

Bottom-Up Approach to Understand Chirality Transfer across Scales in Cellulose Assemblies

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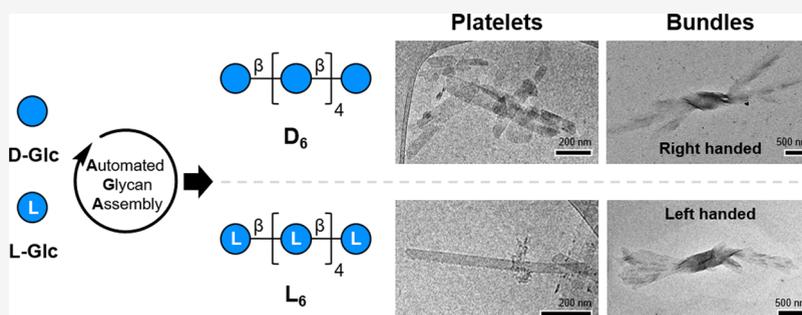
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ABSTRACT: Cellulose is a polysaccharide that displays chirality across different scales, from the molecular to the supramolecular level. This feature has been exploited to generate chiral materials. To date, the mechanism of chirality transfer from the molecular level to higher-order assemblies has remained elusive, partially due to the heterogeneity of cellulose samples obtained *via* top-down approaches. Here, we present a bottom-up approach that uses well-defined cellulose oligomers as tools to understand the transfer of chirality from the single oligomer to supramolecular assemblies beyond the single cellulose crystal. Synthetic cellulose oligomers with defined sequences self-assembled into thin micrometer-sized platelets with controllable thicknesses. These platelets further assembled into bundles displaying intrinsic chiral features, directly correlated to the monosaccharide chirality. Altering the stereochemistry of the oligomer termini impacted the chirality of the self-assembled bundles and thus allowed for the manipulation of the cellulose assemblies at the molecular level. The molecular description of cellulose assemblies and their chirality will improve our ability to control and tune cellulose materials. The bottom-up approach could be expanded to other polysaccharides whose supramolecular chirality is less understood.

INTRODUCTION

Chirality is a key feature of biopolymers and natural products.^{1,2} Molecular chirality based on covalent bonds is well understood, and synthetic methods provide control over asymmetric molecular construction.³ In contrast, the driving forces and key interactions imparting chirality at the supramolecular level are harder to elucidate and reproduce.⁴ Supramolecular chirality has been observed in inorganic materials, polymers, and biological aggregates (*e.g.*, amyloids), and much effort has been devoted to describing the origin of these helices.⁵ Synthetic models offered valuable tools to reveal key molecular features responsible for supramolecular chirality in more complex systems. For instance, well-defined model peptides have contributed to the long-lasting debate on the origin of the twist in amyloid aggregates.^{6–8} Synthetic peptides made of L- and D-amino acid sequences suggested that amino acids located at the termini were responsible for the twist adopted by peptide supramolecular fibers,^{9,10} with a left-handed twist associated with the presence of L-amino acids.^{11,12} Moreover, fine-tuning of these synthetic analogues

generated novel, chiral self-assembled materials with controllable shapes and properties.^{13–15}

The chirality of cellulose, the most abundant organic material on Earth, has not been dissected using a synthetic approach, despite the growing interest in the chiral properties of cellulose nanofibers (CNFs), cellulose nanocrystals (CNCs), and their assemblies.¹⁶ Native CNFs are intrinsically chiral, with nanoscale twists along the fiber direction. Chiral cellulose particles can assemble into larger chiral architectures *in vivo*. Helicoidal arrangements of cellulose fibrils are found in plant cell walls and are capable of generating colors in the absence of any pigment through structural coloration.^{17,18} Left-handed and, in some rare cases, right-handed helicoids have been observed.¹⁷ Still, the mechanism for helicoid formation in

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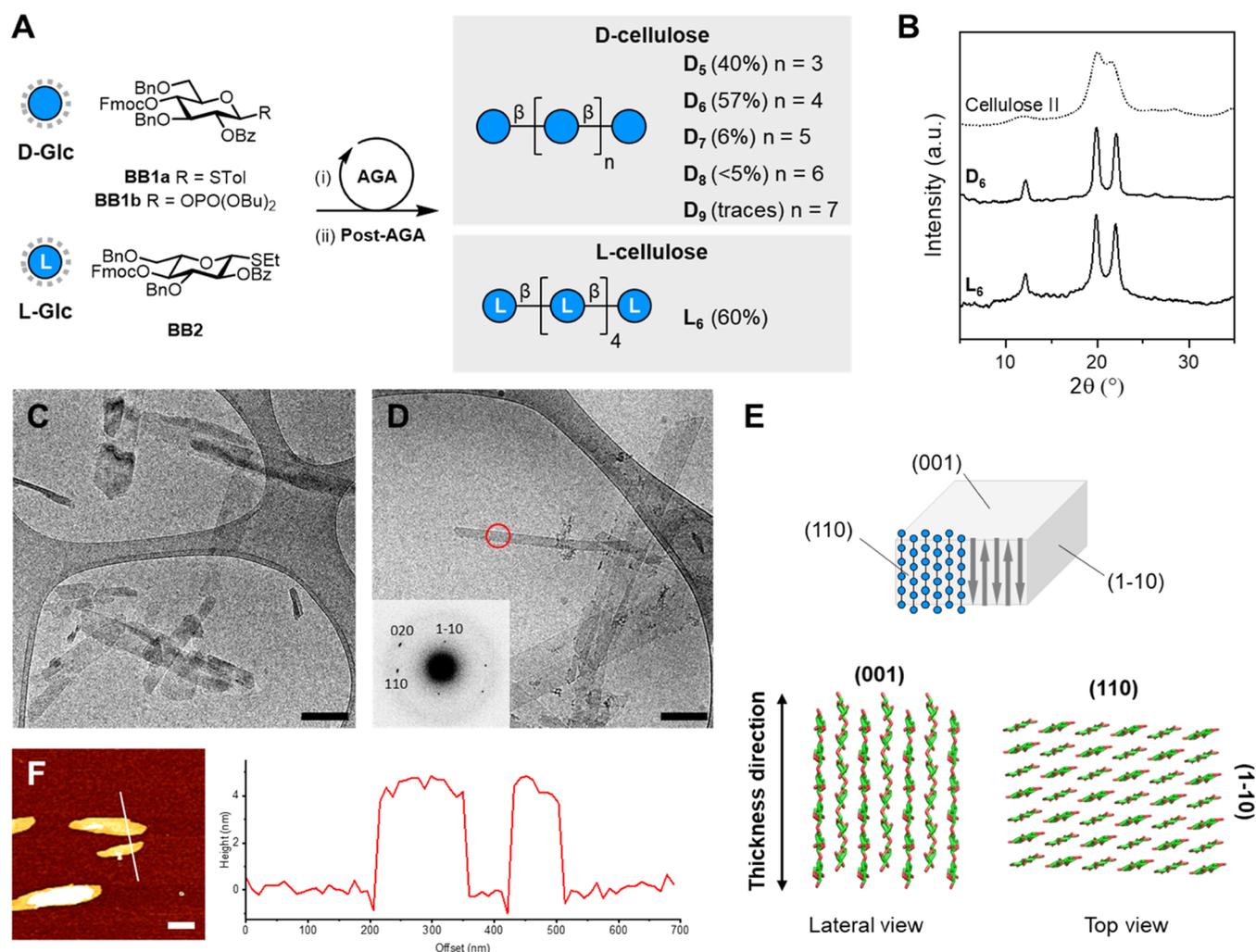


