiment. Summer (16/8 h for day/night, 22°C and photosynthetically active radiation (PAR) 1000 $\mu\text{mol}/\text{m}^2/\text{s}$) and winter (8/12 h for day/night, 5°C and PAR 200 $\mu\text{mol}/\text{m}^2/\text{s}$)

conditions were alternated for 6 months (summer) and 3 months (winter) in the chamber.

Isoprene and MBO Emissions From Trees

Prior to the experiment, the trees were moved to the greenhouse of the Max Planck Institute of Biogeochemistry, Jena, Germany, and kept there for 4 weeks. The greenhouse was set up at 60% humidity and a 12 h day/night cycle (30°C/25°C). LED-lights (ultra violet, <400 nm, 1%; blue, 400–500 nm, 20%; green, 500–600 nm, 39%; red, 600–700 nm, 35%; far-red, 700–800 nm, 5%; Valoya, Finland) illuminated the trees with a PAR of 150 $\mu\text{mol}/\text{m}^2/\text{s}$ and were supplemented by ambient light entering the greenhouse, reaching a PAR level of 300–400 $\mu\text{mol}/\text{m}^2/\text{s}$. Trees were watered daily. Before performing the measurement of isoprene and MBO with the SIFT-MS, trees were put into the chamber for 24 h [setup similar to Huang et al. (2018), scheme in **Supplementary Figure S1**].

The three tree-containing cylindrical chambers plus one reference chamber without a tree were made from FEP-foil. These chambers (height = 50 cm, diameter = 40 cm, volume = 60 L) were mounted in a polyacrylate scaffold. A Teflon tube ring with holes was placed at the bottom of the chambers and connected to an air inlet. Compressed air was dried and purified on adsorber columns, after which CO_2 was added back in to achieve levels of 400 ppm. Rotameters regulated the air flow through the chamber to 3 L/min. 1/4" black PFA tubing of 2 m length connected the chamber to the instrument. The outlets of the VICI-valve on the SIFT-MS were connected via T-pieces and 1 m tubing to a pump flushing the tubes from the chambers to the instrument at all times. Photosynthetically active radiation (PAR) and temperature were tracked in each chamber. Tree emissions were measured via the SIM scan described above for 24 h capturing a full diurnal cycle. After measuring each chamber with the SIM scan described above, additionally, one full mass spectrum was also measured for every chamber plus the instrument's internal background with the settings mentioned above with a single scan.

Mixing ratios were calculated as:

$$\chi_{\text{isoprene}} = 1.0035 \cdot 10^{-10} \cdot \frac{T_{FT}}{p_{FT}} \cdot \left(\frac{\varphi_{\text{carr}}}{\varphi_{\text{samp}}} + 1 \right) \cdot \frac{I(\text{NO}^+, 68 \text{ u}) \cdot \text{ICF}(\text{NO}^+, 68 \text{ u})}{k_{\text{isoprene}, \text{NO}^+} \cdot br_{\text{isoprene}, \text{NO}^+, 68 \text{ u}} \cdot I(\text{NO}^+, 30 \text{ u}) \cdot \text{ICF}(\text{NO}^+, 30 \text{ u})} \quad (1)$$

and

$$\chi_{\text{MBO}} = 1.0035 \cdot 10^{-10} \cdot \frac{T_{FT}}{p_{FT}} \cdot \left(\frac{\varphi_{\text{carr}}}{\varphi_{\text{samp}}} + 1 \right) \cdot \frac{I(\text{O}_2^+, 71 \text{ u}) \cdot \text{ICF}(\text{O}_2^+, 71 \text{ u})}{k_{\text{MBO}, \text{O}_2^+} \cdot br_{\text{MBO}, \text{O}_2^+, 71 \text{ u}} \cdot I(\text{O}_2^+, 32 \text{ u}) \cdot \text{ICF}(\text{O}_2^+, 32 \text{ u})} \quad (2)$$

χ is the mixing ratio in ppb, T_{FT} the flow tube temperature in K, p_{FT} the flow tube pressure in mTorr, φ_{carr} the carrier gas flow in Torr L/s, and φ_{samp} the sample gas flow in Torr L/s. $I()$ is the intensity of the ion ionized by the reagent ion and measured at

the mass stated, $ICF()$ the instrument calibration factor at the ion as specified, k the kinetic rate constant in $\text{cm}^3 \cdot \text{molecule/s}$ of the reaction with isoprene/MBO with the respective reagent ion, and br the branching ratio of the measured ion. The branching ratios were determined from the standard measurements at 100% humidity, which had the most similar 19/37 signal to the samples. The mixing ratios were calculated for each scan omitting the first and last scan of each measurement, and then the mean and standard deviation were calculated from this. The maximum observed ratio of product ion to reagent ion was 0.9% in the case of isoprene emissions from poplar, which is low enough to fulfill the assumption that the reagent ion counts did not change significantly. The maximum ion counts of $\text{NO}^+ \cdot \text{H}_2\text{O}$, an additional reagent ion water cluster, were always below 500 cps, which corresponded to less than 0.25% of the reagent ion counts and was thus not included in the calculation.

From there, the emission rate was calculated as

$$\phi = \frac{\chi \cdot M \cdot \varphi_{\text{air}}}{V_{\text{mol}} \cdot m_{\text{leaves/needles, dry}}} \quad (3)$$

ϕ is the release rate in $\mu\text{mol}/(\text{g h})$, M is the molar mass of the compound, φ_{air} is the air flow through the incubation chamber (cf. **Supplementary Table S3**), V_{mol} is the molar volume, used 24 L/mol as it was 25°C in the chamber, and $m_{\text{leaves/needles, dry}}$ is the leaf or needle dry mass (cf. **Supplementary Table S2**).

The error of the emission rate was calculated as

$$\Delta\phi = \frac{t(95\%, n_{\text{meas}} - 1)}{n_{\text{meas}} - 1} \cdot \sqrt{M \cdot \left(\left(\frac{\varphi_{\text{air}} \cdot \Delta\chi}{m_{\text{leaves/needles, dry}}} \right)^2 + \left(\frac{\chi \cdot \varphi_{\text{air}} \cdot \Delta V_{\text{mol}}}{V_{\text{mol}}^2 \cdot m_{\text{leaves/needles, dry}}} \right)^2 + \left(\frac{\chi \cdot \Delta\varphi_{\text{air}}}{V_{\text{mol}} \cdot m_{\text{leaves/needles, dry}}} \right)^2 + \left(\frac{\chi \cdot \varphi_{\text{air}} \cdot \Delta m_{\text{leaves/needles, dry}}}{V_{\text{mol}} \cdot m_{\text{leaves/needles, dry}}^2} \right)^2 \right)} \quad (4)$$

with $t(95\%, n_{\text{meas}} - 1)$ the result of the t-distribution at 95%. The degrees of freedom are the number of measurements per time point - 1 (21–22, depending on the measurement). $\Delta\chi$ is the standard deviation of the mixing ratios that was calculated based on the 21–22 individual measurements per time point. $\Delta V_{\text{mol}} = 0.72$ L/mol is the error of the molar volume for 5 K and 0.02 bar deviation of the temperature and pressure. $\Delta\varphi_{\text{air}}$ is the reading error of the gas flow measurements, and $\Delta m_{\text{leaves/needles, dry}}$ the reading error of the weight measurement.

