

16 ABSTRACT

17 Although non-genetic inheritance is thought to play an important role in plant ecology and evolution,
18 evidence for adaptive transgenerational plasticity is scarce. Here, we investigated the consequences
19 of copper excess on offspring defences and fitness in the giant duckweed (*Spirodela polyrhiza*) across
20 multiple asexual generations. We found that exposing large monoclonal populations (>10,000
21 individuals) for 30 generations to copper excess decreased plant fitness during the first few
22 generations but increased their fitness in consecutive generations under recurring stress when plants
23 were grown for 5 generations under control conditions prior recurring conditions. Similarly,
24 propagating individual plants as single descendants for 5 or 10 generations under copper excess
25 decreased plant fitness when 5 generations and improved plant fitness when 10 generations passed
26 between initial and recurring stress; thus, transgenerational stress responses likely contributed to
27 the observed variations in offspring fitness of long-term copper exposed populations. Fitness benefits
28 under recurring stress were partially associated with avoidance of excessive copper accumulation,
29 which in turn correlated with transgenerationally modified flavonoid concentrations. Taken together,
30 these data demonstrate time-dependent adaptive transgenerational responses under recurring
31 stress, which highlights the importance of non-genetic inheritance for plant ecology and evolution.

32 INTRODUCTION

33 Dwindling intraspecific genetic diversity in both natural and agroecosystems due to human practices
34 has fuelled interest in the ability of species to resist environmental change in the absence of genetic
35 variation through non-genetic inheritance. Non-genetic inheritance is any effect on offspring
36 phenotype that is attributable to the transmission of factors other than the DNA sequence from
37 ancestors [1], and includes vertical transmission of substances (e.g. nutrients, hormones, proteins,
38 mRNA, toxins) [2, 3], epigenetic marks (DNA methylation, histone modification, non-coding small
39 RNAs) [4, 5] and microbes [6]. Non-genetic inheritance may persist for one (parental), two
40 (“multigenerational”) or more generations (“transgenerational”) [7, 8]. Theoretical work suggests
41 that populations may acquire stress resistance through non-genetic inheritance, particularly when
42 transmission fidelity across generations is high [9-11]. Experimental support of these predictions in
43 multicellular organisms is however scarce [6, 12, 13], as such approaches require large-scale multi-
44 generational studies and the ability to disentangle genetic from non-genetic factors.

45 In plants, a number of studies showed that offspring may benefit from parental stress under recurring
46 conditions (e.g. [4, 14-18]). To date, most of these studies focused on parental or multigenerational
47 effects, and consequently, the observed patterns may partially be attributable to direct effects of the
48 trigger on the organism during its early development [6, 19]. Furthermore, the adaptive value of
49 transmitted traits is often inferred through indirect measurements of plant performance (but see
50 [17, 20]), which may not adequately reflect fitness consequences [21]. Thus, directly assessing plant
51 fitness when initial and recurring stress are separated by multiple generations is critical to progress
52 our understanding in the ecological and evolutionary implications of non-genetic inheritance to
53 mediate stress resistance.

54 Non-genetic inheritance might be particularly relevant in asexually reproducing plants [22], as this
55 inheritance mode may compensate for the lack of genetic recombination [23, 24], may by-pass
56 resetting during meiosis [19, 25, 26] and as the ancestor’s and offspring’s environments likely
57 resemble during short-distance vegetative reproduction. Surprisingly however, research on

58 transgenerational plasticity in clonally reproducing plants accumulated only relatively recently (e.g.
59 [23, 24, 27-32]), despite the prevalence and importance of plant asexual reproduction for natural and
60 agroecosystems [33-39]. In clonal plants, stress exposure may alter DNA methylation profiles across
61 one or more generations [28, 31] and modulate offspring growth and traits [27-30]; however,
62 transgenerational stability and fitness consequences of these responses remain mostly unclear.
63 Furthermore, direct evidence that stress exposure benefits offspring fitness when multiple
64 generations lie in between initial and recurring stress is lacking to date.

65 The giant duckweed, *Spirodela polyrhiza* (L.) Schleid., is a fresh water plant that produces rapidly and
66 almost exclusively asexually through budding. This monocot often grows in proximity to agriculture
67 and thus is often recurrently exposed to copper sulphate used in crop protection. Copper excess
68 results in the formation of reactive oxygen species (ROS), which induce oxidative stress [40, 41].
69 Plants resistance to copper excess may involve either exclusion or neutralization of the metal ions
70 [42-44]. Flavonoids, particularly *ortho*-dihydroxylated B-ring-substituted flavonoids, may help plants
71 to resist copper excess through scavenging ROS and suppressing ROS-formation as chelating agents
72 [45, 46]. *Spirodela polyrhiza* accumulates two *ortho*- dihydroxylated and two monohydroxylated B-
73 ring-substituted flavonoids in high concentrations in its flat, thallus-like plant body (“fronds”) [47].
74 The *ortho*-dihydroxylated but not monohydroxylated B-ring substitute flavonoids are associated with
75 copper resistance in *S. polyrhiza* [48].

76 Here, we investigated whether copper excess affects offspring defences and fitness in *S. polyrhiza*
77 across generations. We found that descendants of large monoclonal *S. polyrhiza* populations that
78 were exposed for 30 generations to copper excess had lower fitness under short and higher fitness
79 during prolonged growth under recurring stress. These patterns resembled transgenerational stress
80 responses, as propagating individual plants for 5 or 10 generations under copper excess had negative
81 effects on plant fitness when 5 generations and positive effects when 10 generations passed
82 between initial and recurring stress. Quantifying flavonoids and copper concentrations suggests that

83 avoidance of excessive copper accumulation rather than enhanced induction of anti-oxidative

84 flavonoids contributed to increased plant fitness under recurring stress.

85

86 METHODS

87 *Plant material*

88 *Spirodela polyrhiza* was cultivated under non-sterile conditions in N-medium (KH₂PO₄ 150 μM,
89 Ca(NO₃)₂ 1 mM, KNO₃ 8 mM, H₃BO₃ 5 μM, MnCl₂ 13 μM, Na₂MoO₄ 0.4 μM, MgSO₄ 1 mM, FeNaEDTA
90 25 μM in deionized water). Genotype 7498 from North Carolina (USA) was used for all experiments.
91 Plants were grown in a climate chamber operating under the following conditions: 26°C constant; 18
92 h light; light intensity 130 – 160 μmol m⁻² s⁻¹, supplied by horizontally arranged neon tubes (Osram
93 Lumilux L36 W/ 865 cool daylight; Osram GmbH, Munich, Germany).

94 *Statistical analysis*

95 All statistical analysis was performed in R version 3.5.1 [49] using Rmsic [50], gridExtra [51], dplyr
96 [52], ggplot2 [53], PerformanceAnalytics [54] and nlme [55]. More details on the statistical analyses
97 are given in the experimental sections below.

98 *Long-term exposure experiment*

99 In order to test whether *S. polyrhiza* acquires copper resistance in the absence of genetic variation,
100 we grew replicated monoclonal *S. polyrhiza* populations in the presence and absence of copper
101 excess for four months (approximately 30 generations; “pre-treatment”). Plants were grown inside
102 52 l transparent plastic ponds (79x58x17.5 cm, Bauhaus, Germany) that were filled with 30 l N-
103 medium with or without 20 μM CuSO₄ (N = 10 each). The ponds were covered with 4 mm transparent
104 plexiglass (UV Gallery100, Sandrock, Germany) with 5 mm distance between pond edges and plates.
105 Every two weeks – when the control populations covered approximately the entire pond – plants
106 covering about 5% of the total pond surface were randomly chosen and transferred into refilled
107 ponds. In the first four weeks of the experiment, the maximum population size per pond reached
108 approximately 27,000 and 16,000 fronds in the control and copper medium, respectively.
109 Subsequently, algae colonized the ponds, which reduced maximum population size in the control but
110 not copper medium to approximately 10,000.