Figure 1. (A) AGA of cellulose analogues using protected monosaccharide BBs (overall yields reported in parentheses. Solid support and modules used for AGA and post-AGA are reported in the Supporting information). (B) Powder X-ray diffraction (XRD) profiles for selected cellulose oligomers indicating a cellulose II-type molecular packing. (C) Cryogenic transmission electron microscope (cryoTEM) image of D₆ platelets (scale bar 200 nm). (D) CryoTEM image of L₆ platelets with the electron diffraction diagram obtained from the red circled area (scale bar 200 nm). (E) Three-dimensional (3D) molecular model of the platelets composed of cellulose oligomers (D₆) arranged in antiparallel fashion according to the cellulose II crystal structure. (F) AFM image of L₆ platelets (left) and height trace measurement (right) (scale bar 200 nm).

plants remains debated, invoking the role of other cell wall matrix constituents such as hemicelluloses.¹⁸

Inspired by nature, CNCs have been used to fabricate optical materials such as films^{16,19,20} and structurally colored pigments.²¹ Due to their chirality, CNCs have found applications as nanosized chiral inducers for liquid crystal assemblies²² and heterogeneous enantioselective palladium catalysis²³ or as chiral templates.^{24,25} *In vitro*, when a suspension of CNCs transitions from a diluted to a concentrated regime, spontaneous self-organization results in the formation of a chiral nematic liquid crystalline phase. Although the intrinsic twisted morphology of single CNCs is considered to govern the chirality of these chiral nematic phases, the mechanism by which nanoscale chirality is transferred to a larger scale is debated.^{26–28} While single particles (*i.e.*, CNCs) are right-handed, upon self-assembly, an inversion of chirality occurs, resulting in a left-handed chiral nematic phase.²⁹ This inversion was quantified using atomic force microscopy (AFM) and electron diffraction methods.^{30–32} Recently, bundles of naturally sourced cellulose crystallites have been suggested to be responsible for the

transfer of chirality across different hierarchical levels.^{28,33} However, heterogeneous degrees of polymerization and crystal sizes of cellulose nanomaterials obtained *via* top-down approaches complicated the description of these systems at the molecular level.³⁴ Computational models have speculated on the origin of the twist in cellulose crystals and are, to date, the only option to study these systems at the molecular level.³⁵ Thus, the molecular origin of this twist and the mechanism of chirality transfer to higher hierarchical assemblies remain open questions.

Herein, we exploit synthetic cellulose oligomers to understand the molecular bases of chirality transfer across scales. Using a bottom-up approach, we synthesized oligomers with well-defined sequences of natural D-glucose (D-Glc) and its enantiomer L-glucose (L-Glc) to manipulate the chirality at the sequence level. These cellulose oligomers self-assembled into platelets with controlled dimensions that further aggregated into bundles, displaying chiral features directly connected to their monosaccharide composition. The insertion of L-Glc units in the sequence of D-Glc cellulose oligomers drastically impacted the macroscopic properties, such as solubility,