RESULTS

Mass spectra of the standards (**Figure 2**) showed that isoprene and MBO react differently with NO^+ and O_2^+ than previously described (Spanel and Smith, 1998; Schoon et al., 2007). The biggest difference lies in the finding of a strong signal for isoprene at $m/z = 67$ u upon reaction with NO^+ . As the carrier gas flow used and moisture level were similar to those in our study (390 ccm at 100% humidity), we attribute the spectral changes to an increased ion energy leading to increased fragmentation

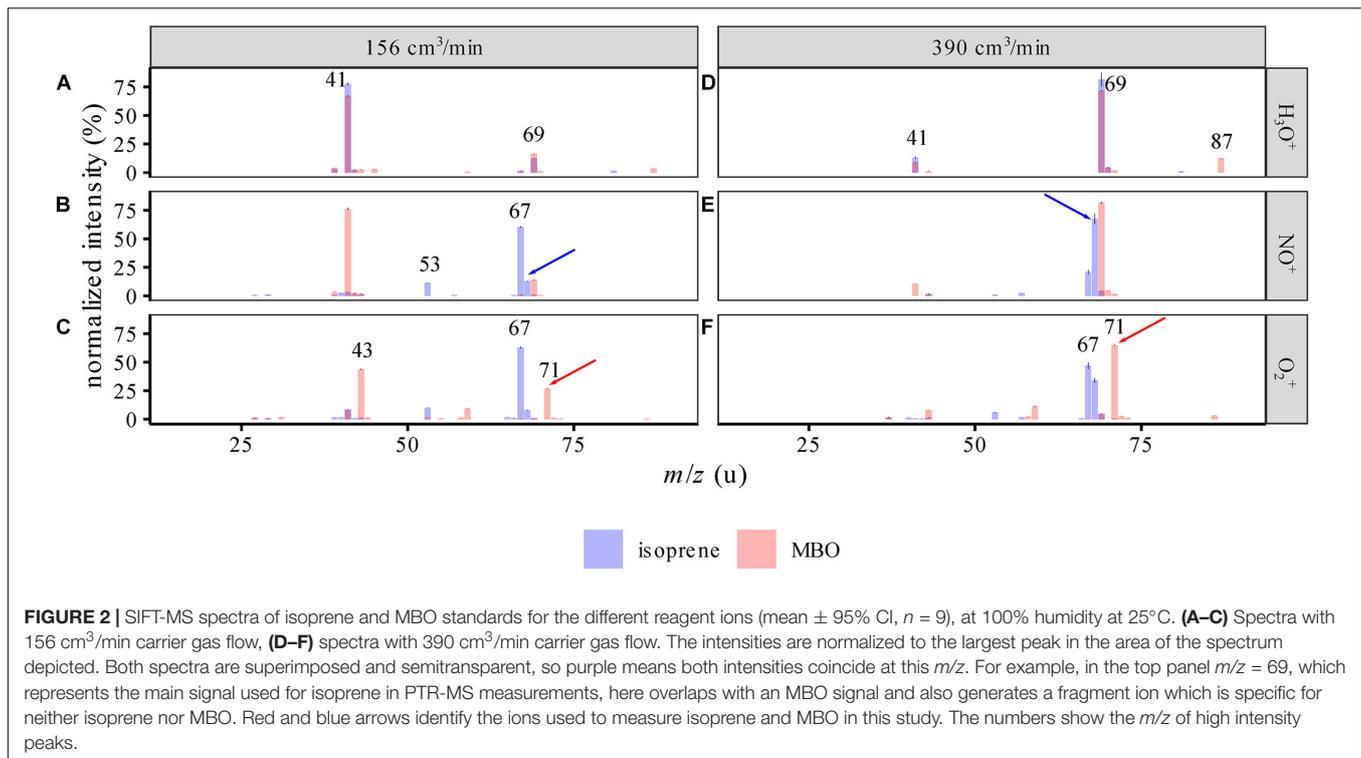
due to the higher flow tube voltage and temperature settings. We observed a decrease in fragmentation when increasing the carrier gas flow (**Figures 2A–C** vs. **D–F**), probably a result of product ions transferring excess energy more rapidly to the carrier gas due to an increased number of collisions.

Upon reaction with NO^+ , one could potentially use the $m/z = 67$ or 68 u signals for measuring isoprene and the $m/z = 69$ u for measuring MBO, as described by Karl et al. (2012). However, limitations in the mass resolution of the quadrupole used in the SIFT-MS resulted in a 5–6% interference [normalized to $m/z(\text{NO}^+) = 68$ u] due to the natural isotopologues of isoprene substituted with a single ^{13}C . When we measured the isoprene standard at different carrier gas flows and humidities, we determined 6% to 8% interference of isoprene with the MBO signal at $m/z(\text{NO}^+) = 69$ u (**Figure 3**). Corrected for the ^{13}C isotope peak, this calculates to a secondary reaction of isoprene to C_5H_9^+ ($m/z = 69$ u) with 1–2% abundance. An explanation for the formation of this ion could be H_2O addition and subsequent OH^\cdot loss, cf. **Figure 1**. Using $m/z(\text{O}_2^+) = 71$ u for measuring MBO was more accurate than using $m/z(\text{NO}^+) = 69$ u, as the interference of isoprene at $m/z(\text{O}_2^+) = 71$ u was below 1% of that at $m/z(\text{O}_2^+) = 67$ u.

We also measured up to 0.7% interference of MBO on the isoprene signal at $m/z(\text{NO}^+) = 68$ u, normalized to $m/z(\text{NO}^+) = 69$ u. This interference was more prominent in dry samples since MBO ionized by NO^+ [$\text{C}_5\text{H}_{10}\text{O}^+$, $m/z(\text{NO}^+) = 86$ u] might not only eliminate OH^\cdot radicals to form C_5H_9^+ [$m/z(\text{NO}^+) = 69$ u], but also eliminate water to form C_5H_8^+ [$m/z(\text{NO}^+) = 68$ u]. In this case, the presence of water would make this side reaction less favorable due to the principle of Le Chatelier. On the other hand, if we had used $m/z(\text{O}_2^+, 67$ u) to measure isoprene, the interference and thus the error would have been much higher, up to 1.9%.