111 Four months after start of the experiment, plant fitness of control and copper pre-treated
112 populations was assessed in both environments across 16 days of growth (approx. 4-6 generations).
113 To avoid growth bias due to direct effects of copper excess, plants were propagated for 5
114 consecutive generations as single descendants under control conditions. To this end, 5 fronds
115 carrying a small daughter (generation 0) of each pond were transferred to transparent 50 ml
116 polystyrene tubes (\varnothing 2.8 cm, height 9.5 cm, Kisker, Germany) covered with foam plugs (Kisker,
117 Germany) and filled with 30 ml control medium, with one frond per tube. The first daughter was
118 separated from the mother and placed inside a new tube once the daughter had fully emerged.
119 Subsequently, to measure plant fitness in the absence of copper excess, a pool of the first daughter
120 from generation four (5 fronds) of each pond was placed inside one half of the 18 L containers that
121 were divided in the middle by a fine mesh, filled with 10 l N-medium and covered with transparent
122 PET lids (Pöppelmann, Lohne, Germany). Each container received the fronds of copper and control
123 pre-treated populations to avoid growth bias due to potentially co-evolved microbes. To measure
124 plant fitness in the presence of copper excess, a pool of the second daughters from generation four
125 of each pond were placed inside containers filled with N-medium containing 20 μ M CuSO₄ as
126 described above. After 8 days of growth, the total number of fronds was counted and the growth
127 medium exchanged. After 16 days, the number of fronds was counted once more, plants were
128 subsequently dried with a tissue paper and fresh weight was determined. Relative growth rates
129 (RGR) were calculated as $RGR = (\ln(N_2) - \ln(N_1)) / (t_2 - t_1)$ [56], with N = population size and t = day. Pre-
130 treatment effects on plant fitness were assessed by comparing RGR between pre-treatments for each
131 offspring environment separately using Kruskal-Wallis rank sum tests. Pre-treatment effects on RGR
132 were expressed as the ratio in RGR of copper to control pre-treated offspring. Resistance was
133 analysed as the difference in these pre-treatment effects on RGR between offspring environments
134 using Kruskal-Wallis rank sum tests. To test for interactions of the pre-treatment and growth interval
135 on RGR, two-way analysis of variance (ANOVAs) were performed. To test for interactions of the pre-
136 treatment and offspring environment on RGR and biomass (log) accumulation, two-way ANOVAs

137 were performed. Biomass accumulation was log-transformed (natural logarithm) prior statistical
138 analysis to account for the exponential growth of *S. polyrhiza*.

139 *Transgenerational experiment*

140 To investigate transgenerational stress responses, we propagated individual *S. polyrhiza* plants as
141 single descendants for different durations under copper and control conditions (Fig S1 in
142 Supplemental Information). Individual mother plants carrying a small daughter were placed inside 50
143 ml polystyrene tubes filled with 30 ml N-medium containing or lacking 20 μ M CuSO₄, and grown as
144 single descendants as described above for 2, 5 or 10 generations (“pre-treatment”; N = 25 each). The
145 starting time points of the control lineages were delayed to allow simultaneous assessment of plant
146 fitness later on. After the pre-treatment phase, propagation continued for 5, 10 or 15 generations in
147 control medium (“recovery”), after which offspring growth assays inside transparent 250 ml
148 polypropylene beakers (Plastikbecher GmbH, Giengen an der Brenz, Germany) filled with 180 ml
149 medium and covered with a perforated and transparent lid were performed. For these assays, the
150 first daughters were transferred to copper-free medium, whereas the second daughters were
151 subjected to medium containing 20 μ M CuSO₄ (“initial fronds”). An image for surface area analysis
152 was taken at a camera installation with a webcam (HD Pro Webcam C920, Logitech; webcam
153 software Yawcam version 0.6.0) and a subjacent adjustable LED light after setting up the experiment.
154 During the growth assays, offspring that directly emerged from the initial fronds were marked
155 (“direct offspring”). The growth medium was exchanged four days after the start of the experiment.
156 After 8 days of growth, the number of direct offspring was counted, an image was taken, fronds were
157 gently dried with a tissue paper and the fresh weight of the initial frond and all other plants
158 (“offspring”) was determined. The initial frond and the offspring were flash-frozen separately inside
159 tubes in liquid nitrogen and stored at -80 °C until further analysis.

160 Frond surface area was measured with ImageJ (version 2.0.0-rc-43/1.51k; Java version 1.6.0_24).
161 Flavonoid concentrations were measured on an HPLC 1100 series equipment (Agilent Technologies)

162 coupled to a photodiode array detector (G1315A DAD, Agilent Technologies) and quantified as
163 described in [48]. To measure plant copper concentration, 20-50 mg ground plant material was
164 extracted with 1 ml 0.01 M MOPS buffer. Samples were centrifuged at 17,000 g and supernatant
165 diluted 1:2 and 1:3 in 0.01 M MOPS buffer for control and copper-subjected plants, respectively.
166 Copper concentration was measured using a Copper Assay Kit (Sigma Aldrich) following the
167 manufacturer's instructions on an Infinite® 200 PRO NanoQuant Plate Reader Spectrophotometers
168 (Tecan Trading AG, Männedorf, Switzerland) at 359 nm. Absolute concentrations were calculated
169 based on an external standard curve.

170 Differences in biomass (log) accumulation, surface area, area- and biomass-based growth rates, the
171 fresh weight of the initial frond and the number of offspring of the initial frond between copper and
172 control pre-treated offspring were analyzed with Kruskal-Wallis rank sum tests for each assay and
173 offspring environment separately. To test for interactions of the pre-treatment and the offspring
174 environment on these parameters, two-way ANOVAs were performed. Growth rates were calculated
175 as described above with N = surface area or fresh weight, respectively. Fitness effect of copper pre-
176 treatment was expressed as the ratio in biomass (log) accumulation of copper to control pre-treated
177 offspring. Resistance was analyzed as the difference in these pre-treatment effects between
178 offspring environments using Kruskal-Wallis rank sum tests. Differences in flavonoid concentrations
179 and plant copper concentration between copper and control pre-treated offspring were analyzed
180 with Student's *t*-tests for each assay and offspring environment separately. To test for interactions of
181 the pre-treatment and offspring environment on flavonoid and copper accumulation, two-way
182 ANOVAs were performed. Pre-treatment effects on flavonoid and copper accumulation were
183 expressed as the ratio in flavonoid or copper concentration of copper to control pre-treated
184 offspring, respectively. The sum of the two major mono- (apigenin 8-C- and 7-O-glucoside) and
185 dihydroxylated (luteolin 8-C- and 7-O-glucoside) B-ring substituted flavonoids were analyzed
186 separately. Due to experimental errors, the number of replicates was reduced in some assays. The
187 correlation among the different fitness parameters, i.e. offspring biomass (log) accumulation, the

188 number of direct offspring and initial frond fresh weight, surface area (log), surface-area and
189 biomass-based growth rates, were calculated using Pearson's moment correlations. Mixed effect
190 models were applied to assess the correlation between pre-treatment effects on offspring copper
191 and flavonoid concentrations, as well as between offspring copper and biomass (log) accumulation
192 using maximum likelihood estimations with assay as a random effect. Significant correlations were
193 assessed by comparing two models with and without flavonoid or biomass (log) accumulation,
194 respectively, as a fixed effect using ANOVA. For the correlation between pre-treatment effects on
195 copper and biomass (log) accumulation, additional models were analyzed in which the data point
196 with extremely high pre-treatment effect on biomass accumulation (> 1.4) was excluded.