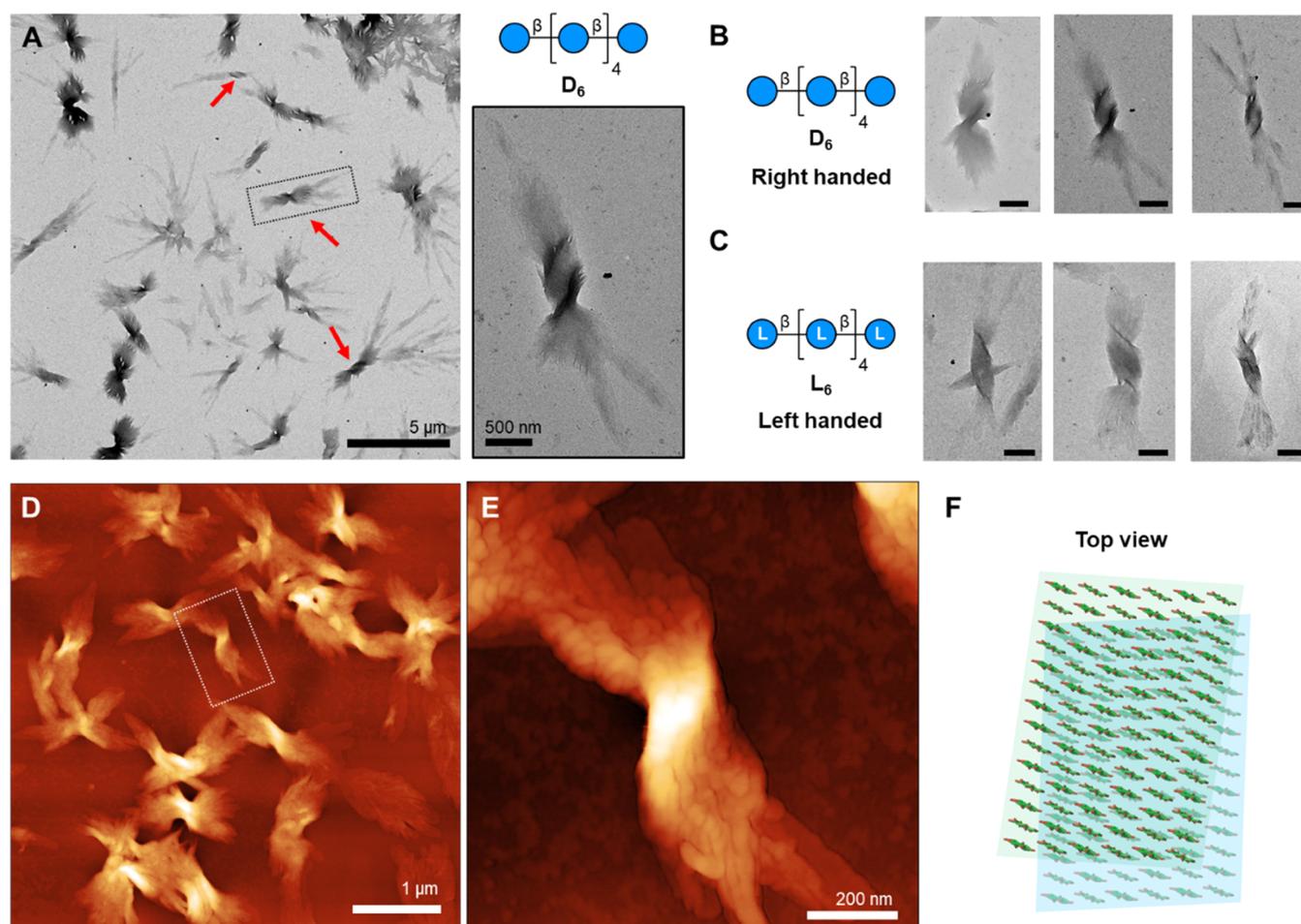


Figure 2. (A) TEM image of D_6 (1 mg/mL aqueous suspension) shows bundles of platelets with intrinsic chirality (red arrows). (B) TEM images of D_6 (1 mg/mL aqueous suspension) bundles showing an intrinsic right-handed chirality (scale bar 500 nm). (C) TEM images of L_6 (1 mg/mL aqueous suspension) bundles showing an intrinsic left-handed chirality (scale bar 500 nm). (D, E) AFM images of D_6 (1 mg/mL aqueous suspension) bundles showing an intrinsic right-handed chirality. AFM samples were prepared on TEM carbon-coated copper grids (see the Supporting information, Figure S37). (F) The fan-like arrangement of the stacking platelets was interpreted as a rotation between the (001) planes.

crystallinity, and chirality of the bundles. The bottom-up approach to study polysaccharide materials highlights the importance of the molecular sequence in dictating supramolecular assembly and chirality.

RESULTS AND DISCUSSION

Cellulose Oligomers Assemble into Nanocrystals. We targeted well-defined oligosaccharides resembling the macroscopic properties of cellulose (*i.e.*, crystallinity and mode of assembly) as a model system to study cellulose assembly. Cellulose oligomers with defined lengths (uniform dispersity, $D = 1$) were synthesized by automated glycan assembly (AGA)^{36,37} on a solid support (functionalized Merrifield resin; see the Supporting information) using protected D-Glc building blocks (BBs) **BB1a** and **BB1b** (Figure 1A). The BBs were equipped with an anomeric thioether or dibutylphosphate leaving group, while most hydroxyl groups were masked as benzyl (Bn) ethers and benzoyl (Bz) esters. Upon glycosylation, the cleavage of the 9-fluorenylmethoxycarbonyl (Fmoc) temporary protecting group liberated the hydroxyl group to be used in the following regioselective chain elongation. Iterative cycles of glycosylation and Fmoc deprotection permitted precise control over the length of the oligomer. Neighboring group participation of the ester group

at C-2 ensured β -stereoselectivity during glycosylation. Each oligosaccharide was assembled overnight following previously reported conditions.³⁷ Post-AGA manipulations included solid-phase methanolysis, photocleavage from the solid support, and hydrogenolysis (see the Supporting information). A single final purification step afforded the target cellulose analogues in overall yields of 6–60%. Longer cellulose analogues (degree of polymerization, DP > 6) were poorly soluble after the hydrogenolysis step and therefore obtained in a drastically decreasing isolated yield. The enantiomeric unnatural analogue L_6 was assembled using protected L-Glc **BB2** (for the synthesis of **BB2**, see the Supporting information).

To evaluate the crystallinity and the aggregation tendency of these oligomers, we employed powder XRD, transmission electron microscopy (TEM), and AFM. Powder XRD indicated that all analogues D_5 , D_6 , and D_7 (DP ranging from 5 to 7) assemble with the cellulose II crystal structure (Figures 1B and S3).³⁸ D_5 was highly soluble, D_6 showed intermediate solubility, while D_7 , D_8 , and D_9 were poorly soluble in water (Table S1). The mirror-image oligomer L_6 showed a cellulose II-type powder XRD profile and solubility similar to D_6 (Figure 1B and Table S1).