To distinguish between isoprene and MBO, we sought signals of each compound that had the least interference from the other compound to minimize error, which scales with signal intensity. Thus, since the interference of isoprene on $m/z(\text{O}_2^+, 71$ u) is much smaller than on $m/z(\text{NO}^+) = 69$ u, we chose the former for measuring MBO. And, since the interference of MBO on $m/z(\text{NO}^+) = 68$ u is smaller than on $m/z(\text{O}_2^+, 67$ u), we chose the former for measuring isoprene. If the differences in mixing ratios between the two compounds are not anticipated to be large, and rapid measurements are needed with just a single reagent ion, it would be best to use both O_2^+ ions, $m/z(\text{O}_2^+, 67$ and 71 u), as the interference is lower than for the two NO^+ ions and one saves the time of measuring both reagent ions.

2-Methyl-3-buten-2-ol is formally an isoprene molecule with the addition of water to the substituted double bond. It could thus be possible that an ionized form of isoprene could react to form MBO in the presence of water. For example, isoprene ionized by NO^+ forms C_5H_8^+ , which could react with water to form $\text{C}_5\text{H}_{10}\text{O}^+$ with the same structure and m/z as ionized MBO. To evaluate the role of water in these proposed flow tube reactions, the standards were measured in air humidified by either H_2O or D_2O (**Figure 4**). If water is involved in the reaction, and a deuterium from water is added to or exchanged with the ion, the measured mass would be 1 u higher due to the higher



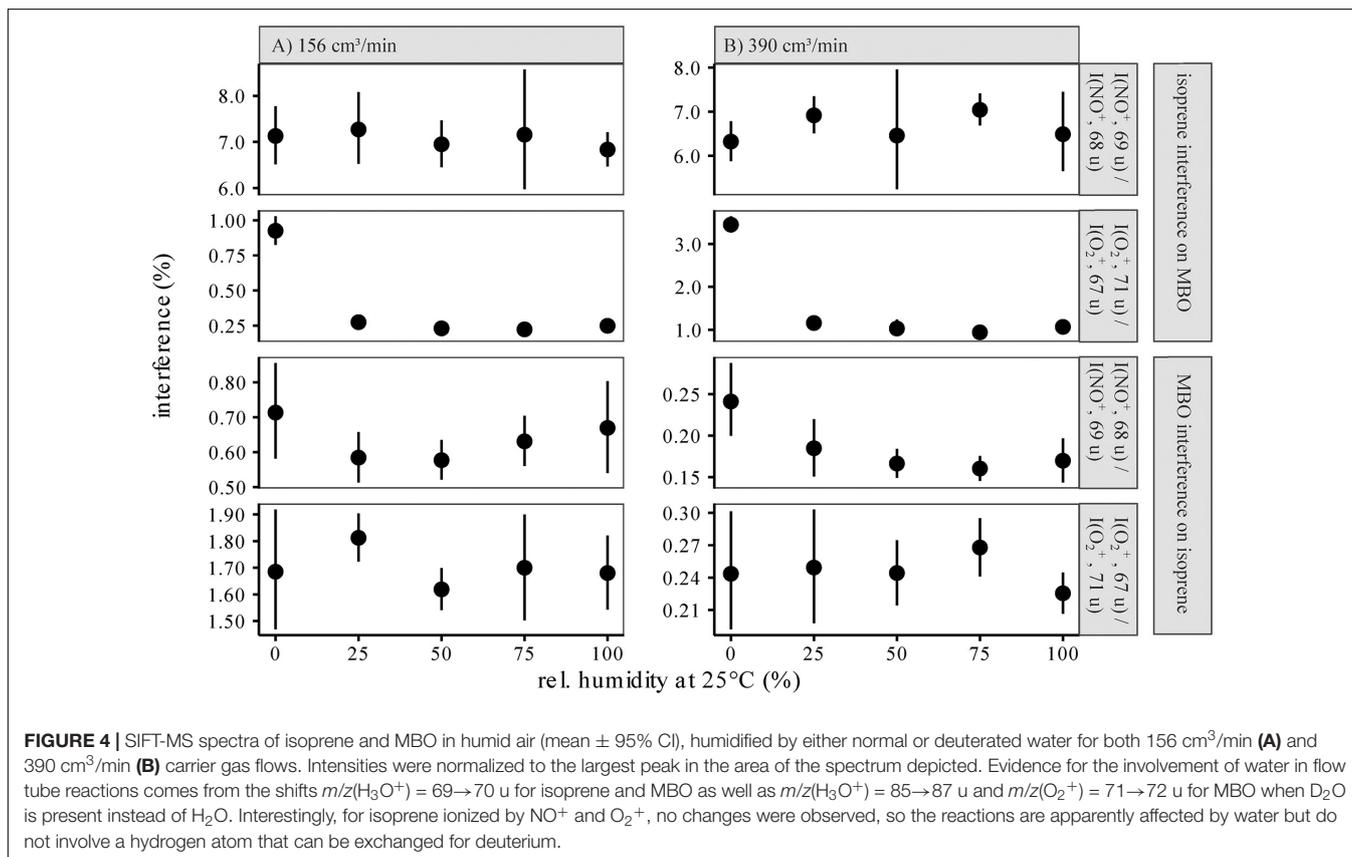
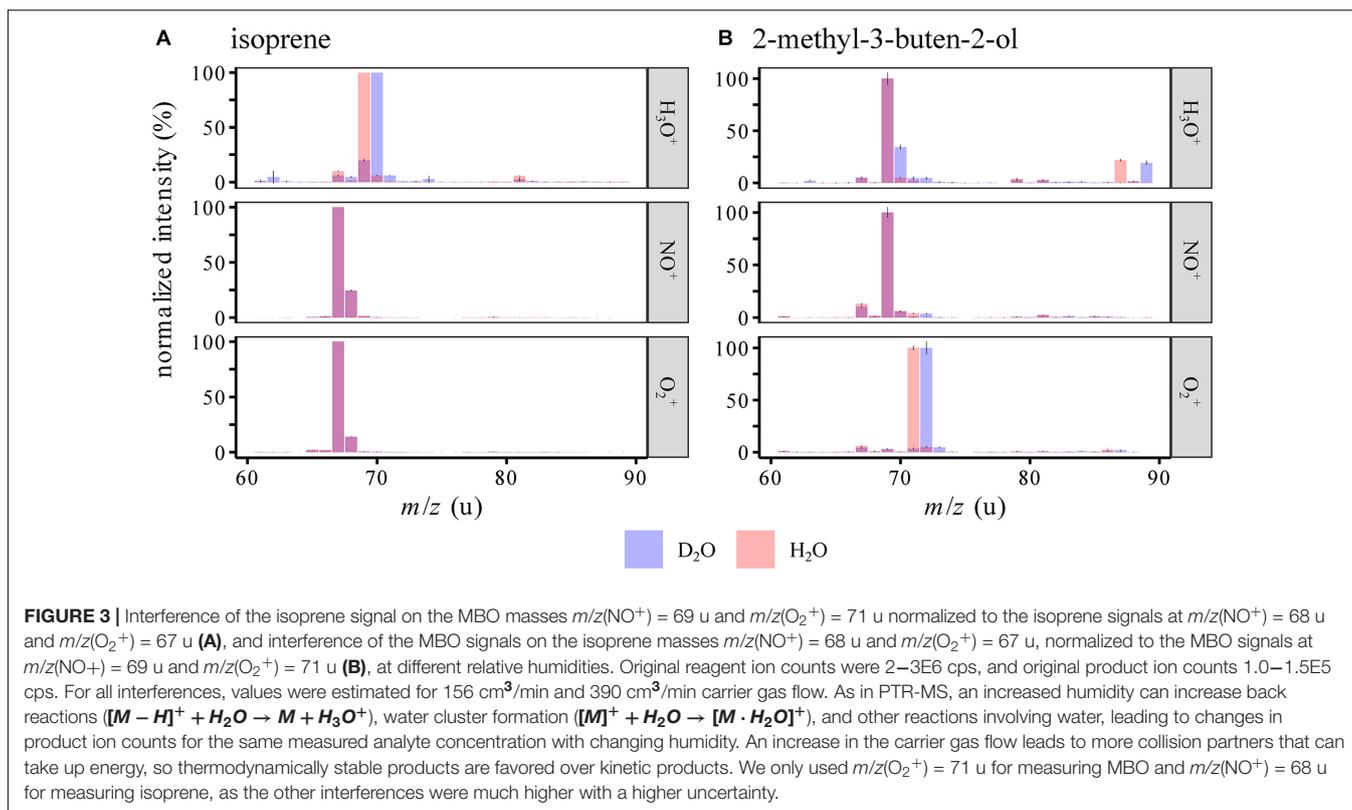
mass of deuterium compared to hydrogen. The reagent ions (see **Supplementary Figure S2**) showed an influence only in the H_3O^+ channel, where H_2DO^+ , HD_2O^+ , and D_3O^+ are detected. As expected, we saw a mass shift from $m/z(\text{H}_3\text{O}^+) = 69$ u to 70 u for both isoprene and MBO, as the reagent ions were both saturated and thus the protonation added a D to the analytes. $m/z(\text{O}_2^+) = 71$ u shifted to 72 u for MBO, so here, also water vapor was involved in forming this ion. Interestingly, for isoprene, the NO^+ and the O_2^+ signals did not change at all, so no proton exchange occurred in the formation of isoprene ions. This contradicts the hypothesis that water is involved in forming $m/z(\text{NO}^+) = 69$ u and $m/z(\text{O}_2^+) = 71$ u. However, we did observe an increase in the relative abundance of those two peaks when switching from dry to wet sample air. This could be due to suppression of the isoprene ions at $m/z(\text{NO}^+) = 68$ u and $m/z(\text{O}_2^+) = 67$ u by excess water.