197 RESULTS

198 To test whether *S. polyrhiza* may acquire stress resistance in the absence of genetic variation, we
199 grew large monoclonal populations for 30 generations in the presence and absence of copper excess
200 (“pre-treatment”) and subsequently propagated offspring for 5 generations under control conditions
201 prior measuring offspring growth rates across 16 days. In the first 8 days of growth, copper pre-
202 treatment reduced offspring growth rates under both control and copper excess to a similar extent (-
203 16%, $P < 0.001$ and -26%, $P = 0.04$, respectively, Kruskal-Wallis rank sum tests; Fig 1 and Fig S2); thus,
204 the pre-treatment did not alter plant resistance (difference in the pre-treatment effects on plant
205 growth rates between offspring environment; $P = 0.20$, Kruskal-Wallis rank sum tests, Fig 1). In the
206 consecutive 8 days, copper pre-treatment enhanced offspring growth rates under copper excess
207 (+36%, $P = 0.005$) while it had no effect under control conditions ($P = 0.56$, Kruskal-Wallis rank sum
208 tests, Fig 1, Fig S2); consequently, plant resistance increased upon copper pre-treatment ($P = 0.003$,
209 Kruskal-Wallis rank sum test, Fig 1). Across the entire 16 days, copper pre-treatment reduced growth
210 rates under control conditions (-14%, $P = 0.002$) and had no effect on growth rates under copper
211 excess ($P = 0.94$, Kruskal-Wallis rank sum tests, Fig S2). Consequently, copper pre-treatment reduced
212 growth depression by copper excess across these 16 days of growth ($P < 0.001$, two-way ANOVA, Fig
213 S2). Assessing plant resistance and fitness based on plant biomass accumulation instead of growth
214 rates exhibited similar patterns (Fig S3). Taken together, these data show that long-term growth of *S.*
215 *polyrhiza* under copper excess may benefit plant fitness and resistance under recurring stress.

216 The observed variation in plant fitness and resistance of long-term pre-treated populations may have
217 been the consequence of selection of newly acquired phenotypes or due to transgenerational stress
218 responses in the absence of selection. To investigate on the latter, we investigated offspring defence
219 and fitness after different durations under copper excess (“pre-treatment”; 2-10 generations) as well
220 as after different durations under control conditions prior recurring stress (“recovery”; 5-15
221 generations) using single descendant propagations, in which no selection takes place (Fig S1). Under
222 5 generations recovery, two generations of copper pre-treatment had positive effects on plant

223 biomass accumulation both in the presence as well as in the absence of recurring stress. Prolonging
224 the pre-treatment to 5 generations eliminated these beneficial effects in both offspring environment.
225 Further expanding the pre-treatment to 10 generations resulted in negative effects on biomass
226 accumulation in the presence of recurring stress, and neutral effects in the absence of recurring
227 stress (Kruskal-Wallis rank sum tests, Fig 2, Fig S4). Thus, under 5 generations recovery, copper pre-
228 treatment had neutral (two and 5 generations pre-treatment) or negative effects (10 generations
229 pre-treatment) on plant resistance (Kruskal-Wallis rank sum tests, Fig 2). Under 10 generations
230 recovery, two generation pre-treatment elicited positive effects on plant biomass accumulation in
231 the presence and absence of recurring stress, similar to the observations under 5 generations
232 recovery. However, prolonging the pre-treatment to 5 and 10 generations had beneficial effects on
233 offspring biomass accumulation in the presence and negative effects in the absence of recurring
234 stress (Kruskal-Wallis rank sum tests, Fig 2, Fig S4). Thus, plant resistance increased under these pre-
235 treatment and recovery combinations (Kruskal-Wallis rank sum tests, Fig 2). Under 15 generations
236 recovery, pre-treatment did not affect biomass accumulation in the presence or absence of stress
237 (Kruskal-Wallis rank sum test, $P = 0.94$, Fig 2, Fig S4). Other fitness parameters, i.e. surface area,
238 growth rates based on surface area and biomass accumulation, as well as the initial plant's fresh
239 weight and offspring number, were all strongly correlated with each other and with offspring
240 biomass accumulation ($R^2 > 0.49$, $P < 0.001$, Pearson's correlation tests, Fig S5), and exhibited similar
241 pre-treatment effects (Figs S6-S10), thus corroborating the above described findings based on
242 offspring biomass accumulation. Taken together, these data demonstrated that copper excess may
243 affect plant fitness and resistance under recurring conditions, and the magnitude and direction of the
244 effects are dependent on the duration of the pre-treatment and recovery phase.

245 To assess whether pre-treatment effects on biomass accumulation were due to altered accumulation
246 of defensive metabolites, we measured the concentrations of the four major flavonoids in offspring.
247 Depending on the pre-treatment and recovery phase combination, copper pre-treatment enhanced
248 the accumulation of the dihydroxylated B-ring substituted flavonoids, including elevated basal and

249 induced levels as well as only elevated induced levels (= primed) after up to 15 generations of
250 recovery (Student's *t*-tests, Fig 3). However, in the assays that exhibited transgenerationally elevated
251 flavonoid levels, no alteration in plant resistance by copper pre-treatment was observed (Fig 3). In
252 contrast, in the two pre-treatment and recovery phase combinations with increased plant fitness and
253 resistance under recurring stress (5 and 10 generations pre-treatment, 10 generations recovery),
254 copper pre-treatment had either neutral (10 generations recovery) or negative (5 generations
255 recovery) effects on the concentration of the dihydroxylated B-ring substituted flavonoids under
256 control conditions, and no effect on the accumulation of these metabolites under recurring stress
257 (Student's *t*-tests, Fig 3). Across all assays, the pre-treatment elicited very similar effects on all four
258 major flavonoids (Figs S11-S15) - albeit with stronger effects in the di - than monohydroxylated B-
259 ring substituted flavonoids - except that the monohydroxylated B-ring substituted flavonoids were
260 not primed ($P = 0.7$, Student's *t*-tests, Fig S13). Taken together, these data show that copper pre-
261 treatment may transgenerationally increase, decrease and prime dihydroxylated B-ring substituted
262 flavonoids when initial and recurring stress were separated by up to 15 generations, and indicate
263 that flavonoids may not be the sole contributor for enhanced offspring fitness under recurring stress.

264 To investigate whether altered copper uptake or excretion efficiency may account for the observed
265 pre-treatment effects in plant fitness and flavonoid accumulation, we measured offspring copper
266 concentration in a subset of these assays (Fig 4, Fig S16). In the assay in which copper pre-treatment
267 benefited plant resistance and reduced basal flavonoid concentrations (5 and 10 generation
268 exposure and recovery, respectively), offspring copper concentrations were lower in copper
269 compared to control pre-treated offspring (pre-treatment: $P = 0.06$, two-way ANOVA), particularly in
270 the presence of recurring stress ($P = 0.049$, Student's *t*-test, Fig 4). In the second assay with beneficial
271 effects of copper pre-treatment on offspring resistance (10 generations pre-treatment and recovery
272 each), pre-treatment did not affect plant copper accumulation (Student's *t*-tests, $P > 0.86$, Fig S16). In
273 the assay with elevated basal and induced flavonoid levels (5 generations exposure and recovery
274 each), copper pre-treated offspring exhibited increased copper concentration ($P = 0.009$, two-way

275 ANOVA), both in the presence ($P = 0.09$) and absence ($P = 0.07$, Student's t -tests) of recurring stress
276 compared to control pre-treated offspring (Fig S16). Across all assays, pre-treatment effects of
277 copper accumulation tended to be negatively correlated with pre-treatment effects of offspring
278 biomass (log) accumulation ($P = 0.06$, mixed effect models, Fig 5A), but only if the assay with
279 extremely high benefits of copper pre-treatment on plant fitness under recurring stress was excluded
280 from the analysis. Furthermore, pre-treatment effects of copper accumulation were positively
281 correlated with pre-treatment effects of total and individual flavonoid accumulation ($P = 0.02$; mixed
282 effect models, Fig 5B, Fig S17). Thus, pre-treatment effects of plant copper accumulation closely
283 reflected pre-treatment effects of plant flavonoid concentrations, and avoidance of excessive copper
284 accumulation in copper pre-treated offspring was partially associated with improved plant fitness
285 and resistance under recurring stress.