Conventional TEM imaging of the negatively stained cellulose oligomers (1 mg/mL in water) indicated the presence

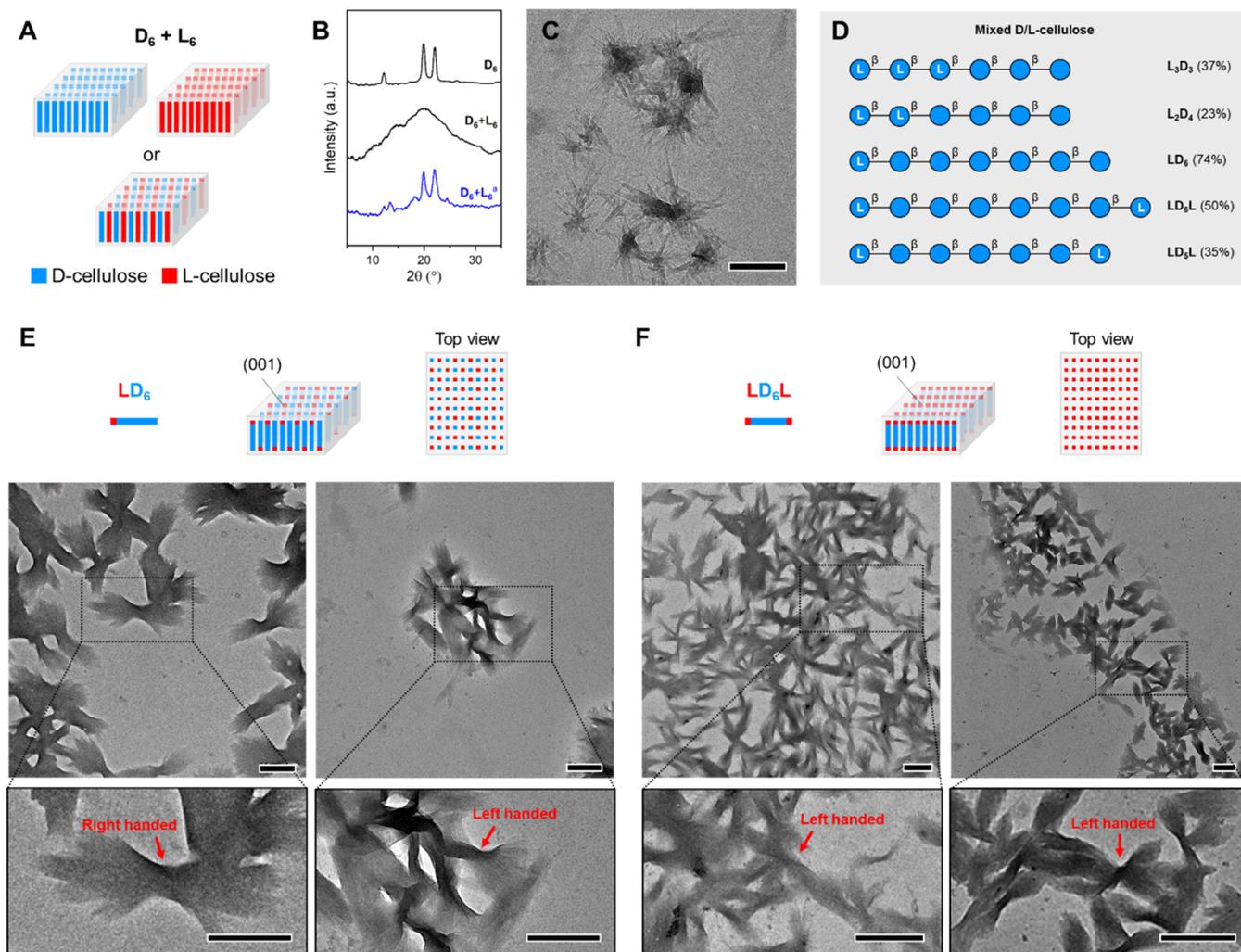


Figure 3. (A) Simplified model of platelets formed by mixing D_6 and L_6 . Self-sorting (top) or coassembly (bottom) of the two enantiomers are two possible scenarios. (B) Powder XRD profiles for $D_6 + L_6$ racemic mixtures. ^aThe sample was prepared by recrystallization from DMSO/MeOH (see the Supporting information). (C) TEM image of $D_6 + L_6$ (MeOH suspension) after recrystallization (scale bar, 500 nm). (D) Hybrid D-/L-cellulose analogues were synthesized using protected monosaccharide BBs (overall yields reported in parentheses). BBs, solid support, and modules used for AGA and post-AGA are reported in the Supporting information. (E) Simplified model of LD_6 platelets and top view of the (001) face (top). TEM images of LD_6 bundles (aqueous suspension, bottom) showing a twisted morphology (red arrows, scale bars 500 nm). (F) Simplified model of LD_6L platelets and top view of the (001) face (top). TEM images of LD_6L bundles (MeOH suspension, bottom) showing a twisted morphology (red arrows, scale bars 500 nm).