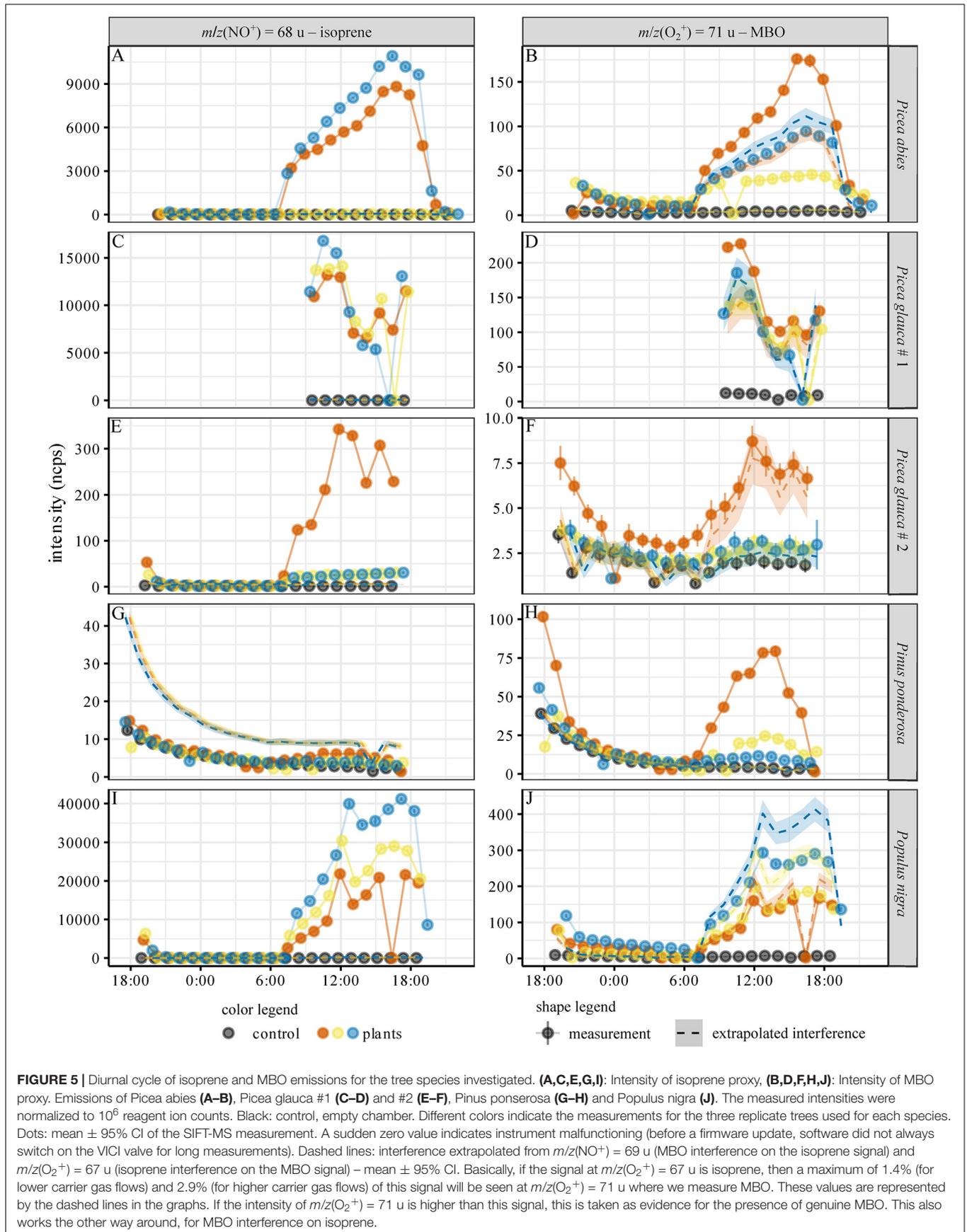
To test our ability to distinguish between isoprene and MBO in an experimental setup with natural sources of these gases, emissions from *Picea glauca*, *Picea abies*, *Populus nigra*, and *Pinus ponderosa* were measured over the course of a day. All trees exhibited a diurnal cycle of BVOC emissions related to the presence of light (**Figure 5**). Isoprene was measured from $m/z(\text{NO}^+) = 68$ u, and MBO was measured from $m/z(\text{O}_2^+) = 71$ u. We calculated the contribution of isoprene to the $m/z(\text{O}_2^+) = 71$ u signal with the equation $I(\text{O}_2^+, 71 \text{ u, isoprene interference}) = x \cdot I(\text{O}_2^+, 67 \text{ u, isoprene})$ (**Figure 3**). If the measured MBO signal is equal or below this value, the signal is not significantly different from the expected isoprene interference and no MBO is actually detected. If the signal is higher, it is measured as MBO. This of course