286

287 DISCUSSION

288 The importance of non-genetic inheritance is a major controversy in plant ecology and evolution [8,
289 26, 57-59], also because clear evidence for adaptive responses across multiple generations is scarce.

290 Here, we showed that in the clonal fresh water plant *S. polyrhiza* copper excess may have positive
291 effects on offspring fitness when initial and recurring stress conditions were separated by multiple
292 generations. Thereby, this study underlines the notion that non-genetic inheritance plays an
293 important role in the ecology and evolution of asexual plants.

294 When large monoclonal populations of *S. polyrhiza* were grown for four months (approximately 30
295 generations) under copper excess, offspring of copper pre-treated populations exhibited under
296 recurring stress lower plant fitness in the first 8 days of growth (2-3 generations) and higher plant
297 fitness in the consecutive 8 days compared to offspring of control pre-treated populations, even
298 though plants were grown for 5 generations in the absence of stress prior fitness assays. Genetic
299 mutations arise very slowly in *S. polyrhiza* (approximately 0.004 point mutation in the protein coding
300 sequence per generation) [60] and thus very unlikely account for the observed patterns. Instead,
301 these variations in fitness may on the one hand be the result of intra-clonal selection of new
302 phenotypes that were either induced by copper excess or randomly occurred through non-genetic
303 processes [61, 62]. On the other hand, such patterns may be due to transgenerational responses, in
304 which the pre-treatment alters offspring fitness without selection of phenotypic variants [11, 17, 20,
305 26, 58]. Indeed, variation in fitness between long-term copper and control pre-treated populations
306 were similar to the observed transgenerational responses during single descendant propagation.
307 Thus, transgenerational responses likely contributed to the observed variation in plant fitness
308 between long-term copper and control pre-treated populations.

309 In our transgenerational experiment, we found that copper pre-treatment may have positive effects
310 on offspring fitness in the presence and negative effects in the absence of recurring stress when 10
311 generations separated initial and recurring stress. To our knowledge, such long-term

312 transgenerational effects on plants fitness have not been reported to date. In most studies,
313 transgenerational effects on plant performance and phenotype vanish after one to two generations
314 [4, 11, 58, 63, 64], but see [28-30, 65, 66]. As most studies however use indirect estimates of plant
315 fitness such as defensive phenotypes and biomass accumulation in sexual plants, which may not
316 adequately reflect fitness consequences [6, 21], the adaptive value of the observed responses are
317 often equivocal. By assessing plant fitness based on biomass accumulation in an asexual, thallus-like
318 plant, we could directly demonstrate that copper excess may have both positive as well as negative
319 effects on offspring fitness when multiple generations separate initial and recurring stress.

320 Interestingly, we observed that copper pre-treatment can have negative plant fitness effects after
321 short (5 generations), and positive effects after long (10 generations) recovery period. Similarly,
322 offspring flavonoid concentrations showed a large variation of different pre-treatment patterns
323 including elevated basal and induced levels, as well as priming, and these effects did not simply
324 weaken with increasing recovery time. Such contrasting effects in plant phenotypes and fitness
325 across generations have been reported previously in both plants and animals [59, 67]. This shows
326 that transgenerational effects may not simply decay over generations, as often predicted and
327 observed [11, 65, 67-69], and highlights the importance of the timing in transgenerational responses
328 [68].

329 Our analysis on offspring phenotypes suggest that reduced copper accumulation under recurring
330 stress may account for beneficial effects of copper pre-treatment on offspring fitness under recurring
331 stress. First, in the only assay in which copper pre-treatment reduced offspring copper concentration,
332 copper pre-treatment benefited offspring fitness under recurring stress. Second, transgenerational
333 effects of copper and biomass accumulation tended to be negatively correlated. Avoidance of
334 excessive metal ion accumulation is a common strategy to resist toxic heavy metal concentrations
335 [42, 43]. In contrast, no association between transgenerational effects on plant fitness and flavonoids
336 were observed, despite that flavonoids, particularly the di-hydroxylated B-ring substituted
337 compounds, are thought to be defensive and are associated with heavy metal resistance in *S.*

338 *polyrhiza* and other plants [46, 48, 70]. Instead, pre-treatment effects of flavonoid and copper
339 accumulation were positively correlated, suggesting that different copper accumulation triggered
340 transgenerational patterns of flavonoid accumulation. Experiments that manipulate flavonoid
341 concentrations as well as plant copper accumulation may provide further mechanistic insights into
342 the relative importance of these physiological responses in modulating plant fitness across
343 generations.

344 While we started to uncover the physiological mechanisms that mediate benefits of copper pre-
345 treatment for offspring fitness under recurring stress, the underlying molecular basis remains
346 unknown. Vertical transmission of substances very unlikely caused the observed pre-treatment
347 effects, as any substance will have reached neglectable concentrations after the multigenerational
348 recovery phase. In contrast, alterations of the microbial community through toxic copper
349 concentrations may have contributed to the observed pre-treatment effects, particularly in our
350 transgenerational experiment, which was performed under non-sterile conditions and without
351 exchange of microbial communities among treatment groups. Transgenerational effects that were
352 mediated by an altered microbial community have been reported in antibiotic-treated *Drosophila*
353 *melanogaster* larvae [71]. In our long-term pre-treatment experiment, offspring of copper and
354 control pre-treated populations were grown together; the observed fitness difference between
355 copper and control pre-treated offspring are thus unlikely due to alterations in the mobile microbial
356 community. Considering that many of the observed variations between copper and control pre-
357 treated offspring were still detected after 10 (plant fitness) and even 15 (priming of flavonoids)
358 generations, an involvement of epigenetic inheritance seems likely. Experiments that assess
359 induction and persistence of epigenetic marks upon copper excess and their relation to plant
360 phenotype and fitness may help to resolve the on-going controversy about the importance of
361 epigenetic inheritance to mediate transgenerational stress resistance [5, 8, 13, 26, 57].

362 Taken together, our data demonstrate time-dependent costs and benefits of copper pre-treatment
363 on offspring fitness when initial and recurring stress conditions were separated by multiple

364 generations. Thereby, this study supports the notion that non-genetic inheritance may modulate
365 plant ecology and evolution in asexually reproducing plants, which may help to explain the ecological
366 and evolutionary success of these organisms.