of thin platelet-like particles for analogues with $DP \geq 6$ (Figures S5, S9, S15, and S17). No platelets were observed for D_5 , suggesting that its high water solubility prevented the assembly at the concentration used for this study (1 mg/mL in water). The platelets obtained from D_6 are *ca.* 50–500 nm long and *ca.* 20–50 nm wide. A similar morphology was observed for the assembly of enzymatically synthesized cellulose oligomers³⁹ and for mercerized cellulose nanocrystals,⁴⁰ both based on the cellulose II-type crystal structure. CryoTEM demonstrated that the platelets existed in aqueous suspension and were not the result of solvent evaporation (Figure 1C,D). Intra- and intermolecular hydrogen bonds between the hydroxyl groups and hydrophobic interactions between the C–H-rich faces of the glucopyranose rings stabilize the platelets through noncovalent interactions.⁴¹ Electron diffraction (ED) analysis of the platelets generated from D_6 and L_6 confirmed the cellulose II-type assembly (Figure 1D) and indicated a molecular packing where the cellulose chains are aligned in an antiparallel manner along the

platelet thickness (Figure 1E). The (001) faces are exposed on the top and bottom sides of the platelets, presenting an alternation of reducing and nonreducing ends at the surface. The (110) face is exposed at the platelet tip and presents the Glc hydrophobic face. In aqueous media, the crystal growth along the hydrophobic [110] direction is faster than along the hydrophilic [1–10] direction, resulting in elongated platelets.³⁹ Owing to the controlled length of the cellulose oligomers, the platelets have a well-defined thickness corresponding to the oligomer length. AFM analysis confirmed the tunable thickness of the platelets (Figure 1F). The increments of about 0.5 nm across the oligomer series are consistent with the addition of a single Glc unit (Figures S32–S36). This finding suggests that chemical synthesis can generate cellulose materials with tunable and controlled subnanometer dimensions.

Nanocrystals Assemble into Chiral Bundles. No evident chiral features were observed when inspecting the single platelets. However, TEM and AFM imaging revealed the

formation of bundles of platelets alongside the isolated ones. These bundles displayed intrinsic chiral features clearly distinguishable by TEM (Figure 2A) and retained the cellulose II crystal structure (Figure S13). D_6 bundles showed a right-handed twist (Figures 2B and S6–S8), while L_6 bundles were left-handed (Figures 2C and S10–S12). The absolute chirality of the bundles was confirmed by AFM imaging (Figure 2D,E). Bundles were observed when preparing samples on different surfaces and confirmed by different imaging techniques (*i.e.*, SEM and AFM; Figure S39). Spherulite-type assemblies have been observed previously for enzymatically synthesized α -chitin⁴² and cellulose II⁴³ aggregates, although no chiral features were recognized. The formation of bundles in our system reflected the strong tendency of the platelets to interact with each other during the drying process, likely due to the increasing concentration.

The analysis of samples at different drying conditions clarified the mechanism of the evaporation-induced assembly. Aqueous suspensions were deposited on TEM grids, and the suspension was blotted with filter paper to a different extent, leading to different amounts of water left on the grid surface at the end of the drying process. The sample prepared with extensive blotting showed mainly flat individual platelets, while short blotting, hence more remaining water, promoted the formation of large bundles (Figure S19). With decreasing blotting strength, multiple platelets stack on top of each other and a twist along the main axis appears (Figure 2E). When enough water remained after blotting, no edge features can be seen by TEM, suggesting that the platelets tend to merge (or fuse). In agreement with this observation, we found that slowing down the evaporation process favored the formation of a large number of bundles, while at fast drying times, the sample is mostly composed of platelets (Figure S20). Varying the concentration of the initial solution did not significantly affect the morphology of the bundles (Figure S21).

Chirality Transfer from the Oligomers to the Bundles.

During the evaporation-induced assembly, the platelets stack on top of each other and rotate in a fan-like manner, suggesting a slight rotation between the (001) crystal faces (Figures 2E and S14). Thus, we hypothesized that the (001) surface of the platelets (or the interaction between these surfaces) plays a major role in the assembly process, determining the chirality of the bundles (Figure 2F). This hypothesis is in agreement with amyloid-type assemblies of peptides, in which amino acids at the termini are responsible for the supramolecular twist.^{9,10} Chirality transfer processes from different parts of the monomeric units (core and/or termini) have also been reported for supramolecular polymer assemblies.^{44–46}

To gain insight into the origin of the twist, we designed three different experiments. First, we analyzed the assembly of a $D_6 + L_6$ racemic mixture (Figure 3A). A 1:1 mixture of D_6 and L_6 was dissolved in water and lyophilized; the XRD profile showed an amorphous arrangement (Figure 3B). No platelets or bundles were observed during TEM analysis of the $D_6 + L_6$ solution in water (1 mg/mL), suggesting an interaction between the two enantiomers that prevented crystallization. When the $D_6 + L_6$ racemic mixture was recrystallized using a DMSO to MeOH solvent switch method (see the Supporting Information), the powder XRD showed a prevalence of the cellulose II pattern and platelets were regenerated, but no chiral features were observed (Figures 3B,C and S22). Mixing suspensions of D_6 and L_6 platelets in different ratios generated

bundles with right- or left-handed chirality depending on the enantiomer in excess (Figure S23).

We then focused on the assembly of cellulose oligomers with a mixed sequence of D- and L-Glc residues. Five new oligomers were synthesized to selectively change the residues placed at the termini of the sequence (Figure 3D). In the second experiment, we analyzed the assembly of L_2D_4 , L_3D_3 , and LD_6 in which a D-Glc core was capped with L-Glc residues. In this scenario, both D- and L-Glc residues are displayed at the (001) surface due to the antiparallel arrangement of cellulose II (Figure 3E, top). When a large portion of D-Glc residues was replaced by their mirror-image L-Glc, such as in L_2D_4 and L_3D_3 , crystallinity was lost and solubility drastically increased (Table S1 and Figures S3 and S4). In contrast, when only one L-Glc unit was placed at the nonreducing end of the oligomer, such as in LD_6 , the solubility was comparable to that of D_6 and the cellulose II crystal packing was preserved (Table S1 and Figure S3). TEM analysis showed LD_6 platelets and bundles with similar sizes and shapes as for the model system D_6 (Figures S24 and S25). However, the introduction of L-Glc weakened the twisting tendency and no clear handedness could be deduced from TEM images (Figures 3E, bottom, and S25).