also applies also for the MBO interference on isoprene, with $I(\text{NO}^+, 68 \text{ u, MBO interference}) = x \cdot I(\text{NO}^+, 69 \text{ u, MBO})$. Again, we used the interference ratio that was determined measuring the standards at 100% humidity, as the trees transpired a substantial amount of water, leading to high air humidity in the chambers. A visualization of the potential isoprene interference with MBO for each species can be found in **Supplementary Figure S4**, where we plotted $I(\text{O}_2^+, 67 \text{ u})$ against $I(\text{O}_2^+, 71 \text{ u})$. Each species shows linear dependence of the two signals, but only *Picea abies* and *Pinus ponderosa* are found significantly above the black line, and so must emit MBO.

Relative humidity does not affect the ability to distinguish between isoprene and MBO except under very dry conditions (**Figure 3**). However, we suggest that interference rates should be determined with standards under conditions as close to the experiment as possible to rule out possible errors.

For each tree species, we analyzed the emission of isoprene and MBO from three individual trees, represented in **Figure 5** by different colors (blue, red, and yellow). The extrapolated interference signals are indicated with a dashed line in the color used for each individual tree. All species except *P. ponderosa* showed isoprene emissions (**Figure 5**), and within the tree species, the emission intensity mostly correlated with the needle or leaf dry mass, cf. **Supplementary Table S2**. For *P. abies*, the isoprene signal for one tree (shown in yellow) was a factor of 150 smaller than the other two (**Figure 5A** and **Supplementary Figure S3**), even though the biomass of this tree individual was the highest (**Supplementary Table S2**). For *P. glauca* (accession #2), one tree (red) had an isoprene signal 10 times higher than the other two, even though the biomass was comparable (**Figure 5E**





and **Supplementary Table S2**). Intraspecific variation was smaller among *P. glauca* (accession #1) and *P. nigra* (**Figures 5C,I**). As the isoprene signal of $m/z(\text{NO}^+) = 68$ u was much higher than the MBO signal of $m/z(\text{NO}^+) = 69$ u, and the calculated interference of MBO based on $m/z(\text{NO}^+) = 69$ u was close to the control in all cases, the isoprene signal did not result from interference of MBO. The isoprene signal for *P. ponderosa* trees was lower than the interference signal extrapolated from the MBO emissions on $m/z(\text{NO}^+) = 69$ u (**Figure 5G**), so we conclude that, as expected, these trees did not emit isoprene, and that the signal at $m/z(\text{NO}^+) = 68$ u in these cases arose from MBO.

A strong signal at $m/z(\text{O}_2^+) = 71$ u was observed from the emissions of *P. ponderosa*. It can be assigned to MBO since no isoprene emission was observed from this plant and thus no interference of this analyte has to be considered. The signals at $m/z(\text{O}_2^+) = 71$ u of both *P. glauca* accessions and *P. nigra* can be attributed to isoprene based on the isoprene signal [$m/z(\text{O}_2^+) = 67$ u]. These trees thus do not produce MBO. Interestingly, two of the *P. abies* individuals emitted MBO as well (**Figure 5B**). The ratio of isoprene and MBO signals differed substantially between the individual trees. Trees indicated with red and blue in **Figure 5** show much higher isoprene emissions at $m/z(\text{NO}^+) = 68$ u than the tree labeled with yellow. In contrast, the MBO emissions at $m/z(\text{O}_2^+) = 71$ u of the three trees are in the same intensity range. Possible interference by other terpenes was considered improbable since no other naturally occurring hemiterpenes are known, and monoterpene emissions measured at $m/z(\text{H}_3\text{O}^+) = 137$ u, $m/z(\text{NO}^+) = 136$ u, and $m/z(\text{O}_2^+) = 136$ u were a factor of 10–100 lower than the measured isoprene and MBO intensities. Only fragment ions from monoterpenes would overlap with isoprene and MBO, and their branching ratio should decrease the intensity even further.

Based on the measured intensities and eq. (1)–(4) above, we calculated the release rates of isoprene and MBO for mid-day (noon), cf. **Table 1** and **Supplementary Figure S4**. Given the low standard deviation, our results give a good idea of relative emission rates for the tree species and individuals involved. Since we could not calibrate the measured intensities as our VOC standard was not concentrated enough to capture the mixing ratio range of the plants, we had to rely on the internal instrument calibration described by Smith and Spanel (2005). As the uncertainty of measurements without external calibration is estimated to be at least $\pm 35\%$ (Langford et al., 2014) including systematic error, our results may not be very accurate. Calculating the isoprene emission rates based on $m/z(\text{O}_2^+) = 67$ u led to fluxes about 2/3 to 1/2 as high as the fluxes calculated from $m/z(\text{NO}^+) = 68$ u (**Supplementary Figure S6**). Thus when rigorous quantification is needed, we strongly recommend calibration.

DISCUSSION

Using SIFT-MS, we developed a method that allows distinguishing between isoprene and MBO in online measurements. For scientific questions where monitoring of both compounds is essential, e.g., for investigation of drought stress and bark beetle infestation or for monitoring BVOC

emissions at the ecosystem level, this is a reliable, easy method. Full scans of isoprene and MBO analytical standards allowed the selection of the ions $m/z(\text{NO}^+) = 68$ u for measuring isoprene and $m/z(\text{O}_2^+) = 71$ u for measuring MBO. These intense ions show the least interference with signals from the other compound and allow a stable and reliable online measurement of the analytes. As proof of concept we applied the method to the determination of isoprene and MBO emissions during the diurnal cycle in five tree species.

SIFT-MS is capable of measuring isoprene and MBO simultaneously under most conditions because of minimal interference between the two compounds for the diagnostic signals we have selected. However, the ratio of these signals depends on the operating conditions of the instrument, especially sample humidity. Thus, these ratios should be determined with standards under identical measurement conditions as used for the sample.

For more accurate quantification of small amounts of isoprene or MBO in the presence of large amounts of the other compound, one could include the humidity-dependence in the interference calculation. In $I(\text{O}_2^+, 71 \text{ u, isoprene interference}) = x \cdot I(\text{O}_2^+, 67 \text{ u, isoprene})$, x could be replaced by a term dependent on the sample humidity, e.g., $x = a \cdot I(\text{O}_2^+, 19 \text{ u}) + b$, which requires a humidity-dependent calibration of all ions. For simplicity, we decided to use the interference factors determined at very high humidity, as this was closest to the humidity in our experiment.