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376 REFERENCES

- 377 [1] Bonduriansky, R. & Day, T. 2009 Nongenetic inheritance and its evolutionary implications. *Annu.*
378 *Rev. Ecol., Evol. Syst.* **40**, 103-125. (doi:10.1146/annurev.ecolsys.39.110707.173441).
- 379 [2] Crean, A.J. & Bonduriansky, R. 2014 What is a paternal effect? *Trends Ecol. Evol.* **29**, 554-559.
380 (doi:10.1016/j.tree.2014.07.009).
- 381 [3] Youngson, N. & Whitelaw, E. 2008 Transgenerational epigenetic effects. *Annu. Rev. Genom. Hum.*
382 *Genet.* **9**, 233-257. (doi:10.1146/annurev.genom.9.081307.164445).
- 383 [4] Holeski, L.M., Jander, G. & Agrawal, A.A. 2012 Transgenerational defense induction and
384 epigenetic inheritance in plants. *Trends Ecol. Evol.* **27**, 618-626. (doi:10.1016/j.tree.2012.07.011).
- 385 [5] Grossniklaus, U., Kelly, B., Ferguson-Smith, A.C., Pembrey, M. & Lindquist, S. 2013
386 Transgenerational epigenetic inheritance: how important is it? *Nat. Rev. Genet.* **14**, 228-235.
387 (doi:10.1038/nrg3435).
- 388 [6] Perez, M.F. & Lehner, B. 2019 Intergenerational and transgenerational epigenetic inheritance in
389 animals. *Nat. Cell Biol.* **21**, 143-151. (doi:10.1038/s41556-018-0242-9).
- 390 [7] Wang, Y., Liu, H. & Sun, Z. 2017 Lamarck rises from his grave: parental environment-induced
391 epigenetic inheritance in model organisms and humans. *Biol. Rev. Camb. Philos. Soc.* **92**, 2084-2111.
392 (doi:10.1111/brv.12322).
- 393 [8] Danchin, E., Pocheville, A., Rey, O., Pujol, B. & Blanchet, S. 2019 Epigenetically facilitated
394 mutational assimilation: epigenetics as a hub within the inclusive evolutionary synthesis. *Biol. Rev.*
395 *Camb. Philos. Soc.* **94**, 259-282. (doi:10.1111/brv.12453).
- 396 [9] Kronholm, I. & Collins, S. 2016 Epigenetic mutations can both help and hinder adaptive evolution.
397 *Mol. Ecol.* **25**, 1856-1868. (doi:10.1111/mec.13296).
- 398 [10] Geoghegan, J.L. & Spencer, H.G. 2012 Population-epigenetic models of selection. *Theor. Popul.*
399 *Biol.* **81**, 232-242. (doi:10.1016/j.tpb.2011.08.001).

- 400 [11] Herman, J. & Sultan, S. 2011 Adaptive transgenerational plasticity in plants: Case studies,
401 mechanisms, and implications for natural populations. *Front. Plant Sci.* **2**.
402 (doi:10.3389/fpls.2011.00102).
- 403 [12] Sentis, A., Bertram, R., Dardenne, N., Ramon-Portugal, F., Espinasse, G., Louit, I., Negri, L.,
404 Haeler, E., Ashkar, T., Pannetier, T., et al. 2018 Evolution without standing genetic variation: change
405 in transgenerational plastic response under persistent predation pressure. *Heredity* **121**, 266-281.
406 (doi:10.1038/s41437-018-0108-8).
- 407 [13] Baugh, L.R. & Day, T. 2020 Nongenetic inheritance and multigenerational plasticity in the
408 nematode *C. elegans*. *eLife* **9**, e58498. (doi:10.7554/eLife.58498).
- 409 [14] Agrawal, A.A., Laforsch, C. & Tollrian, R. 1999 Transgenerational induction of defences in animals
410 and plants. *Nature* **401**, 60-63. (doi:10.1038/43425).
- 411 [15] Uller, T., Nakagawa, S. & English, S. 2013 Weak evidence for anticipatory parental effects in
412 plants and animals. *J. Evol. Biol.* **26**, 2161-2170. (doi:10.1111/jeb.12212).
- 413 [16] Rahavi, M.R., Migicovsky, Z., Titov, V. & Kovalchuk, I. 2011 Transgenerational adaptation to
414 heavy metal salts in *Arabidopsis*. *Front. Plant Sci.* **2**, 91. (doi:10.3389/fpls.2011.00091).
- 415 [17] Galloway, L.F. & Etterson, J.R. 2007 Transgenerational plasticity is adaptive in the wild. *Science*
416 **318**, 1134-1136. (doi:10.1126/science.1148766).
- 417 [18] Luna, E., Bruce, T.J.A., Roberts, M.R., Flors, V. & Ton, J. 2012 Next-generation systemic acquired
418 resistance. *Plant Physiol.* **158**, 844-853. (doi:10.1104/pp.111.187468).
- 419 [19] Paszkowski, J. & Grossniklaus, U. 2011 Selected aspects of transgenerational epigenetic
420 inheritance and resetting in plants. *Curr. Opin. Plant Biol.* **14**, 195-203.
421 (doi:10.1016/j.pbi.2011.01.002).
- 422 [20] Baker, B.H., Sultan, S.E., Lopez-Ichikawa, M. & Waterman, R. 2019 Transgenerational effects of
423 parental light environment on progeny competitive performance and lifetime fitness. *Philos. Trans. R.*
424 *Soc. Lond., Ser. B: Biol. Sci.* **374**, 20180182. (doi:10.1098/rstb.2018.0182).

- 425 [21] Erb, M. 2018 Plant defenses against herbivory: Closing the fitness gap. *Trends Plant Sci.* **23**, 187-
426 194. (doi:10.1016/j.tplants.2017.11.005).
- 427 [22] Latzel, V., Gonzalez, A.P.R. & Rosenthal, J. 2016 Epigenetic memory as a basis for intelligent
428 behavior in clonal plants. *Front. Plant Sci.* **7**, 7. (doi:10.3389/fpls.2016.01354).
- 429 [23] Latzel, V. & Klimešová, J. 2010 Transgenerational plasticity in clonal plants. *Evol. Ecol.* **24**, 1537-
430 1543. (doi:10.1007/s10682-010-9385-2).
- 431 [24] Douhovnikoff, V. & Dodd, R.S. 2015 Epigenetics: a potential mechanism for clonal plant success.
432 *Plant Ecol.* **216**, 227-233. (doi:10.1007/s11258-014-0430-z).
- 433 [25] Tricker, P.J. 2015 Transgenerational inheritance or resetting of stress-induced epigenetic
434 modifications: two sides of the same coin. *Front. Plant Sci.* **6**. (doi:10.3389/fpls.2015.00699).
- 435 [26] Heard, E. & Martienssen, R.A. 2014 Transgenerational epigenetic inheritance: myths and
436 mechanisms. *Cell* **157**, 95-109. (doi:10.1016/j.cell.2014.02.045).
- 437 [27] Münzbergová, Z. & Hadincová, V. 2017 Transgenerational plasticity as an important mechanism
438 affecting response of clonal species to changing climate. *Ecol. Evol.* **7**, 5236-5247.
439 (doi:10.1002/ece3.3105).
- 440 [28] González, A.P.R., Preite, V., Verhoeven, K.J.F. & Latzel, V. 2018 Transgenerational effects and
441 epigenetic memory in the clonal plant *Trifolium repens*. *Front. Plant Sci.* **9**, 1677-1677.
442 (doi:10.3389/fpls.2018.01677).
- 443 [29] González, A.P.R., Chrtek, J., Dobrev, P.I., Dumalasova, V., Fehrer, J., Mraz, P. & Latzel, V. 2016
444 Stress-induced memory alters growth of clonal off spring of white clover (*Trifolium repens*). *Am. J.*
445 *Bot.* **103**, 1567-1574. (doi:10.3732/ajb.1500526).
- 446 [30] González, A.P.R., Dumalasová, V., Rosenthal, J., Skuhrovec, J. & Latzel, V. 2017 The role of
447 transgenerational effects in adaptation of clonal offspring of white clover (*Trifolium repens*) to
448 drought and herbivory. *Evol. Ecol.* **31**, 345-361. (doi:10.1007/s10682-016-9844-5).