In a third experiment, we focused on oligomers bearing L-Glc residues at both termini, such as in LD_5L and LD_6L . In this scenario, only L-Glc residues are displayed at the (001) surface, while the core of the crystal is based on D-Glc units (Figure 3F, top). The solubility of these oligomers was comparable to the model system D_6 (Table S1); however, unexpectedly, LD_5L and LD_6L displayed a cellulose IV_{II}-type XRD profile⁴⁷ (Figure S3) and assembled into square platelets with no bundle-type aggregates (Figures S26 and S30). Recrystallization using a solvent switch method from DMSO to MeOH converted LD_6L to cellulose II, as confirmed by XRD (Figure S4), and generated the typical platelet morphology (Figures S27 and S28). Similar to the second scenario, LD_6L bundles of platelets displayed attenuated chiral features and, in some cases, the bundles appeared to be left-handed (Figure 3F, bottom, and S29).

Overall, the data on LD_6 and LD_6L suggest that perturbing the molecular nature of the surface of cellulose II platelets drastically affects the chirality of their assembly. Bundles with much less pronounced chiral features were observed. No complete inversion of bundle chirality was noticed in the case of LD_6L , suggesting that not only the surface but also the core of the platelets are responsible for inducing the chirality of the homochiral assemblies (*i.e.*, D_6).

CONCLUSIONS

We established a model system for the study of cellulose aggregation using synthetic oligomers produced by AGA. Cellulose oligomers of well-defined lengths and sequences, including both D- and L-Glc, self-assembled into thin platelets with thickness matching the length of a single oligomer chain and arranged in a cellulose II fashion (antiparallel chain arrangement). The thickness of the platelet can be manipulated at the nanoscale by tuning the length of the synthetic cellulose oligomer. The synthetic cellulose platelets further assembled into bundles displaying intrinsic chiral features directly connected to the chirality of the cellulose chain (right-handed for D_6 and left-handed for L_6). Synthetic hybrid D- and L-cellulose oligomers helped to elucidate the origin of the chirality of these bundles. Terminal residues of opposite chirality drastically weakened the twisting tendency of the

bundles, suggesting that the surface of the platelets plays an important role in determining the chirality of the bundles. Still, the surface does not solely determine the twist, as complete inversion of chirality was not observed for analogues displaying only L-Glc at the extremities.

Bundles of naturally sourced cellulose crystallites have been recently proposed to be a key element in the transfer of chirality across different hierarchical levels.^{28,33} Our model system offers a well-defined approach to validate this hypothesis and could be extended to other biopolymers, such as chitin, a polysaccharide composed of β -1,4-linked N-acetyl glucosamine. In contrast to cellulose, the chiral nature of chitin nanocrystals (ChNCs) remains elusive³² even though the *in vitro* formation of chiral nematic phases⁴⁸ and helicoidal organization of chitin microfibrils have been reported.

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/jacs.2c04522>.

All experimental details regarding building block synthesis, AGA, XRD, TEM, AFM, and SEM are reported in the Supporting information (PDF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Lv, J.; Gao, X.; Han, B.; Zhu, Y.; Hou, K.; Tang, Z. Self-Assembled Inorganic Chiral Superstructures. *Nat. Rev. Chem.* **2022**, *6*, 125–145.
- (2) Brandt, J. R.; Salerno, F.; Fuchter, M. J. The Added Value of Small-Molecule Chirality in Technological Applications. *Nat. Rev. Chem.* **2017**, *1*, No. 0045.
- (3) Noyori, R. Asymmetric Catalysis: Science and Opportunities (Nobel Lecture). *Angew. Chem., Int. Ed.* **2002**, *41*, 2008–2022.
- (4) Morrow, S. M.; Bisette, A. J.; Fletcher, S. P. Transmission of Chirality through Space and across Length Scales. *Nat. Nanotechnol.* **2017**, *12*, 410–419.
- (5) Yashima, E.; Ousaka, N.; Taura, D.; Shimomura, K.; Ikai, T.; Maeda, K. Supramolecular Helical Systems: Helical Assemblies of Small Molecules, Foldamers, and Polymers with Chiral Amplification and Their Functions. *Chem. Rev.* **2016**, *116*, 13752–13990.
- (6) Volpatti, L. R.; Vendruscolo, M.; Dobson, C. M.; Knowles, T. P. J. A Clear View of Polymorphism, Twist, and Chirality in Amyloid Fibril Formation. *ACS Nano* **2013**, *7*, 10443–10448.
- (7) Harper, J. D.; Lieber, C. M.; Lansbury, P. T. Atomic Force Microscopic Imaging of Seeded Fibril Formation and Fibril Branching by the Alzheimer's Disease Amyloid- β Protein. *Chem. Biol.* **1997**, *4*, 951–959.
- (8) Nyström, G.; Arcari, M.; Mezzenga, R. Confinement-Induced Liquid Crystalline Transitions in Amyloid Fibril Cholesteric Tactoids. *Nat. Nanotechnol.* **2018**, *13*, 330–336.
- (9) Wang, M.; Zhou, P.; Wang, J.; Zhao, Y.; Ma, H.; Lu, J. R.; Xu, H. Left or Right: How Does Amino Acid Chirality Affect the Handedness of Nanostructures Self-Assembled from Short Amphiphilic Peptides? *J. Am. Chem. Soc.* **2017**, *139*, 4185–4194.
- (10) Lara, C.; Reynolds, N. P.; Berryman, J. T.; Xu, A.; Zhang, A.; Mezzenga, R. ILQINS Hexapeptide, Identified in Lysozyme Left-Handed Helical Ribbons and Nanotubes, Forms Right-Handed Helical Ribbons and Crystals. *J. Am. Chem. Soc.* **2014**, *136*, 4732–4739.
- (11) Ke, P. C.; Zhou, R.; Serpell, L. C.; Riek, R.; Knowles, T. P. J.; Lashuel, H. A.; Gazit, E.; Hamley, I. W.; Davis, T. P.; Fändrich, M.; Otzen, D. E.; Chapman, M. R.; Dobson, C. M.; Eisenberg, D. S.; Mezzenga, R. Half a Century of Amyloids: Past, Present and Future. *Chem. Soc. Rev.* **2020**, *49*, 5473–5509.
- (12) Luo, Z.; Zhang, S. Designer Nanomaterials Using Chiral Self-Assembling Peptide Systems and Their Emerging Benefit for Society. *Chem. Soc. Rev.* **2012**, *41*, 4736–4754.
- (13) Nagy-Smith, K.; Beltramo, P. J.; Moore, E.; Tycko, R.; Furst, E. M.; Schneider, J. P. Molecular, Local, and Network-Level Basis for the Enhanced Stiffness of Hydrogel Networks Formed from Coassembled Racemic Peptides: Predictions from Pauling and Corey. *ACS Cent. Sci.* **2017**, *3*, 586–597.
- (14) Swanekamp, R. J.; DiMaio, J. T. M.; Bowerman, C. J.; Nilsson, B. L. Coassembly of Enantiomeric Amphipathic Peptides into Amyloid-Inspired Rippled β -Sheet Fibrils. *J. Am. Chem. Soc.* **2012**, *134*, 5556–5559.
- (15) Clover, T. M.; O'Neill, C. L.; Appavu, R.; Lokhande, G.; Gaharwar, A. K.; Posey, A. E.; White, M. A.; Rudra, J. S. Self-Assembly of Block Heterochiral Peptides into Helical Tapes. *J. Am. Chem. Soc.* **2020**, *142*, 19809–19813.