With SIFT-MS, isoprene and MBO can be determined in a single run. In previous approaches described for this analytical problem, a rather laborious measurement of the analytes with GC-MS for identification and PTR-MS for quantification was employed (Jardine et al., 2020). Using PTR-MS, complex calculations were required for a semiquantitative determination of the analytes. The SIFT-MS method introduced here represents a substantial simplification. With the Eqs (1) and (2), no tedious, humidity-dependent calibration is necessary as for PTR-MS. For increased accuracy, a calibration is advised for SIFT-MS as well (Langford et al., 2014; Lehnert et al., 2019).

Isoprene can even be determined in a 50-fold excess of MBO with SIFT-MS, as the MBO interference signal on the isoprene signal is only 0.5%. MBO determination can be accomplished in the presence of a 20-fold excess of isoprene. Limitations to the method are only to be expected if other analytes with the same mass to charge ratios as used for quantification of isoprene and MBO are present in the VOC mixture of the samples. Isoprene and MBO are abundant in natural BVOC samples (Penuelas and Staudt, 2010), thus this limitation should rarely be a major problem.

The ionization mechanism of the two structurally related analytes was investigated by using deuterated water for air humidification. If $m/z(\text{NO}^+) = 69$ u could form from isoprene by addition of water and than elimination of an OH-radical, in a D_2O atmosphere, we should see a mass shift to $\text{NO}^+/70$ u, and likewise for the O_2^+ ion. As we did not see any deuterated product ions forming when the analytes were ionized with NO^+ and O_2^+ in a deuterium-oxide saturated gas stream, the hydration-dehydration mechanism indicated by the dashed lines in **Figure 1** was not substantiated. Thus, the suppression of

TABLE 1 | Isoprene and MBO emission rates in $\mu\text{g}/(\text{g}_{\text{dryweight}} \times \text{h})$ at 12:00 noon.

	Isoprene			MBO		
	1	2	3	1	2	3
<i>P. abies</i>	8.8 ± 0.2	0.040 ± 0.002	19.5 ± 0.4	0.125 ± 0.006	0.029 ± 0.002	n.s.
<i>P. glauca</i> 1	41.7 ± 0.8	13.3 ± 0.2	64 ± 1	n.s.	n.s.	n.s.
<i>P. glauca</i> 2	25.2 ± 0.7	2.9 ± 0.2	3.3 ± 0.2	n.s.	n.s.	n.s.
<i>P. nigra</i>	175 ± 7	174 ± 4	126 ± 3	n.s.	n.s.	n.s.
<i>P. ponderosa</i>	n.s.	n.s.	n.s.	1.42 ± 0.05	0.79 ± 0.05	0.95 ± 0.05

Values listed represent the mean ± 95% CI. n.s.: After correcting for interference of the other analyte, the signal was not significantly different from 0 ($p = 95\%$). The confidence intervals are calculated based on the measured standard deviations of the intensities, however, these values can only be considered as estimates of the emissions, as the instrument is reported to have an actual error of ±34% (Langford et al., 2014) for the mixing ratio calculation employed.

the major ion under higher humidity-conditions is not caused by a formation of the detected side products, but possibly by suppressing the ionization reaction of the analyte itself.

The SIFT-MS measurement of volatiles from several tree species mostly confirmed previous literature reports of isoprene and MBO production (Kesselmeier and Staudt, 1999). *Picea glauca*, *Picea abies* and *Populus nigra* were found to emit isoprene but not MBO, and *Pinus ponderosa* to emit MBO but not isoprene (Figure 5). The relative release rates we measured allow qualitative comparisons among species and individuals over the entire diurnal cycle with a frequency of 15 min per measurement. If desired, this frequency can even be increased by reducing the number of scans per measurement.

In relation to previous measurements of isoprene, the emission rates determined with SIFT-MS were typically higher than those in the literature, though still of the same magnitude (Evans et al., 1982; Steinbrecher, 1989; Janson, 1993; Kempf et al., 1996; Staudt, 1997; Niinemets et al., 2011). Together they confirm previous observations that poplar trees are higher isoprene emitters than conifers (Sharkey et al., 2008; Laothawornkitkul et al., 2009). For MBO, our measurements of *Pinus ponderosa* were lower (Supplementary Figure S5H) than in the literature (Harley et al., 1998). Differences in isoprene and MBO emission between this study and others can be explained at least in part by natural genetic variation of the trees as well as the environmental conditions of measurement. The instrument was calibrated daily using a one-point calibration with a 2 ppm VOC standard as suggested by the manufacturer. This updated the reaction time and the instrument calibration function used in Eqs. (1) and (2) and ensured stable instrument performance. To avoid systematic errors, we recommend calibrating the SIFT-MS under conditions as close to those of the intended experiment as possible. In particular, matching relative humidity is necessary if precise, quantitative values are required.

Among the individuals of *P. glauca* accession #2, one tree emitted isoprene at much higher rates than the other two trees (Supplementary Figure S5E). Genetic variation in isoprenoid formation is very commonly observed within species of *Picea* and other conifers (Martin et al., 2003; Kännaste et al., 2012). Higher isoprene emission could also originate from exposure to slightly different environmental conditions. The high isoprene-emitting tree also had a greater number of flushing buds compared to the other two trees, which could also translate into a higher isoprene emission rate.

In *P. abies*, two of the three experimental trees (entries labeled red and yellow, Table 1) emitted MBO in addition to isoprene, confirming a previous report on simultaneous emission of both volatiles (Hakola et al., 2017) from a species usually considered to be an exclusive isoprene emitter. Since the young *P. abies* trees measured were reared under controlled conditions, MBO is unlikely to have arisen from bark beetle activity. MBO and isoprene are both biosynthesized from dimethylallyl diphosphate but by different terpene synthases (Gray et al., 2011). Neither enzyme has yet been identified in *P. abies*. Since the *P. ponderosa* MBO synthase also produces a trace amount of isoprene in *in vitro* assays (Zeidler and Lichtenthaler, 2001; Gray et al., 2011), one enzyme could in principle produce both isoprene and MBO. Further work is needed on the genetic and biochemical basis of hemiterpene formation in *P. abies*.

In conclusion, we demonstrated that SIFT-MS is suitable for the simultaneous quantification of isoprene and MBO. We introduced a robust easy-to-use online method that requires minimum data treatment. In a proof of principle study, we measured the diurnal cycle of volatile emission of five different tree species with high time resolution. Single 30 cm trees were sufficient to generate robust signals. This method should be useful in applications in plant sciences, entomology, chemical ecology, and atmospheric sciences.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/Supplementary Material. Code and data are published as Lehnert, A., Perreca, E., Gershenson, J., Pohnert, G., Trumbore, S., doi: 10.17617/3.43,2020.

AUTHOR CONTRIBUTIONS

EP and A-SL planned and conducted the tree experiment together. EP organized the trees. A-SL conducted the standard measurements and evaluated all experimental data. EP and A-SL wrote the manuscript together. All authors assisted with data interpretation, discussion of results and helped to improve the quality of the manuscript.

