



Special issue: Multicomponent lipid membranes—how molecular organisation leads to function

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From 2009 to 2020, a group of scientists, with great expertise in the area of membrane biophysics, closely worked together in the Collaborative Research Centre 803 (CRC 803, *Sonderforschungsbereich* 803), with the common goal to unravel the interactions between the large number of different lipids and specialized proteins in cellular membranes on the molecular level. The mission of the CRC 803 was to clarify how the spatial and temporal organisation of membrane components influences their function and vice versa. The findings of the CRC 803 enabled us to draw molecular pictures of how peptides and proteins in the lipid membrane as well as between two membranes lead to structures that are responsible for transport processes across membranes, and substantial topological changes comprising the fusion of two lipid bilayers. To obtain the required high spatial and temporal resolution to follow these molecular processes combined with precise control of composition, altogether, 19 projects pursued bottom-up approaches utilizing

tailor-made, peptide-/protein containing model membrane systems paired with experimental and theoretical methods developments, such as high-resolution microscopy and computer simulations.

We had the great opportunity to be part of this team and decided to act as guest editors for a special issue highlighting major parts of our results. We collected 14 articles, a blend of original articles and reviews. In the first article of this special issue, the question is addressed how the chemical structure of the globoside Gb₃ influences the phase behaviour of membranes and the binding of Shiga toxin (Sibold et al. 2020). In the second original article, the influence of different arginine derivatives on the integrity of lipid membranes is dissected by means of experimental and theoretical approaches (Verbeek et al. 2021). Three articles impressively show how EPR- and NMR-spectroscopy contribute to our understanding how simple transmembrane peptides as well as large proteins are organised in lipid membranes and interact with small molecules and membrane lipids (Tkach et al. 2021; Naibauer et al. 2021; Riviere et al. 2020). The article of de Groot and co-worker shows the potential of atomistic simulations for our understanding of transmembrane proteins and their interaction with lipids (Daday and de Groot 2020). Besides these insights into the biophysics of interactions within membranes, the CRC 803 was also able to develop new methods, such as the model-free multi-scale analysis of patch-clamp recordings (Pein et al. 2021) and microscopy tools to visualize membrane interactions (Grothe et al. 2021). A large part of the CRC 803 was dedicated to elucidate the process of membrane fusion. Here, five articles contribute to our understanding of how the process of neuronal fusion takes place, identifying prominent intermediate states of fusion and quantifying the energetics of fusion (Witt et al. 2021; Mühlenbrock et al. 2020; Smirnova et al. 2021; Scheu et al. 2021; Risselada et al. 2020). In the last article of this special issue, protein-dependent membrane remodelling is discussed (Tarasenko and Meinecke 2021).

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(Guest Editors).

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References

- Daday C, de Groot BL (2020) Lipid–protein forces predict conformational changes in a mechanosensitive channel. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01488-z>
- Grothe T, Nowak J, Jahn R et al (2021) Selected tools to visualize membrane interactions. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01516-6>
- Mühlenbrock P, Sari M, Steinem C (2020) In vitro single vesicle fusion assays based on pore-spanning membranes: merits and drawbacks. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01479-0>
- Najbauer EE, Becker S, Giller K et al (2021) Structure, gating and interactions of the voltage-dependent anion channel. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01515-7>
- Pein F, Eltzner B, Munk A (2021) Analysis of patchclamp recordings: model-free multiscale methods and software. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01506-8>
- Risselada HJ, Grubmüller H (2020) How proteins open fusion pores: insights from molecular simulations. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01484-3>
- Rivière G, Jaipuria G, Andreas LB et al (2020) Membrane-embedded TSPO: an NMR view. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01487-0>
- Scheu M, Komorowski K, Shen C et al (2021) A stalk fluid forming above the transition from the lamellar to the rhombohedral phase of lipid membranes. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01493-2>
- Sibold J, Ahadi S, Werz DB et al (2020) Chemically synthesized Gb3 glycosphingolipids: tools to access their function in lipid membranes. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01461-w>
- Smirnova YG, Müller M (2021) How does curvature affect the free-energy barrier of stalk formation? Small vesicles vs apposing, planar membranes. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01494-1>
- Tarasenko D, Meinecke M (2021) Protein-dependent membrane remodeling in mitochondrial morphology and clathrin-mediated endocytosis. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01501-z>
- Tkach I, Diederichsen U, Bennati M (2021) Studies of transmembrane peptides by pulse dipolar spectroscopy with semi-rigid TOPP spin labels. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01508-6>
- Verbeek SF, Awasthi N, Teiwes NK et al (2021) How arginine derivatives alter the stability of lipid membranes: dissecting the roles of side chains, backbone and termini. *Eur Biophys J*. <https://doi.org/10.1007/s00249-021-01503-x>
- Witt H, Savić F, Verbeek S et al (2021) Membrane fusion studied by colloidal probes. *Eur Biophys J*. <https://doi.org/10.1007/s00249-020-01490-5>

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