- (16) Frka-Petescic, B.; Vignolini, S. So Much More than Paper. *Nat. Photonics* **2019**, *13*, 365–367.
- (17) Vignolini, S.; Rudall, P. J.; Rowland, A. V.; Reed, A.; Moyroud, E.; Faden, R. B.; Baumberg, J. J.; Glover, B. J.; Steiner, U. Pointillist Structural Color in Pollia Fruit. *Proc. Natl. Acad. Sci. U.S.A.* **2012**, *109*, 15712–15715.
- (18) Chang, Y.; Middleton, R.; Ogawa, Y.; Gregory, T.; Steiner, L. M.; Kovalev, A.; Karanja, R. H. N.; Rudall, P. J.; Glover, B. J.; Gorb, S. N.; Vignolini, S. Cell Wall Composition Determines Handedness Reversal in Helicoidal Cellulose Architectures of Pollia Condensata Fruits. *Proc. Natl. Acad. Sci. U.S.A.* **2021**, *118*, No. e2111723118.
- (19) Parker, R. M.; Guidetti, G.; Williams, C. A.; Zhao, T.; Narkevicius, A.; Vignolini, S.; Frka-Petescic, B. The Self-Assembly of Cellulose Nanocrystals: Hierarchical Design of Visual Appearance. *Adv. Mater.* **2018**, *30*, No. 1704477.
- (20) Parker, R. M.; Frka-Petescic, B.; Guidetti, G.; Kamita, G.; Consani, G.; Abell, C.; Vignolini, S. Hierarchical Self-Assembly of Cellulose Nanocrystals in a Confined Geometry. *ACS Nano* **2016**, *10*, 8443–8449.
- (21) Droguet, B. E.; Liang, H.-L.; Frka-Petescic, B.; Parker, R. M.; De Volter, M. F. L.; Baumberg, J. J.; Vignolini, S. Large-Scale Fabrication of Structurally Coloured Cellulose Nanocrystal Films and Effect Pigments. *Nat. Mater.* **2022**, *21*, 352–358.
- (22) Gonçalves, D. P. N.; Hegmann, T. Chirality Transfer from an Innately Chiral Nanocrystal Core to a Nematic Liquid Crystal: Surface-Modified Cellulose Nanocrystals. *Angew. Chem., Int. Ed.* **2021**, *60*, 17344–17349.
- (23) Kaushik, M.; Basu, K.; Benoit, C.; Cirtiu, C. M.; Vali, H.; Moores, A. Cellulose Nanocrystals as Chiral Inducers: Enantioselective Catalysis and Transmission Electron Microscopy 3D Characterization. *J. Am. Chem. Soc.* **2015**, *137*, 6124–6127.
- (24) Majoinen, J.; Hassinen, J.; Haataja, J. S.; Rekola, H. T.; Kontturi, E.; Kostiaainen, M. A.; Ras, R. H. A.; Törmä, P.; Ikkala, O. Chiral Plasmonics Using Twisting along Cellulose Nanocrystals as a Template for Gold Nanoparticles. *Adv. Mater.* **2016**, *28*, 5262–5267.
- (25) Shopsowitz, K. E.; Qi, H.; Hamad, W. Y.; MacLachlan, M. J. Free-Standing Mesoporous Silica Films with Tunable Chiral Nematic Structures. *Nature* **2010**, *468*, 422–425.
- (26) Ogawa, Y. Release of Internal Molecular Torque Results in Twists of Glaucoyctis Cellulose Nanofibers. *Carbohydr. Polym.* **2021**, *251*, No. 117102.
- (27) Araki, J.; Kuga, S. Effect of Trace Electrolyte on Liquid Crystal Type of Cellulose Microcrystals. *Langmuir* **2001**, *17*, 4493–4496.
- (28) Chiappini, M.; Dussi, S.; Frka-Petescic, B.; Vignolini, S.; Dijkstra, M. Modeling the Cholesteric Pitch of Apolar Cellulose Nanocrystal Suspensions Using a Chiral Hard-Bundle Model. *J. Chem. Phys.* **2022**, *156*, No. 014904.
- (29) Nyström, G.; Arcari, M.; Adamcik, J.; Usov, I.; Mezzenga, R. Nanocellulose Fragmentation Mechanisms and Inversion of Chirality from the Single Particle to the Cholesteric Phase. *ACS Nano* **2018**, *12*, 5141–5148.
- (30) Ogawa, Y. Electron Microdiffraction Reveals the Nanoscale Twist Geometry of Cellulose Nanocrystals. *Nanoscale* **2019**, *11*, 21767–21774.
- (31) Usov, I.; Nyström, G.; Adamcik, J.; Handschin, S.; Schütz, C.; Fall, A.; Bergström, L.; Mezzenga, R. Understanding Nanocellulose Chirality and Structure–Properties Relationship at the Single Fibril Level. *Nat. Commun.* **2015**, *6*, No. 7564.