FUNDING

A-SL was financially supported by the Deutsche Forschungsgemeinschaft (DFG) in the frame of the collaborative research center CRC 1076 AquaDiva Project no. 218627073. EP was financially supported by the Max Planck Society and a Max Planck Society-Fraunhofer Society cooperation grant.

ACKNOWLEDGMENTS

Thanks to Agnes Fastnacht and the gardeners of the MPI-CÖ for taking care of the trees in the greenhouse before the experiment, setting up the greenhouse for the experiment, and advice on

REFERENCES

- Atkinson, R., and Arey, J. (2003). Atmospheric degradation of volatile organic compounds. *Chem. Rev.* 103, 4605–4638. doi: 10.1021/cr0206420
- Baader, E. J. (1989). *Pityogenes* Spp.(Col., Scolytidae): investigations on semiochemicals and their application in forest protection. *J. Appl. Entomol.* 107, 1–31.
- Bakke, A., Froyen, P., and Skattebol, L. (1977). Field response to a new pheromonal compound isolated from *Ips typographus*. *Naturwissenschaften* 64, 98–99. doi: 10.1007/bf00437364
- Carlton, A. G., Wiedinmyer, C., and Kroll, J. H. (2009). A review of secondary organic aerosol (SOA) formation from isoprene. *Atmos. Chem. Phys.* 9, 4987–5005. doi: 10.5194/acp-9-4987-2009
- Evans, R. C., Tingey, D. T., Gumpertz, M. L., and Burns, W. F. (1982). Estimates of isoprene and monoterpene emission rates in plants. *Bot. Gaz.* 143, 304–310. doi: 10.1086/botanicalgazette.143.3.2474826
- Fantechi, G., Jensen, N. R., Hjorth, J., and Peeters, J. (1998). Determination of the rate constants for the gas phase reactions of methyl butenol with OH radicals, ozone, NO₃ radicals, and Cl atoms. *Int. J. Chem. Kinet.* 30, 589–594. doi: 10.1002/(sici)1097-4601(1998)30:8<589::aid-kin8>3.0.co;2-o
- Ferronato, C., Orlando, J. J., and Tyndall, G. S. (1998). Rate and mechanism of the reactions of OH and Cl with 2-methyl-3-buten-2-ol. *J. Geophys. Res. Atmos.* 103, 25579–25586. doi: 10.1029/98jd00528
- Goldan, P. D., Kuster, W. C., and Fehsenfeld, F. C. (1993). The observation of a C5 alcohol emission in a North American pine forest. *Geophys. Res. Lett.* 20, 1039–1042. doi: 10.1029/93gl00247
- Gray, D. W., Breneman, S. R., Topper, L. A., and Sharkey, T. D. (2011). Biochemical characterization and homology modeling of methylbutenol synthase and implications for understanding hemiterpene synthase evolution in plants. *J. Biol. Chem.* 286, 20582–20590. doi: 10.1074/jbc.m111.237438
- Gray, D. W., Lerdau, M. T., and Goldstein, A. H. (2003). Influences of temperature history, water stress, and needle age on methylbutenol emissions. *Ecology* 84, 765–776. doi: 10.1890/0012-9658(2003)084[0765:iothws]2.0.co;2
- Guenther, A. B., Jiang, X., Heald, C. L., Sakulyanontvittaya, T., Duhl, T., Emmons, L. K., et al. (2012). The model of emissions of gases and aerosols from nature version 2.1 (MEGAN2.1): an extended and updated framework for modeling biogenic emissions. *Geosci. Model Dev.* 5, 1471–1492. doi: 10.5194/gmd-5-1471-2012
- Hakola, H., Tarvainen, V., Praplan, A. P., Jaars, K., Hemmilä, M., Kulmala, M., et al. (2017). Terpenoid and carbonyl emissions from Norway Spruce in Finland during the growing season. *Atmos. Chem. Phys.* 17, 3357–3370. doi: 10.5194/acp-17-3357-2017
- Hanson, D. T., Swanson, S., Graham, L. E., and Sharkey, T. D. (1999). Evolutionary Significance of isoprene emission from mosses. *Am. J. Bot.* 86, 634–639. doi: 10.2307/2656571
- Harley, P., Fridd-Stroud, V., Greenberg, J., Guenther, A., and Vasconcellos, P. (1998). Emission of 2-methyl-3-buten-2-ol by pines: a potentially large natural source of reactive carbon to the atmosphere. *J. Geophys. Res. Atmos.* 103, 25479–25486. doi: 10.1029/98jd00820

how to treat the trees so that they are not stressed during the experiment. Thanks to Olaf Kolle for help with the PAR and temperature measurements and the whole tree measurement setup. Thanks to Hasan Mahmudul for helping to pick the trees. Thanks to Axel Schmidt, Thomas Fabisch, Danny Kessler, and Almuth Hammerbacher for providing the trees.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2020.578204/full#supplementary-material>