- 449 [31] Verhoeven, K.J.F., Jansen, J.J., van Dijk, P.J. & Biere, A. 2010 Stress-induced DNA methylation
450 changes and their heritability in asexual dandelions. *New Phytol.* **185**, 1108-1118.
451 (doi:10.1111/j.1469-8137.2009.03121.x).
- 452 [32] Verhoeven, K.J.F. & van Gurp, T.P. 2012 Transgenerational effects of stress exposure on offspring
453 phenotypes in apomictic dandelion. *Plos One* **7**. (doi:10.1371/journal.pone.0038605).
- 454 [33] Tiffney, B.H. & Niklas, K.J. 1985 Clonal growth in land plants: A paleobotanical perspective. In
455 *Population Biology and Evolution fo Clonal Organisms* (eds. J.B.C. Jackson, L.W. Buss & R.E. Cook), pp.
456 35-66. New Haven, Yle University Press.
- 457 [34] Song, Y.-B., Yu, F.-H., Keser, L.H., Dawson, W., Fischer, M., Dong, M. & van Kleunen, M. 2013
458 United we stand, divided we fall: a meta-analysis of experiments on clonal integration and its
459 relationship to invasiveness. *Oecologia* **171**, 317-327. (doi:10.1007/s00442-012-2430-9).
- 460 [35] Liu, J., Dong, M., Miao, S.L., Li, Z.Y., Song, M.H. & Wang, R.Q. 2006 Invasive alien plants in China:
461 role of clonality and geographical origin. *Biol. Invasions* **8**, 1461-1470. (doi:10.1007/s10530-005-
462 5838-x).
- 463 [36] Pyšek, P. 1997 Clonality and plant invasions: can a trait make a difference? In *Ecology and*
464 *evolution of clonal plants*. (eds. d.K. H & v.G. J), pp. 405–427. Leiden, Netherlands, Backhuys
465 Publishers.
- 466 [37] McKey, D., Elias, M., Pujol, B. & Duputié, A. 2010 The evolutionary ecology of clonally
467 propagated domesticated plants. *New Phytol.* **186**, 318-332. (doi:10.1111/j.1469-
468 8137.2010.03210.x).
- 469 [38] Hutchinson, G.E. 1975 *A treatise on limnology, Limnological botany*. New York, John Wiley; 660
470 p.
- 471 [39] Klimeš, L., Klimešová, J., Hendriks, R. & van Groenendael, J. 1997 *Clonal plant architecture: a*
472 *comparative analysis of form and function* 1-29 p.
- 473 [40] Yruea, I. 2009 Copper in plants: acquisition, transport and interactions. *Funct. Plant Biol.* **36**,
474 409-430. (doi:10.1071/fp08288).

- 475 [41] Dietz, K.-J., Baier, M. & Krämer, U. 1999 Free radicals and reactive oxygen species as mediators
476 of heavy metal toxicity in plants. In *Heavy Metal Stress in Plants: From Molecules to Ecosystems* (pp.
477 73-97. Berlin, Heidelberg, Springer Berlin Heidelberg.
- 478 [42] Baker, A.J.M. 1981 Accumulators and excluders -strategies in the response of plants to heavy
479 metals. *J. Plant Nutr.* **3**, 643-654. (doi:10.1080/01904168109362867).
- 480 [43] Woolhouse, H.W. 1983 Toxicity and tolerance in the responses of plants to metals. In
481 *Physiological Plant Ecology III: Responses to the Chemical and Biological Environment* (eds. O.L.
482 Lange, P.S. Nobel, C.B. Osmond & H. Ziegler), pp. 245-300. Berlin, Heidelberg, Springer Berlin
483 Heidelberg.
- 484 [44] Arrivault, S., Senger, T. & Krämer, U. 2006 The Arabidopsis metal tolerance protein AtMTP3
485 maintains metal homeostasis by mediating Zn exclusion from the shoot under Fe deficiency and Zn
486 oversupply. *The Plant Journal* **46**, 861-879. (doi:10.1111/j.1365-313x.2006.02746.x).
- 487 [45] Rice-Evans, C.A., Miller, N.J. & Paganga, G. 1996 Structure-antioxidant activity relationships of
488 flavonoids and phenolic acids. *Free Radic Biol Med* **20**, 933-956.
- 489 [46] Brown, J.E., Khodr, H., Hider, R.C. & Rice-Evans, C.A. 1998 Structural dependence of flavonoid
490 interactions with Cu²⁺ ions: implications for their antioxidant properties. *Biochem. J.* **330**, 1173-1178.
491 (doi:10.1042/bj3301173).
- 492 [47] Qiao, X., He, W.-n., Xiang, C., Han, J., Wu, L.-j., Guo, D.-a. & Ye, M. 2011 Qualitative and
493 quantitative analyses of flavonoids in *Spirodela polyrrhiza* by high-performance liquid
494 chromatography coupled with mass spectrometry. *Phytochem. Anal.* **22**, 475-483.
495 (doi:10.1002/pca.1303).
- 496 [48] Böttner, L., Grabe, V., Gablenz, S., Böhme, N., Appenroth, K.J., Gershenzon, J. & Huber, M. 2020
497 Differential localization of flavonoid glucosides in an aquatic plant implicates different functions
498 under abiotic stress. *Plant Cell Environ.* (doi:10.1111/pce.13974).
- 499 [49] R Core Team. 2018 *R: A language and environment for statistical computing*. Vienna, R
500 Foundation for Statistical Computing.

- 501 [50] Hope, R.M. 2013 Rmisc: Ryan Miscellaneous.
- 502 [51] Auguie, B. 2012 gridExtra: functions in Grid graphics.
- 503 [52] Wickham, H., François, R., Henry, L. & Müller, K. 2018 dplyr: a grammar of data manipulation.
- 504 [53] Wickham, H. 2009 *ggplot2: elegant graphics for data analysis*, Springer New York.
- 505 [54] Peterson, B.G. & Carl, P. 2020 PerformanceAnalytics: econometric tools for performance and risk
506 analysis.
- 507 [55] Pinheiro, J., Bates, D., DebRoy, S., Sarkar, S. & team, R.c. 2014 nlme: Linear and Nonlinear Mixed
508 Effects Models.
- 509 [56] Hunt, R. 1982 *Plant growth curves. the functional approach to plant growth analysis*. London,
510 UK, Edward Arnold Ltd.
- 511 [57] Quadrana, L. & Colot, V. 2016 Plant transgenerational epigenetics. *Annu. Rev. Genet.* **50**, 467-
512 491. (doi:10.1146/annurev-genet-120215-035254).
- 513 [58] Sánchez-Tójar, A., Lagisz, M., Moran, N.P., Nakagawa, S., Noble, D.W.A. & Reinhold, K. 2020 The
514 jury is still out regarding the generality of adaptive 'transgenerational' effects. *Ecol. Lett.* **23**, 1715-
515 1718. (doi:10.1111/ele.13479).
- 516 [59] Bell, A.M. & Hellmann, J.K. 2019 An integrative framework for understanding the mechanisms
517 and multigenerational consequences of transgenerational plasticity. *Annu. Rev. Ecol., Evol. Syst.* **50**,
518 97-118. (doi:10.1146/annurev-ecolsys-110218-024613).
- 519 [60] Xu, S., Stapley, J., Gablenz, S., Boyer, J., Appenroth, K.J., Sree, K.S., Gershenzon, J., Widmer, A. &
520 Huber, M. 2019 Low genetic variation is associated with low mutation rate in the giant duckweed.
521 *Nat. Commun.* **10**, 1243. (doi:10.1038/s41467-019-09235-5).
- 522 [61] Becker, C., Hagmann, J., Müller, J., Koenig, D., Stegle, O., Borgwardt, K. & Weigel, D. 2011
523 Spontaneous epigenetic variation in the *Arabidopsis thaliana* methylome. *Nature* **480**, 245-249.
524 (doi:10.1038/nature10555).