- (32) Bai, L.; Kämäräinen, T.; Xiang, W.; Majoinen, J.; Seitsonen, J.; Grande, R.; Huan, S.; Liu, L.; Fan, Y.; Rojas, O. J. Chirality from Cryo-Electron Tomograms of Nanocrystals Obtained by Lateral Disassembly and Surface Etching of Never-Dried Chitin. *ACS Nano* **2020**, *14*, 6921–6930.
- (33) Parton, T. G.; Parker, R. M.; van de Kerkhof, G. T.; Narkevicius, A.; Haataja, J. S.; Frka-Petescic, B.; Vignolini, S. Chiral Self-Assembly of Cellulose Nanocrystals Is Driven by Crystallite Bundles. *Nat. Commun.* **2022**, *13*, No. 2657.
- (34) Gray, D. G. Chiral Nematic Ordering of Polysaccharides. *Carbohydr. Polym.* **1994**, *25*, 277–284.
- (35) Conley, K.; Godbout, L.; Whitehead, M. A.; van de Ven, T. G. M. Origin of the Twist of Cellulosic Materials. *Carbohydr. Polym.* **2016**, *135*, 285–299.
- (36) Guberman, M.; Seeberger, P. H. Automated Glycan Assembly: A Perspective. *J. Am. Chem. Soc.* **2019**, *141*, 5581–5592.
- (37) Yu, Y.; Tyrikos-Ergas, T.; Zhu, Y.; Fittolani, G.; Bordoni, V.; Singhal, A.; Fair, R. J.; Grafmüller, A.; Seeberger, P. H.; Delbianco, M. Systematic Hydrogen-Bond Manipulations To Establish Polysaccharide Structure-Property Correlations. *Angew. Chem., Int. Ed.* **2019**, *58*, 13127–13132.
- (38) Langan, P.; Nishiyama, Y.; Chanzy, H. X-Ray Structure of Mercerized Cellulose II at 1 Å Resolution. *Biomacromolecules* **2001**, *2*, 410–416.
- (39) Wada, M.; Wakiya, S.; Kobayashi, K.; Kimura, S.; Kitaoka, M.; Kusumi, R.; Kimura, F.; Kimura, T. Three-Dimensional Alignment of Cellulose II Microcrystals under a Strong Magnetic Field. *Cellulose* **2021**, *28*, 6757–6765.
- (40) Hirota, M.; Tamura, N.; Saito, T.; Isogai, A. Cellulose II Nanoelements Prepared from Fully Mercerized, Partially Mercerized and Regenerated Celluloses by 4-Acetamido-TEMPO/NaClO₂ Oxidation. *Cellulose* **2012**, *19*, 435–442.
- (41) Medronho, B.; Romano, A.; Miguel, M. G.; Stigsson, L.; Lindman, B. Rationalizing Cellulose (in)Solubility: Reviewing Basic Physicochemical Aspects and Role of Hydrophobic Interactions. *Cellulose* **2012**, *19*, 581–587.
- (42) Sakamoto, J.; Sugiyama, J.; Kimura, S.; Imai, T.; Itoh, T.; Watanabe, T.; Kobayashi, S. Artificial Chitin Spherulites Composed of Single Crystalline Ribbons of α -Chitin via Enzymatic Polymerization. *Macromolecules* **2000**, *33*, 4155–4160.
- (43) Kobayashi, S.; Hobson, L. J.; Sakamoto, J.; Kimura, S.; Sugiyama, J.; Imai, T.; Itoh, T. Formation and Structure of Artificial Cellulose Spherulites via Enzymatic Polymerization. *Biomacromolecules* **2000**, *1*, 168–173.
- (44) Aparicio, F.; Nieto-Ortega, B.; Nájera, F.; Ramírez, F. J.; López Navarrete, J. T.; Casado, J.; Sánchez, L. Inversion of Supramolecular Helicity in Oligo-*p*-Phenylene-Based Supramolecular Polymers: Influence of Molecular Atropisomerism. *Angew. Chem.* **2014**, *126*, 1397–1401.
- (45) Hifsudheen, M.; Mishra, R. K.; Vedhanarayanan, B.; Praveen, V. K.; Ajayaghosh, A. The Helix to Super-Helix Transition in the Self-Assembly of Π -Systems: Superseding of Molecular Chirality at Hierarchical Level. *Angew. Chem.* **2017**, *129*, 12808–12812.
- (46) Greciano, E. E.; Mataranz, B.; Sánchez, L. Pathway Complexity Versus Hierarchical Self-Assembly in *N*-Annulated Perylenes: Structural Effects in Seeded Supramolecular Polymerization. *Angew. Chem.* **2018**, *130*, 4787–4791.
- (47) Buleon, A.; Chanzy, H. Single Crystals of Cellulose IVII: Preparation and Properties. *J. Polym. Sci. Polym. Phys. Ed.* **1980**, *18*, 1209–1217.
- (48) Narkevicius, A.; Steiner, L. M.; Parker, R. M.; Ogawa, Y.; Frka-Petescic, B.; Vignolini, S. Controlling the Self-Assembly Behavior of Aqueous Chitin Nanocrystal Suspensions. *Biomacromolecules* **2019**, *20*, 2830–2838.