- Huang, J., Hartmann, H., Hellen, H., Wisthaler, A., Perreca, E., Weinhold, A., et al. (2018). New perspectives on CO₂, temperature and light effects on BVOC emissions using online measurements by PTR-MS and cavity ring-down spectroscopy. *Environ. Sci. Technol.* 52, 13811–13823. doi: 10.1021/acs.est.8b01435
- Janson, R. W. (1993). Monoterpene emissions from Scots pine and Norwegian spruce. *J. Geophys. Res.* 98, 2839–2850. doi: 10.1029/92jd02394
- Jardine, K. J., Zorzaneli, R. F., Gimenez, B. O., Oliveira Piva, L. R., Teixeira, A., Fontes, C. G., et al. (2020). Leaf isoprene and monoterpene emission distribution across hyperdominant tree genera in the Amazon basin. *Phytochemistry* 175:112366. doi: 10.1016/j.phytochem.2020.112366
- Jordan, A., Haidacher, S., Hanel, G., Hartungen, E., Herbig, J., Märk, L., et al. (2009). An online ultra-high sensitivity proton-transfer-reaction mass-spectrometer combined with switchable reagent ion capability (PTR+SRI-MS). *Int. J. Mass Spectrom.* 286, 32–38. doi: 10.1016/j.ijms.2009.06.006
- Kännaste, A., Zhao, T., Lindström, A., Stättin, E., Långström, B., and Borg-Karlson, A.-K. (2012). Odors of Norway spruce (*Picea abies* L.) seedlings: differences due to age and chemotype. *Trees* 27, 149–159. doi: 10.1007/s00468-012-0783-7
- Karl, T., Hansel, A., Cappellin, L., Kaser, L., Herdinger-Blatt, I., and Jud, W. (2012). Selective measurements of isoprene and 2-methyl-3-buten-2-ol based on NO⁺ ionization mass spectrometry. *Atmos. Chem. Phys.* 12, 11877–11884. doi: 10.5194/acp-12-11877-2012
- Kempf, K., Allwine, E. J., Westberg, H., Clariborn, C., and Lamb, B. K. (1996). Hydrocarbon emissions from spruce species using environmental chamber and branch enclosure methods. *Atmos. Environ.* 30, 1381–1389. doi: 10.1016/1352-2310(95)00462-9
- Kesselmeier, J., and Staudt, M. (1999). Biogenic volatile organic compounds (VOC): an overview on emission, physiology and ecology. *J. Atmos. Chem.* 22, 23–88.
- Krechmer, J., Lopez-Hilfiker, F., Koss, A., Hutterli, M., Stoermer, C., Deming, B., et al. (2018). Evaluation of a new reagent-ion source and focusing ion-molecule reactor for use in proton-transfer-reaction mass spectrometry. *Anal. Chem.* 90, 12011–12018. doi: 10.1021/acs.analchem.8b02641
- Langford, V. S., Graves, I., and Mcewan, M. J. (2014). Rapid monitoring of volatile organic compounds: a comparison between gas chromatography/mass spectrometry and selected ion flow tube mass spectrometry. *Rapid Commun. Mass Spectrom.* 28, 10–18. doi: 10.1002/rcm.6747
- Laothawornkitkul, J., Taylor, J. E., Paul, N. D., and Hewitt, C. N. (2009). Biogenic volatile organic compounds in the earth system. *New Phytol.* 183, 27–51. doi: 10.1111/j.1469-8137.2009.02859.x
- Lehnert, A.-S., Behrendt, T., Ruecker, A., Pohnert, G., and Trumbore, S. E. (2020). SIFT-MS optimization for atmospheric trace gas measurements at varying humidity. *Atmos. Meas. Tech.* 13, 3507–3520.
- Loreto, F. (2015). Reconciling functions and evolution of isoprene emission in higher plants. *New Phytol.* 206, 578–582. doi: 10.1111/nph.13242
- Martin, D. M., Gershenzon, J., and Bohlmann, J. (2003). Induction of volatile terpene biosynthesis and diurnal emission by methyl jasmonate in foliage of Norway spruce. *Plant Physiol.* 132, 1586–1599. doi: 10.1104/pp.103.021196
- Niinemetts, Ü, Kuhn, U., Harley, P. C., Staudt, M., Arneth, A., Cescaati, A., et al. (2011). Estimations of isoprenoid emission capacity from enclosure

- studies: measurements, data processing, quality and standardized measurement protocols. *Biogeosciences* 8, 2209–2246. doi: 10.5194/bg-8-2209-2011
- Penuelas, J., and Staudt, M. (2010). Bvocs and global change. *Trends Plant Sci.* 15, 133–144. doi: 10.1016/j.tplants.2009.12.005
- Perreca, E., Gershenzon, J., and Eberl, F. (2020). “Tree volatiles: effects of biotic and abiotic factors on emission and biological roles,” in *Biology of Plant Volatiles*, 2nd Edn, eds E. Pichersky and N. Dudareva (Boca Raton, FL: CRC Press), 361–375.
- Schade, G. W., and Goldstein, A. H. (2001). Fluxes of oxygenated volatile organic compounds from a ponderosa pine plantation. *J. Geophys. Res. Atmos.* 106, 3111–3123. doi: 10.1029/2000jd900592
- Schade, G. W., Goldstein, A. H., Gray, D. W., and Lerdau, M. T. (2000). Canopy and leaf level 2-methyl-3-buten-2-ol fluxes from a ponderosa pine plantation. *Atmos. Environ.* 34, 3535–3544. doi: 10.1016/s1352-2310(00)00120-5
- Schoon, N., Amelynck, C., Debie, E., Bultinck, P., and Arijis, E. (2007). A selected ion flow tube study of the reactions of H_3O^+ , NO^+ , and O_2^+ with a series of C5, C6 and C8 unsaturated biogenic alcohols. *Int. J. Mass Spectrom.* 263, 127–136. doi: 10.1016/j.ijms.2007.01.007
- Sharkey, T. D., Wiberley, A. E., and Donohue, A. R. (2008). Isoprene emission from plants: why and how. *Ann. Bot.* 101, 5–18. doi: 10.1093/aob/mcm240
- Smith, D., and Spanel, P. (2005). Selected ion flow tube mass spectrometry (SIFT-MS) for on-line trace gas analysis. *Mass Spectrom. Rev.* 24, 661–700. doi: 10.1002/mas.20033
- Smith, D., and Spanel, P. (2011). Direct, rapid quantitative analyses of BVOCs Using SIFT-MS and PTR-MS obviating sample collection. *TrAC Trends Analyt. Chem.* 30, 945–959. doi: 10.1016/j.trac.2011.05.001
- Spanel, P., and Smith, D. (1998). Selected ion flow tube studies of the reactions of H_3O^+ , NO^+ , and O_2^+ with several aromatic and aliphatic hydrocarbons. *Int. J. Mass Spectrom.* 181, 1–10. doi: 10.1016/s1387-3806(98)14114-3
- Staudt, M. (1997). *Untersuchungen Der Monoterpen-Abgabe an Europäischen Nadelbaumarten in Abhängigkeit Von Umweltfaktoren*. Ph.D. dissertation, Universität Hohenheim, Stuttgart.
- Steinbrecher, R. (1989). *Gehalt Und Emission Von Monoterpenen in Oberirdischen Organen Von Picea Abies*. Ph.D. dissertation, Technische Universität München, München.
- Steiner, A. L., Tonse, S., Cohen, R. C., Goldstein, A. H., and Harley, R. A. (2007). Biogenic 2-methyl-3-buten-2-ol increases regional ozone and HOx sources. *Geophys. Res. Lett.* 34, L15806–L15812.
- Tingey, D. T., Evans, R. C., Bates, E. H., and Gumpertz, M. L. (1987). Isoprene emissions and photosynthesis in three ferns – the influence of light and temperature. *Physiol. Plant* 69, 609–616. doi: 10.1111/j.1399-3054.1987.tb01974.x
- Zeidler, J., and Lichtenthaler, H. K. (2001). Biosynthesis of 2-methyl-3-buten-2-ol emitted from needles of *Pinus ponderosa* via the non-mevalonate DOXP/MEP pathway of isoprenoid formation. *Planta* 213, 323–326. doi: 10.1007/s004250100562
- Zhang, Q.-H., Schlyter, F., and Birgersson, G. (2012). 2-methyl-3-buten-2-ol: a pheromone component of conifer bark beetles found in the bark of nonhost deciduous trees. *Psyche* 2012:414508.
- Zuo, Z., Weraduwege, S. M., Lantz, A. T., Sanchez, L. M., Weise, S. E., Wang, J., et al. (2019). Isoprene acts as a signaling molecule in gene networks important for stress responses and plant growth. *Plant Physiol.* 180, 124–152. doi: 10.1104/pp.18.01391

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Lehnert, Perreca, Gershenzon, Pohnert and Trumbore. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.