- 525 [62] Jiang, C., Mithani, A., Belfield, E.J., Mott, R., Hurst, L.D. & Harberd, N.P. 2014 Environmentally
526 responsive genome-wide accumulation of de novo *Arabidopsis thaliana* mutations and epimutations.
527 *Genome Res.* **24**, 1821-1829. (doi:10.1101/gr.177659.114).
- 528 [63] Suter, L. & Widmer, A. 2013 Phenotypic effects of salt and heat stress over three generations in
529 *Arabidopsis thaliana*. *Plos One* **8**. (doi:10.1371/journal.pone.0080819).
- 530 [64] Suter, L. & Widmer, A. 2013 Environmental heat and salt stress induce transgenerational
531 phenotypic changes in *Arabidopsis thaliana*. *Plos One* **8**. (doi:10.1371/journal.pone.0060364).
- 532 [65] Groot, M.P., Kooke, R., Knobens, N., Vergeer, P., Keurentjes, J.J.B., Ouborg, N.J. & Verhoeven,
533 K.J.F. 2016 Effects of multi-generational stress exposure and offspring environment on the
534 expression and persistence of transgenerational effects in *Arabidopsis thaliana*. *Plos One* **11**, 16.
535 (doi:10.1371/journal.pone.0151566).
- 536 [66] Yang, X., Sanchez, R., Kundariya, H., Maher, T., Dopp, I., Schwegel, R., Viridi, K., Axtell, M.J. &
537 Mackenzie, S.A. 2020 Segregation of an MSH1 RNAi transgene produces heritable non-genetic
538 memory in association with methylome reprogramming. *Nat. Commun.* **11**, 2214.
539 (doi:10.1038/s41467-020-16036-8).
- 540 [67] Alvarez, M., Bleich, A. & Donohue, K. 2020 Genotypic variation in the persistence of
541 transgenerational responses to seasonal cues. *Evolution* **74**, 2265-2280. (doi:10.1111/evo.13996).
- 542 [68] Auge, G.A., Leverett, L.D., Edwards, B.R. & Donohue, K. 2017 Adjusting phenotypes via within-
543 and across-generational plasticity. *New Phytol.* **216**, 343-349. (doi:10.1111/nph.14495).
- 544 [69] Groot, M.P., Kubisch, A., Ouborg, N.J., Pagel, J., Schmid, K.J., Vergeer, P. & Lampei, C. 2017
545 Transgenerational effects of mild heat in *Arabidopsis thaliana* show strong genotype specificity that
546 is explained by climate at origin. *New Phytol.* **215**, 1221-1234. (doi:10.1111/nph.14642).
- 547 [70] Keilig, K. & Ludwig-Müller, J. 2009 Effect of flavonoids on heavy metal tolerance in *Arabidopsis*
548 *thaliana* seedlings. *Bot. Stud.* **50**, 311-318.

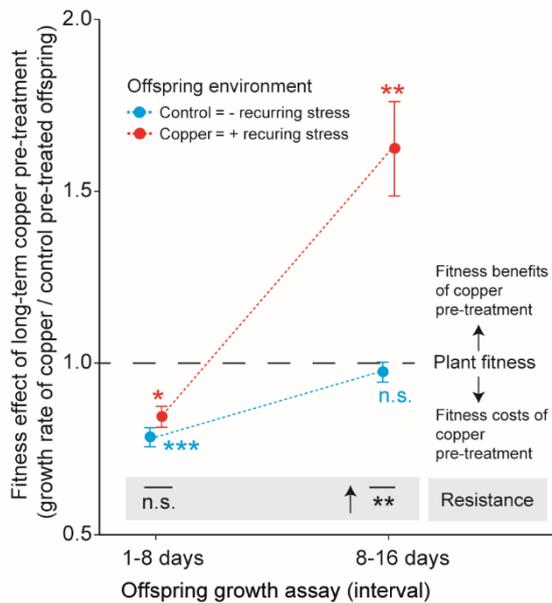
549 [71] Fridmann-Sirkis, Y., Stern, S., Elgart, M., Galili, M., Zeisel, A., Shental, N. & Soen, Y. 2014 Delayed
550 development induced by toxicity to the host can be inherited by a bacterial-dependent,
551 transgenerational effect. *Front Genet* **5**, 27. (doi:10.3389/fgene.2014.00027).

552

553

554 FIGURES

Figure 1



555

556 **Figure 1. Growth of large monoclonal *S. polyrhiza* populations for 30 generations under copper**

557 **excess had negative effects on offspring fitness during short and positive effects during prolonged**

558 **growth under recurring copper excess.** Fitness effects of long-term copper pre-treatment are the

559 ratios in growth rates of copper to control pre-treated offspring. Asterisks beside data points depict

560 *P*-values of Kruskal-Wallis rank sum tests comparing growth rates of copper and control pre-treated

561 offspring for each offspring environment and growth interval separately. Resistance is the difference

562 in fitness pre-treatment effects between offspring environments (Kruskal-Wallis rank sum tests,

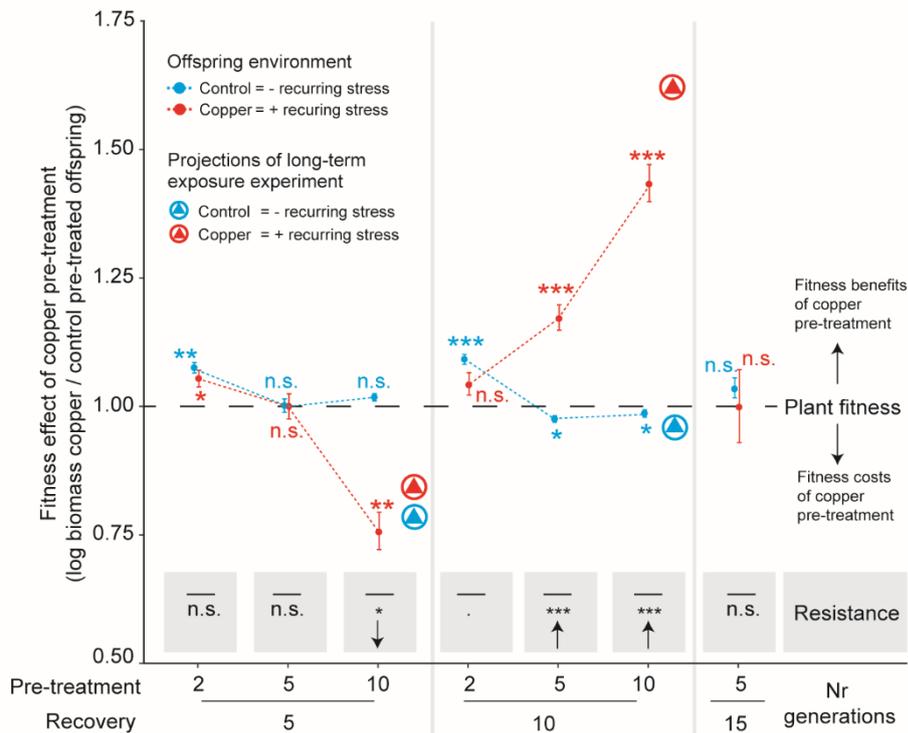
563 arrow indicates positive effect). Descendants from copper and control pre-treated populations were

564 grown for 5 generations under control conditions prior assessing offspring growth. (**P* < 0.05, ***P* <

565 0.01, ****P* < 0.001; n.s. = non-significant). Data display mean values and standard errors. N = 9-10.

566

Figure 2



567

568 **Figure 2. Copper pre-treatment under single descendant propagation can elicit positive fitness and**

569 **resistance effects under recurring stress, similar to pre-treatment effects observed after long-term**

570 **growth of monoclonal *S. polyrhiza* populations.** Fitness effects of copper pre-treatment are the

571 ratios of biomass (log) accumulation of copper to control pre-treated offspring after 8 days of growth

572 for both offspring environments separately. Asterisks next to data points indicate P -values of Kruskal-

573 Wallis rank sum tests comparing biomass (log) accumulation of copper and control pre-treated

574 offspring within each offspring environment. Resistance is the difference in these pre-treatment

575 effects between offspring environments (Kruskal-Wallis rank sum tests, arrows indicate positive and

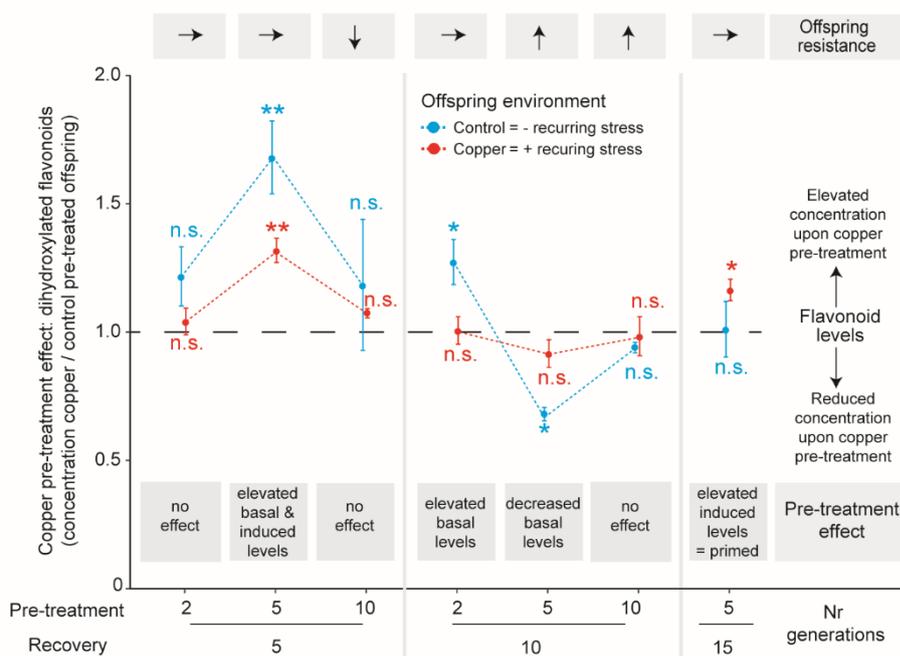
576 negative effects). Mean values of the long-term copper exposure are projected as encircled triangles.

577 ($*P < 0.05$, $**P < 0.01$, $***P < 0.001$; n.s. = non-significant). Data display mean values and standard

578 errors. $N = 3-20$.

579

Figure 3

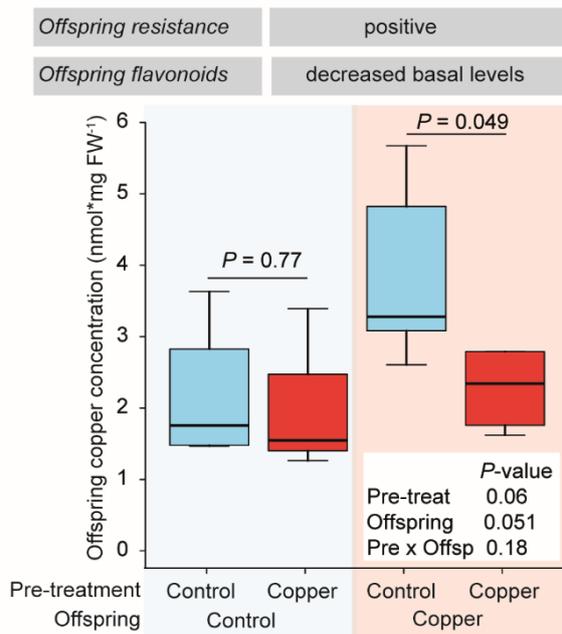


580

581 **Figure 3. Copper pre-treatment may enhance offspring flavonoid accumulation, but these**
 582 **increments were not associated with enhanced offspring resistance.** Copper pre-treatment effects
 583 on di-hydroxylated B-ring substituted flavonoid accumulation were expressed as the ratio in the sum
 584 of the two major luteolin glucosides of copper to control pre-treated offspring after 8 days of growth,
 585 for both offspring environments separately. Asterisks display *P*-values of Student's *t*-tests comparing
 586 dihydroxylated B-ring substituted flavonoid levels of copper and control pre-treated offspring within
 587 each offspring environment. Offspring resistance is display above the panel as arrows and refer to
 588 results of figure 2 (**P* < 0.05, ***P* < 0.01, ****P* < 0.001; n.s. = non-significant). Data display mean
 589 values and standard errors. N = 3-6.

590

Figure 4

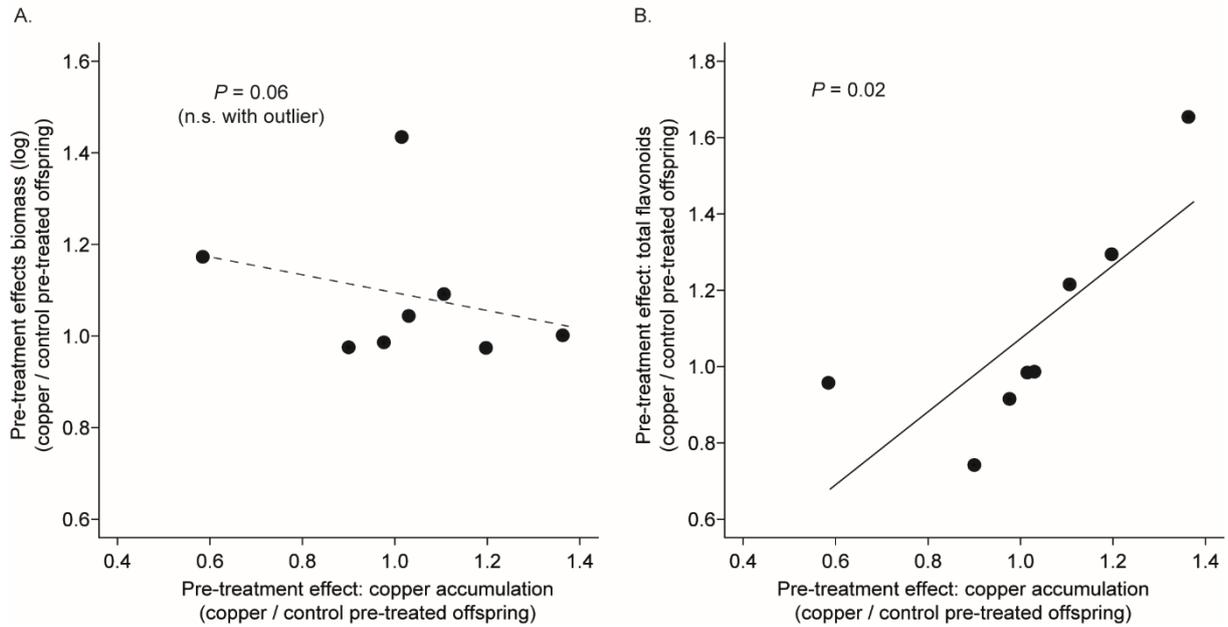


591

592 **Figure 4. Avoidance of excessive copper accumulation was partially associated with enhanced**
593 **offspring resistance under recurring stress.** Offspring copper concentration was measured after 8
594 days of growth in the presence and absence of copper excess in the assay with 5 and 10 generations
595 of pre-treatment and recovery, respectively. *P*-values of Student's *t*-tests and two-way ANOVAs are
596 shown. Offspring resistance and flavonoid levels are display above the panel and refer to results of
597 figure 2 and 3. N = 4-5.

598

Figure 5



599

600 **Figure 5. Pre-treatment effects of copper accumulation correlated weakly negatively with pre-**
601 **treatment effects of biomass accumulation and positively with pre-treatment effects of flavonoid**
602 **concentrations.** Pre-treatment effects are the ratio in copper concentration and offspring biomass
603 accumulation (log) (A), as well as total flavonoids concentration (B), of copper to control pre-treated
604 offspring. Each data point displays the mean value of a pre-treatment and recovery combination and
605 offspring environment. *P*-values of the comparison of linear mixed effect models using one-way
606 ANOVA are shown. Results of models including and excluding the extreme pre-treatment effect in
607 biomass accumulation (> 1.4) are shown in (B). N = 7-8. n.s. = not significant.